Association of plasma and CSF cytochrome P450, soluble epoxide hydrolase and ethanolamides metabolism with Alzheimer's disease

Kamil Borkowski (✉ kborkowski@ucdavis.edu)

University of California Davis <https://orcid.org/0000-0003-3637-3450>

Theresa L. Pedersen

University of California Davis

Nicholas T. Seyfried

Emory University School of Medicine

James J. Lah

Emory University School of Medicine

Allan I. Levey

Emory University School of Medicine

Chadwick M. Hales

Emory University School of Medicine

Eric B. Dammer

Emory University School of Medicine

Colette Blach

Duke University

Gregory Louie

Duke University

Rima Kaddurah-Daouk

Duke University

John W. Newman

United States Department of Agriculture Agriculture research service

Research

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Abstract

Background

Alzheimer's disease, cardiovascular disease and other cardiometabolic disorders may share inflammatory origins. Lipid mediators, including oxylipins, endocannabinoids, bile acids and steroids regulate inflammation, energy metabolism and cell proliferation with well-established involvement in cardiometabolic diseases. However, their roles in Alzheimer's disease and their potential as biomarkers are poorly understood. Here we describe the analysis of plasma and cerebrospinal fluid lipid mediators in a case-control comparison of individuals with Alzheimer's disease and healthy controls to investigate these knowledge gaps.

Methods

Lipid mediators were measured using targeted quantitative mass spectrometry. Data were analyzed using analysis of covariates, adjusting for sex, age, and ethnicity. Partial least square discriminant analysis identified plasma and cerebrospinal fluid lipid mediator discriminates of Alzheimer's disease. Alzheimer's disease predictive models were constructed using machine learning combined with stepwise logistic regression.

Results

In both plasma and cerebrospinal fluid, individuals with Alzheimer's disease had elevated cytochrome P450/soluble epoxide hydrolase pathway components and decreased fatty acid ethanolamides compared to healthy controls. Circulating metabolites of soluble epoxide hydrolase and ethanolamides provide Alzheimer's disease predictors with areas under receiver operator characteristic curves ranging from 0.82 to 0.92 for cerebrospinal fluid and plasma metabolites, respectively.

Conclusions

Previous studies report Alzheimer's disease-associated soluble epoxide hydrolase upregulation in the brain and that endocannabinoid metabolism provides an adaptive response to neuroinflammation. This study supports the involvement of P450-dependent and endocannabinoid metabolism in Alzheimer's disease, yielding potential biomarkers of the disorder. The results further suggest that combined pharmacological intervention targeting both metabolic pathways may have therapeutic benefits for Alzheimer's disease.

Introduction

Risk factors for Alzheimer's disease (AD) include cardiometabolic disorders, low-grade systemic inflammation and altered lipid and energy metabolism (1–3). Lipid mediators are lipid derived signaling molecules that regulate both acute and low-grade systemic inflammation and energy metabolism along with other processes (4). Circulating lipid mediators have been implicated in the pathogenesis of cardiometabolic diseases (5–7), however, their involvement in AD is still not well described.

Four important families of such lipid mediators readily detected in the circulation are the oxygenated polyunsaturated fatty acids (i.e. oxylipins), the endogenous cannabinoid receptor activators and their structural equivalents (i.e. endocannabinoids, including monoacylglycerols and ethanolamides), bile acids and steroids. Oxylipins are mainly oxygenated products of polyunsaturated fatty acids (PUFA) generated *via* 4 main pathways: cyclooxygenases (COX), generating mainly prostaglandins; lipoxygenases (LOX), generating mainly hydroxy-fatty acids, including leukotrienes and lipoxins; cytochrome P450 (CYP) and soluble epoxide hydrolase (sEH), generating epoxy-fatty acids and dihydroxy fatty acids respectively (4), and reactive oxygen-mediated formation of prostanoids and hydroxy-fatty acids (8). Products from these pathways exhibit both pro- and anti-inflammatory actions (4). Endocannabinoids are mainly fatty acids esters and amides, which are ligands for the cannabinoid receptors CB1 and CB2, the transient potential vanilloid receptor TRPV1 and G- protein-coupled receptor GPR55 (9), all highly expressed in the central nervous system (10). Endocannabinoids regulate energy metabolism (9) and are generally considered anti-inflammatory (11). Bile acids (BA) are generated by the liver as primary bile acids and secreted into the gut to aid in lipid digestion, and they are further metabolized by the gut microbiome form secondary bile acids (12). After reabsorption from the gut into the blood stream, BAs regulate energy metabolism with different potency between primary and secondary species (13).

Changes in circulating lipid mediators in relation to AD were previously reported. Particularly, several oxylipins of the acute inflammation pathway were reportedly elevated in AD (14, 15) and pro-resolving (quenching activated inflammatory signal) lipid mediators have been suggested as potential treatment for AD (16). Specific changes in bile acids metabolism, including a decrease in primary and an increase in secondary metabolites were also observed in AD subjects (17) and differences in bile acids clearance for cholesterol pathway were reported in AD (18). Notably, bile acids and some steroids manifest neuroprotective functions through activation of steroid receptors (19). However, a comprehensive analysis of the AD related changes in circulating lipid mediators is lacking and reports of lipid mediators in cerebrospinal fluid (CSF) are minimal. In the current work we utilize plasma and CSF samples of AD patients and cognitively normal controls from the Emory Goizueta Alzheimer's Disease Research Center (ADRC). Using a case control approach, we provide comprehensive analysis of AD-associated changes in both circulating and CSF lipid mediators, including major classes of oxylipins, endocannabinoids, bile acids and some steroids, covering multiple aspects of inflammatory cascades and regulators of energy metabolism. Moreover, to minimize the bias caused by postprandial fluctuation in plasma lipids, the fasting state of opportunistically collected samples was estimated using a novel predictive model that uses levels of circulating lipid mediators (20).

Materials And Methods

Subjects

All participants from whom plasma and CSF samples were collected provided informed consent under protocols approved by the Institutional Review Board at Emory University.

Cohorts included the Emory Healthy Brain Study (IRB00080300), Cognitive Neurology Research (IRB00078273), and Memory @ Emory (IRB00079069).

All protocols were reviewed and approved by the Emory University Institutional Review Board.. All patients received standardized cognitive assessments (including Montreal Cognitive Assessment (MoCA)) in the Emory Cognitive Neurology clinic, the ADRC and affiliated Emory Healthy Brain Study (EHBS) (21).

All diagnostic data were supplied by the ADRC and the Emory Cognitive Neurology Program. CSF was collected by lumbar puncture and banked according to 2014 ADC/NIA best practices guidelines.

All CSF samples collected from research participants in the ADRC, Emory Healthy Brain Study, and Cognitive Neurology clinic were assayed using the INNO-BIA AlzBio3 Luminex assay at AKESOgen (Peachtree Corners, GA).

AD cases and healthy individuals were defined using established biomarker cutoff criteria for AD for each assay platform (22, 23). In total, plasma samples were available for 148 AD patients and 133 healthy controls and CSF samples were available for 150 AD patients and 139 healthy controls. Plasma and CSF sample collection overlap (both plasma and CSF collected at the same day) was 145 for AD group and 133 for the control group. Cohort information is provided in **Table S1**.

Quantification of lipid mediators

Plasma concentrations of non-esterified PUFA, oxylipins, endocannabinoids, a group of non-steroidal anti-inflammatory drugs (NSAIDs) including ibuprofen, naproxen, acetaminophen, a suite of conjugated and unconjugated bile acids, and a series of glucocorticoids, progestins and testosterone were quantified in 50µL of plasma by liquid chromatography tandem mass spectrometry (LC-MS/MS) after protein precipitation in the presence of deuterated metabolite analogs (i.e. analytical surrogates) (24). CSF analyses were performed with 100µL samples prepared as previously reported for the analyses of sweat (25) and analyzed as reported for plasma. All samples were processed with rigorous quality control measures including case/control randomization, and the analysis of batch blanks, pooled matrix replicates and NIST Standard Reference Material 1950 – Metabolites in Human Plasma (Sigma-Aldrich, St Louis, MO). Extraction batches were re-randomized for acquisition, with method blanks and reference materials and calibration solutions scattered regularly throughout the set. Instrument limits of detection (LODs) and limits of quantification (LOQs) were estimated according to the Environmental Protection Agency method (40 CFR, Appendix B to Part 136 revision 1.11, U.S. and EPA 821-R-16-006 Revision 2). These values were then transformed into sample nM concentrations by multiplying the calculated concentration by the final sample volume and dividing by the volume of sample extracted. A complete analyte list with plasma LODs and LOQs have been reported (24). The majority of analytes were quantified against analytical standards with the exception of eicosapentaenoyl ethanolamide (EPEA),

palmitoleoyl ethanolamide (POEA), and the measured PUFAs [i.e. linoleic acid (LA); alpha-linolenic acid (aLA); arachidonic acid (AA); eicosapentaenoic acid (EPA); docosahexaenoic acid (DHA)].

For these compounds, area counts were recorded, adjusted for deuterated-surrogate responses and the relative response factors were expressed as the relative abundance across all analyzed samples. Reported monoacylglycerols (MAGs) are the sum of 1- and 2-acyl isomers, due to isomerization during sample processing.

Fasting state assessment and sample selection

Many of the CSF and plasma samples from AD patients were collected following additional research consent in the course of patients' clinical evaluations. Lumbar puncture procedures were nearly all scheduled in the morning, but fasting was not mandated in these individuals. Therefore, fasting state of the samples was estimated using previously published predictive equation, based on the raw plasma levels of 12(13)-EpOME, GCDCA and NO-Gly (20). Only subjects with the probability of the fasting state > 60% were used for the plasma analysis. All subjects were used to compare lipid mediators level in CSF. CSF was reported not to manifest postprandial lipid fluctuations (26), additionally, comparing predicted fasted to predicted non-fasted AD subjects reveal minimal differences in only 2 metabolites (**Table S2**).

Statistical analysis

All statistical tests were performed using JMP Pro 14 (JMP, SAS institute, Carry, NC). Prior to analysis, data were tested for outliers using the robust Huber M test and missing data were imputed using multivariate normal imputation for variables which were at least 75% complete. The imputed numbers constitute less than 3% of the data for both plasma and CSF. Imputation facilitated multivariate data analysis and non-imputed data were used for univariate approaches. Additionally, variables were normalized, centered, and scaled using Johnson's transformation, with normality verification using the Shapiro-Wilk test. The difference between the control and the AD group were assessed using t-test with gender, age, and race as covariates. Additionally, two-way ANOVA was used to test for the gender x group and race x group interactions. In case of significant interaction, the group effect was tested separately for the interacting factor. Correlations between MoCA score and lipid mediators were assessed using Spearman's rank order correlation, to account for non-linear associations. This analysis was performed using only AD subjects, stratified by the assessed fasting state for plasma. CSF samples were analyzed without fasting state stratification. Multiple comparison control was accomplished with the false discovery rate (FDR) correction method of Benjamini and Hochberg with a $q = 0.2$ (27).

Predictive models for AD were prepared using a combination of bootstrap tree and stepwise logistic regression modeling. Prior to analysis, subjects were randomly split into training (70%) and validation (30%) cohorts. Variables most frequently appearing in the models were identified by bootstrap tree: Number of layers = 50; split per tree = 3, bootstrap sampling rate for variables and subjects = 1. A variable contribution scree plot was generated using variable rank and the likelihood ratio of chi-square. The scree plot was used to determine a likelihood ratio of chi-square cutoff value for variables contributing to the

model. Selected variables were then subjected to stepwise logistic regressions. A stepwise analysis was performed with the maximal validation r^2 as the model stopping criteria, or if an additional step increased the Bayesian information criteria (BIC). Variables selected by the stepwise approach were then used to build the model using logistic regression. Metabolites that the model contribution p-value < 0.05 were excluded, to ensure the strongest model with the minimal number of predictors.

Partial least square discriminant analysis (PLA-DA) was used to integrate AD related differences in metabolite levels between plasma and CSF. PLS-DA model was built using the nonlinear iterative partial least squares algorithm with K-Fold variation method ($k = 7$) and included 235 variables from plasma and CSF, including metabolite levels and informative metabolite ratios. For the clarity purpose, only variables with a variable importance in projection (VIP) score > 1.4 we displayed on the loading plot.

Correlation between CSF and plasma metabolites was assessed using Spearman's rank order correlation.

Results

Fasting state assessment

Analysis of opportunistically collected samples brings a challenge of the unknown fasting state. The control cohort contained samples collected in the fasted state per ADRC and EHBS protocols, but the AD cohort included many who had no collected fasting state information and consist of samples collected in both fasted and non-fasted states. Therefore, to allow a direct comparison of the control and AD groups, we assessed the estimated subject fasting state using our previously published predictive model (20). As expected, control group was predicted to contain mostly fasted subjects (**Figure S1**). Out of 133 control subjects, 105 (i.e. 79%) were predicted as fasted, while 17 (i.e. 12%) as non-fasted, with a probability of > 60% and 11 (i.e. 8%) had a fasting state probability of < 60%. Out of 148 AD subjects, 60 (i.e. 40%) were predicted as fasted and 81 (i.e. 55%) as non-fasted, with a probability of > 60% and 7 (5%) had a fasting state probability of < 60%. Fifty percent of detected metabolites manifested difference between predicted fasted and non-fasted AD subjects in plasma and only minimal differences were observed in CSF (**Table S2**).

Cytochrome P450/soluble epoxide hydrolase metabolism is elevated in AD subjects

We compared plasma and CSF lipid mediator concentrations between the control and the AD groups, using only estimated fasted subjects with probability > 60% for plasma. In plasma, we detected 42 oxylipins (85 measured), 5 PUFAs (5 measured), 17 endocannabinoids (22 measured), 3 NSAIDs (4 measured), 19 bile acids (23 measured) and 8 steroid hormones (8 measured). The mean values and p-values for t-tests and two-way ANOVA interactions for all detected metabolites are provided in **Table S3**. Plasma group-fold differences in the oxylipin, endocannabinoids and PUFAs, projected onto their metabolic pathway, are presented in Fig. 1. The largest differences were observed in the long chain omega-3 PUFA metabolism. Both EPA and DHA enzyme derived mono-alcohols (5-LOX-derived 5-HEPE and 4-HDoHE and 12-LOX-derived 12-HEPE and 14-HDoHE) were lower (1.5-fold in average) in the AD

group, when compared to the control. On the other hand, the sEH EPA metabolite 17,18-DiHETE, was 3-fold higher in the AD group. In the AD group, the AA pathway manifested lower levels of the COX derived prostaglandins PGF2 α and PGD2 (1.6-fold average). Additionally, the AD group showed lower levels of acylethanolamides (1.5-fold in average) derived from dihomo-gamma-linolenic acid (DGLA), AA (AEA), docosatetraenoic acid (DEA), DHA (DHEA) and oleic acid (OEA). Notable are also lower levels of autooxidation markers, particularly the EPA-derived 9-HEPE (2-fold), linoleic acid (LA) -derived TriHOMEs (1.65-fold) and AA-derived isoprostanes (1.3-fold) in AD group.

Fewer lipid mediators were detected in CSF than in plasma. Detected CSF lipid mediators included 17 oxylipins, 5 PUFAs, 3 endocannabinoids, 14 bile acids and 6 steroids. The mean values and p-values for ttests and two-way ANOVA interactions are provided in **Table S4**. CSF significant group-fold differences in the level of oxylipin, endocannabinoids and PUFAs, projected onto their metabolic pathway, are presented in Fig. 2. In this matrix, the largest differences were observed in the LA CYP metabolic pathway, where both epoxy and dihydroxy FA, products of CYP and subsequent sEH metabolism, were higher in the AD group when compared to the control: epoxide average 1.5-fold; diol average 1.3-fold. All PUFAs from both omega-3 and omega-6 pathway were lower in the AD group, although the difference was only 1.2-fold on average. Additionally, AD group manifested 1.5-folds lower level of OEA and 1.3-fold lower level of the EPA-derived 14,15-DiHETE.

While few differences were observed in plasma and CSF bile acid levels between control and AD subjects, numerous differences were present in the specific bile acid ratios (Table 1). **Figure S2** shows bile acids metabolic pathway together with their median plasma levels to help understand the biological aspects of specific bile acid ratios. In plasma, the AD group was characterized by lower levels of cholic acid (CA), a product of the neutral bile acids synthesis pathway, while chenodeoxycholic acid (CDCA), a product of the acidic pathway was unchanged. This difference becomes even more pronounced when looking at the CA/CDCA ratio. On the other hand, the difference between neutral and acidic pathway was not present in downstream metabolites, when comparing the secondary unconjugated bile acids ratio, like deoxycholic acid/(lithocholic acid + ursodeoxycholic acid) (DCA/(LCA + UDCA)) or the most abundant primary conjugated derivatives glycocholic acid/glycochenodeoxycholic acid (GCA/GCDCA); **Table S3**. Of note, small differences between the neutral and acidic pathway were observed in the low abundance taurine conjugates of the secondary bile acids taurodeoxycholic/tauro lithocholic acid (TDCA/TLCA). Difference in conjugation ratio (more conjugates than the substrate) was observed in the neutral synthesis pathway (GDCA/DCA and GCA/CA) but not in the acidic synthesis pathway. Differences between the neutral and acidic synthesis pathway were also observed in the conversion of the primary to secondary bile acids. The ratio of downstream products to their precursor in the neutral pathway was higher in the AD group in the case of DCA/CA, TDCA/CA and GDCA/CA, but not in parallel acidic pathway metabolites (i.e. LCA/CDCA, UDCA/CDCA).

Table 1
Differences in bile acids metabolites and their specific ratios between control and AD group. Means are expressed in nM or as a ratio of the concentrations. Metabolites and their ratio are stratified by the metabolic affiliations. All tested bile acids and their ratios are presented in the **Table S3**.

Metabolite	P value	Mean [95%CI]	
		Control	AD
Neutral vs acidic synthesis pathway			
CA	0.0321	27.4 [20.8–36.2]	19.3 [13.6–27.4]
CDCA	0.978	38.7 [29.2–51.5]	46.7 [32.6–66.9]
CA/CDCA	0.0008	0.707 [0.571–0.876]	0.363 [0.271–0.487]
GCDCA/GDCA	0.5	2.08 [1.66–2.61]	1.97 [1.51–2.57]
TDCA/TLCA	0.0114	1.33 [1.1–1.62]	1.81 [1.49–2.21]
Conjugation, neutral synthesis pathway			
GDCA/DCA	0.0031	0.719 [0.604–0.856]	1.05 [0.836–1.33]
TDCA/DCA	0.1	0.0605 [0.0467–0.0783]	0.0777 [0.0556–0.109]
GCA/CA	0.0104	2.52 [1.86–3.39]	4.13 [2.84–6.01]
TCA/CA	0.076	0.506 [0.358–0.716]	0.711 [0.434–1.16]
Conjugation, acidic synthesis pathway			
GUDCA/UDCA	0.7	0.546 [0.341–0.876]	0.515 [0.334–0.793]
TUDCA/UDCA	0.746	2.83 [1.93–4.17]	2.04 [1.33–3.13]
GCDCA/CDCA	0.491	8.15 [6.21–10.7]	8.36 [5.68–12.3]
TCDC/CDCA	0.644	0.734 [0.53–1.01]	0.687 [0.433–1.09]
TLCA/LCA	0.961	0.282 [0.2–0.396]	0.252 [0.182–0.347]
GLCA/LCA	0.721	0.711 [0.53–0.955]	0.749 [0.552–1.02]
Conversion of primary to secondary. Gut metagenome activity			
DCA/CA	0.0459	8.01 [5.8–11.1]	12 [7.89–18.1]
TDCA/CA	0.0056	0.469 [0.309–0.711]	0.906 [0.517–1.59]
GDCA/CA	0.0007	5.76 [4.01–8.27]	12.5 [7.71–20.2]
LCA/CDCA	0.451	0.66 [0.419–1.04]	0.589 [0.42–0.825]
UDCA/CDCA	0.691	1.39 [0.815–2.36]	1.94 [1.23–3.07]

Metabolite	P value	Mean [95%CI]	
		Control	AD
TUDCA	0.0275	1.07 [0.78–1.48]	1.92 [1.53–2.41]

CSF manifested few differences in bile acids and their ratios. The AD group had 1.3-fold higher levels of GLCA and 1.4-fold higher level of T-a-MCA. Additionally, AD group had lower ratio of GCDCA/GLCA (1.3-folds, **Table S4**).

Of those measured, only a few steroid hormones showed different levels between AD and the control. In plasma, dehydroepiandrosterone sulfate (DHEAS) and progesterone were lower in AD group (1.9 and 1.7-folds respectively). Additionally, testosterone and the testosterone/progesterone ratio showed significant gender x group interaction. Females AD subjects showed 1.4-fold lower testosterone, when compared to females controls, but no differences were observed in males. On the other hand, the testosterone/progesterone ratio was 2-fold higher in AD male subjects compared male controls. Testosterone/progesterone ratio differences were not observed in females.

In CSF, only corticosterone showed a significant difference between AD and the control group, however, the magnitude of the-fold difference only ~ 1.1.

Relation between CSF and plasma AD markers.

In the current study, matched plasma and CSF samples were collected, allowing an assessment of the relationships between metabolites in these pools. Spearman's ρ rank order correlation between plasma and CSF lipid mediator levels are shown in Table 2. The associations were distinct by metabolite classes, with oxylipins showing only 2 of 15 significant correlations, while bile acids and steroids showing 14 of 18 significant correlations. Correlations within PUFA and PUFA ethanolamide were also apparent for the long chain omega 3 species (DHA EPA and DHA ethanolamide) but not others.

Table 2
Spearman's rank order correlation between plasma and CSF
metabolites. Significant p values are bolded.

Metabolite class	Metabolite	Spearman's ρ	P value
Oxylipins	PGF2a	-0.2	0.0746
	F2-IsoP	0.34	0.0019
	12_13-DiHOME	0.26	0.0223
	9_10-DiHOME	0.15	0.1810
	15_16-DiHODE	0.21	0.0625
	14_15-DiHETrE	0.062	0.5830
	11_12-DiHETrE	0.11	0.3340
	17_18-DiHETE	0.33	0.0027
	19_20-DiHDoPE	0.16	0.1450
	13-HODE	0.029	0.7970
	9-HODE	-0.054	0.6380
	13-HOTE	-0.014	0.9010
	9-HOTE	-0.051	0.6520
	12(13)-EpOME	0.0089	0.9380
	9(10)-EpOME	-0.028	0.8070
Acyl-EA	OEA	-0.031	0.7820
	LEA	0.062	0.5860
	DHEA	0.51	0.0001
PUFA	LA	0.15	0.1860
	ALA	0.2	0.0826
	AA	-0.049	0.6670
	EPA	0.45	0.0001
	DHA	0.42	0.0001
Bile acids	CDCA	0.51	0.0001
	UDCA	0.78	0.0001
	DCA	0.71	0.0001

Metabolite class	Metabolite	Spearman's ρ	P value
	TCA	0.62	0.0001
	TCDCA	0.43	0.0001
	TUDCA	0.16	0.1570
	TDCA	0.54	0.0001
	GCA	0.36	0.0012
	GCDCA	0.19	0.0925
	GUDCA	0.47	0.0001
	GDCA	0.54	0.0001
	GLCA	-0.083	0.4670
Steroids	17OH-PROG	0.58	0.0001
	Cortisol	0.3	0.0071
	Cortexolone	0.089	0.4340
	corticosterone	0.51	0.0001
	Testosterone	0.81	0.0001

Next, we used partial least square discriminant analysis (PLS-DA) to illustrate the relationship between plasma and CSF AD markers (Fig. 3). The analysis showed that discrimination between control and AD was dominated by the plasma metabolites. Fifteen plasma metabolites (and their ratios) manifested variable importance in projection (VIP) score > 1.4 compared to only 4 CSF metabolites. The discrimination between AD and the control group was characterized by higher plasma 17,18-DiHETE (VIP = 2.16) and CSF EpOMEs (VIP = 1.95 and 1.58 for the 12(13) and 9(10) isoforms respectively) and lower levels of the acylethanolamide ratios including both DHEA/LEA and DEA/LEA in plasma and both plasma and CSF OEA/LEA. Plasma and CSF OEA/LEA manifest similar discriminatory power based on their proximity on the loading plot. On the other hand, plasma 17,18-DiHETE and CSF EpOMEs occupied distinct parts of the loading plot, suggesting distinct discriminatory properties. The VIPs for each metabolite are provided in the **Table S5**.

Fatty acid ethanolamides and CYP/sEH metabolites are strong AD predictors in both plasma and CSF. We used predictive modeling to investigate how well plasma and CSF metabolites can report AD status. Plasma lipid mediators generated stronger models than those in CSF with area under the receiver operator characteristic curves (ROC AUC) of 0.924 vs. 0.824, with the two models consisting of distinct metabolites (Fig. 4). However, in both matrices, the strongest predictors belonged to the same two metabolic pathways, the acyl ethanolamides and CYP/sEH pathway. Plasma predictors included ethanolamides (OEA and DEA normalized to the LEA level), the 12,13-DiHOME/EpOME an indicator of

sEH activity (28) and sEH metabolite of AA (14,15-DiHETrE). In CSF, the strongest predictors included OEA/LEA and the linoleate-derived epoxides 12(13)-EpOME and 9(10)-EpOME. When plasma and CSF markers were combined in predictive model efforts, the resulting model consisted uniquely of ethanolamides, including plasma long chain PUFA ethanolamides (DEA/LEA and DHEA/LEA) and CSF OEA/LEA. This model resulted in the ROC AUC of 0.889.

For all 3 models, ethanolamides OEA, DEA and DHEA were stronger predictors when used as a ratio to LA derived ethanolamide – LEA. LEA itself was not different between AD and the control group in either plasma or CSF (Fig. 1 and Fig. 2) unlike OEA, DEA and DHEA. Therefore, LEA likely serves as a surrogate for the general acyl-ethanolamides level and adjustment of other ethanolamides by LEA lowers intra-individual variability.

Lipid mediator–cognitive score associations in AD. The AD cohort is characterized by a high log(t-Tau/A β 42) ratio and MoCA scores ranging from normal cognitive function to severe cognitive impairment (**Figure S3**). Taking advantage of the broad MoCA range, we investigated lipid mediator associations with cognitive function in this group pathological levels of t-Tau/A β 42. Additionally, since the AD cohort was represented by subjects in both fasted and non-fasted states, we stratified the analysis by fasting state for plasma samples (Table 3). In the fasting state, PUFA oxidation markers, 5,15-DiHETE and 9-HETE were negatively associated with the MoCA score (although only 5,15-DiHETE passed FDR correction). 5,15-DiHETE can have enzymatic or autooxidative origin, whereas 9-HETE is a strictly an autooxidative product. 5,15-DiHETE correlated with 9-HETE in fasted subjects with an $R^2 = 0.415$ ($n = 60$; $p < 0.001$). In non-fasted AD subjects, a strong positive association between the MoCA score and EPA-derived ethanolamide (EPEA) as well as the levels of EPA and DHA were observed. Additionally, a positive correlation was detected between MoCA and the EPA-derived 17,18-DiHETE, the DHA-derived 14-HDoHE and the 18 carbon PUFAs (LA and ALA), however, these did not pass FDR correction.

Table 3

Spearman's ρ rank order correlation between MoCA score and plasma lipid mediators in AD patients. Analysis stratified by predicted fasted state. Only correlations with the $p < 0.05$ are shown. P-values that passed FDR correction at $q = 0.2$ are bolded.

Fasted			Non-fasted		
Metabolite	Spearman ρ	P value	Metabolite	Spearman ρ	P value
5,15-DiHETE	-0.448	0.0005	EPEA	0.424	0.0003
9-HETE	-0.338	0.0102	EPA	0.386	0.001
13-KODE	-0.299	0.0238	DHA	0.338	0.0043
DCA	-0.273	0.0398	17,18-DiHETE	0.3	0.0117
			4-HDoHE	0.269	0.0246
			LA	0.267	0.0254
			ALA	0.249	0.0373
			9-HETE	-0.246	0.0405
			8-HETE	-0.24	0.0459

In CSF, the linoleic acid derived epoxides 12(13)- and 9(10)-EpOMEs showed weak but significant positive correlations with MoCA ($p > 0.2$, $p < 0.005$; Table 4). Additionally, positive associations were observed between MoCA and DHA and DHA derived diol (19,20-DiHDoPE) and conjugated bile acids GCA (and the ratio of GCA to GDCA and GCDCA), TCDCA and the conjugated to unconjugated ratio for DCA and CDCA (GCA/GCDCA, GCDCA/CDCA and TCDCA/CDCA). However, only linoleic acid epoxides passed the FDR correction.

Table 4
Spearman's ρ rank order correlation between MoCA score and CSF lipid mediators in AD patients. Only correlations with the $p < 0.05$ are shown. P-values that passed FDR correction at $q = 0.2$ are bolded.

Metabolite	Spearman ρ	P value
12(13)-EpOME	0.279	0.0009
9(10)-EpOME	0.24	0.0047
19,20-DiHDoPE	0.21	0.0138
GCA	0.207	0.0152
DHA	0.203	0.0174
GCA/GDCA	0.202	0.0182
GCDCA/CDCA	0.19	0.026
TDCA/DCA	0.19	0.0261
GCA/GCDCA	0.18	0.0352
TCDCa/CDCA	0.172	0.0441
TCDCa	0.17	0.0468

Discussion

Metabolic disruptions influencing vascular physiology, inflammation and energy metabolism have been reported to increase the risk of Alzheimer's disease, however whether these changes are independent risk factors or how they may interact has not been well established. If novel biomarkers of AD can be identified within these domains, they could not only provide useful screening and risk assessment tools but may also provide insight into connections between metabolism and neurodegenerative diseases. To this end, we performed a comprehensive analysis of plasma and CSF lipid mediators, endogenous regulators of multiple processes including inflammation and energy metabolism and described their associations with AD and cognitive function. In the process, we identify clear differences between AD and healthy controls in two metabolic pathways, CYP/sEH and fatty acids-derived ethanolamides, and subtle differences in bile acids and steroids.

AD-associated differences in plasma bile acid were in agreement with previously reported analyses of The Religious Orders Study and the Rush Memory and Aging Project (ROS/MAP) cohort (17). These included lower levels of CA and the CA:CDCA ratio in AD, suggesting that the neutral bile acid synthesis pathway could be affected in AD. There are few studies regarding the shift between neutral and acidic BA synthesis, one of them reporting an increase in neutral/acidic pathway products ratio in nonalcoholic steatohepatitis (29). However, the biological relevance of this difference in terms of AD is

yet to be determined. Interestingly, we previously reported associations between postprandial bile acids and cognition, with few associations in the fasting state (20). Considering that the current manuscript focuses on AD related differences in the fasting state, further studies probing postprandial bile acid metabolism and AD are suggested. Interestingly, several bile acid and steroid differences in AD were gender specific. Few studies reports gender specific action of TUDCA and UDCA on ER stress markers in rodent model for the prion disease (30). These findings further support the importance of gender focused approaches when investigating cholesterol-derived metabolism in the context of AD, as established with regards to the links between ApoE4 and AD risk in post-menopausal women (31).

With respect to fatty acid metabolism, our study identified substantial AD-associated elevations in CYP/sEH pathway products and lower levels of acylethanolamides in both plasma and CSF, although different elements of these pathways were affected in plasma and CSF. Oxylin and endocannabinoid levels show no association between plasma and CSF, suggesting independent regulation of these pools. Likewise, CYP/sEH metabolites of plasma and CSF manifest distinct discriminatory power in PLS-DA models of AD. On the other hand, CSF and plasma acylethanolamides seem to manifest similar AD discriminatory power and can be substituted in the AD predictive model. These findings are consistent with previous reports implicating both CYP/sEH metabolites and acylethanolamides as important regulators of inflammation in neurodegenerative disorders (11, 32, 33).

In the CYP/sEH pathway, polyunsaturated fatty acids are converted to anti-inflammatory and vasodilating epoxy fatty acids by CYPs, which are further metabolized to pro-inflammatory and vasoconstricting diols by sEH, a process primarily recognized in cardiovascular disease (34). In the current study, we found higher plasma level of the EPA sEH metabolite 17,18-DiHETE in AD patients, but only slight difference in the parallel AA metabolites were observed. Notably, EPA metabolites derived from LOX pathways were lower in AD patients, suggesting that the observed differences are not a result of differential omega-3 fatty acid intake, but rather specific enhancement in sEH-dependent EPA metabolism. Omega-3 sEH metabolites are particularly potent regulators of the cardiovascular system, especially blood vessel tone and vascular inflammation (35) and sEH inhibitors have been suggested to improve outcomes for both cardiovascular (36) and neurodegenerative diseases (37). Clinical associations between cardiovascular disease and AD have been reported, where the regulation of a vascular tone and blood flow play a role in both pathologies (38). Additionally, we previously reported plasma EPA sEH metabolites to be negatively associated with perceptual speed in cognitively normal subjects (20). Therefore, the current findings further support involvement of vascular dysfunction in AD, perhaps through alterations to the blood brain barrier and vascular-related inflammatory signaling, with overlapping molecular mechanisms leading to cardiovascular and neurodegenerative pathologies. When considering these shifts in oxylin profiles, it is important to remember that an epoxides reservoir is generated by esterification into phospholipid membranes (39), whereas diols are not readily reincorporated into the membranes and rapidly appear in the free pool and are actively excreted from cells (40). Since tissue esterified lipid mediators were not evaluated in these samples, it is difficult to know whether the observe difference in EPA diol was due to an increased production of CYP/sEH metabolites or increased clearance of membrane bound EPA epoxides, and future studies are needed to resolve this issue.

In contrast to plasma, CSF showed higher levels of LA-derived epoxides, along with a moderate increase in LA-diols, but not CYP/EH metabolites of longer chain PUFAs. The source of the CSF metabolites is likely tied to the central nervous system, and linoleate-derived oxylipins have been identified as the dominant form in the developing rat brain (41). Like long chain PUFAs, LA-derived epoxides and diols can also modulate vascular tone (42) and multiple studies point towards their cytotoxic and pro-inflammation nature. However most of these studies used concentrations greatly exceeding physiological levels (43) and cytotoxic effects were sEH-dependent, pointing towards LA diols as cytotoxic agents (44). Interestingly, LA CYP/sEH metabolites elevated in the spinal tissue of burn victims were shown to activate the transient vanilloid receptor type 1 (TRPV1) (45). Activation of the TRPV1 can rescue neuronal function from A β -induced impairment (46) and can alleviate cognitive and synaptic plasticity impairments in the *APP23/PS45* mouse model of AD (47). Considering that acylethanolamides are also potent activators of the TRPV1 (48), increased LA CYP metabolites may compensate for the AD-related decrease in these ethanolamides. This hypothesis is supported by positive correlation of CSF LA-epoxides with the MoCA score in the AD patients, suggesting elevation of epoxy fatty acids in the central nervous system being potentially beneficial in AD. It is important to mention that the transportation of oxylipins in CSF is poorly understood. In plasma, the majority of oxylipins are transported as complex lipid esters in lipoproteins, with different lipoproteins manifesting distinct oxylipin compositions (49). Therefore, the potential for lipoprotein-dependent oxylipin transport within CSF specific HDL particles (50) is particularly intriguing and warrants further investigation.

Together the CSF and plasma results implicate changes in both peripheral and central CYP/sEH metabolism in association with AD and cognitive impairment. These conclusions are consistent with previous reports of single nucleotide polymorphisms (SNPs) in the CYP2J2 promoter region that reduces gene expression by ~ 50% that appears to increase the ApoE4-independent AD risk (51). Several functional SNPs are also known to influence sEH activity and/or expression and influence disease risk (52). Relevant to AD-associated pathologies, loss of function sEH mutations protect neurons from ischemia-induced death (53) and may alter the risk of vascular cognitive impairment (54). Additionally, postmortem brains from human subjects with AD show higher sEH levels, when compared to the healthy controls, and sEH inhibitors can reverse microglia and astrocyte reactivity and immune pathway dysregulation in mouse AD models (32). Additionally, brain sEH was positively associated with AD in a replicated protein wide-association study of AD (55). Therefore, reducing sEH function appears to be protective, and supports sEH as a valuable therapeutic target for the treatment and investigation of neuro-inflammatory pathologies including AD.

Both plasma and CSF acylethanolamides were lower in AD, with both PLS-DA and predictive model identifying OEA as the strongest predictor of AD in the current cohort. Acylethanolamides are generally considered antiinflammatory (56) and neuroprotective (57) and were previously implicated in neuroinflammatory processes (58, 59). Their neuroprotective action is mediated by activation of the CB1 and CB2 receptors (60) and TRPV1, involved in the acute and inflammatory pain signals in periphery (61). Some acylethanolamides, like OEA, are also peroxisome proliferator-activated receptor (PPAR) α agonist (60) and regulate satiety and sleep with both central and peripheral anorexigenic effects. Notably, sleep

disturbances themselves have been reported to be a risk factor for AD (62), and the identified reductions in CSF OEA would be consistent with such a physiological manifestation (63). A recent study also suggested that the EPA-derived ethanolamide (EPEA) is a potential PPAR γ agonist (64), a transcription factor known for its neuroprotective and anti-inflammatory action (65). Interestingly, non-fasting levels of EPEA showed positive association with MoCA in AD patients. This is in agreement with our previous findings of acylethanolamides in non-fasted individuals, including EPEA, being positively associated with perceptual speed in cognitively normal elderly individuals (20). Literature provides conflicting results regarding both the levels of acylethanolamides in biological fluids, as well as the expression of CB1 and CB2 receptors in the context of AD (66). Nevertheless, the body of literature suggests that exogenous cannabinoids are potent activators of the CB1 and CB2 receptors with potential therapeutic benefit for AD treatment, due to their neuroprotective and anti-inflammatory activity (67). Our data suggest acylethanolamide biology is altered in relation to both AD pathology as well as cognition. However, future studies are needed to fully elaborate the role of these endocannabinoids in AD pathology.

Limitations

This study was conducted using opportunistically collected samples with the fasting state estimated using a previously developed predictive model (20) with an $\sim 17\%$ inherent misclassification rate.

Conclusions

In conclusion, the current study shows AD related differences in CYP/sEH and acylethanolamide metabolism observed in both plasma and CSF. Strong predictive and discriminant models suggest their potential as biomarkers of AD-associated metabolic disruptions. This further supports the contention that a combination therapy reducing sEH activity while increasing acylethanolamide tone by either promoting production or reducing degradation could be a more effective strategy than targeting either pathway independently in treating multifactorial inflammatory diseases like AD (32). Important questions remain regarding the metabolic changes in the lipid mediators preceding pathological changes in tau and cognitive decline. We have previously reported that plasma sEH metabolites of the long chain omega-3 PUFA were negatively associated and PUFA ethanolamides positively associated with perceptual speed (20), mimicking the currently described AD related associations. While these data suggest early alterations in these important regulatory pathways, a comprehensive analysis of longitudinal metabolome changes in relation to cognition and tauopathies is warranted. Combining assessments of dietary, lifestyle and genetic factors promoting these metabolic changes offers the opportunity for novel risk factor discovery and the development of targeted preventive measures.

Declarations

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Conflicts of Interest: none

Kaddurah-Daouk is an inventor on several patents in the field of metabolomics and holds equity in Metabolon Inc which was not involved in this study.

Author contribution:

KB, JWN and **TLP** contributed to metabolomic data generation and quality assessment and quality control. **KB** and **JWN** analyzed the data and wrote the manuscript. **RKD, NTS, JJL, AIL** and **CMH** designed the study and contributed to data interpretation. **EBD** contributed to data interpretation. **NTS, JJL, AIL, GL** and **CB** contributed to the subject selection and collected the clinical data and the samples. **RKD, NTS, JJL, AIL** revised the manuscript. All authors read and approved the final manuscript.

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Figures

Figure 1

Plasma differences in oxylipins and endocannabinoids, between control and AD group. Fold changes projected on fatty acids, oxylipin and endocannabinoids metabolic pathway. Only differences with the t-test $p < 0.05$ and FDR correction at $q = 0.2$ are shown. Network presents fatty acids metabolic pathway, including saturates and monounsaturates (SFA and MUFA) and omega 3 and omega 6 fatty acids with oxylipins and endocannabinoids synthesis pathway. Both detected (black font) and not-detected (gray font) metabolites are shown to visualize the coverage of the metabolic pathway in our targeted assay

and facilitate data interpretation. Although, for the clarity of the figure, not all assay metabolites are displayed. The complete list of measured metabolites was previously reported (68). Oxylin metabolizing enzymes are colored by their class: red – lipoxygenase (LOX) and autoxidation pathway; blue – cytochrome p450 (CYP) epoxygenase; green – cyclooxygenase (COX); yellow – N-acylphosphatidylethanolamide-phospholipase D. Node size represents the fold difference, and the color represents the directionality of the difference: orange – higher in AD; light blue – lower in AD. Key enzymes involved in metabolic step are abbreviated next to the edge. Fads – fatty acid desaturase; Elov – fatty acids elongase; sEH – soluble epoxide hydrolase; DH – dehydrogenase. Saturated and monounsaturated fatty acids were not measured in this assay and are indicated only to visualize precursors for measured oxylin and endocannabinoids. Group means for all metabolites together with corresponding t-test p-values are provided in the Table S3.

Figure 2

CSF differences in oxylin and endocannabinoids, between control and AD group. Fold changes projected on fatty acids, oxylin and endocannabinoids metabolic pathway. Only differences with the t-test $p < 0.05$ and FDR correction at $q = 0.2$ are shown. Network presents fatty acids metabolic pathway, including saturates and monounsaturates (SFA and MUFA) and omega 3 and omega 6 fatty acids with oxylin and endocannabinoids synthesis pathway. For the clarity purposes, only metabolites detected in CSF are shown. Oxylin metabolizing enzymes are colored by their class: red – lipoxygenase (LOX) and autoxidation pathway; blue – cytochrome p450 (CYP) epoxygenase; green – cyclooxygenase (COX); yellow – N-acylphosphatidylethanolamide-phospholipase D. Node size represents the fold difference, and the color represents the directionality of the difference: orange – higher in AD; light blue – lower in AD. Key enzymes involved in metabolic step are abbreviated next to the edge. Fads – fatty acid desaturase; Elov – fatty acids elongase; sEH – soluble epoxide hydrolase; DH – dehydrogenase. Saturated and monounsaturated fatty acids were not measured in this assay and are indicated only to visualize

precursors for measured oxylipins and endocannabinoids. Group means for all metabolites together with corresponding t-test p-values are provided in the Table S4.

Figure 3

Relation between plasma and CSF predictors of AD. Partial least square discriminant analysis (PLS-DA) of AD vs control, utilizing metabolites from both plasma and CSF in predicted fasted samples. Treatment group discrimination is shown by the SCORES (inset) with a plane of discrimination indicated by dashed red line, while metabolites weighting in group discrimination are shown by the LOADINGS. Loading node color indicates metabolite origin (pink for plasma and blue for CSF). Loading node size indicates metabolite variable importance in projection (i.e. VIP). Analysis was performed with all measured metabolites, including specific ratios, but only those with $VIP \geq 1.4$ are displayed for clarity purpose.

Figure 4

Predictive model for AD with plasma and CSF metabolites. Predictive model built independently for plasma (left) and CSF (middle) and plasma+CSF (right). Effect summary shows metabolite model components, sorted by their contribution to the model, with key pathway colored in yellow (fatty acids ethanolamides) and blue (cytochrome p450/soluble epoxide hydrolase pathway). Receiver operating characteristics (ROC) curve for the training set, together with the area under the curve (AUC) and the n for the training (T) and the validation (V) cohorts, are showed in the bottom panel. Metabolite selection through the stepwise logistic regression model together with validation cohort parameters are provided in the Table S6.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [FigureS1.pdf](#)
- [TableS1Biometrics.xlsx](#)
- [TableS2Fastingvsnonfasting.xlsx](#)
- [TableS3TTestplasma.xlsx](#)
- [TableS4TTestCSF.xlsx](#)
- [FigureS3.pdf](#)

- [FigureS2.pdf](#)
- [TableS5VIPs.xlsx](#)
- [TableS6Predictivemodeling.xlsx](#)