

Comparing the Effect of Dexmedetomidine and Etomidate Administration with Acute Height Stress on Spatial Memory in Male Mice

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Research

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Abstract

Background: Nowadays, anesthetic drugs are widely used in anesthesia and surgical procedures and their effects on memory have been the focus of attention for a very long time. The effects of these common drugs include Dexmedetomidine (DEX) and Etomidate (ETO), on memory are controversial. In this study, the effects of these two drugs, co-administrated with heights stress, were evaluated on short-term and long-term spatial memory. 48 male mice were divided into 6 experimental groups consisting of Control, Control+heightstress (H.S), ETO, ETO+H.S, DEX+H.S. Drugs were administered Intra-peritoneal with doses of 0.3-0.4 mg/kg and 11 mg/kg for DEX and ETO respectively, and spatial memory was assessed using the Barnes Model.

Results: DEX improved acquisition and retention of spatial reference memory, whereas ETO showed no such effects. In addition, DEX and ETO showed excitatory effects on short-term spatial memory, however DEX was more effective than ETO.

Conclusion: the results suggested the neuoprotective, synaptic plasticity and memory improving effects of DEX on spatial reference and working memory. However, the precise neuronal and molecular mechanisms of these effects and their relation to the anti-stress system is still unknown and requires further research.

Background:

Anesthetic drugs are widely used in anesthesia and surgical procedures and their effects on memory have been the focus of attention for a very long time. The experience of height stress may result in stress-related psychiatric disorders such as Post-traumatic stress disorder (PTSD). Which is a psychiatric disease that occurs after serious accidents, anxiety and fright(1). The symptoms of PTSD include increased sensitivity to stress, intrusive memory, hyperarousal, and avoidance (1, 2).

During recent years, improvement of behavioral tasks which assess spatial learning and memory in rodents has us comprehend the role of the hippocampus in these cognitive functions (3). Allocentric navigation involves different parts such as the hippocampus, entorhinal cortex, and its surrounding structures while in the human body this system encodes allocentric, semantic, and episodic memory. This form of memory is assessed in laboratory animals in many ways, but one form of assessment is the Barnes maze (4).

The primary aim of the pharmacotherapeutic agents for PTSD is to disrupt the memory consolidation and the associated fear conditioning response after exposure to an event (5, 6).

The risk of PTSD occurrence in patients exposed to stressful medical conditions associated with perioperative awareness or Intensive Care Unit (ICU) treatment is moderately high (7-9). Patients treated in ICU for life-threatening medical events have multiple traumatic experiences and often develop severe

PTSD symptomatology, including vivid memory of traumatic experiences, anxiety, panic, hallucinations and nightmares (10, 11).

Commonly used anesthetics/sedatives, such as ETO and the α 2-adrenoceptor agonist Dexmedetomidine (DEX) interact with several neurotransmission systems importantly involved in the regulation of emotional memory formation (12).

DEX is an α 2 adrenoceptor agonist that has been used as an anesthetic agent and sedative for several years (13). DEX produces dose-dependent sedation, anxiolysis, and analgesia without respiratory depression. DEX triggers and maintains natural sleeping status without eye movement by stimulating the locus coeruleus in the brain stem, so it increases the activity of inhibitory gamma aminobutyric acid (GABA) neurons in the ventrolateral preoptic nucleus (14). Experimental and clinical practices have revealed that DEX is neuroprotective on delirium, stress and inflammatory response (15-18). Based on previous studies, DEX may suppress memory formation only at doses reducing central nervous system activity in response to sensory inputs(19). Previous studies reported that DEX reduced fear memory consolidation in an inhibitory avoidance task (20). DEX treatment after trauma at sub-anesthetic doses reduced fear memory and anxiety behavior in a rat PTSD model (21). Use of DEX after experiencing a traumatic event might prevent the occurrence of emotional distress(12).

GABAA receptors (GABAARs) are heteropentameric ligand-gated anion channels responsible for the majority of inhibitory synaptic transmission in the brain (22). GABAARs are considered important targets of a variety of agents, including Etomidate (ETO) (23-25).

ETO is an ultra short-acting agent which in contrast to ketamine, induces only transient apnea with good cardiovascular stability in animals and humans (26-28). It also can cause nausea, vomiting, myoclonus, pain on injection and impairment of endocrine system function (29). Cardiovascular stability is attributed to the stimulation of central adrenergic receptors, with a lack of myocardial depression and little change in the heart rate or blood pressure (30). Because of its favorable hemodynamic profile, ETO is used in the clinical setting to induce anesthesia in patients at risk for cardiovascular compromise, and in other select circumstances such as electroconvulsive therapy (31). The inverse agonist effects of α 5-containing GABA_ARs were also found to reduce the amnestic and intraoperative-awareness effects experienced with some anesthetic drugs such as ETO, but without affecting the long-term plasticity and sedative-hypnotic side effects of anesthesia (32).

In this study, the effects of these two drugs, in addition to fear of heights stress, using Barnes maze were evaluated on short-term and long-term spatial memory in mice.

Results:

3 – 1: Effect of acute stress with DEX and ETO on reference spatial learning in Barnes maze model:

Figure 1A shows the comparison between number of errors on learning days in experimental groups. ANOVA showed significant statistical difference between groups. Post hoc test shows significant difference between DEX + H.S with CON + H.S and CON groups ($P < 0.05$, $p < 0.01$ respectively).

Figure 1B shows the comparison of the time spent on finding the target hole on learning days between experimental groups. ANOVA showed significant statistical difference between groups. Post hoc test shows significant difference between DEX and DEX + H.S with CON groups ($P < 0.05$, $p < 0.01$ respectively) and also DEX + H.S with CON + H.S and ETO + H.S ($P < 0.05$).

Figure 1C shows the comparison of Distance traveled to finding target hole on learning days between experimental groups. ANOVA showed no significant statistical difference between groups ($P < 0.05$).

These results indicate that DEX could facilitate spatial reference learning while ETO had no such effect; in addition, DEX shows anxiolytic effect which enhanced learning.

3 – 2: Effect of acute stress with DEX and ETO on spatial reference memory in Barnes maze model:

Figure2A shows the comparison of number of errors on spatial reference memory between experimental groups. ANOVA showed significant statistical difference between groups. Post hoc test shows significant difference between DEX and DEX + H.S with CON groups ($P < 0.01$, $p < 0.001$ respectively), between DEX and DEX + H.S with CON + H.S groups ($P < 0.01$, $p < 0.05$ respectively), between DEX and DEX + H.S with ETO groups ($P < 0.01$), between DEX and DEX + H.S with ETO + H.S groups ($P < 0.01$).

Figure2B shows the comparison of distance traveled to finding target hole on spatial reference memory between experimental groups. ANOVA showed significant statistical difference between groups. Post hoc test shows significant difference between DEX and DEX + H.S with CON groups ($P < 0.05$), between DEX and DEX + H.S with CON + H.S groups ($P < 0.01$), between DEX and DEX + H.S with ETO groups ($P < 0.05$), between DEX and DEX + H.S with ETO + H.S groups ($P < 0.001$).

Our results indicate that DEX could facilitate spatial reference memory whereas ETO had no effect, height stress had no significant effect on spatial reference memory.

3–3: Effect of acute stress with DEX and ETO on acquisition of spatial working memory in Barnes maze model:

Figure3A shows the comparison of number of errors on acquisition spatial working memory between experimental groups. ANOVA showed no significant statistical difference between groups ($P < 0.05$).

Figure 3B shows the comparison of distance traveled to finding target hole on acquisition spatial working memory between experimental groups. ANOVA showed significant statistical difference between groups. Post hoc test shows significant difference between DEX with CON groups ($P < 0.05$), between DEX and DEX + H.S with CON + H.S groups ($P < 0.05$), between DEX with ETO groups ($P < 0.05$).

In this protocol animals should extinct the former goal place and acquire new place. There was no significant difference of number of errors between groups but traveled distance was significantly different between experimental groups and showed that in the DEX group the mice immediately looked for a new place after finding the replacement of the goal box, which indicates facilitating effects of DEX on acquisition of spatial working memory.

3–4: Effect of acute stress with DEX and ETO on retention spatial working memory in Barnes maze model:

Figure 4A shows the comparison of number of errors on retention spatial working memory between experimental groups. ANOVA showed significant statistical difference between groups. Post hoc test shows significant difference between DEX and DEX + H.S with CON groups ($P < 0.05$), between DEX and DEX + H.S with CON + H.S groups ($P < 0.001$), between ETO and ETO + H.S with CON + H.S groups ($P < 0.001$, $p < 0.01$ respectively), between CON with CON + H.S groups ($P < 0.05$).

Figure 4B shows the comparison of distance traveled to finding target hole on retention spatial working memory between experimental groups. ANOVA showed significant statistical difference between groups. Post hoc test shows significant difference between DEX with CON groups ($P < 0.01$), difference between DEX with CON + H.S groups ($P < 0.05$), DEX, DEX + H.S and ETO + H.S with ETO groups ($P < 0.001$, $P < 0.001$, $p < 0.01$ respectively).

Our results indicate that both DEX and ETO have facilitating effect on spatial working memory.

Discussion:

Our main findings areas follow: A) DEX strengthened the reference spatial learning in Barnes maze model whereas ETO had no effect. B) DEX also improved the recall of long-term and Short-term spatial memory. C) Both DEX and ETO improved short-term spatial memory recall, although the effect of DEX was relatively greater than ETO. D) DEX showed facilitating effects on acquisition of spatial working memory. E) In our study, the used height stress model was less effective on most spatial memory parameters, which seems to be because of the sensitivity of our experimental model.

The Barnes maze is broadly used for the evaluation of learning and memory function in mice, particularly spatial learning and memory (33, 34). In the present study, the Barnes maze test was used to determine the effect of DEX and ETO on Spatial memory. GABA_A receptors (GABA_ARs) are heteropentameric ligand-gated anion channels responsible for the majority of inhibitory synaptic transmission in the brain (22). GABA_ARs are considered to be important targets of a variety of agents, including Etomidate (ETO) (23–25). Because the majority of $\alpha 5$ -GABA_ARs are located extra-synaptically on pyramidal cells, where they mediate a persistent conductance termed “tonic inhibition” (35), and tonic inhibition is strongly enhanced by amnesic drugs, it was proposed that the effect of ETO on synaptic plasticity is due to its enhancement of tonic inhibition (36–38). It is also possible that ETO controls synaptic plasticity and memory by targeting interneurons (39). Based on these findings we can conclude that the effect of ETO

to improve short-term spatial memory can be explained through these receptors and their role on synaptic plasticity, which is the fundamental mechanism for memory traces.

Cheng and colleagues, using whole cell patch clamp technique on hippocampal pyramidal cells, have shown the ETO-induced amnesia through selective modulation of $\alpha 5\beta 2$ -containing GABAARs (38). In addition, it has been shown that ETO at doses that impair memory may causes sedation, as indicated by a decrease in exploratory activity (40). Considering that sedation is attributed to the modulation of $\beta 2$ -containing GABAARs (41, 42), it seems likely that it has contributed to the $\beta 2$ -GABAAR-mediated learning impairment (43). In the present study, we could not observe any memory impairment in ETO group compare with control group. This discrepancy may be because of the different experimental protocol and dosage of ETO compared to other studies.

Previous studies demonstrated that pretreatment with DEX could reduce learning and memory impairment, and decrease neuronal cell apoptosis, inflammation and oxidative stress. Moreover, the neuroprotective effects of DEX was reversed by yohimbine, an $\alpha 2$ adrenoceptor antagonist, suggesting that the effects of DEX may be mediated by $\alpha 2$ adrenoceptors(13). DEX increases extracellular signal regulated kinases (ERK) $\frac{1}{2}$ phosphorylation, a key mitogen-activated protein kinase involved in cell survival and memory by the activation of protein kinase C, and probably imidazoline receptors (44, 45). DEX also increases the expression of growth factors such as epidermal growth factor and brain-derived neurotrophic factors, and then participates in neuroprotection. These molecular properties of DEX may underlie its anti-inflammatory and anti-apoptotic effect (46). Some of these effects of DEX may be explained by the anti-inflammatory and neuroprotective action of this agent. The anti-inflammatory effect of DEX is achieved by inhibiting the TLR4/NF- κ B (47), JAK2-STAT3 (48, 49), and NF- κ B/COX-2 pathways (50, 51); activating the ERK1/2 pathway (52); and releasing acetylcholine (ACh) through anti-sympathetic effects via the cholinergic pathway (53). DEX also reduces neuronal apoptosis via myriad mechanisms that enhance the viability of the neurocyte. The brain is quite sensitive to the annihilation of oxygen-free radicals, and ischemia reperfusion injuries produce an intracorporal antioxidant-peroxidation state imbalance. The excessive free radicals in the body are eliminated by DEX and this pathological chain-reaction is reduced by decreasing malondialdehyde and improving the activity of superoxide dismutase(54, 55).

In addition, it has been reported that DEX reduces fear memory consolidation without altering symptoms associated with PTSD in animal models. Also, it is reported that DEX reduces fear memory consolidation in an inhibitory avoidance task (12, 20) which is suggested that this effect may result from anxiolytic effect of DEX. Furthermore, it has been shown that DEX treatment after acute stress at sub-anesthetic doses reduces fear memory and anxiety behavior in a mice stress full model(21). In our results, we found facilitating effects of DEX on short term and long term spatial memory which may be because of anxiolytic effect of DEX at sedative dose which was reported in previous studies. In this study, for the first time, we compared the effect of DEX and ETO on reference and working spatial memory together with height stress as an acute stress model. In this study our acute stress model could not affect spatial memory and it means that this point should be examined using different stress models such as

immobility stress or swimming stress which remain to be elucidated. On the other hand, some discrepancies with previous findings concerning the role of this agent in learning and memory may be explained by the different behavioral task models which were used (fear memory task or passive avoidance task versus Barnes spatial memory).

Conclusion

Our results suggest that DEX administration after experiencing a stress event might prevent the occurrence of emotional distress and subsequent stress disorder development. The results showed that the neuron protection, synaptic formation and memory improving effects of DEX positively affect on learning and remembering reference spatial memory and also remembering working memory. However, the precise neuronal and molecular mechanisms of these effects and their relation to the anti-stress system is still unknown and requires further research.

Materials And Methods:

Animals:

Animal care and experimentation were performed in accordance with protocols approved by the Science and Animal Experimentation ethics Committee no. IR.UMSHA.REC.1397.064 (Hamadan University of medical sciences).

Forty-eight mice (25–30 g) were purchased from the Neuroscience Research Center of Laboratory Animals, Each group contained 8 animals. Only males were chosen to reduce variability due to endocrine cycling. All mice were kept in a controlled air-conditioned room with food and water available ad-libitum with a controlled temperature ($22 \pm 2^\circ\text{C}$) under a 12-hr light/dark cycle with lights on 7:00 A.M.–7:00 P.M..The adaptability phase for the Mice was at least 7 days before the onset of experiments. Training and testing was performed during the light phase of the cycle between 9:00 AM and 5:00 PM

Drug preparation and administration

Etomidate (11mg/kg; [B.Braun Medical S.R.L](#)) and Dexmedetomidine (0.3 or 0.4 mg/kg; Exir, Tehran, Iran) were dissolved in saline solution (0.9%). Drugs solutions were freshly prepared the day of the experiment and were administered by intraperitoneal injection 30 minutes before trial (43). Drug doses were chosen based on previous studies performed (12, 40).

Experiment protocol

Barnes Maze model:

Spatial learning and working memory were evaluated using the Barnes maze (56). The maze consists of a circular platform (92-cm diameter) surrounded by 20 holes (5- cm diameter, equidistant), one providing

an escape route and was elevated 90 cm above the floor. The mice were placed in a dark chamber in the center of the maze and aversive stimuli subsequently triggered (bright light and noise). The animal was then given the opportunity to leave the maze by crawling through the hole allowing the escape. There was a small dark recessed "goal box" (20x 9 x 9 cm) located under one of the holes where mice can escape the aversive stimulus (bright light) and hide. Charts were pasted on the walls to act as visual cues and were kept permanently in their positions throughout the days of the study. A videocamera was positioned about 150 cm above the platform to record the activities (57).

Behavioral procedures:

In the spatial acquisition phase, animals were subjected to five trials per day for four consecutive days, with a constant inter-trial interval time of 20 min. The number of holes explored and the time required for the mice to locate the escape hole, were quantified. For the reference memory phase or probe trials, animals were tested on the 24hr and 7th day (short-term retention) following the first trials, omitting training sessions between test trials. At some point in the probe trial, the escape hole was removed and replaced with a standard hole which the animal could not enter through. The cut off time was 300-s period and mice were located in the box for 30s. The latency to reach the virtual target hole and number of errors were recorded, the outcome measure corresponding to the spatial memory capacity of the animal (33). For the purpose of the spatial working memory test, the escape hole was moved 180 degrees from its original location and the first working trial were done. In the second trial (working memory test) escape latency were measured when animal reached to the escape hole.

Elevated platform (EPF) exposure (Height stress):

For height stress, animal were exposed to EPF before Barnes maze session for 5 min (58). Each mouse was individually placed in the center of the EPF (diameter 18 cm; elevation 75 cm; room light 160 lux) and then the Barnes maze test was carried out.

Experimental groups:

For the study a total of 48 male mice (age 8-10 weeks) were obtained from the Experimental Animal Center of Hamadan University of Medical Sciences. The study groups included control 1, control 2 (Control+H.S), DEX, ETO, DEX+H.S and ETO+H.S; the control group received saline, DEX received Dexmedetomidine and ETO received Etomidate intraperitoneally.

Statistical analysis

The tests were carried out by a person who was blinded to the condition of the animals. The results are presented as the means \pm standard error of the mean (SEM). Comparisons between the experimental and control groups were performed using two-way analysis of variance (ANOVA), followed by Bonferroni or Tukeys post hoc tests when appropriate. A value of $p < 0.05$ was considered as significant.

Declarations

Ethics approval and consent to participate:

all experimental procedures using mice were conducted in accordance with the animal care and use guidelines approved by the institutional ethics committee at Hamadan University of Medical Sciences (Code of Ethics Committee: Grant Number: IR.UMSHA.REC.1397.064) and were performed in accordance with the National Institutes of Health Guide for Care and Use of Laboratory Animals

Consent for publication:

Not applicable.

Availability of data and materials:

The data are available for any scientific use with kind permission.

Competing interests:

The authors declare that they have no competing interests.

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Authors' contributions:

A.Emam and A. Sarihi conceived of the presented idea. M. Tarbiat. developed the theory and performed the computations. M. Rezaei. verified the analytical methods. A. Emam Drafted or provided critical revision of the article and A. Sarihi Provided final approval of the version to publish. All authors discussed the results and contributed to the final manuscript.

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References

1. Yehuda R. Post-traumatic stress disorder. *New England journal of medicine*. 2002;346(2):108–14.
2. Zoladz PR, Fleshner M, Diamond DM. Differential effectiveness of tianeptine, clonidine and amitriptyline in blocking traumatic memory expression, anxiety and hypertension in an animal model of PTSD. *Prog Neuropsychopharmacol Biol Psychiatry*. 2013;44:1–16.
3. Cimadevilla JM, Fenton AA, Bures J. Functional inactivation of dorsal hippocampus impairs active place avoidance in rats. *Neurosci Lett*. 2000;285(1):53–6.
4. Vorhees CV, Williams MT. Assessing spatial learning and memory in rodents. *ILAR journal*. 2014;55(2):310–32.
5. Parsons RG, Ressler KJ. Implications of memory modulation for post-traumatic stress and fear disorders. *Nature neuroscience*. 2013;16(2):146–53.
6. Johansen JP, Cain CK, Ostroff LE, LeDoux JE. Molecular mechanisms of fear learning and memory. *Cell*. 2011;147(3):509–24.
7. Shemesh E, Yehuda R, Milo O, Dinur I, Rudnick A, Vered Z, et al. Posttraumatic stress, nonadherence, and adverse outcome in survivors of a myocardial infarction. *Psychosom Med*. 2004;66(4):521–6.
8. Osterman JE, Hopper J, Heran WJ, Keane TM, van der Kolk BA. Awareness under anesthesia and the development of posttraumatic stress disorder. *Gen Hosp Psychiatry*. 2001;23(4):198–204.
9. Kapfhammer HP, Rothenhäusler HB, Krauseneck T, Stoll C, Schelling G. Posttraumatic stress disorder and health-related quality of life in long-term survivors of acute respiratory distress syndrome. *Am J Psychiatry*. 2004;161(1):45–52.
10. Schelling G, Stoll C, Haller M, Briegel J, Manert W, Hummel T, et al. Health-related quality of life and posttraumatic stress disorder in survivors of the acute respiratory distress syndrome. *Critical care medicine*. 1998;26(4):651–9.
11. Hauer D, Kaufmann I, Strewe C, Briegel I, Campolongo P, Schelling G. The role of glucocorticoids, catecholamines and endocannabinoids in the development of traumatic memories and posttraumatic stress symptoms in survivors of critical illness. *Neurobiol Learn Mem*. 2014;112:68–74.
12. Morena M, Berardi A, Peloso A, Valeri D, Palmery M, Trezza V, et al. Effects of ketamine, dexmedetomidine and propofol anesthesia on emotional memory consolidation in rats: Consequences for the development of post-traumatic stress disorder. *Behav Brain Res*. 2017;329:215–20.
13. Zhang Y, Li M, Cui E, Zhang H, Zhu X, Zhou J, et al. Dexmedetomidine attenuates sevoflurane-induced neurocognitive impairment through α_2 -adrenoceptors. *Mol Med Rep*. 2021;23(1):1-.
14. Liu S, Wang Y, Zhu Y, Yu T, Zhao H. Safety and sedative effect of intranasal dexmedetomidine in mandibular third molar surgery: a systematic review and meta-analysis. *Drug Des Devel Ther*. 2019;13:1301.

15. Zhang X, Wang J, Qian W, Zhao J, Sun L, Qian Y, et al. Dexmedetomidine inhibits tumor necrosis factor-alpha and interleukin 6 in lipopolysaccharide-stimulated astrocytes by suppression of c-Jun N-terminal kinases. *Inflammation*. 2014;37(3):942–9.
16. Zhang Y, Xing Z, Xu Y, Xu S. Effects of different doses of dexmedetomidine on cognitive dysfunction in elderly patients early after laparoscopic surgery for colorectal cancer. *Nan fang yi ke da xue xue bao = Journal of Southern Medical University*. 2014;34(5):743–6.
17. Bekker A, Haile M, Kline R, Didehvar S, Babu R, Martiniuk F, et al. The effect of intraoperative infusion of dexmedetomidine on quality of recovery after major spinal surgery. *J Neurosurg Anesthesiol*. 2013;25(1):16.
18. Yang S, Lee H. A dose-finding study of preoperative intravenous dexmedetomidine in children's emergence delirium after epiblepharon surgery. *Eur J Ophthalmol*. 2014;24(3):417–23.
19. van Oostrom H, Stienen PJ, Doornenbal A, Hellebrekers LJ. The α_2 -adrenoceptor agonist dexmedetomidine suppresses memory formation only at doses attenuating the perception of sensory input. *Eur J Pharmacol*. 2010;629(1–3):58–62.
20. Sirviö J, Riekkinen P Jr, Ekonsalo T, Lammintausta R, Riekkinen P. The effects of dexmedetomidine, an α_2 agonist, on learning and memory, assessed using passive avoidance and water maze tasks in rats. *Neuropharmacology*. 1992;31(2):163–8.
21. Ji M-H, Jia M, Zhang M-Q, Liu W-X, Xie Z-C, Wang Z-Y, et al. Dexmedetomidine alleviates anxiety-like behaviors and cognitive impairments in a rat model of post-traumatic stress disorder. *Prog Neuropsychopharmacol Biol Psychiatry*. 2014;54:284–8.
22. Olsen RW, Sieghart W. International Union of Pharmacology. LXX. Subtypes of γ -aminobutyric acidA receptors: classification on the basis of subunit composition, pharmacology, and function. *Update Pharmacological reviews*. 2008;60(3):243–60.
23. Uchida I, Kamatchi G, Burt D, Yang J. Etomidate potentiation of GABAA receptor gated current depends on the subunit composition. *Neurosci Lett*. 1995;185(3):203–6.
24. Jones MV, Brooks PA, Harrison NL. Enhancement of gamma-aminobutyric acid-activated Cl-currents in cultured rat hippocampal neurones by three volatile anaesthetics. *J Physiol*. 1992;449(1):279–93.
25. Jones MV, Harrison NL. Effects of volatile anesthetics on the kinetics of inhibitory postsynaptic currents in cultured rat hippocampal neurons. *J Neurophysiol*. 1993;70(4):1339–49.
26. Djuric M, Kostic S, Turnic TN, Stankovic S, Skrbic R, Djuric D, et al. The comparison of the effects of ketamine and etomidate on cardiodynamics, biochemical and oxidative stress parameters in Wistar male rats. *Mol Cell Biochem*. 2020;474(1):125–34.
27. GOODING JM, CORSSSEN G. Effect of etomidate on the cardiovascular system. *Anesthesia Analgesia*. 1977;56(5):717–9.
28. GOLDBERG AH, KEANE PW, Phear W. Effects of ketamine on contractile performance and excitability of isolated heart muscle. *J Pharmacol Exp Ther*. 1970;175(2):388–94.
29. Reves J, Glass P, Lubarsky D, McEvoy M. Intravenous nonopioid anesthetics In: Miller RD, ed. *Miller's anesthesia 6th ed Philadelphia: Churchill Livingstone*. 2005:317 – 78.

30. Flynn G, Shehabi Y. Pro/con debate: Is etomidate safe in hemodynamically unstable critically ill patients? *Crit Care*. 2012;16(4):1–5.
31. Forman SA. Clinical and molecular pharmacology of etomidate. *Anesthesiology: The Journal of the American Society of Anesthesiologists*. 2011;114(3):695–707.
32. Mohamad FH, Has ATC. The $\alpha 5$ -Containing GABA A Receptors—a Brief Summary. *J Mol Neurosci*. 2019;67(2):343–51.
33. Antunes MS, Ladd FVL, Ladd AABL, Moreira AL, Boeira SP, Souza LC. Hesperidin protects against behavioral alterations and loss of dopaminergic neurons in 6-OHDA-lesioned mice: the role of mitochondrial dysfunction and apoptosis. *Metab Brain Dis*. 2021;36(1):153–67.
34. Kurawa MI, Magaji RA, Yusuf T, Magaji MG. Effects of Mosquito coil Smoke Inhalation on Spatial Memory in Mice. *Nigerian Journal of Physiological Sciences*. 2020;35(1):68–76.
35. Caraiscos VB, Elliott EM, You-Ten KE, Cheng VY, Belelli D, Newell JG, et al. Tonic inhibition in mouse hippocampal CA1 pyramidal neurons is mediated by $\alpha 5$ subunit-containing γ -aminobutyric acid type A receptors. *Proceedings of the National Academy of Sciences*. 2004;101(10):3662-7.
36. Martin LJ, Oh GH, Orser BA. Etomidate targets $\alpha 5\gamma$ -aminobutyric acid subtype A receptors to regulate synaptic plasticity and memory blockade. *The Journal of the American Society of Anesthesiologists*. 2009;111(5):1025–35.
37. Orser BA. Lifting the fog around anesthesia. *Sci Am*. 2007;296(6):54–61.
38. Cheng VY, Martin LJ, Elliott EM, Kim JH, Mount HT, Taverna FA, et al. $\alpha 5$ GABAA receptors mediate the amnestic but not sedative-hypnotic effects of the general anesthetic etomidate. *J Neurosci*. 2006;26(14):3713–20.
39. Banks MI, White JA, Pearce RA. Interactions between distinct GABAA circuits in hippocampus. *Neuron*. 2000;25(2):449–57.
40. Benkwitz C, Liao M, Laster MJ, Sonner JM, Eger EI, Pearce RA. Determination of the EC50 Amnesic Concentration of Etomidate and Its Diffusion Profile in Brain Tissue: Implications for In Vitro Studies. *The Journal of the American Society of Anesthesiologists*. 2007;106(1):114–23.
41. Reynolds DS, Rosahl TW, Cirone J, O'Meara GF, Haythornthwaite A, Newman RJ, et al. Sedation and anesthesia mediated by distinct GABAA receptor isoforms. *J Neurosci*. 2003;23(24):8608–17.
42. Jurd RAM, Lambert S, Drexler B, Siegwart R, Crestani F, Zaugg M, Vogt KE, Ledermann B, Antkowiak B, Rudolph U. General anesthetic actions in vivo strongly attenuated by a point mutation in the GABAA receptor $\beta 3$ subunit. *FASEB J*. 2003;17(2):250–2.
43. Zarnowska E, Rodgers F, Oh I, Rau V, Lor C, Laha K, et al. Etomidate blocks LTP and impairs learning but does not enhance tonic inhibition in mice carrying the N265M point mutation in the $\beta 3$ subunit of the GABAA receptor. *Neuropharmacology*. 2015;93:171–8.
44. Xue L, Murray JH, Tolkovsky AM. The Ras/phosphatidylinositol 3-kinase and Ras/ERK pathways function as independent survival modules each of which inhibits a distinct apoptotic signaling pathway in sympathetic neurons. *J Biol Chem*. 2000;275(12):8817–24.

45. Dahmani S, Paris A, Jannier V, Hein L, Rouelle D, Scholz J, et al. Dexmedetomidine Increases Hippocampal Phosphorylated Extracellular Signal-regulated Protein Kinase 1 and 2 Content by an α 2-Adrenoceptor-independent Mechanism: Evidence for the Involvement of Imidazoline I1 Receptors. *The Journal of the American Society of Anesthesiologists*. 2008;108(3):457–66.
46. Qian X-L, Zhang W, Liu M-Z, Zhou Y-B, Zhang J-M, Han L, et al. Dexmedetomidine improves early postoperative cognitive dysfunction in aged mice. *Eur J Pharmacol*. 2015;746:206–12.
47. Smrcka AV, Fisher I. G-protein $\beta\gamma$ subunits as multi-functional scaffolds and transducers in G-protein-coupled receptor signaling. *Cell Mol Life Sci*. 2019;76(22):4447–59.
48. Lavon H, Matzner P, Benbenishty A, Sorski L, Rossene E, Haldar R, et al. Dexmedetomidine promotes metastasis in rodent models of breast, lung, and colon cancers. *Br J Anaesth*. 2018;120(1):188–96.
49. Lankadeva YR, Ma S, Iguchi N, Evans RG, Hood SG, Farmer DG, et al. Dexmedetomidine reduces norepinephrine requirements and preserves renal oxygenation and function in ovine septic acute kidney injury. *Kidney international*. 2019;96(5):1150–61.
50. Bao N, Tang B, Wang J. Dexmedetomidine preconditioning protects rats from renal ischemia-reperfusion injury accompanied with biphasic changes of nuclear factor-kappa B signaling. *Journal of immunology research*. 2020;2020.
51. Yao H, Chi X, Jin Y, Wang Y, Huang P, Wu S, et al. Dexmedetomidine inhibits TLR4/NF- κ B activation and reduces acute kidney injury after orthotopic autologous liver transplantation in rats. *Scientific reports*. 2015;5(1):1–12.
52. de Castro Abrantes H, Briquet M, Schmuziger C, Restivo L, Puyal J, Rosenberg N, et al. The lactate receptor HCAR1 modulates neuronal network activity through the activation of $G\alpha$ and $G\beta\gamma$ subunits. *J Neurosci*. 2019;39(23):4422–33.
53. Gu J, Sun P, Zhao H, Watts HR, Sanders RD, Terrando N, et al. Dexmedetomidine provides renoprotection against ischemia-reperfusion injury in mice. *Critical care*. 2011;15(3):1–11.
54. Li Y, Liu S. The effect of dexmedetomidine on oxidative stress response following cerebral ischemia-reperfusion in rats and the expression of intracellular adhesion molecule-1 (ICAM-1) and S100B. *Medical science monitor: international medical journal of experimental clinical research*. 2017;23:867.
55. Yeh Y-C, Wu C-Y, Cheng Y-J, Liu C-M, Hsiao J-K, Chan W-S, et al. Effects of dexmedetomidine on intestinal microcirculation and intestinal epithelial barrier in endotoxemic rats. *Anesthesiology*. 2016;125(2):355–67.
56. Walker J, Fowler S, Miller D, Sun A, Weisman G, Wood W, et al. Spatial learning and memory impairment and increased locomotion in a transgenic amyloid precursor protein mouse model of Alzheimer's disease. *Behav Brain Res*. 2011;222(1):169–75.
57. Barnes CA. Memory deficits associated with senescence: a neurophysiological and behavioral study in the rat. *Journal of comparative physiological psychology*. 1979;93(1):74.
58. Uschold-Schmidt N, Nyuyki KD, Fuchsl AM, Neumann ID, Reber SO. Chronic psychosocial stress results in sensitization of the HPA axis to acute heterotypic stressors despite a reduction of adrenal

Figures

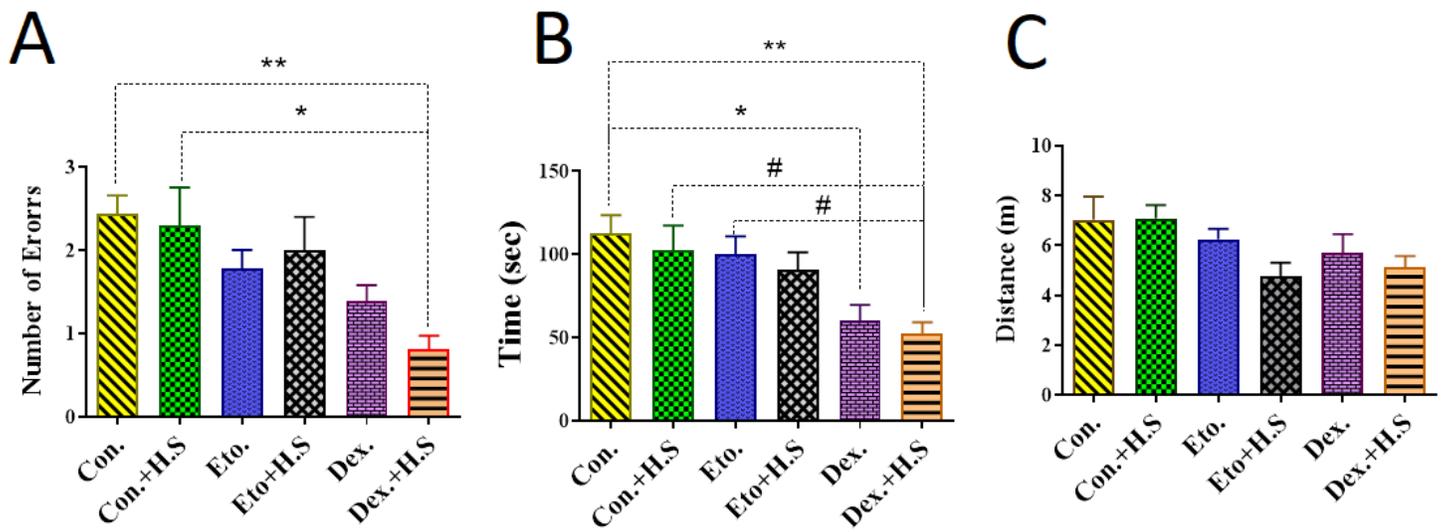


Figure 1

Effect of acute stress with DEX and ETO on Reference spatial learning A: comparison number of errors (* $P < 0.05$, ** $P < 0.01$), B: comparison of spent time to finding target hole (*, # $P < 0.05$, ** $P < 0.01$) C: comparison of Distance traveled.

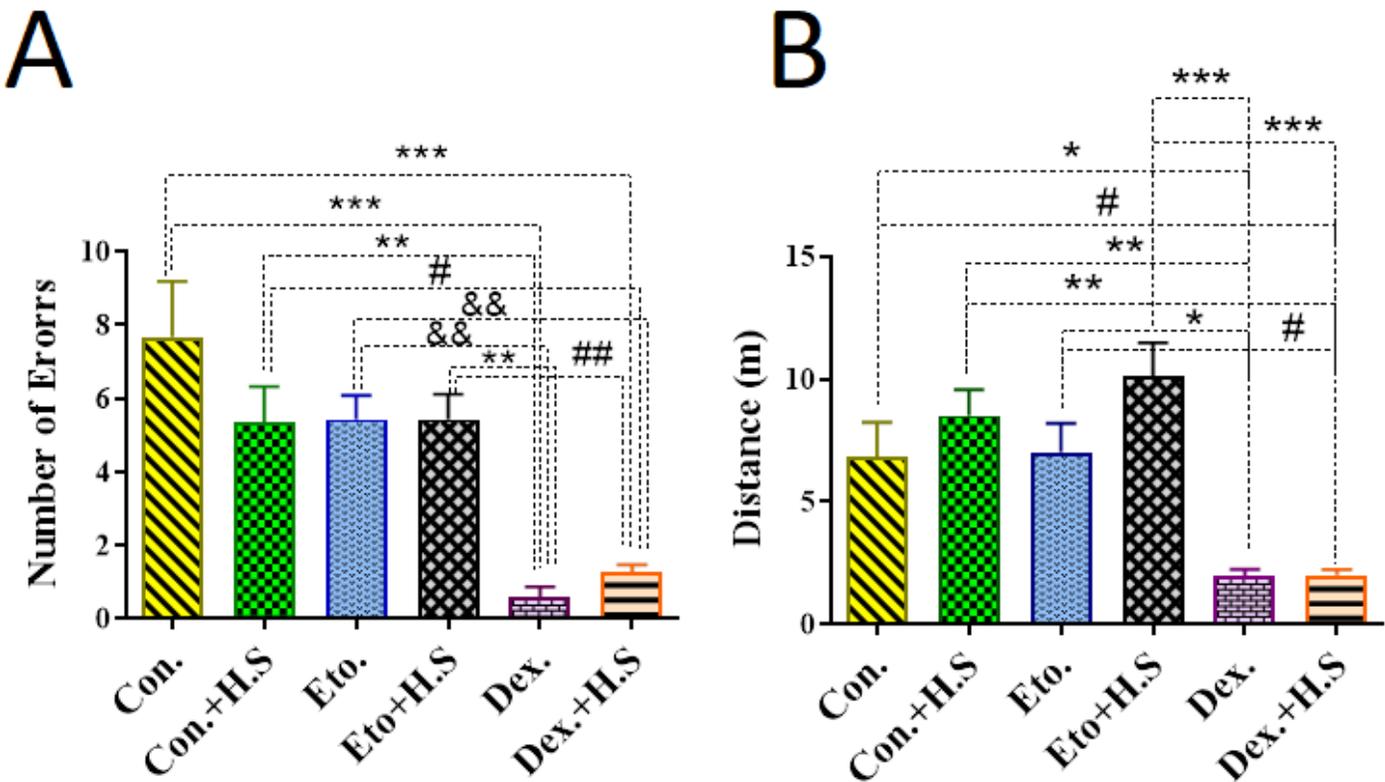


Figure 2

Effect of acute stress with DEX and ETO on recalling reference spatial memory A: comparison number of errors (# P<0.05 , ***,**,##,&& P<0.01) B: comparison of distance traveled to finding target hole (#,* P<0.05 ** P<0.01 *** P<0.001).

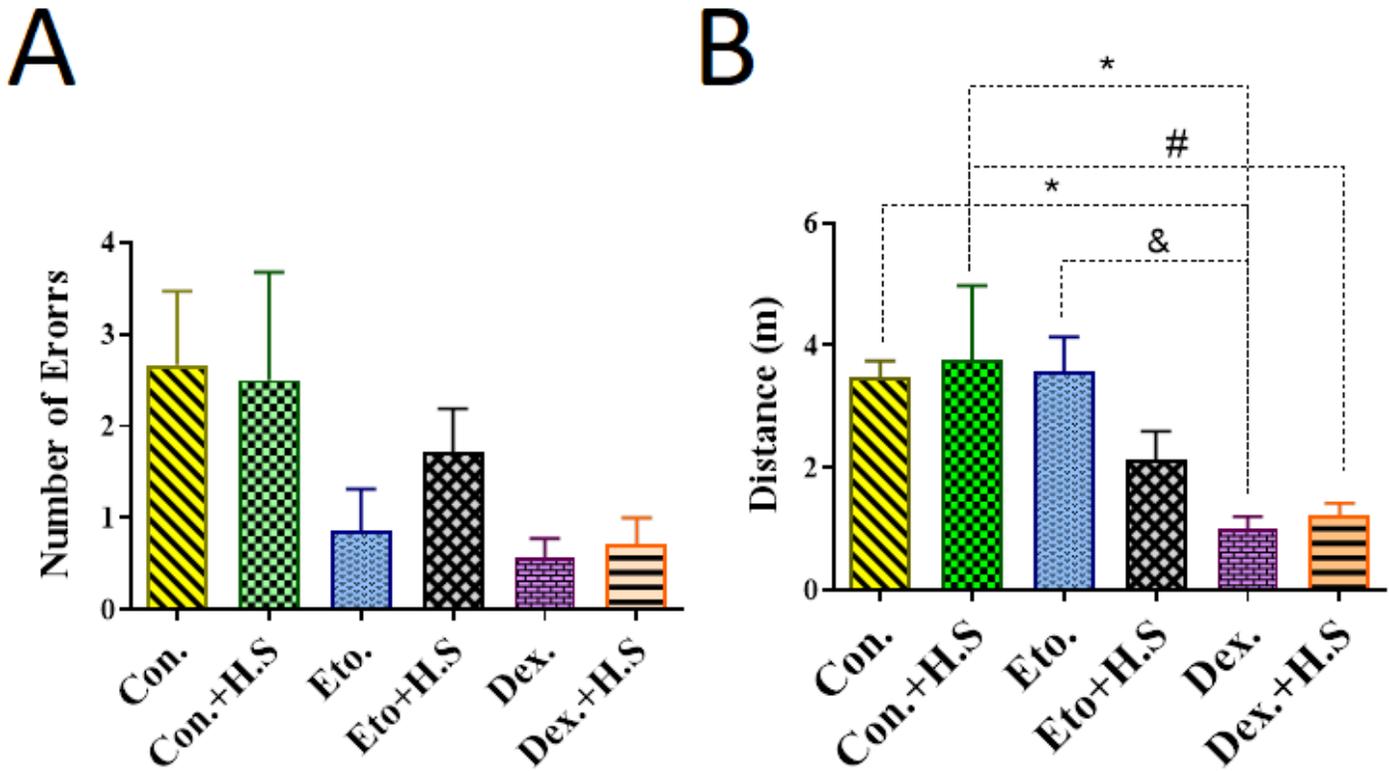
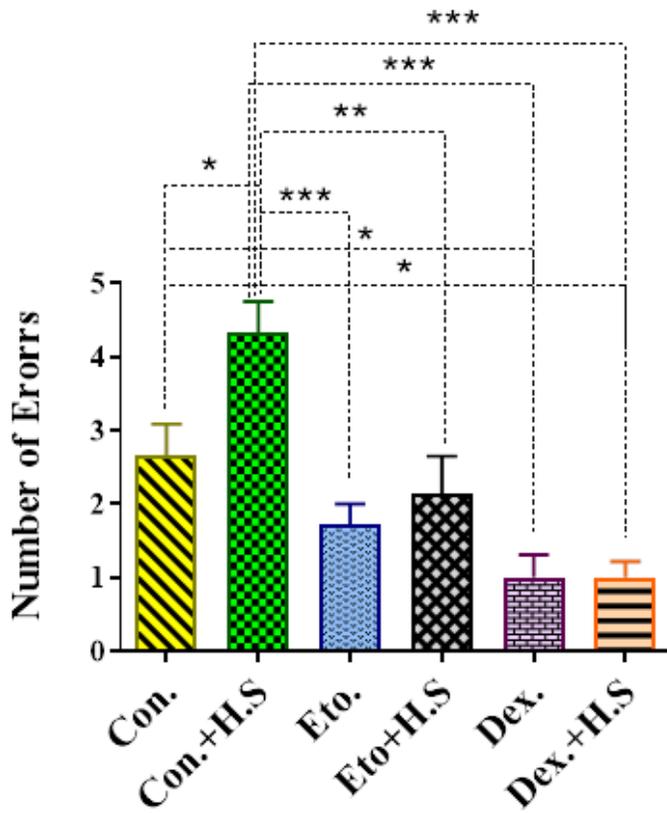
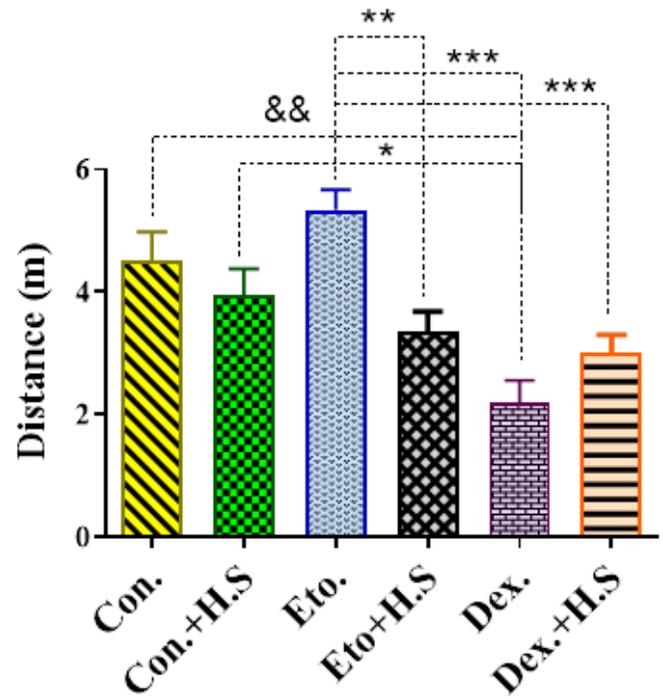


Figure 3

Effect of acute stress with DEX and ETO on learn spatial working memory A: comparison number of errors B: comparison of distance traveled to finding target hole (#,* & P<0.05).

A**B****Figure 4**

Effect of acute stress with DEX and ETO on recalling spatial working memory A: comparison number of errors (* $P < 0.05$ □ ** $P < 0.01$ □ *** $P < 0.001$) B: comparison of distance traveled to finding target hole (* $P < 0.05$ □ &&, ** $P < 0.01$, *** $P < 0.001$).