

Genetic polymorphisms of *PGF* and *TNFAIP2* genes related to cervical cancer risk among Uygur females from China

Zumurelaiti Ainiwaer

People's Hospital of Xinjiang Uygur Autonomous

Reyilanmu Maisaidi

People's Hospital of Xinjiang Uygur Autonomous

Jing Liu

People's Hospital of Xinjiang Uygur Autonomous

Lili Han (✉ hanlili19941226@163.com)

People's Hospital of Xinjiang Uygur Autonomous Region <https://orcid.org/0000-0002-8664-0820>

Sulaiya Husaiyin

People's Hospital of Xinjiang Uygur Autonomous

Jing Lu

People's Hospital of Xinjiang Uygur Autonomous

Mayinuer Niyazi

People's Hospital of Xinjiang Uygur Autonomous

Research article

Keywords: PGF, TNFAIP2, Polymorphism, Cervical cancer

Posted Date: September 15th, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-30116/v2>

License: © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License. [Read Full License](#)

Version of Record: A version of this preprint was published at BMC Medical Genetics on October 27th, 2020. See the published version at <https://doi.org/10.1186/s12881-020-01144-5>.

Abstract

Background: *PGF* and *TNFAIP2* are important angiogenic factors, which were abnormal expression in cervical cancer (CC). However, there is currently no report investigating the relationship of *PGF* and *TNFAIP2* gene polymorphisms to CC risk.

Methods: We conducted a case-control study of 342 CC patients and 498 cancer-free controls in a Chinese Uyur female population. Three SNPs (*PGF* rs8019391, *PGF* rs2268615, and *TNFAIP2* rs710100) were selected and genotyped to assess the possible association of *PGF* and *TNFAIP2* polymorphisms with CC susceptibility. Logistic regression analysis adjusted by age was used.

Results: *PGF* rs2268615 (OR = 1.39, 95% CI = 1.04-1.86, $p = 0.024$) and *TNFAIP2* rs710100 (OR = 1.44, 95% CI = 1.07-1.95, $p = 0.018$) polymorphisms were associated with the increased risk of CC. Moreover, T allele of *PGF* rs8019391 was highly represented in patients with stage III–IV compared with stage I–II (OR = 2.17, $p = 4.58 \times 10^{-4}$). MDR analysis revealed a positive interaction between the SNPs.

Conclusion: Our data indicated that *PGF* rs2268615, and *TNFAIP2* rs710100 polymorphisms might be risk factors for CC susceptibility, which contributed to the increased risk of CC.

Trail registration: Not applicable.

Background

Worldwide, cervical cancer (CC) is the fourth most common cancer, with an estimated 570,000 cases and 311,000 deaths in 2018(1). Cervical cancer is the most common cancer of female genital system in China and the incidence of CC tends to be younger (2). A wide range of inter-individual genetic variability and other pathogeneses might contribute to cervical carcinogenesis. Accumulating evidence suggested that single nucleotide polymorphisms (SNPs) in tumor-associated genes played important role in the genetic susceptibility to CC, such as *FAS*, *IL-17A*, *IL-17F* and *HOTAIR* polymorphisms (3-5).

PGF (placental growth factor) gene, also named *PLGF*, encodes a homologous of vascular endothelial growth factor. *PGF* has been reported as a potent stimulator in cancer invasion by activating angiogenesis(6). In addition, the overexpression of *PGF* is correlated with tumor stage, cancer progression and metastasis(7). *TNFAIP2* (TNF alpha induced protein 2) is a primary response gene of TNF α , and the expression of *TNFAIP2* is regulated by various transcription factors and signaling pathways, including NF- κ B, *KLF5* and retinoic acid pathways (8). *TNFAIP2* is an important angiogenic factor, and is significantly associated with intratumoral microvessel density(9). In addition, previous studies reported the abnormal expression of *PGF* and *TNFAIP2* in human cancer, including CC(6, 10). However, there is no report about the associations between *PGF* and *TNFAIP2* polymorphisms and CC risk. We hypothesized that genetic variants in *PGF* and *TNFAIP2* could contribute to CC susceptibility.

Here, *PGF* rs8019391 and rs2268615, and *TNFAIP2* rs710100 were selected as candidate SNPs from the NCBI dbSNP database (<http://www.ncbi.nlm.nih.gov/projects/SNP>) and the 1000 Genomes Project data (<http://www.internationalgenome.org/>), based on minor allele frequency (MAF) of at least 5 % in Chinese populations, with a pairwise $r^2 > 0.80$ and the call rate of genotyping $> 95\%$. The aim of our study was to investigate the possible association between three SNPs (*PGF* rs8019391, *PGF* rs2268615, and *TNFAIP2* rs710100) and the risk of CC in Chinese Uyur female population.

Subjects And Methods

Study participants

This study protocol was approved by the Ethics Committee of the People's Hospital of Xinjiang Uygur Autonomous Region and was in accordance with the guidelines of the Declaration of Helsinki. Written informed consents were obtained from all participants.

342 cervical cancer patients (cases, 43.27 ± 11.78 years) and 498 age-matched cancer-free individuals (controls, 43.46 ± 13.03 years) were enrolled from the People's Hospital of Xinjiang Uygur Autonomous Region, as shown in Table 1. All recruited subjects were unrelated ethnic Han Chinese females. Patients were newly diagnosed and histopathologically confirmed primary cervical cancer according to the clinical staging standards of the International Federation of Gynecology and Obstetrics (FIGO). Among patients, 132 cases were stage I–II, 80 cases were stage III–IV and 130 cases were missing. Patients with any history of other cancers, radiotherapy, chemotherapy, or surgery and inflammatory diseases were excluded. The controls were recruited from the health checkup in the same hospital during the same period. Controls had no any history of cancers and diseases of the liver, kidneys, heart, brain, and vascular system, as well as gynecological inflammatory diseases. Demographic characteristics and clinical information were collected from the standardized questionnaires.

SNP selection and genotyping

Peripheral venous blood samples (5 mL) were obtained from all subjects and stored in EDTA-coated tubes. Genomic DNA was extracted using the GoldMag DNA Purification Kit (GoldMag Co. Ltd, Xi'an City, China) according to the manufacturer's protocol, then quantified by NanoDrop 2000 (Thermo Scientific, Waltham, MA, USA), and stored at -20°C for further experiments. *PGF* rs8019391 and rs2268615, and *TNFAIP2* rs710100 were selected as candidate from the NCBI dbSNP database (<http://www.ncbi.nlm.nih.gov/projects/SNP>) and the 1000 Genomes Project data (<http://www.internationalgenome.org/>), based on minor allele frequency (MAF) of at least 5 % in Chinese populations, with a pairwise $r^2 > 0.80$, and the call rate of genotyping $> 95\%$. SNPs. miRNASNP_v2 database (<http://bioinfo.life.hust.edu.cn/miRNASNP2/index.php>) and HaploReg v4.1 (<https://pubs.broadinstitute.org/mammals/haploreg/haploreg.php>) were used to predict the potential function of these polymorphisms (Supplementary Table 1). These candidate SNPs were genotyped with Agena MassARRAY system (Agena, San Diego, CA, U.S.A.) as described previously(11, 12), and performed by two laboratory technicians in a double-blinded fashion. The primers for PCR amplification and single base extension were designed using the Assay Design 3.0 software (Supplementary Table 2). For quality control, approximately 10% of the samples were randomly selected and repeated genotyping, and 100% concordance rate was observed.

Data analysis

All statistical analysis was performed using SPSS version 18.0 (SPSS Inc., Chicago, IL, USA) and PLINK software. A student's t-test was performed to analyze the differences in the age distribution between patients and controls. The Hardy–Weinberg equilibrium (HWE) was tested by a goodness-of-fit χ^2 test for each SNP among the control subjects. The genotype and allele frequencies between two groups were compared using χ^2 test. The association of candidate SNPs with CC risk was assessed by odds ratios (ORs) and 95% confidence intervals (CIs) using logistic regression analysis for both combined and respective genotype(13). Stratification analysis by demographic and clinic variables was also performed to assess the genetic association. Multifactor dimensionality reduction (MDR) (version 3.0.2) was used to evaluate the SNP–SNP interactions in CC risk (14). A two-tailed p -value < 0.05 was considered to be statistically significant for all the analyses.

Results

In Supplementary table 3, the MAF of *PGF* rs8019391 and rs2268615, and *TNFAIP2* rs710100 between the case and control groups were listed. The genotype distribution of these SNPs in controls were in accordance with the Hardy-Weinberg equilibrium ($p > 0.05$). The call rate for rs8019391, rs2268615 and rs710100 were 100%, 99.7% and 99.1%, respectively. The MAFs distribution of *PGF* rs2268615-A allele and *TNFAIP2* rs710100-A allele were higher in the case group, which increased the risk of CC (rs2268615, A vs C, OR = 1.27, 95% CI = 1.03-1.58, $p = 0.029$; and rs710100, A vs G, OR = 1.23, 95% CI = 1.01-1.50, $p = 0.043$).

The results of multiple genetic model adjusted by age revealed *PGF* rs2268615 and *TNFAIP2* rs710100 conferred to the increased CC risk (Table 2). *PGF* rs2268615 was associated with an increased risk of CC under heterozygote (OR = 1.39, 95% CI = 1.04-1.86, $p = 0.024$), dominant (OR = 1.40, 95% CI = 1.06-1.84, $p = 0.018$) and log-additive (OR = 1.29, 95% CI = 1.03-1.61, $p = 0.027$) models. For rs710100 in *TNFAIP2*, compared with GG genotype, GA genotype (OR = 1.44, 95% CI = 1.07-1.95, $p = 0.018$) and GA+AA genotype (OR = 1.42, 95% CI = 1.07-1.89, $p = 0.016$) increased 1.44-fold and 1.42-fold CC risk, respectively. Moreover, the result of the additive model also showed an increased risk of CC (rs710100, OR = 1.23, 95% CI = 1.00-1.50, $p = 0.046$). However, there was no significant association between *PGF* rs8019391 and CC susceptibility.

Age stratification displayed that *PGF* rs2268615 and *TNFAIP2* rs710100 increased the risk of CC among women at age ≥ 43 years (Table 3). After calculating the ORs for the allele (OR = 1.38, $p = 0.041$ and OR = 1.42, $p = 0.018$, respectively), genotype (CA vs CC, OR = 1.55, $p = 0.039$; and AA vs GG, OR = 1.97, $p = 0.031$, respectively), dominant (OR = 1.56, $p = 0.030$; and OR = 1.57, 95%, $p = 0.034$, respectively), and log-additive (OR = 1.40, $p = 0.042$; and OR = 1.42, $p = 0.020$) genetic models, they all displayed the genetic association of *PGF* rs2268615 and *TNFAIP2* rs710100 with CC susceptibility.

Subsequently, stratification analysis by tumor stage showed that the risk effect for *PGF* rs8019391 appeared to be more prominent in the subset of patients with stage III+IV (Table 4). Compared with the C allele, rs8019391 T allele was highly represented in patients with stage III–IV as compared to patients with stage I–II under the allele (OR = 2.17, $p = 4.58 \times 10^{-4}$), heterozygote (OR = 2.34, $p = 0.005$), homozygote (OR = 5.76, $p = 0.015$), dominant (OR = 2.59, $p = 0.001$), recessive (OR = 4.13, $p = 0.045$), and log-additive models (OR = 2.36, $p < 0.001$).

Subsequently, MDR analysis was implemented to assess the impact of SNP-SNP interaction. Association of higher order interaction with CC risk was summarized in Figure 1. The result revealed the additive effect between *TNFAIP2* rs710100-GA, *PGF* rs2268615-CA, and *PGF* rs8019391-CT on conferring risk towards the susceptibility to CC. The result of dendrogram and the Fruchterman-Reingold (Figure 2) showed that *PGF* rs2268615, *TNFAIP2* rs710100, *PGF* rs8019391 exhibited a strong synergy effect on CC risk. Table 5 showed that *TNFAIP2* rs710100 was the best single-locus model to predict the risk of CC (testing accuracy = 0.508, CVC = 6/10, $p = 0.014$). The best two-locus model was the combination of *PGF* rs2268615 and *TNFAIP2* rs710100 (testing accuracy = 0.536, CVC = 9/10, $p < 0.0001$). The three-locus model included *TNFAIP2* rs710100, *PGF* rs2268615, and *PGF* rs8019391 (testing accuracy = 0.550, CVC = 10/10, $p < 0.0001$).

Discussion

In this case–control study, we assessed the association between three SNPs (*PGF* rs8019391, *PGF* rs2268615, and *TNFAIP2* rs710100) and CC susceptibility in the Chinese Uyghur female population. We found that *PGF* rs2268615, and *TNFAIP2* rs710100 were associated with the increased risk of CC. Our findings also suggested some possible interaction between genetic variations (*PGF* rs2268615 and *TNFAIP2* rs710100) and age in CC risk. Moreover, the risk effect for *PGF* rs8019391 appeared to be more prominent in patients with stage III+IV. To the best of our knowledge, this is the first report describing that *PGF* and *TNFAIP2* polymorphisms might be risk factor for CC.

PGF, located at 14q24.3, belongs to the vascular endothelial growth factor family and presents on various cell types. The expression of *PGF* in tissue or plasma of cancer patients was upregulated in most human tumor types, including gallbladder, gastric and prostate cancers. *PGF* regulates certain cellular processes such as survival, vascular endothelial cell growth, invasiveness, and *PGF* is also involved in pathological angiogenesis and metastasis (15-17). Previous researches indicated that *PGF* was overexpressed in CC tissues, serum and vaginal lavage compared with adjacent normal tissues or normal women group(6, 18). *PGF* promotes migration by regulating the expression of epithelial-mesenchymal transition-related protein in CC(6). These evidence led us to propose the hypothesis that *PGF* could be of importance in CC occurrence. In our study, we firstly assess the association of *PGF* polymorphisms (rs8019391 and rs2268615) with CC susceptibility, and found that rs2268615 conferred the increased risk of CC, and rs8019391 was a risk factor for patients with stage III-IV. A retrospective population-based study showed that 5-year relative survival rates of CC were 90.9%, 71.0%, 41.7%, and 7.8% for the stage I, II, III, and IV, respectively(19). Therefore, it is highly speculated that *PGF* rs8019391 polymorphism may affect CC progression. Further large-scale studies are needed to verify our findings.

TNFAIP2 gene (also named B94) is located on chromosome 14q32, and encodes TNF α -inducible protein 2. *TNFAIP2* participates in the NF κ B and KLF5 signaling pathway to regulate cell inflammatory, angiogenesis, cell proliferation, migration and invasion (20, 21). The expression of *TNFAIP2* was found to be abnormal in various cancers, including breast cancer, esophageal squamous cell carcinoma, and glioma (20, 22, 23). The expression of *TNFAIP2* was significantly increased in CC tissues compared with normal tissues based on TCGA (The Cancer Genome Atlas) database(8). In addition, the disruption of *TNFAIP2* cytokine/retinoic acid-inducible gene through viral integration contributed to the rapid progression of CC(10). These evidence suggested that *TNFAIP2* might play an important role in the progression of CC. In this study, we found that *TNFAIP2* rs710100 was associated with an increased risk of CC. Rs710100 in the 3'UTR of *TNFAIP2*, located at the predicted miRNAs-binding sites, was related to an increased risk of CC. Specifically, rs1064607 putatively affects the binding of miR-155(24), whose abnormal expression in CC was correlated with FIGO stage, lymph nodes metastasis, and vascular invasion(25). Therefore, we proposed that rs710100 in 3'UTR of *TNFAIP2* might affect *TNFAIP2* expression in CC by disturbing mRNA stability or miRNA binding activity, thus causing a higher risk of CC. However, it should be confirmed in further functional studies.

In spite of interesting findings on the association of *PGF* and *TNFAIP2* polymorphisms with CC risk, several limitations need to be addressed. First, there may be selection and information bias since the retrospective study was designed as a hospital-based case-control study. Second, due to insufficient data of HPV screening results and lifestyle data (e.g., smoking), we could not evaluate HPV infection and lifestyle as the potential confounder in the risk estimation of CC.

Conclusion

In conclusion, our findings indicated a relationship between *PGF* rs2268615, and *TNFAIP2* rs710100 and the increased CC susceptibility in the Chinese Uygur females. Considering that this is the first report on the association of *PGF* and *TNFAIP2* polymorphisms with CC risk, well-designed large and prospective studies are required to validate our findings.

Declarations

Ethics approval and consent to participate

This study protocol was approved by the Ethics Committee of the People's Hospital of Xinjiang Uygur Autonomous Region and under the guidelines of the Declaration of Helsinki. Written informed consent was obtained from all participants.

Consent for publication

Not applicable.

Data Availability Statement

The data that support the findings of this study are available on request from the corresponding author.

Competing interests

The authors declare that they have no competing interests

Funding

This article was financially supported by the National Natural Science Foundation of China (No. 81760467)

Author's contributions

ZA, RM and LLH designed this study protocol and drafted the manuscript; JL1 and SH performed the DNA extraction and genotyping; JL2 and RM performed the data analysis; RM and JL2 performed the sample collection and information recording. MN and LLH conceived and supervised the study. All authors read and approved the final manuscript.

Acknowledgements

The authors thank all the participants in this study.

Abbreviation

CC, cervical cancer; SNPs, single nucleotide polymorphisms; *PGF*, placental growth factor; *TNFAIP2*, TNF alpha induced protein 2.

References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 2018;68(6):394-424.
2. Jiang X, Tang H. Epidemiology of gynecologic cancers in China. 2018;29(1):e7.
3. Tan SC, Ismail MP, Duski DR, Othman NH, Ankathil R. FAS c.-671A>G polymorphism and cervical cancer risk: a case-control study and meta-analysis. *Cancer Genet.* 2017;211:18-25.
4. Lv Q, Zhu D, Zhang J, Yi Y, Yang S, Zhang W. Association between six genetic variants of IL-17A and IL-17F and cervical cancer risk: a case-control study. *Tumour Biol.* 2015;36(5):3979-84.
5. Guo L, Lu X, Zheng L, Liu X, Hu M. Association of Long Non-Coding RNA HOTAIR Polymorphisms with Cervical Cancer Risk in a Chinese Population. *PLoS One.* 2016;11(7):e0160039.
6. Huang W, Zhu S, Liu Q, Li C, Li L. Placenta growth factor promotes migration through regulating epithelial-mesenchymal transition-related protein expression in cervical cancer. *Int J Clin Exp Pathol.* 2014;7(12):8506-19.
7. Dewerchin M, Carmeliet P. Placental growth factor in cancer. *Expert Opin Ther Targets.* 2014;18(11):1339-54.

8. Jia L, Shi Y, Wen Y, Li W, Feng J, Chen C. The roles of TNFAIP2 in cancers and infectious diseases. 2018;22(11):5188-95.
9. Chen LC, Chen CC, Liang Y, Tsang NM, Chang YS, Hsueh C. A novel role for TNFAIP2: its correlation with invasion and metastasis in nasopharyngeal carcinoma. *Mod Pathol.* 2011;24(2):175-84.
10. Einstein MH, Cruz Y, El-Awady MK, Popescu NC, DiPaolo JA, van Ranst M, et al. Utilization of the human genome sequence localizes human papillomavirus type 16 DNA integrated into the TNFAIP2 gene in a fatal cervical cancer from a 39-year-old woman. *Clin Cancer Res.* 2002;8(2):549-54.
11. Xia P, Li B, Geng T, Deng Z, Dang C, Chang D, et al. FGFR2 gene polymorphisms are associated with breast cancer risk in the Han Chinese population. *Am J Cancer Res.* 2015;5(5):1854-61.
12. Liang J, Kang X, Halifu Y, Zeng X, Jin T, Zhang M, et al. Secreted frizzled-related protein promoters are hypermethylated in cutaneous squamous carcinoma compared with normal epidermis. *BMC Cancer.* 2015;15:641.
13. Li S, Jin T, Zhang J, Lou H, Yang B, Li Y, et al. Polymorphisms of TREH, IL4R and CCDC26 genes associated with risk of glioma. *Cancer Epidemiol.* 2012;36(3):283-7.
14. Leem S, Park T. An empirical fuzzy multifactor dimensionality reduction method for detecting gene-gene interactions. *BMC Genomics.* 2017;18(Suppl 2):115.
15. Li H, Jin Y, Hu Y, Jiang L, Liu F, Zhang Y, et al. The PLGF/c-MYC/miR-19a axis promotes metastasis and stemness in gallbladder cancer. 2018;109(5):1532-44.
16. Mahmoodi F, Akrami H. PIGF Knockdown Decreases Tumorigenicity and Stemness Properties of Spheroid Body Cells Derived from Gastric Cancer Cells. *J Cell Biochem.* 2017;118(4):851-9.
17. Zins K, Thomas A, Lucas T, Sioud M, Aharinejad S, Abraham D. Inhibition of stromal PIGF suppresses the growth of prostate cancer xenografts. *Int J Mol Sci.* 2013;14(9):17958-71.
18. Yang S, Cheng H, Cai J, Cai L, Zhang J, Wang Z. PIGF expression in pre-invasive and invasive lesions of uterine cervix is associated with angiogenesis and lymphangiogenesis. *Apmis.* 2009;117(11):831-8.
19. Cheung FY, Mang OW, Law SC. A population-based analysis of incidence, mortality, and stage-specific survival of cervical cancer patients in Hong Kong: 1997-2006. *Hong Kong Med J.* 2011;17(2):89-95.
20. Jia L, Zhou Z, Liang H, Wu J, Shi P, Li F, et al. KLF5 promotes breast cancer proliferation, migration and invasion in part by upregulating the transcription of TNFAIP2. *Oncogene.* 2016;35(16):2040-51.
21. Thair SA, Topchiy E, Boyd JH, Cirstea M, Wang C, Nakada TA, et al. TNFAIP2 Inhibits Early TNFalpha-Induced NF-x03BA;B Signaling and Decreases Survival in Septic Shock Patients. *J Innate Immun.* 2016;8(1):57-66.
22. Xie Y, Wang B. Downregulation of TNFAIP2 suppresses proliferation and metastasis in esophageal squamous cell carcinoma through activation of the Wnt/beta-catenin signaling pathway. *Oncol Rep.* 2017;37(5):2920-8.
23. Cheng Z, Wang HZ, Li X, Wu Z, Han Y, Li Y, et al. MicroRNA-184 inhibits cell proliferation and invasion, and specifically targets TNFAIP2 in Glioma. *J Exp Clin Cancer Res.* 2015;34:27.
24. Liu Z, Wei S, Ma H, Zhao M, Myers JN, Weber RS, et al. A functional variant at the miR-184 binding site in TNFAIP2 and risk of squamous cell carcinoma of the head and neck. *Carcinogenesis.* 2011;32(11):1668-74.
25. Fang H, Shuang D, Yi Z, Sheng H, Liu Y. Up-regulated microRNA-155 expression is associated with poor prognosis in cervical cancer patients. *Biomed Pharmacother.* 2016;83:64-9.

Tables

Table 1. Characteristics of patients with cervical cancer and controls

Characteristics		Cases (n = 342)	Controls (n = 498)	<i>p</i>
Age	Mean ± SD (years)	43.27 ± 11.78	43.46 ± 13.03	0.832
	> 43	176 (51.5%)	263 (52.8%)	
	≤ 43	166 (48.5%)	235 (47.2%)	
HPV status	Negative	51 (14.9%)		
	Positive	195 (57.0%)		
	Missing	96 (28.1%)		
Stage	I-II	132 (38.6%)		
	III-IV	80 (23.4%)		
	Missing	130 (38.0%)		

Table 2. Relationships between the candidate SNPs and cervical cancer risk

Gene SNP ID	Model	Genotype	Case	Control	Adjusted by age and gender	
					OR (95%CI)	<i>p</i>
PGF rs8019391	Genotype	CC	208	327	1.00	
		CT	119	145	1.29 (0.96-1.74)	0.093
		TT	15	26	0.91 (0.47-1.76)	0.777
	Dominant	CC	208	327	1.00	0.150
		CT-TT	134	171	1.23 (0.93-1.64)	
	Recessive	CC-CT	327	472	1.00	0.585
		TT	15	26	0.83 (0.43-1.6)	
Log-additive	--	--	--	1.13 (0.89-1.42)	0.324	
PGF rs2268615	Genotype	CC	160	273	1.00	
		CA	156	191	1.39 (1.04-1.86)	0.024
		AA	26	31	1.43 (0.82-2.49)	0.209
	Dominant	CC	160	273	1.00	0.018
		CA-AA	182	222	1.40 (1.06-1.84)	
	Recessive	CC-CA	316	464	1.00	0.453
		AA	26	31	1.23 (0.72-2.11)	
Log-additive	--	--	--	1.29 (1.03-1.61)	0.027	
TNFAIP2 rs710100	Genotype	GG	118	210	1.00	
		GA	171	211	1.44 (1.07-1.95)	0.018
		AA	53	69	1.37 (0.89-2.08)	0.150
	Dominant	GG	118	210	1.00	0.016
		GA-AA	224	280	1.42 (1.07-1.89)	
	Recessive	GG-GA	289	421	1.00	0.576
		AA	53	69	1.12 (0.76-1.65)	
Log-additive	--	--	--	1.23 (1.00-1.50)	0.046	

SNP, single nucleotide polymorphism; OR, odds ratio; 95% CI, 95% confidence interval.

p values were calculated by logistic regression analysis with adjustments for age and gender.

p < 0.05 means the data is statistically significant.

Table 3. Relationships between the candidate SNPs and cervical cancer risk according to the stratification by age

SNP ID	Model	Genotype	> 43 years				≤ 43 years			
			Case	Control	OR (95%CI)	<i>p</i>	Case	Control	OR (95%CI)	<i>p</i>
PGF rs2268615	Allele	C	253	393	1.00	0.303	223	344	1.00	0.041
		A	99	131	1.17 (0.87-1.59)		109	122	1.38 (1.01-1.88)	
	Genotype	CC	89	147	1.00		71	126	1.00	
		CA	75	99	1.25 (0.84-1.86)	0.282	81	92	1.55 (1.02-2.36)	0.039
		AA	12	16	1.22 (0.55-2.70)	0.626	14	15	1.62 (0.74-3.56)	0.228
	Dominant	CC	89	147	1.00	0.269	71	126	1.00	0.030
		CA-AA	87	115	1.24 (0.85-1.82)		95	107	1.56 (1.05-2.33)	
	Recessive	CC-CA	164	246	1.00	0.795	152	218	1.00	0.481
		AA	12	16	1.11 (0.51-2.41)		14	15	1.31 (0.61-2.81)	
	Log-additive	--	--	--	1.17 (0.86-1.60)	0.317	--	--	1.40 (1.01-1.92)	0.042
TNFAIP2 rs710100	Allele	G	217	331	1.00	0.597	190	300	1.00	0.018
		A	135	191	1.08 (0.82-1.43)		142	158	1.42 (1.06-1.90)	
	Genotype	GG	64	111	1.00		54	99	1.00	
		GA	89	109	1.41 (0.93-2.14)	0.105	82	102	1.46 (0.94-2.27)	0.092
		AA	23	41	0.96 (0.53-1.74)	0.890	30	28	1.97 (1.07-3.63)	0.031
	Dominant	GG	64	111	1.00	0.208	54	99	1.00	0.034
		GA-AA	112	150	1.29 (0.87-1.91)		112	130	1.57 (1.03-2.38)	
	Recessive	GG-GA	153	220	1.00	0.418	136	201	1.00	0.103
		AA	23	41	0.80 (0.46-1.38)		30	28	1.59 (0.91-2.79)	
	Log-	--	--	--	1.07	0.635	--	--	1.42	0.020

additive

(0.81-
1.41)(1.06-
1.90)

SNP, single nucleotide polymorphism; OR, odds ratio; 95% CI, 95% confidence interval.

p values were calculated by logistic regression analysis with adjustments for age.

$p < 0.05$ indicates statistical significance.

Table 4. Relationship of clinical stage with *PGF* rs8019391 polymorphism in cervical cancer patients adjusted by age

SNP ID	Model	Genotype	II	III-IV	OR (95%CI)	p
rs8019391	Allele	C	110	220	1.00	4.58$\times 10^{-4}$
		T	50	44	2.27 (1.43-3.62)	
	Codominant	CC	37	91	1.00	
		CT	36	38	2.34 (1.29-4.25)	0.005
		TT	7	3	5.76 (1.41-23.52)	0.015
	Dominant	CC	37	91	1.00	0.001
		CT-TT	43	41	2.59 (1.46-4.60)	
	Recessive	CC-CT	73	129	1.00	0.045
		TT	7	3	4.13 (1.04-16.45)	
	Log-additive	—	—	—	2.36 (1.45-3.86)	<0.001

SNP, single nucleotide polymorphism; OR, odds ratio; 95% CI, 95% confidence interval.

p values were calculated by logistic regression analysis with adjustments for age.

$p < 0.05$ indicates statistical significance.

Table 5. SNP–SNP interaction models of the *PGF* and *TNFAIP2* genes analyzed by the MDR method

Model	Training Bal. Acc.	Testing Bal. Acc.	CVC	OR (95% CI)	p
<i>TNFAIP2</i> rs710100	0.544	0.508	6/10	1.46 (1.08–1.97)	0.014
<i>PGF</i> rs2268615, <i>TNFAIP2</i> rs710100	0.564	0.536	9/10	1.91 (1.41–2.59)	<0.0001
<i>PGF</i> rs2268615, <i>TNFAIP2</i> rs710100, <i>PGF</i> rs8019391	0.587	0.550	10/10	2.11 (1.56–2.84)	<0.0001

MDR, multifactor dimensionality reduction; Bal. Acc., balanced accuracy; CVC, cross-validation consistency; OR, odds ratio; CI, confidence interval.

p values were calculated using c^2 tests.

$p < 0.05$ indicates statistical significance.

Figures

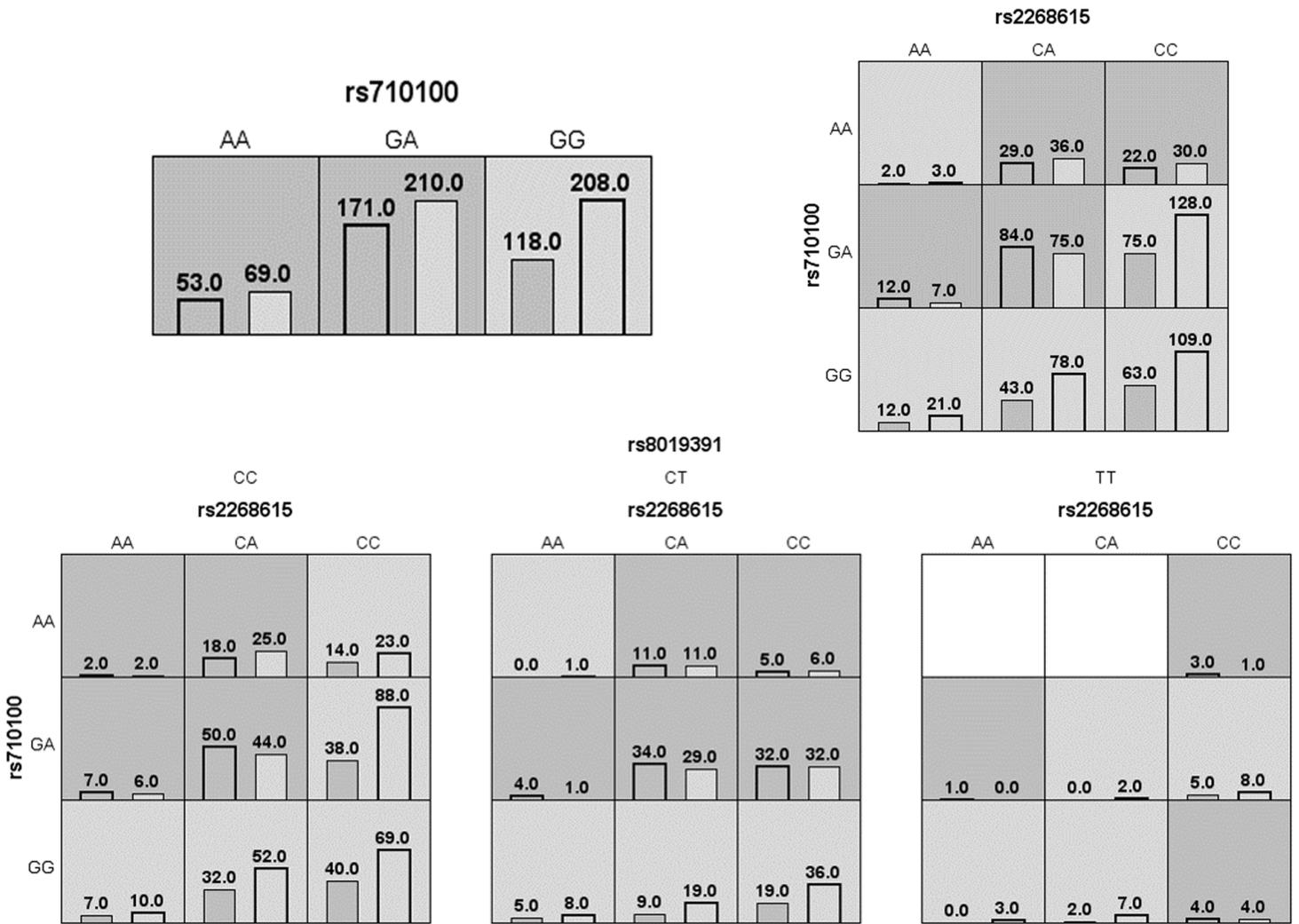


Figure 1

Summary of MDR gene-gene interaction. Each cell shows counts of "case" on left and "control" on right.

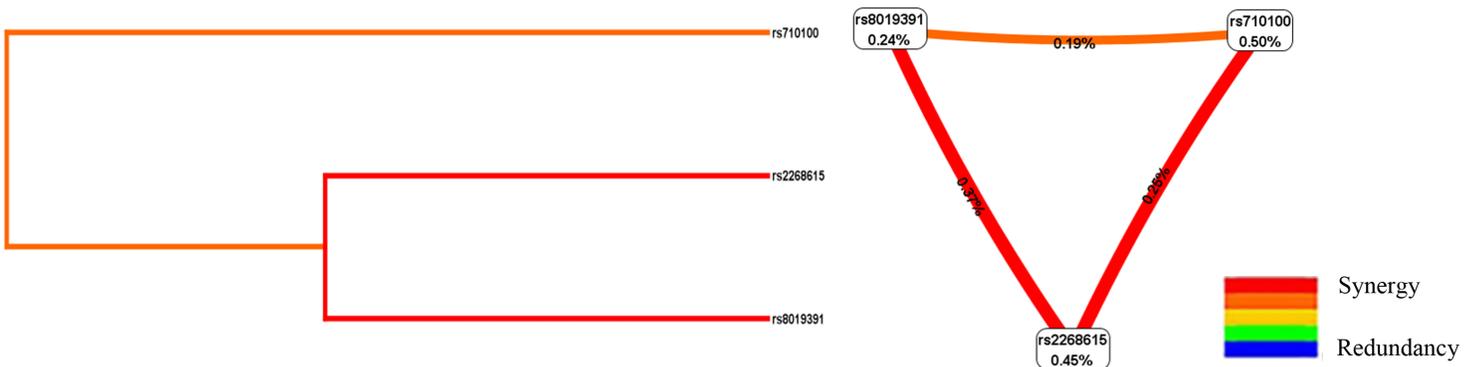


Figure 2

SNP-SNP interaction dendrogram and Fruchterman-Reingold.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [SupplementaryTableunmarked.docx](#)