

Influence of Slaughter Methods That Are Indigenous to Nguni People on Meat Physico-Chemical Characteristics of Goat Meat

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1 **Influence of slaughter methods that are indigenous to Nguni people on meat physico-**
2 **chemical characteristics of goat meat**

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25 **Abstract**

26 **Background:** Resource-limited households in smallholder farming systems slaughter goats
27 use indigenous methods for performing traditional ceremonies and meat consumption.
28 Although extensive research has been done to determine the effect of slaughter methods on
29 meat physico-chemical characteristics, there is paucity of information on methods which are
30 indigenous to Nguni people. Therefore, the objective of the study was to determine meat
31 quality of Nguni goats slaughtered using indigenous slaughter methods.

32 **Methods:** Thirty 15-18-month old wethers were randomly assigned to three slaughter
33 methods; transverse neck incision (TNI), suprasternal notch piercing (SNP) and under
34 shoulder blade chest floor point of elbow piercing (CFP) to the direction of the heart. Post-
35 mortem, the *m. longissimus thoracis et lumborum* (LTL) was sampled for meat quality
36 measurements.

37 **Results:** Wethers slaughtered using the SNP method had greater ultimate pH values when
38 compared with TNI and CFP slaughter methods. Wethers slaughtered using SNP method had
39 greater rate of pH decline when compared with TNI and CFP slaughter methods. Wethers
40 slaughtered using the SNP method had lower meat redness (a*), yellowness (b*), and chroma
41 (C*) values when compared with TNI and CFP slaughter methods. Slaughter method had no
42 effect ($P \geq 0.05$) on drip loss, water holding capacity, cooking loss and shear force.

43 **Conclusions:** Overall, Nguni wethers slaughtered using the TNI and CFP methods produced
44 chevon with fresh meat appearance.

45

46 **Keywords:** chroma, suprasternal notch piercing, transverse neck incision, redness, ultimate
47 pH, yellowness.

48

49

50

51 **1. Introduction**

52 In developing countries (e.g. Africa and the Middle East) where more than 90 % of the
53 world's goat population is found [1], goats are ranked as the second most important and
54 abundant livestock species following cattle. Such importance is due to their ability to graze
55 and browse poor quality forage, survive drought and saline conditions [2]. Furthermore, goats
56 have high prolificacy making them a short term investment [3]. Goats are owned and kept by
57 farmers for meat (chevon), milk, manure, skins and hides [4]. Although goats are kept for
58 socio-economic purposes, the primary reason for keeping them is to use them for religious
59 and cultural puporses [5].

60

61 In 2018, the world's total goat population was estimated at 1 billion with 40 % of these goats
62 found in Africa [1]. Southern Africa's total goat population contributes approximately 2 %,
63 where 50 % these goats are found in South Africa [6]. In South Africa, where the total goat
64 population is equally distributed between smallholder and commercial farming systems, more
65 than 95 % of goats are sold informally in private markets for slaughter during cultural beliefs
66 [7]. When performing cultural practices, goats are slaughtered informally using indigenous
67 slaughter methods [8; 9]. Indigenous slaughter methods include the transverse neck incision
68 (TNI), under shoulder blade chest floor point of elbow piercing (CFP) and suprasternal notch
69 piercing (SNP) to the direction of the heart using a short spear [8]. Goats slaughtered using
70 indigenous slaughter methods (TNI, CFP and SNP) had different bleeding time and
71 efficiency, blood volume in the thoracic cavity, time to loss of sensitivity and cardiac arrest
72 and comparable behavioral responses and dressing pecentages and similar behavioural
73 responses [10]. These slaughter methods are used when when performing traditional
74 ceremonies [4] such as connecting the living and the dead, clensing the deceased, and

75 celebration of marriages and births. Although these indigenous slaughter methods are
76 permitted under the provision of the South African Meat Safety Act 40 of 2000 [9], animal
77 welfare advocates consider these methods as inhumane as no stunning is involved. Animal
78 rights activists trivialise the religious and cultural functions of these slaughter techniques.
79 Immediately after slaughter and dressing, in most cases, the carcasses are stored under room
80 temperature conditions for 24 hours where they are hanged using hocks for them to cool
81 slowly, dry and allow maximum blood loss. This is important because consuming meat with
82 blood is prohibited as ancestors spirits do not accept meat with blood [11]. Offals are cleaned
83 and consumed on the day of slaughter as they are highly perishable.

84

85 Cultural beliefs invoked when celebrating circumcision, marriages and births, venerating
86 ancestors, avenging evil spirits, and performing a ritual during funerals are integral to most
87 African cultures [11; 12]. Meat consumed after performing traditional ceremonies leads to
88 increased protein intake resulting in to a decrease of protein-energy malnutrition and dietary
89 deficiency [13]. Although several studies has been done to assess the effect of slaughter
90 methods on chevon quality [14; 15; 16], however, the effect of effect of TNI, SNP and CFP
91 methods on chevon quality is poorly understood. Therefore, understanding the effect of TNI,
92 SNP and CFP methods on chevon quality is crucial for communities to improveg food
93 security within local cultural pactices.

94

95 The effects of indigenous slaughter methods on chevon quality has, however, not been
96 investigated since resource-limited households interpret carcass characteristics as physical-
97 chemical. Understanding the effect of slaughter methods on meat physico-chemical
98 characteristics could assist farmers to select a slaughter method that promote their culture and
99 enhance modern consumer acceptability without compromising animal welfare and meat

100 quality. The objective of the current study was, therefore, to assess the effect of TNI, SNP
101 and CFP methods on meat physico-chemical characteristics from Nguni goats. It was
102 hypothesized that meat quality from Nguni goats slaughtered using indigenous slaughter
103 methods are comparable to that of goats slaughtered using conventional methods.

104

105 **2. Material and methods**

106 2.1. Goats and experimental design

107 Thirty clinically healthy Nguni wethers (about 15-18 months old based on dentition) with
108 body weight averaging 16.8 ± 1.84 kg where bought from the local rural farmers of
109 Nongoma (27°53'S 31°38'E) were they were managed on communal rangelands dominated
110 by *Vachellia karroo* browse species. Goats were kept in the the same kraal and randomly
111 assigned to each slaughter treatment. Classification of goats as Nguni breed was based on
112 their multiple coat colour patterns, small and compact frame size [17]. Goats were
113 slaughtered randomly after 24 hours of fasting where clean water was provided *ad libitum*.
114 Slaughtering of goats was completed within a day where the process began at 05h00 in
115 morning and ended at 09h00.

116

117 2.2. Treatments

118 Thirty wethers were randomly assigned to three slaughter treatments (n=10/treatment) and
119 subjected to; transverse neck incision (TNI), under shoulder blade chest floor point of elbow
120 piercing (CFP; Figure 2B) and suprasternal notch piercing with a short spear (SNP; Figure
121 2C). Briefly, goats were slaughtered without stunning using either a sharp knife or a short
122 spear specifically designed for slaughtering of goats. Transverse neck incision slaughter
123 technique involved the use of a sharp knife for cutting the skin, muscles (brachiocephalic,
124 sternocephalic, sternohyoid, and sternothyroid), trachea, oesophagus, carotid arteries, jugular

125 veins and the major, superficial and deep nerves of the cervical region [18]. Cuts were
126 defined as a change in the direction of movement of the knife (e.g. the forward movement of
127 the knife would count as one cut, while the corresponding reverse movement would be
128 recorded as a second cut).

129

130 Suprasternal notch piercing (SNP) targeting the heart was performed by two experienced
131 slaughtermen using short spears. During slaughter, each goat was allowed to stand upright
132 using rear/hind legs. One slaughter man held the left front leg and the head (using horns)
133 while the second slaughter man held the right front leg and the spear which was used for
134 piercing the goat in the suprasternal notch in the direction of the heart. Goats were allowed to
135 bleed into a 5 litre (L) water bucket.

136

137 Under shoulder blade chest floor point of elbow (CFP), piercing involved the use of a short
138 spear and five slaughtermen. These slaughtermen held each goat in dorsal recumbent position
139 by holding all legs sideways. The fifth slaughter man was responsible for piercing each goat
140 on the heart girth position, next to the chest floor and point elbow to the direction of the heart.
141 Goats were also allowed to bleed into a 5 litre (L) water bucket.

142

143 2.3. Meat sampling and storage

144 Following exsanguination, carcasses were dressed as described by [10]. Forty-five minutes
145 after slaughter, *M. longissimus thoracis* (LTL) muscles were removed from the left and right
146 sides of each carcass for meat quality analyses. The LTL muscles were then vacuum-packed
147 and stored in polystyrene cooler boxes and transported to the animal science laboratory at
148 University of KwaZulu Natal, Pietermaritzburg, South Africa for meat quality analyses. The
149 slaughter point and animal science laboratory are 380 km apart. Eleven hours later on arrival

150 at the lab, meat samples were unpacked from the cooler box and stored at room temperature
151 (23 °C) until 24 hours after slaughter.

152

153 2.4. Measurements

154 2.4.1. Meat pH

155 Post-mortem pH were measured 45 minutes after slaughter and thereafter for a period of 24
156 hours (11, 13, 15, 17, and 24). Post mortem rate of decline of pH in meat were measured
157 using a portable pH meter probe (CRISON pH25, CRISON instrument SA, Spain).

158

159 2.4.2. Meat colour

160 Meat colour was measured 24 hours after slaughter using a colour meter (HunterLab,
161 ColorFlex EZ Spectrophotometer). The parameters used to evaluate meat colour followed
162 colour CIE (1976) coordinates which measured: lightness (L^*), redness (a^*) and yellowness
163 (b^*) from three locations on the cut surface of individual meat samples. Three replicate
164 measurements were done. Areas of connective tissue and intramuscular fat per sample were
165 avoided. Colour saturation was calculated as the square root of the sum of a^{*2} and b^{*2} .

166

167 2.4.3. Drip loss and water holding capacity

168 Drip loss was determined by the standard bag method [19]. Drip loss was measured as the
169 weight loss during the suspension of a standardized muscle sample (40–50 g and
170 approximately $30 \times 60 \times 25$ mm) in an airtight transparent plastic bag over 48 h at 4°C. Drip
171 loss was expressed as a percentage of the weight loss in 48 hours over the initial weight
172 sample.

173

174 Water holding capacity (WHC) was determined by compressing approximately 3 – 4 gram of
175 meat with 30 kg of weight for 5 minutes using a texture analyser (Stable Micro System,
176 Model TA.XT 2i/25, UK). The water content of meat was determined by multiplying the
177 initial weight of meat with 0.7. Water loss was determined by subtracting final weight from
178 the initial weight. Water holding capacity was therefore calculated by subtracting water loss
179 from water content, dividing by water content and multiplying by 100.

180

181 2.4.4. Cooking loss and shear force

182 Fresh LTL meat samples were cut and weighed (initial weight) to form individual
183 standardized slices of approximately 50 mm thick. Prepared meat samples were then placed
184 in a RATIONAL Granite-enamelled container 20 mm deep. A RATIONAL SCC 61E self-
185 cooking centre (Landsberg, Munich, Germany) was used to roast *M. longissimus thoracic*
186 (LTL). Briefly, the hot plate was preheated for 5 min to 205 °C. Immediately after preheating,
187 RATIONAL Granite-enamelled container 20 mm deep was placed onto a RATIONAL Grid,
188 stainless steel 1/IGN were meat was roasted for 4 min. After completing the cooking process,
189 meat samples were cooled at room temperature and weighed. Therefore, the cooking loss was
190 calculated as the percentage difference in weight before and after cooking.

191

192 Following cooking, sub-samples of specified core diameter parallel to the grain of the meat
193 was used. Samples were sheared perpendicular to the fibre direction using a texture analyser
194 model TA.XTplus, texture analyser (Stable Micro System, Model TA.XT 2i/25, UK) as
195 outlined by [20]. The mean maximum load recorded for the three cores were represented as
196 the average of peak force in Newton's (N) for each sample.

197

198 2.5. Statistical analyses

199 All data were analysed using [21]. A general linear model (GLM) procedure with repeated
200 measures analysis was used to test the effect of indigenous slaughter method on the rate of
201 pH decline of meat in hours.

202

203 The model used was the following:

$$Y_{ijk} = \mu + S_i + T_j + (S \times T)_{ij} + b_l W_k + \varepsilon_{ijkl}$$

204 Where:

205 Y_{ijk} = Response variables (pH, a^* , b^* , L^* , water holding capacity, shear force, and cooking
206 loss);

207 μ = population mean common to all observation;

208 S_i = effect of indigenous slaughter methods;

209 $(S \times T)_{ij}$ = the interaction between slaughter method and time (hours);

210 $b_l W_k$ = co-variant (initial temperature or pH); and

211 ε_i = residual error.

212

213 A general linear model was used to test the effect of the slaughter method on meat physico-
214 characteristics of Nguni goats. Comparisons of least-square means were done using the
215 PDIFF option of [21]. The significance threshold was set at $P \leq 0.05$.

216

217 PROC REG was also used to determine relationships between slaughter methods and meat
218 pH over time. The slope of each curve was tested if it was significantly different from each
219 other using the TEST statement of regression procedure for each slaughter method.

220

221 **3. Results**

222 Slaughter method had no effect ($P \geq 0.05$) on initial meat pH (Table 1) and pH changes
223 (Figure 1), however its interaction with time was significant ($P \leq 0.05$). Initial meat pH was
224 highest for TNI followed by SNP and CFP slaughter methods. Ultimate meat pH was highest
225 for SNP followed by TNI and CFP methods. Ultimate meat pH was greater ($P \leq 0.05$) for the
226 SNP slaughter method when compared with TNI and CFP slaughter methods (Table 1). Rate
227 of pH decline in meat was highest for SNP followed by TNI and CFP slaughter methods.
228 Rate of pH decline in meat was higher ($P \leq 0.05$) for SNP when compared with TNI and CFP
229 slaughter methods (Table 2).

230

231 Slaughter method had an effect ($P \leq 0.05$) on meat redness and yellowness (Table 1). Meat
232 redness was highest for TNI followed by CFP and SNP slaughter methods. Redness (a^*)
233 values of meat from goats slaughtered using SNP method were lower ($P \leq 0.05$) compared
234 values of meat from goats slaughtered using TNI and CFP methods (Table 1). Meat
235 yellowness (b^*) values of goats slaughtered using SNP method were lower ($P \leq 0.05$)
236 compared values of meat from goats slaughtered using TNI and CFP methods (Table 1).
237 Slaughter method had no effect ($P \leq 0.05$) on the lightness (L^*) and hue (H^*) coordinates of
238 meat (Table 1). Slaughter method had an effect ($P \leq 0.05$) on chroma coordinates. Chroma
239 coordinates were highest for TNI followed by CFP and SNP slaughter methods. Chroma
240 coordinates for meat from goats slaughtered using SNP was lower ($P \leq 0.05$) than those of
241 meat from TNI and CFP slaughter methods (Table 1). Slaughter method had no effect ($P >$
242 0.05) on drip loss, WHC, cooking loss and shear force.

243

244 **4. Discussion**

245 The high ultimate pH value reported for SNP slaughter treatment could be explained by
246 prolonged stress before and during slaughter, which may have lead to reduction in glycogen

247 levels, and therefore low post-mortem lactic acid production [22; 23; 15]. The high pHu
248 observed in meat from animals slaughtered using the SNP method suggest these animals were
249 more stressed than those slaughtered using other methods. The the causes of stress are,
250 however, not immediately clear. The prolonged stress could be related to the nature,
251 frequency, strength, severity, intensity, and/or duration of stressor(s) before and during
252 slaughter. Such stressors include animal handling before and during slaughter and pain
253 experienced by the animal during sticking and exsanguination. These factors were not
254 measured in the current study and merit investigation.

255

256 The lower meat redness (a^*) observed for whethers slaughtered using the SNP slaughter
257 method may be related to the high pH observed for this slaughter method. The pH effect on
258 a^* has been associated with oxygen consumption [24; 25]. High pH often results in low
259 protein denaturation, which creates a more closed tissue structure. The closed structure of
260 meat reduce the diffusion of oxygen into the meat from the surface and any oxygen that do
261 not reach the interior is then utilised by high cytochrome activity encouraged by high pH
262 [26]. As a result less oxygenated myoglobin is formed, and consequently meat looks less red.

263

264 The observation that the SNP method had lower meat yellowness (b^*) values than other
265 methods could also be explained by the effect of high muscle pH on oxygenation of the
266 myoglobin [27; 28; 29]. The lower chroma (C^* , color intensity) values observed for the SNP
267 slaughter method compared to other methods also corresponds high pH values, which
268 negatively correlates with low oxymyoglobin content in the meat [30]. It is well known that a
269 decrease in oxymyoglobin content in meat is accompanied by lower values of a^* , b^* and C^*
270 [31; 32].

271

272 The finding that slaughter method had no effect on drip loss, WHC, cooking loss and shear
273 force agree with earlier reports [33; 34; 16]. Overall, the high pHu (6.4-6.8), WHC, and low
274 drip loss, a*, b*, and chroma values reported across slaughter methods are a characteristic of
275 dark firm and dry (DFD) meat [26]. This implies that all the goats across treatments had
276 prolonged stress before and/or during slaughter and that could be related to the high
277 temperamental behaviour of the Nguni goat breed among other stress factors [35]. Further
278 studies to determine the pre-, peri- and post-mortem glycogen reserves, lactic acid
279 concentration in muscles and stress hormones in Nguni goats could be important in
280 explaining the effect of their temperamental behaviour on meat quality. Causes of pre- and
281 peri-mortem stress which could be important in minimising in DFD meat from Nguni goats
282 also merit investigation.

283

284 **5. Conclusions**

285 Nguni goats slaughter using SNP slaughter method had higher pHu which resulted in lower
286 meat redness, yellowness and chroma values .. It was concluded that Nguni goats slaughtered
287 using TNI and CFP methods produce chevon with better meat colour than those slaughterd
288 using the SNP method. The high meat pHu, WHC, and low drip loss and colour (a*, b*, and
289 chroma) are indicative of DFD meat. Further studies to determine causes of DFD meat in
290 Nguni goats slaughtered using indigenous methods could be important.

291

292 **List of abbreviations**

293 TNI - transverse neck incision

294 SNP - suprasternal notch piercing

295 CFP - under shoulder blade chest floor point of elbow piercing

296 LTL - *m. longissimus thoracis et lumborum*

297 a* - Redness
298 b* - Yellowness
299 C* - chroma values
300 FAOSTAT - Food and Agriculture Organization Corporate Statistical
301 L – litre
302 WHC - Water holding capacity
303 GLM – general linear model
304 SAS – statistical analysis software
305 PDIFF - Requests that p-values for differences
306 DFD - dark firm and dry

307

308 **Declarations**

309 Ethics approval
310 Ethical clearance for this study was granted by the Animal Ethics Committee of the
311 University of KwaZulu-Natal (AREC/001/018D).

312

313 Consent for publication

314 Not applicable.

315

316 Availability of data and material

317 The datasets used and/or analysed during the current study are available from the
318 corresponding author on reasonable request.

319

320 Competing interests

321 The authors declare that they have no competing interests

322

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325

326 Authours contributions

327 ZMM designed, collected and analysed data and wrote the manuscript. MC supervised the
328 study and the writing of the manuscript. CM read and approved the final version of the
329 manuscript.

330

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333 goats.

334

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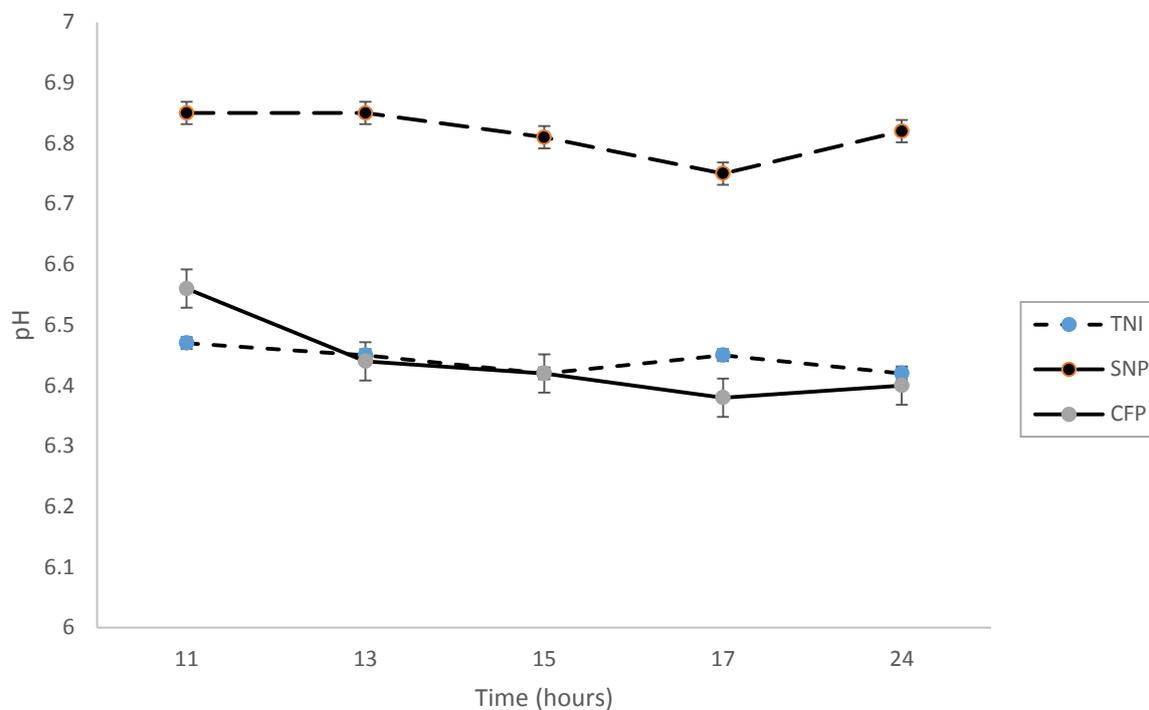
427



428

429 Figure 2: Visual pictures of a spear (A), chest-floor point-of-elbow (B), and suprasternal
430 notch piercing (C).

431 Adapted from [10].



432

433 Figure 1: The relationship between meat pH and time for TNI, SNP and CFP slaughter
434 methods.

435 Table 1: Effects of TNI, SNP and CFP slaughter methods on colour parameters (a*, b*, L*, h and C), drip loss, cooking loss, shear force and
 436 water holding capacity of goat meat

Variable	Slaughter method			Significance
	TNI	SNP	CFP	
pH_{45 min}	7.8 ± 0.102	7.7 ± 0.102	7.6 ± 0.102	NS
pH_{24h}	6.42 ± 0.13 ^a	6.82±0.13 ^b	6.40 ± 0.13 ^a	*
Drip loss (%)	3.0 ± 0.75	1.75 ± 0.87	1.88 ± 0.75	NS
Water holding capacity (%)	74.18 ± 1.28	75.64 ± 2.21	72.88 ± 1.57	NS
Colour parameters				
a*	16.4 ± 0.56 ^a	14.7 ± 0.56 ^b	16.3 ± 0.56 ^a	*
b*	13.5 ± 0.61 ^a	11.5 ± 0.61 ^b	13.2 ± 0.61 ^a	*
L*	29.5 ± 1.02	28.3 ± 1.02	30.6 ± 1.02	NS
H*	0.69 ± 0.01	0.66 ± 0.01	0.68 ± 0.01	NS
C*	21.24 ± 0.78 ^a	18.68 ± 0.78 ^b	21.02 ± 0.78 ^a	*
Cooking loss (%)	22.73 ± 2.3	21.6 ± 3.0	19.4 ± 2.6	NS
Shear force (N)	9.52 ± 0.72	10.73 ± 0.83	10.55 ± 0.72	NS

437 ^{a, b, c}Means in the same row with different superscripts are significantly different at P≤0.05; *p ≤ 0.05; **p ≤ 0.01; NS-p > 0.05. NS: not
 438 significant; L*: lightness; a*: redness; b*: yellowness

439 Table 2: The relationship between meat pH and temperature for TNI, SNP and CFP slaughter
 440 methods

Independent variable	Parameter estimates (time)				
	Intercept	<i>P</i> intercept	Slope	<i>P</i> slope	RMSE
pH					
TNI	6.31 ± 0.65	≤.0001	0.02 ± 0.03 ^a	0.591	0.54
SNP	6.7 ± 0.13	≤.0001	0.04 ± 0.033 0.017 ^b		0.44
CFP	9.44 ± 1.39	≤.0001	-0.20 ± 0.10 ^a	0.055	0.49

441 ^{a, b, c}Means in the same column with different superscripts are significantly different at $p \leq$
 442 0.05; * $p \leq 0.05$; ** $p \leq 0.01$; NS- $p > 0.05$.

443

Figures

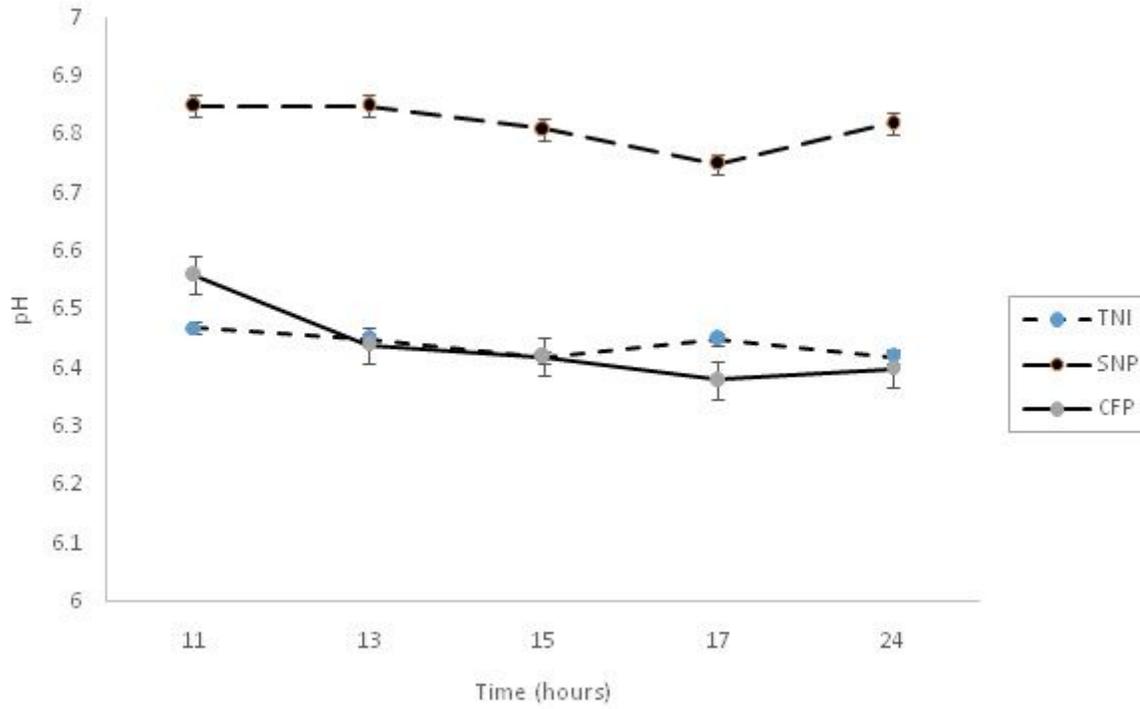


Figure 1

The relationship between meat pH and time for TNI, SNP and CFP slaughter methods.



Figure 2

Visual pictures of a spear (A), chest floor point of elbow (B), and suprasternal notch piercing (C).

Supplementary Files

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