

# Uptake of phosphorus by epilithon in three oligo- to mesotrophic post-mining lakes

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## Research Article

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13      **Abstract**

14        Epilithon contributes to phosphorus (P) cycling in lakes, but its P uptake traits have been  
15        rarely studied. We measured the chemical composition of epilithon and its inorganic P uptake  
16        kinetics using isotope  $^{33}\text{P}$  in three deep oligo- to mesotrophic post-mining lakes in April, July,  
17        and October 2019. Over the sampling period, epilithon biomass doubled, while the P content  
18        in biomass dropped to 60% of the April values. High epilithic C:P molar ratios (677 on average)  
19        suggested strong P deficiency in all investigated lakes.

20        Regarding the kinetic parameters of phosphorus uptake, maximum uptake velocity  
21        ( $V_{max}$ , seasonal range  $0.9\text{--}101 \text{ mg P g OM}^{-1} \text{ h}^{-1}$ ) decreased by an order of magnitude from April  
22        to October, while half-saturation constant ( $K_s$ ,  $1.6\text{--}103 \text{ mg P L}^{-1}$ ) did not show any consistent  
23        temporal trend. We found a general decrease in the specific P uptake affinity ( $0.1\text{--}2.2 \text{ L g OM}^{-1}$   
24         $\text{h}^{-1}$ ) and the estimated *in-situ* P uptake ( $0.04\text{--}2.3 \text{ } \mu\text{g P g OM}^{-1} \text{ h}^{-1}$ ) of epilithon over the  
25        season, which might have resulted from changes in epilithon community composition, a  
26        decreasing ratio of living biomass to extracellular polymers and detritus, rapid internal  
27        recycling, and/or thickening of the epilithic biofilm. Additionally, we observed a phenomenon  
28        of reversible abiotic P adsorption on epilithon.

29     **Introduction**

30         Periphyton is an assemblage of autotrophic and heterotrophic organisms that  
31         produces extracellular polymeric substances, all together forming a biofilm attached to a  
32         solid substrate<sup>1,2</sup>. Epilithon is a subgroup of periphyton covering submerged surfaces of  
33         stones<sup>1</sup>. Periphyton represents an important component of aquatic food webs and  
34         biodiversity<sup>1,3</sup> and can contribute considerably to nutrient cycling<sup>4,5</sup>. Along with  
35         phytoplankton and submerged macrophytes, periphyton can account for up to 92% of the  
36         total primary production in oligotrophic lakes<sup>6</sup>.

37         Periphyton productivity may be limited by the availability of inorganic nutrients,  
38         especially phosphorus (P). Phosphorus is an essential constituent of microbial cells.  
39         Integrated into phospholipids, nucleic acids, coenzymes and various metabolites, it is  
40         considered the most common element limiting the growth rates and metabolic activities of  
41         photoautotrophs in temperate freshwater ecosystems<sup>7,8</sup>. Whereas P uptake by  
42         phytoplankton has been widely studied<sup>9–12</sup>, only few studies have addressed the kinetics of P  
43         uptake by periphyton in lentic waters<sup>13,14</sup> and no study ever quantified the P uptake of  
44         periphyton in temperate freshwaters. Given the above-mentioned importance of periphyton  
45         for total primary production of lakes, the periphyton P uptake might be significant as well. A  
46         study of P fluxes in the shallow subtropical Lake Okeechobee found a significantly greater P  
47         uptake in phytoplankton than in periphyton. However, in an oligotrophic marsh area in the  
48         lake, due to the well-developed periphytic biomass, the cumulative P uptake by periphyton  
49         was twice as high as P uptake by phytoplankton<sup>14</sup>.

50         A thorough analysis of periphyton and phytoplankton contributions to P cycling in lake  
51         ecosystems requires the determination of kinetic parameters of P uptake for both  
52         periphyton and phytoplankton. In osmotrophic organisms, phosphorus can only be taken up

53 in substantial amount in the form of inorganic orthophosphate ( $P_i$ )<sup>15</sup>. The relationship  
54 between  $P_i$  concentration in water and its uptake rate can be often described by a  
55 hyperbolic function analogous to Michaelis-Menten equation for enzymatic reaction kinetics  
56 possessing two parameters: maximum velocity ( $V_{max}$ ) and half-saturation constant ( $K_s$ ,  
57 concentration at which  $V_{max} = 1/2$ ). However, at extremely low  $P_i$  concentrations, typical for  
58 oligotrophic waters, P uptake velocity is far below  $V_{max}$ , as  $P_i$  is much smaller than  $K_s$ <sup>16,17</sup>. A  
59 relevant parameter describing P uptake and the competitive ability of microorganisms at low  
60  $P_i$  is the specific P uptake affinity (*SPUA*), calculated as a proportion of  $V_{max}$  and  $K_s$ , equal to  
61 the initial slope of the function at the beginning of the curve<sup>18,19</sup>. Interpretation of the *SPUA*  
62 values is not straightforward due to their unusual units (L mg OM<sup>-1</sup> h<sup>-1</sup>); however, it is  
63 analogous to the clearance rate of zooplankton – the volume of water cleared of phosphate  
64 per unit biomass per unit time<sup>20</sup>.

65 Large lakes newly emerge during the hydric recultivation of post-mining pits.  
66 Understanding the nutrient cycling and energy fluxes in the unique ecosystems of post-  
67 mining lakes is urgently needed since their number is expected to rise in the upcoming  
68 decades due to the coal mining suppression associated with the European Union's Green  
69 Deal<sup>21,22</sup>. Post-mining lakes are often oligotrophic with very low P concentrations (total  
70 phosphorus,  $TP < 15 \mu\text{g L}^{-1}$  and soluble reactive phosphorus concentration,  $SRP \sim 1 \mu\text{g L}^{-1}$ )<sup>23</sup>.  
71 However, no detailed report on P cycling in these lakes and about the organisms involved is  
72 available.

73 In the present study we sampled three recently flooded oligo- to mesotrophic post-  
74 mining lakes in the north-west region of the Czech Republic (Milada, Medard and Most  
75 Lakes) to evaluate the role of dissolved inorganic phosphorus acquisition for periphyton  
76 nutrition and in-lake nutrient cycling. Despite very low P concentration in water, dense mats

77 of periphyton, mainly represented by epilithon, have developed in all investigated lakes  
78 (Řeháková, unpublished data).

79 We sought to investigate P uptake traits of epilithon in these three lakes during the  
80 whole vegetation season, using  $^{33}\text{P}$ -labelled orthophosphate. Our specific objectives were (i)  
81 to determine kinetic parameters of the epilithon P uptake, namely the specific P uptake  
82 affinity ( $SPUA_E$ ), maximum P uptake velocity ( $V_{max,E}$ ), and half-saturation constant ( $K_S$ ); (ii) to  
83 estimate *in-situ* P uptake by epilithon; (iii) to decipher the environmental factors affecting  
84 the specific P uptake affinity; and (iv) to compare specific P uptake affinities of epilithon and  
85 phytoplankton.

86 **Results**

87 **Background limnological parameters**

88 All lakes (see **Fig. 1** for their geographic position) had high water transparency  
89 corresponding to the euphotic depth (the depth of 1% of the surface irradiance) between 10  
90 and 18 m (**Table 1**) and were stratified with the mixed depth varying from 3 to 11 m. In April,  
91 the sampling was performed shortly after the spring overturn, thus the stratification was  
92 weak, nutrient concentrations were higher, and the temperature lower (7–11 °C), than in  
93 summer and autumn (13–24 °C, **Table 1**). Lakes Medard and Most were characterised by a  
94 low chlorophyll *a* (*Chla*) and total phosphorus (*TP*) concentrations (0.9 and 1.9 µg L<sup>-1</sup>; 4.2  
95 and 11 µg L<sup>-1</sup> respectively in their seasonal averages). Lake Milada's *Chla* and *TP*  
96 concentrations were slightly higher (5.1 and 14.1 µg L<sup>-1</sup>, respectively).

97

98 **Epilithon characterisation**

99 Parameters related to epilithon biomass and chemical composition, as well as their  
100 differences among lakes and seasonal development with statistical differences are  
101 summarised in **Fig. 2** and in **Table 2** (for the full two-way ANOVA table see **Supplementary**  
102 **Table 1**). Epilithon biomass expressed as organic matter (*OM*) content and normalised per  
103 area on average doubled from April (1.8 mg cm<sup>-2</sup>) to October (3.6 mg cm<sup>-2</sup>), being about  
104 three times higher in Lake Milada (4.8 mg cm<sup>-2</sup>) than in the two other lakes (**Table 2**, **Fig. 2a**);  
105 the corresponding dry-weight figures showed a similar pattern (**Table 2**, **Fig. 2a**). Average  
106 biomass P content dropped from 3.6 mg P g OM<sup>-1</sup> in April to 2.2 mg P g OM<sup>-1</sup> in October  
107 independently of the lake (**Table 2**). In contrast, average epilithic P content per area  
108 considerably differed among the lakes, being 2.5 times higher in Milada Lake (11 µg P cm<sup>-2</sup>)

109 than in the two other lakes and showing contrasting seasonal trends - a decrease in Most  
110 Lake and an increase in the other lakes.

111 Epilithon nutrient molar element ratios were generally high (**Table 2**) - C:P ratios  
112 ranged from 160 to 1260, N:P ratios ranged from 13 to 68 (**Fig. 2b**), and C:N ratios ranged  
113 from 12 to 25 (**Fig. 2c**), being on average 1.3–2.0 times lower in Lake Medard than in the two  
114 other lakes (**Table 2**, **Fig. 2b,c**). All the ratios were significantly (up to 2-fold) higher in June  
115 and October than in April (**Table 2**, **Fig. 2b, c**).

116

117 Epilithon P uptake - laboratory measurements

118 There was no systematic variability in any epilithon P uptake parameter within lakes  
119 (north shore vs. south shore; a depth of 0.5 m vs. 1.5 m; paired t-tests,  $p = 0.12\text{--}0.81$ ).  
120 Hence, all these in-lake values were treated as replicates ( $n = 4$ , see, e.g., **Figs. 3–5**). Time  
121 course data on P uptake by epilithon (i.e.  $^{33}\text{P}$  disappearance from water) were fitted well to a  
122 simple monoexponential model (**Supplementary Fig. 1**, **Supplementary Table 2**). Fourteen  
123 out of the 348 progress curves were excluded due to a lack of fit (high scatter or an  
124 increasing trend in the data). Calculated P uptake velocities ( $v_{upt}$ ) varied widely from 0.04 to  
125 112 mg P g OM $^{-1}$  h $^{-1}$  and were strongly concentration-dependent (**Supplementary Table 2**).  
126 The relationship between  $v_{upt}$  and  $P_{add}$  followed the Michaelis-Menten kinetics in 28 out of  
127 the 35 cases, while the remaining seven followed the simplified linear model (**Table 3**,  
128 **Supplementary Figs. 2–4**). Maximum P uptake velocity ( $V_{max}$ ) ranged between 0.9–101 mg P  
129 g OM $^{-1}$  h $^{-1}$ . The corresponding estimates of the half-saturation constant ( $K_S$ ) ranged from 1.6  
130 to 103 mg P L $^{-1}$  (**Table 3**, **Fig. 3b, c**). Both  $V_{max}$  and  $K_S$  significantly decreased over the season  
131 by around one order of magnitude, while the observed differences among lakes became less  
132 prominent and were statistically significant only for  $V_{max}$  (**Table 2**, **Fig. 3b, c**).

133        The specific P uptake affinity of epilithon ( $SPUA_E$ ), representing the initial slope of the  
134         $v_{upt}$  vs.  $P_{add}$  plot, ranged from 0.11 to 2.22 L g OM $^{-1}$  h $^{-1}$  (**Table 3, Fig. 4a**). There was a  
135        significant seasonal decrease in  $SPUA_E$  from 1.3 to 0.37 L g OM $^{-1}$  h $^{-1}$  independently of the  
136        lake (**Table 2**). The generalised linear mixed-effect model with season and lake treated as  
137        random factors indicated the C:N molar ratio of the epilithon biomass ( $p < 0.001$ ) and the  
138        lake-water pH ( $p < 0.002$ ) as significant drivers of  $SPUA_E$ .

139        All values of P uptake parameters reported throughout this paper have not been  
140        corrected for abiotic P uptake (see Discussion). Abiotic  $^{33}\text{P}$  adsorption occurring in the  
141        presence of 4-% formaldehyde was relatively high, accounting on average for 8% (range 1–  
142        18%), 24% (7–61%), and 54% (21–90%) of the formaldehyde-untreated samples in Lakes  
143        Milada, Most and Medard, respectively (**Fig. 3a**). In three additional experiments, we applied  
144        formaldehyde at a wide range of P concentrations (0.1–10 mg P L $^{-1}$ ). In two of those we  
145        observed a roughly two-fold increase in P adsorption with increasing P concentration,  
146        whereas no concentration dependence was found in the remaining experiment  
147        (**Supplementary Fig. 5**). We discovered that more than one half of the adsorbed P (50–70%)  
148        could be readily released back to the dissolved phase during a one-hour treatment of the  
149        samples with an oxalacetate-based "surface wash reagent" (**Supplementary Table 3**), which  
150        indicates a weak and reversible abiotic P adsorption.

151

152        Epilithon P uptake - calculated *in-situ* P dynamics

153        *In-situ* epilithon P uptake velocities ( $EPUV$ ,  $\mu\text{g P g OM}^{-1} \text{h}^{-1}$ ) calculated from the  
154        specific P uptake affinity and  $SRP_{corr}$  (corrected soluble reactive phosphorus concentration,  
155        see Methods for explanation) were rather low and variable (0.04 to 2.3  $\mu\text{g P g OM}^{-1} \text{h}^{-1}$ ). On

156 average, the *in-situ* epilithon P uptake was roughly twice as high in Lake Most compared to  
157 the other lakes, with a generally decreasing seasonal trend (**Table 2, Fig. 4b**).

158 Epilithon P doubling time (*EPDT*) – i.e. the theoretical time necessary for doubling the  
159 actual P content in the biomass assuming the *in-situ* P uptake velocity estimated as above –  
160 ranged from 30 to 3,940 days (**Fig. 5a**). Average *EPDT* increased over the season from 140  
161 days in April to 800 days in October. In Lake Medard, it was three to four times as high as in  
162 Lakes Most and Milada (800 days vs. 200–300 days, respectively; **Table 2, Fig. 5a**).

163 Finally, theoretical amounts of the P that epilithon could have sequestered from the  
164 water column during the periods between the two successive sampling campaigns were  
165 calculated (63–105 days) assuming the estimated *in-situ* P uptake velocity. These theoretical  
166 values were compared with observed changes in P content determined in the biomass (**Fig.**  
167 **5b**). The observed and the calculated changes in the epilithon P content did not differ  
168 statistically in Lakes Milada and Medard. In contrast, the theoretical values greatly exceeded  
169 the observed ones in Lake Most, where a decrease or minor change in the P content was  
170 measured (**Fig. 5b**).

171

172 Seston P dynamics and comparison with epilithon

173 Turnover time of dissolved orthophosphate measured with  $^{33}\text{P}$  ranged from 7 minutes  
174 to 3.6 hours indicating moderate (in spring) to extreme (in summer and autumn) P deficiency  
175 in all lakes (**Supplementary Table 5**). Seston P uptake kinetics followed the Michaelis-  
176 Menten model (**Supplementary Fig. 6**), yielding  $V_{max}$  estimates ranging from 0.29 to 1.3  $\mu\text{g P}$   
177  $\text{L}^{-1} \text{h}^{-1}$  and  $K_s$  from 0.4 to 6.1  $\mu\text{g P L}^{-1}$  (**Supplementary Table 5**). The specific P uptake  
178 affinities in seston ranged from 230 to 24,000  $\text{L g OM}^{-1} \text{h}^{-1}$ , being thus 2–4 orders of

179 magnitude higher (**Supplementary Table 5**) than the epilithon-specific values (0.11–2.22 L g

180  $\text{OM}^{-1} \text{ h}^{-1}$ , **Table 3**).

181      **Discussion**

182      Data on periphyton P uptake kinetics in lentic systems are extremely scarce in the  
183      literature<sup>13,14,24</sup>. Our study represents the first report on P uptake kinetics by epilithon  
184      assemblages naturally growing on stones in temperate oligo- to mesotrophic lakes. The only  
185      comparable data are available from very different freshwater habitats (a subtropical lake  
186      and wetlands)<sup>13,14</sup>. The values of specific P uptake affinity ( $SPUA_E$ ) measured in this study  
187      ( $0.11\text{--}2.22 \text{ L g OM}^{-1} \text{ h}^{-1}$ ) were roughly one order of magnitude lower than those presented  
188      for periphyton by Scinto and Reddy in subtropical wetlands<sup>13</sup>. While  $V_{max}$  estimates  
189      presented in our study ( $0.9\text{--}101 \text{ mg P g OM}^{-1} \text{ h}^{-1}$ ) roughly match the values available in the  
190      literature, our  $K_S$  estimates ( $1.6\text{--}103 \text{ mg P L}^{-1}$ ) are 1–3 orders of magnitude higher than those  
191      previously reported<sup>13,14</sup>. The among-system differences in kinetic parameters might be  
192      attributed to the differences in abiotic and biotic drivers affecting P uptake in compared  
193      ecosystems, in the species composition of periphyton, environmental conditions, and  
194      morphological and limnological variances between studied lakes. In freshwater wetlands,  
195      extreme periphytic N:P ratios (152–231) have been reported<sup>13</sup>. Such high values suggest a  
196      strong P limitation of these wetlands and may explain the ten times higher  $SPUA_E$  compared  
197      to our estimates. A positive relationship between the degree of P deficiency and  $SPUA_E$  is  
198      well documented in algae<sup>10,25</sup>. The wide range of linear increase in P uptake velocity with  
199      concentration of dissolved P, reflected by high  $K_S$  values or even by the absence of  
200      saturation, might represent an adaptation of the epilithon community to episodic increases  
201      or pulses of dissolved P, ensuring optimal utilisation of available resources.

202      Variability in  $SPUA_E$  was statistically related to both epilithon C:N molar ratio and lake-  
203      water pH. The negative relation between  $SPUA_E$  and epilithon C:N could indicate a tendency  
204      to a decrease in specific P uptake affinity under conditions of N+P co-limitation of the

epilithic microbial community<sup>26</sup>. Notably, the highest epilithon C:N ratios (around 22) were observed at Lake Milada in July and October (**Fig. 2c**), concomitantly with low  $SPUA_E$  values (**Fig. 4a**) and an extremely low lake-water nitrate concentration (0.02 mg L<sup>-1</sup>; **Table 1**), generally considered to be limiting for algal growth<sup>27</sup>. Since the naturally growing epilithon consist of a variety of species, some of them might have been N-limited, whereas the others might have been P-limited. Following this logic, this could have led to a lower P demand and lower  $SPUA_E$  values of the epilithic community. However, the relation of  $SPUA_E$  to species composition, was not a subject of this study. The relationship between  $SPUA_E$  and pH remains rather obscure.

In the lakes studied, epilithon biomass per area and nutrient stoichiometry corresponded with the values reported for comparable oligotrophic lakes<sup>28</sup>. Two slightly different stoichiometric definitions of growth-limiting conditions were suggested in the literature. According to Hillebrand and Sommer, P limitation of benthic algae is indicated when C:P > 180 and N:P > 22<sup>26</sup>. Kahlert reported the threshold for P limitation of periphyton when C:P > 369 and N:P > 32<sup>29</sup>. Applying both definitions, the values found in the investigated post-mining lakes (average C:P 680 and N:P 39) indicate P limitation of epilithic growth which tends to strengthen over the season (**Fig. 2b**). A comparably high epilithic C:P and N:P ratios were reported also for deep oligotrophic lakes in Sweden (C:P and N:P ratios up to 800 and 74, respectively)<sup>28</sup> and an oligotrophic lake in north-western Ontario (C:P ratio 820)<sup>30</sup>.

We observed a discrepancy between the seasonal development of  $SPUA_E$  and of the other P limitation indices. Epilithon biomass cover (OM per cm<sup>2</sup>) roughly doubled, while biomass P content roughly halved from April to July and October. One would expect a simultaneous increase in  $SPUA_E$  in response to strengthened P limitation<sup>10,25</sup> implied by an

observed decrease in P content and increase in C:P ratio. However, the opposite was found, i.e.  $SPUA_E$  decreased roughly ten times during the same period (**Table 2, Fig. 4a**). There are several possible explanations for this apparent contradiction: (1) Seasonal changes in the epilithic microbial community might favour species with lower P requirements: Seasonal shifts in the phototrophic assemblage composition were in fact observed; however, the significance of their relationship with  $SPUA_E$  has not been proven (Řeháková et al. in prep.). It has been reported in the literature that higher periphyton C:P ratios can be associated with shifts towards a higher contribution of heterotrophic over autotrophic organisms<sup>31</sup>. (2) The decreasing ratio of living biomass to structural (mainly extracellular) polymers or detritus during the seasonal succession:  $SPUA_E$  was normalised to the total *OM* content, thus including both living and dead cells as well as extracellular polymeric substances. This explanation appears to be supported by the facts that the C:P ratio doubled from spring (~410) to the extreme values in summer and autumn (~820, **Fig. 2b**). (3) The high efficiency of P internal recycling in epilithon. P released out from the dying biomass may be taken up by the living cells in the epilithon without leaving the biofilm thus lowering the demand for external P input. (4) The decreased accessibility of P to epilithic microorganisms due to the thickening of the epilithic biofilm on the stone over the season: If true, this would markedly constrain P diffusion through the epilithon matrix as described by Riber and Wetzel<sup>32</sup>. As a result, the cells situated closer to the stone substrate could exhibit lower P uptake than the cells situated near the surface of the biofilm even if their individual P uptake affinities were high. Consequently, this would lead to lower values of overall P uptake and of the related parameters ( $SPUA_E$ ,  $V_{max}$  etc.) of the epilithon.

*In-situ* epilithon P uptake velocities were calculated by multiplying  $SPUA_E$  by the corrected soluble reactive phosphorus values,  $SRP_{corr}$  to elucidate how much our kinetic

253 measurement of P uptake by epilithon can explain the changes in epilithic P content that  
254 actually occurred between sampling campaigns. Although the calculated *in-situ* P uptake  
255 velocities were extremely low, the observed and calculated (potential) changes in epilithon P  
256 content were roughly in agreement in Lakes Milada and Medard. However, in Lake Most,  
257 where epilithon P uptake velocity was the highest, the calculated changes greatly exceeded  
258 the observed ones (**Fig. 5b**), indicating high community losses due to e.g. fragile parts  
259 removal, disturbances<sup>33</sup>, intensive grazing<sup>34</sup> or parasite-induced cell lysis<sup>35</sup>.

260 Our *in-situ* P uptake calculations suffer from an important but unavoidable limitation:  
261 generally,  $P_i$  concentrations in the lake-water cannot be reliably determined under P limiting  
262 conditions because of their extremely low values<sup>9</sup>. *In-situ* epilithon P uptake velocities were  
263 calculated by multiplying the  $SPUA_E$  values by  $SRP_{corr}$  (Eq. 7). The calculation is only plausible  
264 if  $SRP_{corr}$  is a reliable estimate of  $P_i$  concentration, which is extremely unlikely as the  
265 correction relies on  $SRP$  concentration determined with molybdenum blue method<sup>36</sup> known  
266 to highly (up to 100 times) overestimate the actual  $P_i$  concentration at low  $SRP$  levels (< 5 µg  
267 P L<sup>-1</sup>) due to the interference with arsenates and hydrolysed organic phosphorus<sup>9</sup>. One  
268 possibility how to approach the actual  $P_i$  concentration would be an application of the  
269 radiobioassay suggested by Rigler<sup>37</sup>. However, the assay is biased by a subjective decision.  
270 Instead, the conservative correction was used, which is unambiguous (Eq. 7) and based on  
271 the fact that  $P_i$  concentration in lake water cannot exceed  $K_{ssest}$  estimated by measurement  
272 of saturation kinetics of sestonic P uptake if the experiment is carried out in the presence of  
273  $P_i$  naturally occurring in the water sample<sup>38</sup>. The values of  $SRP_{corr}$ , however, still represent  
274 only the upper limit of the real  $P_i$  concentration in lake water and consequently also the  
275 estimates of any derived parameters are on their upper limits.

276 Our values of epilithic specific P uptake affinity,  $SPUA_E$ , were 2–4 orders of magnitude  
277 lower than the corresponding values for seston,  $SPUA_{sest}$  (**Supplementary Table 5**). Similar  
278 differences were also reported for a shallow subtropical lake in Florida<sup>14</sup>. The differences  
279 between  $SPUA_{sest}$  and  $SPUA_E$  likely result from the mostly unicellular arrangement of  
280 sestonic bacterio- and phytoplankton that tends to maximise surface to volume ratio<sup>39</sup>, while  
281 epilithic P uptake is affected by a limited surface for nutrients entering the biofilm and the  
282 presence of an extracellular matrix that limits their transport from water into the cells<sup>32,40</sup>.

283 The use of naturally growing epilithon assemblage in the present study is challenging,  
284 yet provides more ecologically relevant data compared to the commonly used artificial  
285 substrates<sup>41,42</sup>. An obvious disadvantage is, however, an inherent heterogeneity of individual  
286 samples, resulting in a high scatter of P uptake data (see **Supplementary Figs. 2–4**). The  
287 average difference in duplicates was 30%, but the differences were frequently several-fold.  
288 Nevertheless, variation coefficients of  $SPUA_E$  and  $V_{max}$  estimates were on average 23%  
289 (range 7–55%) and 25% (range 8–47%), respectively, (**Table 3**), documenting that these P-  
290 uptake-related kinetic parameters were measured with acceptable error. The estimates of  $K_s$   
291 were, however, less reliable (on average 44% with range 15–80%).

292 Values of P uptake parameters reported throughout this paper have not been  
293 corrected for abiotic P uptake (i.e. uptake occurring in the presence of 4% formaldehyde),  
294 since the results of abiotic adsorption experiments were inconclusive, showing variable  
295 concentration dependency of the formaldehyde effect (**Supplementary Fig. 5**), which does  
296 not allow P uptake to be corrected properly. Nevertheless, the abiotic P uptake was  
297 substantial, averaging at 54% of total P uptake in Lake Medard, 24% in Lake Most, and 8% in  
298 Lake Milada (**Fig. 4B**). In additional experiments, oxalate-based surface wash reagent  
299 released 50–70% of such abiotically adsorbed P, indicating a weak and reversible P binding.

300 (Supplementary Table 3). Similarly, Sañudo-Wilhelmy et al. reported surface abiotic P  
301 adsorption ranging from 60 to 90% of the apparent total P uptake in the phytoplankton<sup>43</sup>.  
302 Surface P adsorption has been suggested to be a mechanism for storing the temporary  
303 excess of P before it can be transported into the algal cells<sup>44</sup>. Our data indicate that the  
304 phenomenon of temporary abiotic P adsorption could be important also in periphyton, but  
305 so far this has not been considered and thus requires further attention in both theoretical  
306 and experimental studies.

307 **Methods**

308 **Study sites**

309 Three oligo- to mesotrophic post-mining lakes, Milada (area: 252 ha, max/average  
310 depth: 25/16 m, flooded in 2010), Medard (493 ha, 59/28 m, flooded in 2016), and Most  
311 (311 ha, 75/22 m, flooded in 2014), situated in the northwestern Bohemia, the Czech  
312 Republic, were chosen as study sites (**Fig. 1**, **Supplementary Figs. 7–9**). All lakes possess well-  
313 developed epilithon assemblages in their littoral zone (**Supplementary Fig. 10**). Basic  
314 limnological parameters of the lakes are summarised in **Table 2**.

315

316 **Sampling**

317 In each lake, epilithon was sampled at two opposite shores of each lake, designated as  
318 North (N) and South (S) (see **Fig. 1** and **Supplementary Table 6** for more details). Twelve  
319 small epilithon-covered stones (approx. diameter 4 cm) were gently collected by scuba  
320 diving at each site from 0.5 and 1.5 m depths in April, July, and October 2019. Lake water for  
321 chemical analyses and epilithon P uptake experiments was collected with a Friedinger  
322 sampler above the deepest points of the lakes from the depth of 0.5 m. Epilithon samples  
323 were kept in the lake water at *in-situ* temperature and a photoperiod of 16:8 (light:dark)  
324 until the P uptake experiments (< 2 days). The cell-free filtrate of lake water was prepared by  
325 passing it through a PCTE filter (0.22-µm porosity; Sterlitech Corporation, Seattle, USA) and  
326 stored at 4 °C until the start of the experiment (< 2 days).

327

328 **Background limnological parameters**

329 Vertical profiles of water temperature, pH, and O<sub>2</sub> concentration were measured with  
330 YSI EXO 2 multiparametric probe (YSI Inc., Yellow Springs, USA). The euphotic depth was

331 estimated as the depth of 1 % of surface irradiance calculated from vertical light profiles  
332 obtained using LICOR LI-1400 datalogger with a spherical quantum underwater sensor LI 193  
333 SA (Licor, Lincoln, NE, USA). Chlorophyll a (*Chl**a*) was determined according to ISO  
334 10260:1992<sup>45</sup>, total organic carbon (*TOC*) and total nitrogen (*TN*) were determined using  
335 Shimadzu *TOC/TN* analyser. Total phosphorus (*TP*) was measured spectrophotometrically  
336 after nitric-perchloric acid digestion<sup>46</sup> using Flow Injection Analyser. Dissolved reactive silica  
337 (*DSi*)<sup>47</sup> and soluble reactive phosphorus (*SRP*)<sup>36</sup> were determined spectrophotometrically.  
338 Nitrate-nitrogen (*NO*<sub>3</sub>-*N*) was determined following the procedure of Procházková<sup>48</sup>.

339

340 Epilithon chemical characterisation

341 After termination of <sup>33</sup>P uptake experiments (see below), epilithon was quantitatively  
342 removed from each stone, dried at 110°C to constant weight and the dry weight (*DW*) was  
343 determined. To determine the organic matter (*OM*) content, the dry sample was combusted  
344 in a muffle furnace at 450°C for 12 h. For determination of epilithon elemental (C, N, P)  
345 composition and biomass (*DW* and *OM* per area), we used different stones than those used  
346 for uptake experiments, albeit collected from the same sampling site. Samples were frozen  
347 till the analysis at -20 °C and homogenised using glass beads and an automatic homogeniser.  
348 *TN*, *TOC*, and *TP* were analysed, as described above.

349

350 Epilithon P uptake kinetics

351 Before each experiment, the lake water was removed from containers containing  
352 epilithon-covered stones. Stones were gently rinsed with 50 mL of the 0.2-µm filtered (cell-  
353 free) lake water, and 100 mL of the filtered water was added. The containers were  
354 supplemented with five different concentrations of orthophosphate (*P*<sub>add</sub>, 1, 3, 10, 30, 100

355 mg P L<sup>-1</sup>, added as KH<sub>2</sub>PO<sub>4</sub>) in duplicates. Incubations started by the addition of carrier-free  
356 <sup>33</sup>P-labelled H<sub>3</sub>PO<sub>4</sub> (50–100 kBq; American Radiolabelled Chemicals, Inc., St. Louis, USA)  
357 along with unlabelled orthophosphate into testing containers placed in a gently shaken (120  
358 rev/min) water bath tempered at *in-situ* temperature. Samples were incubated for 180–240  
359 min under laboratory illumination (cca 10 µE m<sup>-2</sup> s<sup>-1</sup> PAR). At 15–30 min intervals, 0.6-mL  
360 aliquots were transferred into Eppendorf tubes containing 0.6 mL of scintillation cocktail  
361 (Packard Ultima Gold) and vortexed vigorously. Radioactivity (typically 20–40 thousand  
362 c.p.m. per aliquot) was determined with a liquid scintillation counter (Tri-Carb 2900TR,  
363 Packard, USA). Control for abiotic uptake was performed by adding formaldehyde (final  
364 concentration of 4%) into two containers ( $P_{add} = 1 \text{ mg P L}^{-1}$ ) overnight before an experiment.

365 P incorporation into the epilithon biomass was considered to be proportional to the  
366 <sup>33</sup>P disappearance from incubation water, as confirmed in an experiment with simultaneous  
367 spectrophotometric and scintillation detection of dissolved P (not shown). The P uptake rate  
368 constant,  $k_{upt}$  (h<sup>-1</sup>), which reflects the initial uptake rate, was calculated using the two-  
369 parameter exponential function:

$$370 A_t = A_0 \times e^{(-k_{upt} \times t)}, \quad (1)$$

371 where  $t$  (hours) is time and  $A_t$  and  $A_0$  are <sup>33</sup>P activities (c.p.m.) at the time  $t$  and zero,  
372 respectively. The velocity of P uptake by epilithon,  $v_{upt}$  (mg P g OM<sup>-1</sup> h<sup>-1</sup>) was calculated as:

$$373 v_{upt} = \frac{k_{upt} \times P_{add} \times V_{inc}}{OM}, \quad (2)$$

374 where  $V_{inc}$  (L) is incubation volume, and  $OM$  (g) is organic matter content in the epilithon  
375 sample. To study concentration dependence of the P uptake velocity, it was plotted as a  
376 function of  $P_{add}$  and fitted either to a Michaelis-Menten model:

$$377 v_{upt} = \frac{V_{max} \times P_{add}}{K_S + P_{add}}, \quad (3)$$

378 or, if the P uptake velocity did not show saturation, to a simplified linear model:

379  $v_{upt} = v_{slope} \times P_{add},$  (4)

380 where  $V_{max}$  (the maximum P uptake velocity; mg P g OM<sup>-1</sup> h<sup>-1</sup>) and  $K_S$  (half-saturation  
381 constant; mg P L<sup>-1</sup>) are the parameters of the Michaelis-Menten model (Eq. 3); the linear  
382 model (Eq. 4) had only one parameter,  $v_{slope}$ , (L g OM<sup>-1</sup> h<sup>-1</sup>). The more complex model (Eq. 3)  
383 was selected in preference to the simpler one (Eq. 4) when improving the fit at the  
384 probability level of p<0.05 (F test). Parameter estimations and model selection were  
385 performed in PRISM v.7 (GraphPad Software). For saturation data (Eqs. 3, 4), relative  
386 weighting by  $1/(v_{upt})^2$  was applied as the error of the  $v_{upt}$  was heteroscedastic, accounting for  
387 around 30% (average difference between duplicates), irrespectively of  $P_{add}$ . No weighting  
388 was applied to the time-course data. Outliers were excluded with the PRISM v.7 built-in  
389 algorithm using the recommended settings<sup>49</sup>.

390 Epilithon specific P uptake affinity,  $SPUA_E$  (L g OM<sup>-1</sup> h<sup>-1</sup>) for the Michaelis-Menten  
391 model (Eq. 3) was calculated as:

392  $SPUA_E = V_{max}/K_S,$  (5)

393 while for the linear model (Eq. 4) it was directly equal to  $v_{slope}$ . To estimate *in-situ* epilithon P  
394 uptake velocity ( $EPUV$ , µg P g OM<sup>-1</sup> h<sup>-1</sup>),  $SPUA_E$  was multiplied with  $SRP_{corr}$  (corrected soluble  
395 reactive phosphorus concentration; µg P L<sup>-1</sup>):

396  $EPUV = SPUA_E \times SRP_{corr},$  (6)

397 where  $SRP_{corr}$  is:

398  $SRP_{corr} = \min (SRP, K_{Sest}).$  (7)

399 As Eq. 7 suggests,  $SRP_{corr}$  was defined as directly measured  $SRP$  when  $SRP < K_{Sest}$ . Vice  
400 versa, where  $SRP > K_{Sest}$ ,  $SRP_{corr}$  was defined as  $K_{Sest}$  (see “Seston P uptake kinetics”).  
401 Further, the theoretical time necessary for doubling of the actual epilithon P content – i.e.

402 epilithon phosphorus doubling time,  $EPDT$  (h) – was calculated applying the estimated P  
403 uptake velocity:

404 
$$EPDT = EPC/EPUV. \quad (8)$$

405 In eq. 8,  $EPC$  ( $\mu\text{g P g OM}^{-1}$ ) is epilithon phosphorus content.

406

407 Abiotic P adsorption

408 To distinguish the P uptake by cells from the abiotic P adsorption, two samples with 4%  
409 formaldehyde (final conc.) were pre-incubated overnight (>12 hours) in each experiment.

410 These samples received the lowest concentration of  $P_{add}$  used (i.e.  $1 \text{ mg L}^{-1}$ ) and the effect of  
411 formaldehyde was expressed relative to uninhibited uptake velocity. Additional experiments  
412 were performed to elucidate concentration dependency of the abiotic adsorption over a  
413 wide range of P concentrations ( $P_{add} = 0.01, 0.1, 1, \text{ and } 10 \text{ mg P L}^{-1}$ ), and reversibility of the  
414 abiotic P adsorption using the surface wash reagent<sup>44,50</sup> applied immediately after  
415 termination of  $^{33}\text{P}$  uptake experiments for 1 hour. The amount of  $^{33}\text{P}$  released by the surface  
416 wash reagent was expressed relative to the  $^{33}\text{P}$  removed from the water in the presence of  
417 4% formaldehyde.

418

419 Seston P uptake kinetics

420 Lake-water samples (30 mL) were supplemented simultaneously with 50–100 kBq of  
421 carrier-free  $^{33}\text{P}$ -labelled  $\text{H}_3\text{PO}_4$  and seven concentrations of unlabelled orthophosphate ( $P_{add}$ ,  
422 range  $0.1\text{--}60 \mu\text{g P L}^{-1}$ , added as  $\text{KH}_2\text{PO}_4$ ), and incubated at *in-situ* temperature for 2–3 hours.  
423 At appropriate time intervals (2–30 min), 0.5-mL subsamples were filtered through  
424 polycarbonate filters (Poretics, 0.2  $\mu\text{m}$  mean porosity). The filter-retained activity was  
425 measured with the scintillation counter. The P uptake rate constants,  $k_{upt}$ , were determined

426 from the time courses of  $^{33}\text{P}$  incorporation with the method of Currie and Kalff<sup>51</sup> applying  
427 the second-order polynomial equation:

428 
$$P_t/T = P_0 + k_{upt} \times t + a \times t^2, \quad (9)$$

429 where  $P_t$  is filter-retained  $^{33}\text{P}$  activity (c.p.m.) at time  $t$ ,  $T$  is total  $^{33}\text{P}$  activity added, and  $P_0$  is  
430 filter-retained  $^{33}\text{P}$  activity at zero time, to the initial part of data ( $P_t/T < 0.7$ )<sup>52</sup>. The P uptake  
431 velocity,  $v_{upt}$  ( $\mu\text{g P L}^{-1} \text{ h}^{-1}$ ) was calculated as:

432 
$$v_{upt} = k_{upt} \times P_{add}. \quad (10)$$

433 The maximum velocity of P uptake,  $V_{maxsest}$  ( $\mu\text{g P L}^{-1} \text{ h}^{-1}$ ), and half-saturation constant,  
434  $K_{ssest}$ , ( $\mu\text{g P L}^{-1}$ ) were determined by non-linear regression as described above (Eq. 3). To  
435 compare specific P uptake affinities of epilithon and seston, expressed in the same units, the  
436 seston specific P uptake affinity,  $SPUA_{sest}$  ( $\text{L g OM}^{-1} \text{ h}^{-1}$ ) was calculated as:

437 
$$SPUA_{sest} = \frac{V_{maxsest}}{K_{ssest} \times OM}, \quad (11)$$

438 where  $OM$  is seston organic matter content ( $\text{g L}^{-1}$ ) estimated from  $Chla$  concentration in lake  
439 water assuming  $OM:Chla$  (w/w) ratio of 200<sup>53</sup>. Turnover time of dissolved orthophosphate,  
440 TT-PO<sub>4</sub> (h), was determined as  $1/k_{upt}$  measured in the absence of added P and used for  
441 epilimnion P deficiency assessment<sup>54</sup>.

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568

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575

576 **Author contributions**

577 E.K. and J.N. designed and performed experiments, interpreted results, and wrote the  
578 manuscript. K.C., P.Z., M.R., E.K., J.N. and K.R. performed field work. P.Z., P.C., K.C., K.R. and  
579 M.P. reviewed the early version of manuscript. P.C. analysed the data and made statistics.  
580 K.C. designed map of lakes and took photo of epilithon. P.Z. took aerial photos of the lakes  
581 (presented in Supplement). All authors reviewed and approved the final version of  
582 manuscript.

583

584 **Additional Information**

585 **Competing Interests:** The authors declare that they have no competing interests.

586 **Supplementary Information** is available for this paper at xxxxx

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588

589 **Figures**

590 **Figure 1.** Maps of the investigated post-mining lakes and location of the sampling sites  
591 (asterisks).

592

593 **Figure 2.** Biomass and elemental composition of epilithon in three post-mining lakes  
594 sampled in the Czech Republic in 2019. **(a)** epilithon dry weight and organic matter  
595 content expressed per overgrown area, **(b)** relationship between epilithon molar N:P and  
596 C:P molar ratios (error bars omitted for the sake of clarity), **(c)** epilithon C:N ratios.  
597 Columns show averages of 4 (2×2) values from two sampling sites and two depths ± SD.  
598 Different lowercase letters indicate significant seasonal differences within lakes ( $p < 0.05$ ,  
599 Two-way ANOVA with Tukey's multiple comparisons post-test). Shaded areas (b, c) show  
600 values indicating P and N limitation, respectively, given by Hillebrand and Sommer<sup>26</sup>.

601

602 **Figure 3.** Epilithon P uptake in three post-mining lakes sampled in the Czech Republic in  
603 2019. **(a)** Abiotic P uptake (adsorption). Columns show averages of 4 values from two  
604 sampling sites and two depths ± SD. **(b)** Maximum P uptake capacity,  $V_{max}$ . **(c)** Half-  
605 saturation constant,  $K_s$ . Symbols in two lower panels show individual values. Different  
606 lowercase letters indicate significant seasonal differences within lakes ( $p < 0.05$ , Two-way  
607 ANOVA and Tukey's multiple comparisons post-test).

608

609 **Figure 4.** P uptake parameters of epilithon in three post-mining lakes sampled in the Czech  
610 Republic in 2019. **(a)** Epilithon specific phosphorus uptake affinity ( $SPUA_E$ ), **(b)** calculated  
611 *in-situ* P uptake by epilithon. Symbols show individual values. Different lowercase letters  
612 indicate significant seasonal differences within lakes ( $p < 0.05$ , Two-way ANOVA and  
613 Tukey's multiple comparisons post-test).

614

615 **Figure 5.** Calculated epilithon internal P dynamics based on measured epilithon P uptake  
616 properties in three post-mining lakes sampled in the Czech Republic in 2019. **(a)** Epilithon  
617 phosphorus doubling time. Symbols show individual values; different lowercase letters  
618 indicate significant seasonal differences within lakes ( $p < 0.05$ ; Two-way ANOVA and  
619 Tukey's multiple comparisons post-test), **(b)** comparison of observed (blue columns) and

620 theoretical (red columns) changes in epilithon P content. Columns show averages of 4  
621 (2×2) values from two sampling sites and two depths ± SD.

622

623 **Tables**

624 **Table 1.** Chemical and limnological parameters of the three post-mining lakes in the Czech  
625 Republic sampled from the depth of 0.5 m at their deepest points in 2019. <sup>1</sup>Corrected SRP  
626 value - see Discussion for explanation, <sup>2</sup>mixed layer, <sup>3</sup>euphotic depth, the depth of 1% of  
627 the surface photosynthetically active radiation, <sup>4</sup>unstable stratification, <sup>5</sup>b.d.l. = below  
628 detection limit of 0.5 µg L<sup>-1</sup> for SRP.

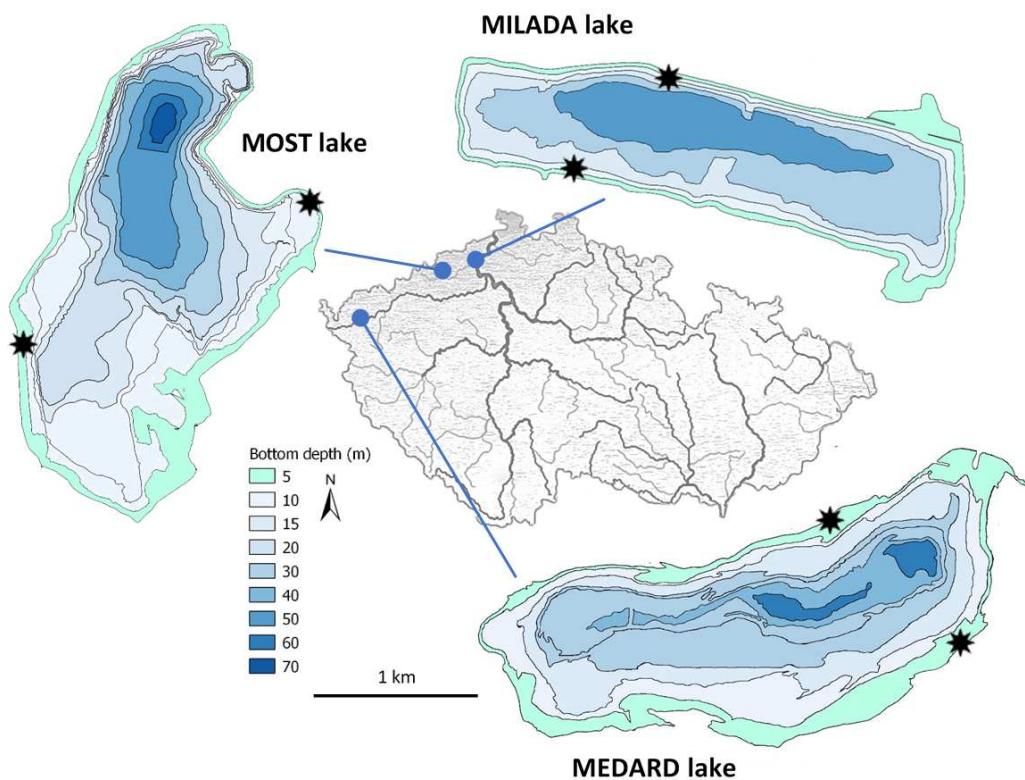
629

630 **Table 2.** Average values of epilithon biomass- and P uptake-related variables measured in  
631 three post-mining lakes in the Czech Republic in 2019. Differences were tested with two-  
632 way ANOVA with Lake and Sampling season as factors. <sup>1</sup>Different lowercase letters  
633 indicate significant difference between lakes or sampling seasons ( $p < 0.05$ ; Tukey's  
634 multiple comparisons post-test), p is the significance of the difference (for full ANOVA  
635 table see **Supplementary Table 3**). <sup>2</sup>Data with variability exceeding one order of  
636 magnitude were log-transformed.

637

638 **Table 3.** A summary of estimates of kinetic parameters of P uptake by epilithon in three  
639 post-mining lakes in the Czech Republic in 2019 (parameters of Michaelis-Menten kinetics  
640  $V_{max}$  and  $K_s$ , and specific P uptake affinity,  $SPUA_E$ ) with indices of the goodness of fit. <sup>1</sup>N, S  
641 denote the North and South sampling site, respectively (see **Fig. 1** and **Supplementary**  
642 **Table 6**), <sup>2</sup>the most parsimonious model selected with F-test, MM - Michaelis-Menten  
643 kinetics (Eq. 3), L - linear model (Eq. 4), <sup>3</sup>best-fit parameter ± standard error of the  
644 estimate.

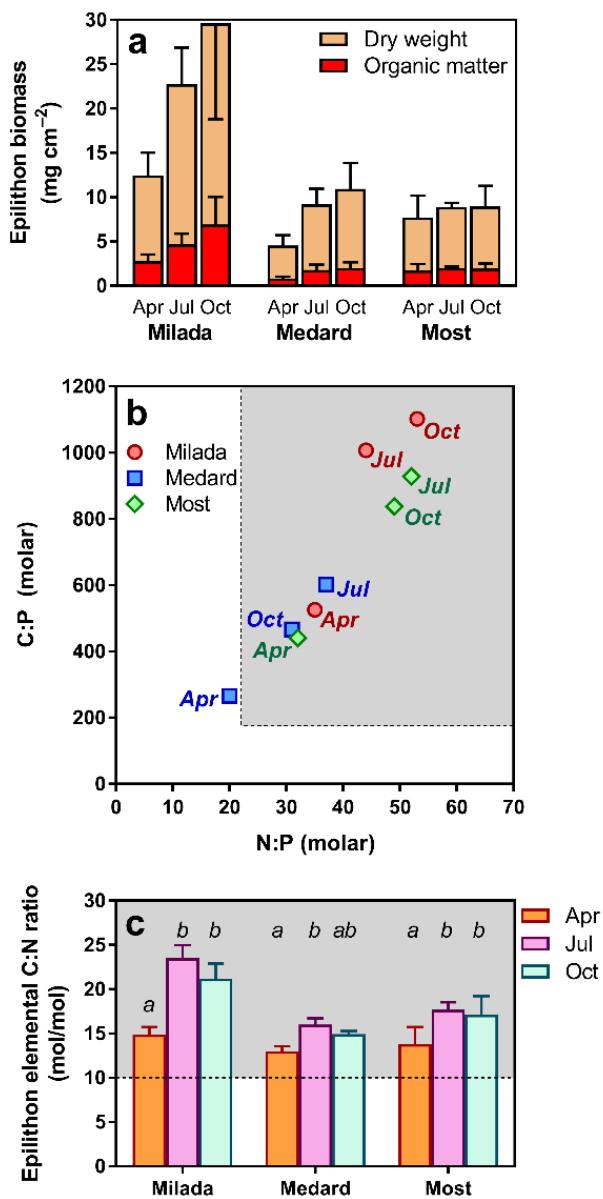
645 **Figure 1.**



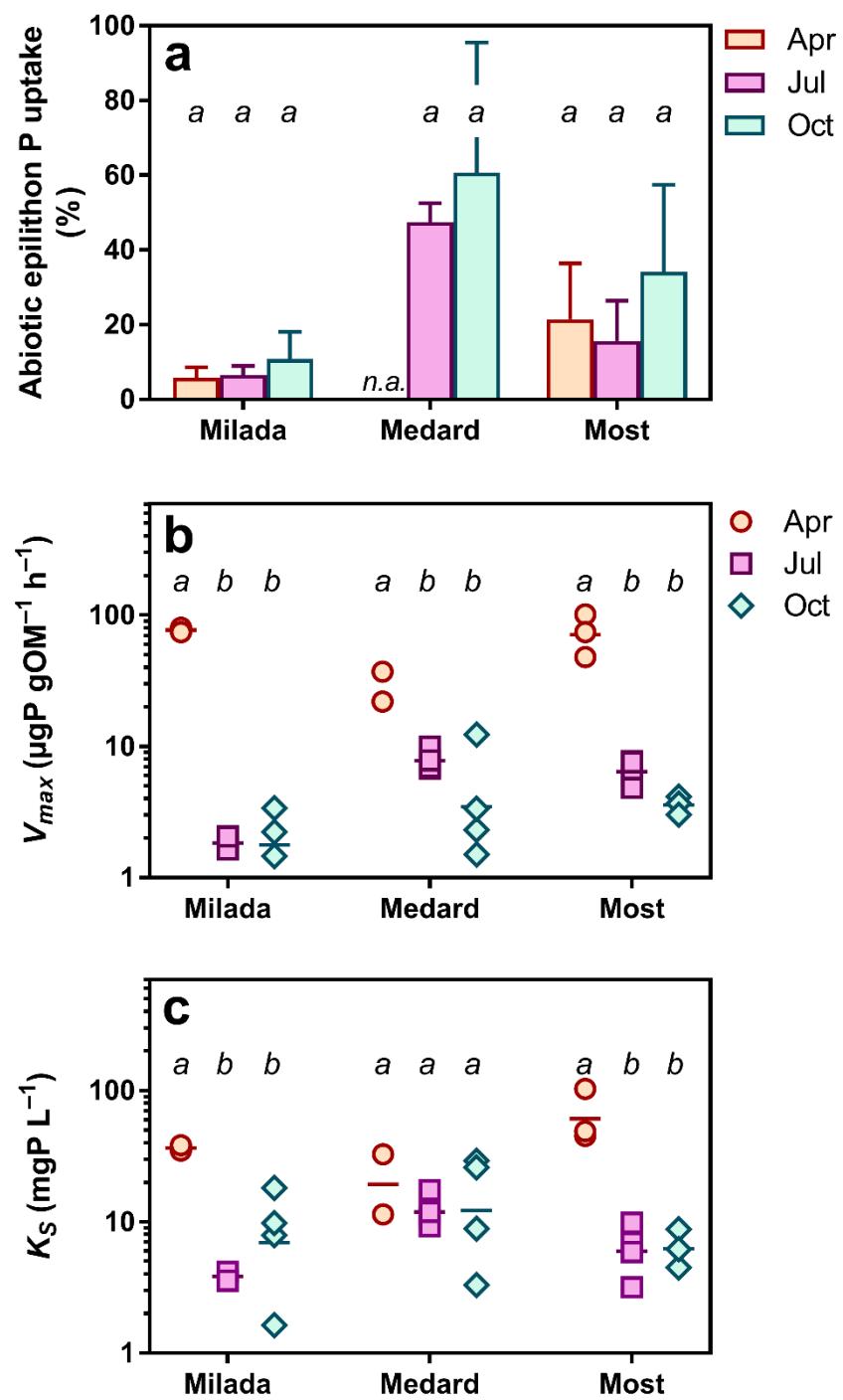
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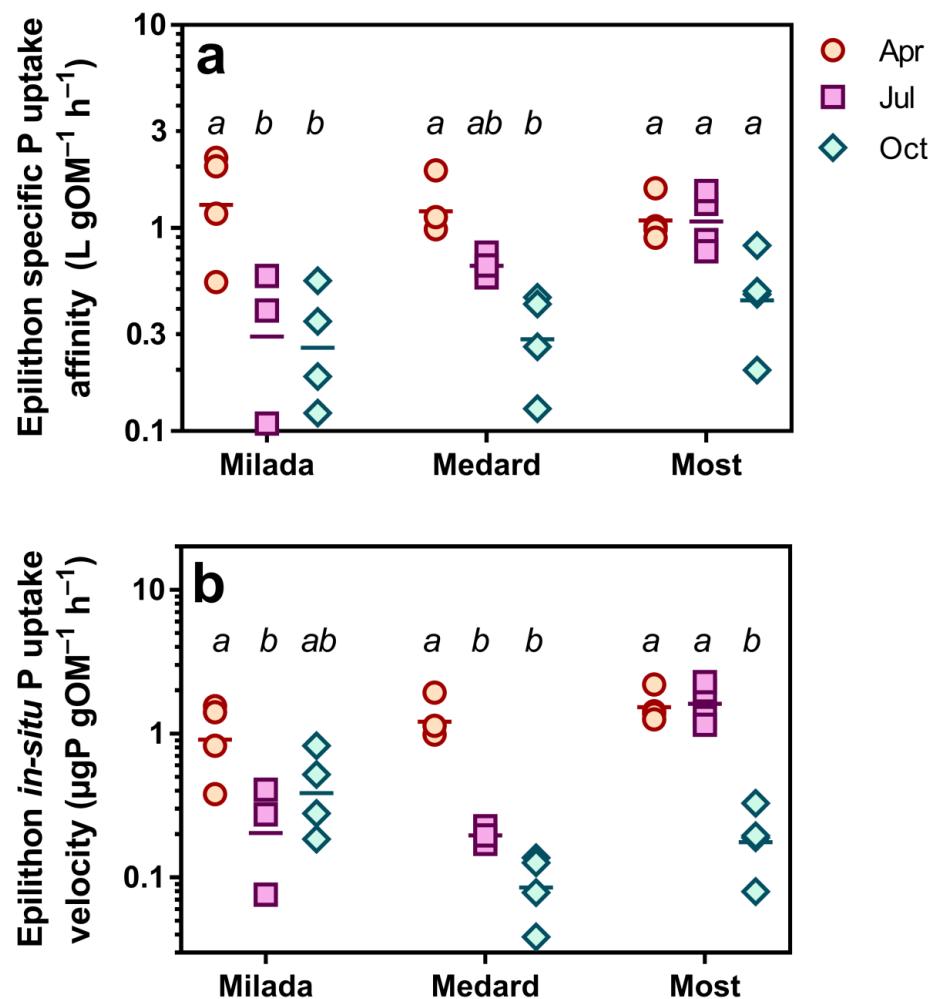
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648 **Figure 2.**

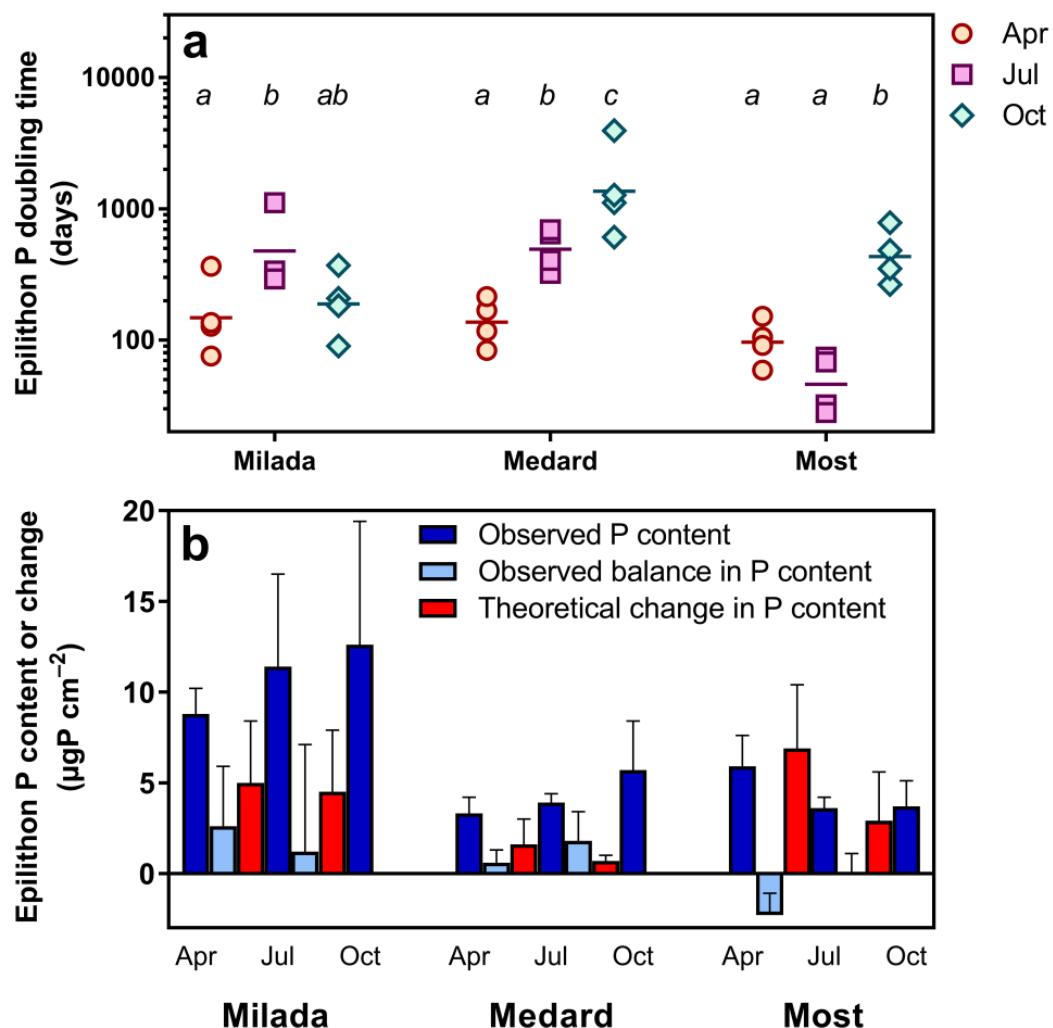


649

**Figure 3.**

**Figure 4.**

654 **Figure 5.**



655

656 **Tables**657 **Table 1.**

658

Lake	Month	T <sub>w</sub> (°C)	pH	Conductivity (µS cm <sup>-1</sup> )	TP (µg L <sup>-1</sup> )	SRP (µg L <sup>-1</sup> )	SRP <sub>corr</sub> <sup>1</sup> (µg L <sup>-1</sup> )	Si (mg L <sup>-1</sup> )	NO <sub>3</sub> (mg L <sup>-1</sup> )	DOC (mg L <sup>-1</sup> )	Chla (µg L <sup>-1</sup> )	Z <sub>mix</sub> <sup>2</sup> (m)	Z <sub>eu</sub> <sup>3</sup> (m)
<b>Milada</b>	April	7.7	9.1	1037	17.1	3.2	0.7	0.8	0.13	6.8	8.2	~8 <sup>4</sup>	9.9
	July	22.4	9.1	1030	10.2	1.6	0.7	0.9	0.02	8.1	2.1	6	11.7
	October	14.8	9.1	1035	13.9	1.5	1.5	0.7	0.02	8.9	4.3	10	15.0
<b>Medard</b>	April	7.5	7.7	1093	4.5	1.0	1.0	2.7	1.26	3.0	0.6	~4 <sup>4</sup>	9.4
	July	21.9	7.9	1080	4.0	b.d.l. <sup>5</sup>	0.3	2.9	1.12	3.1	1.0	7	17.6
	October	12.7	7.8	1087	4.0	b.d.l.	0.3	2.4	1.12	3.1	1.2	12	14.5
<b>Most</b>	April	11.7	8.6	540	7.9	1.6	1.4	1.2	0.92	4.6	1.0	5	16.7
	July	24.1	8.7	524	13.7	2.0	1.5	0.7	0.64	4.6	1.5	3	13.6
	October	17.0	8.8	533	14.2	2.1	0.4	0.5	0.56	5.0	3.1	10	14.6

659

660

**Table 2.**

661

<b>Epilithon variable</b>	<b>Lake averages<sup>1</sup></b>				<b>Sampling-season averages<sup>1</sup></b>			
	<b>Milada</b>	<b>Medard</b>	<b>Most</b>	<b>p</b>	<b>April</b>	<b>July</b>	<b>October</b>	<b>p</b>
Biomass (mg OM cm <sup>-2</sup> )	4.80 <sup>a</sup>	1.54 <sup>b</sup>	1.89 <sup>b</sup>	<0.0001	1.79 <sup>a</sup>	2.65 <sup>ab</sup>	3.63 <sup>b</sup>	0.0042
Biomass (mg DW cm <sup>-2</sup> )	16.7 <sup>a</sup>	6.7 <sup>b</sup>	6.6 <sup>b</sup>	<0.0001	6.4 <sup>a</sup>	10.1 <sup>ab</sup>	12.9 <sup>b</sup>	0.0036
OM/DW (%)	29 <sup>a</sup>	23 <sup>b</sup>	29 <sup>a</sup>	0.0001	27 <sup>a</sup>	26 <sup>a</sup>	28 <sup>a</sup>	0.5668
P content (mg P g OM <sup>-1</sup> )	2.49 <sup>a</sup>	3.14 <sup>a</sup>	2.42 <sup>a</sup>	0.058	3.64 <sup>a</sup>	2.21 <sup>b</sup>	2.18 <sup>b</sup>	0.0001
P content (µg P cm <sup>-2</sup> )	10.9 <sup>a</sup>	4.3 <sup>b</sup>	4.4 <sup>b</sup>	<0.0001	6.0 <sup>a</sup>	5.8 <sup>a</sup>	7.4 <sup>a</sup>	0.5314
C:P (molar ratio)	866 <sup>a</sup>	444 <sup>b</sup>	736 <sup>a</sup>	0.0001	411 <sup>a</sup>	831 <sup>b</sup>	802 <sup>b</sup>	<0.0001
C:N (molar ratio)	19.5 <sup>a</sup>	14.7 <sup>b</sup>	16.2 <sup>c</sup>	<0.0001	13.9 <sup>a</sup>	18.7 <sup>b</sup>	17.8 <sup>b</sup>	<0.0001
N:P (molar ratio)	43.8 <sup>a</sup>	29.4 <sup>b</sup>	44.4 <sup>a</sup>	0.0086	29.2 <sup>a</sup>	44.4 <sup>b</sup>	44.1 <sup>b</sup>	0.0075
<i>V</i> <sub>max</sub> (mg P g OM <sup>-1</sup> h <sup>-1</sup> ) <sup>2</sup>	20.7 <sup>a</sup>	11.0 <sup>ab</sup>	26.0 <sup>b</sup>	0.0446	62.3 <sup>a</sup>	6.1 <sup>b</sup>	3.5 <sup>c</sup>	<0.0001
<i>K</i> <sub>S</sub> (mg P L <sup>-1</sup> ) <sup>2</sup>	14.8 <sup>a</sup>	16.0 <sup>a</sup>	24.1 <sup>a</sup>	0.5495	44.8 <sup>a</sup>	8.3 <sup>b</sup>	11.3 <sup>b</sup>	0.0002
Specific P uptake affinity (L g OM <sup>-1</sup> h <sup>-1</sup> ) <sup>2</sup>	0.75 <sup>a</sup>	0.74 <sup>a</sup>	0.91 <sup>a</sup>	0.0534	1.29 <sup>a</sup>	0.74 <sup>b</sup>	0.37 <sup>c</sup>	<0.0001
<i>In-situ</i> P uptake (µg P g OM <sup>-1</sup> h <sup>-1</sup> ) <sup>2</sup>	0.61 <sup>ab</sup>	0.52 <sup>a</sup>	1.15 <sup>b</sup>	0.0002	1.29 <sup>a</sup>	0.75 <sup>b</sup>	0.25 <sup>c</sup>	<0.0001
<i>In-situ</i> P uptake (µg P cm <sup>-2</sup> d <sup>-1</sup> ) <sup>2</sup>	0.058 <sup>a</sup>	0.013 <sup>b</sup>	0.054 <sup>a</sup>	0.002	0.054 <sup>a</sup>	0.041 <sup>a</sup>	0.029 <sup>a</sup>	0.1218
Doubling time of P in biomass (days) <sup>2</sup>	299 <sup>a</sup>	799 <sup>b</sup>	207 <sup>c</sup>	<0.0001	141 <sup>a</sup>	364 <sup>a</sup>	806 <sup>b</sup>	<0.0001

662

663

**Table 3.**

664

Date	Lake	Site <sup>1</sup>	Depth (m)	Best model <sup>2</sup>	F-test P	R <sup>2</sup> <sub>adj</sub>	V <sub>max</sub> (mg P g OM <sup>-1</sup> h <sup>-1</sup> )	K <sub>s</sub> (mg P L <sup>-1</sup> )	SPUA <sub>E</sub> (L g OM <sup>-1</sup> h <sup>-1</sup> )
15 Apr 2019	Milada	N	0.5	L	0.1124	0.6572	—	—	1.18 ± 0.16 <sup>3</sup>
15 Apr 2019	Milada	N	1.5	MM	0.0132	0.7805	77.4 ± 30.4 <sup>3</sup>	34.9 ± 19.1 <sup>3</sup>	2.22 ± 0.52
15 Apr 2019	Milada	S	0.5	L	>0.9999	0.7237	—	—	0.543 ± 0.073
15 Apr 2019	Milada	S	1.5	MM	0.0125	0.8451	76.8 ± 25.4	38.2 ± 17.4	2.01 ± 0.38
22 Jul 2019	Milada	N	0.5	MM	0.0006	0.7778	1.64 ± 0.31	4.17 ± 1.79	0.393 ± 0.117
22 Jul 2019	Milada	N	1.5	L	0.1096	-0.2178	—	—	0.109 ± 0.026
22 Jul 2019	Milada	S	0.5	MM	0.001	0.5423	2.07 ± 0.51	3.55 ± 1.88	0.582 ± 0.209
14 Oct 2019	Milada	N	0.5	MM	0.0078	0.5802	1.46 ± 0.6	7.88 ± 5.54	0.186 ± 0.074
14 Oct 2019	Milada	N	1.5	MM	0.004	0.7236	3.39 ± 0.92	9.8 ± 4.55	0.347 ± 0.094
14 Oct 2019	Milada	S	0.5	MM	0.0003	0.9506	2.23 ± 0.35	18.13 ± 4.25	0.123 ± 0.014
14 Oct 2019	Milada	S	1.5	MM	0.0001	0.7764	0.9 ± 0.11	1.63 ± 0.63	0.55 ± 0.164
08 Apr 2019	Medard	N	0.5	L	>0.9999	0.7264	—	—	0.987 ± 0.147
08 Apr 2019	Medard	N	1.5	MM	0.0018	0.9254	37 ± 8.4	32.6 ± 9.6	1.13 ± 0.12
08 Apr 2019	Medard	S	0.5	MM	0.0003	0.787	21.9 ± 4.6	11.4 ± 4.1	1.93 ± 0.41
08 Apr 2019	Medard	S	1.5	L	no MM fit	0.5267	—	—	0.99 ± 0.164
08 Jul 2019	Medard	N	0.5	MM	0.0052	0.8073	6.74 ± 1.56	10.58 ± 4.44	0.637 ± 0.162
08 Jul 2019	Medard	N	1.5	MM	0.0001	0.9759	9.95 ± 1.09	17.38 ± 3.06	0.572 ± 0.051
08 Jul 2019	Medard	S	0.5	MM	<0.0001	0.944	6.98 ± 0.68	9.23 ± 1.56	0.757 ± 0.076

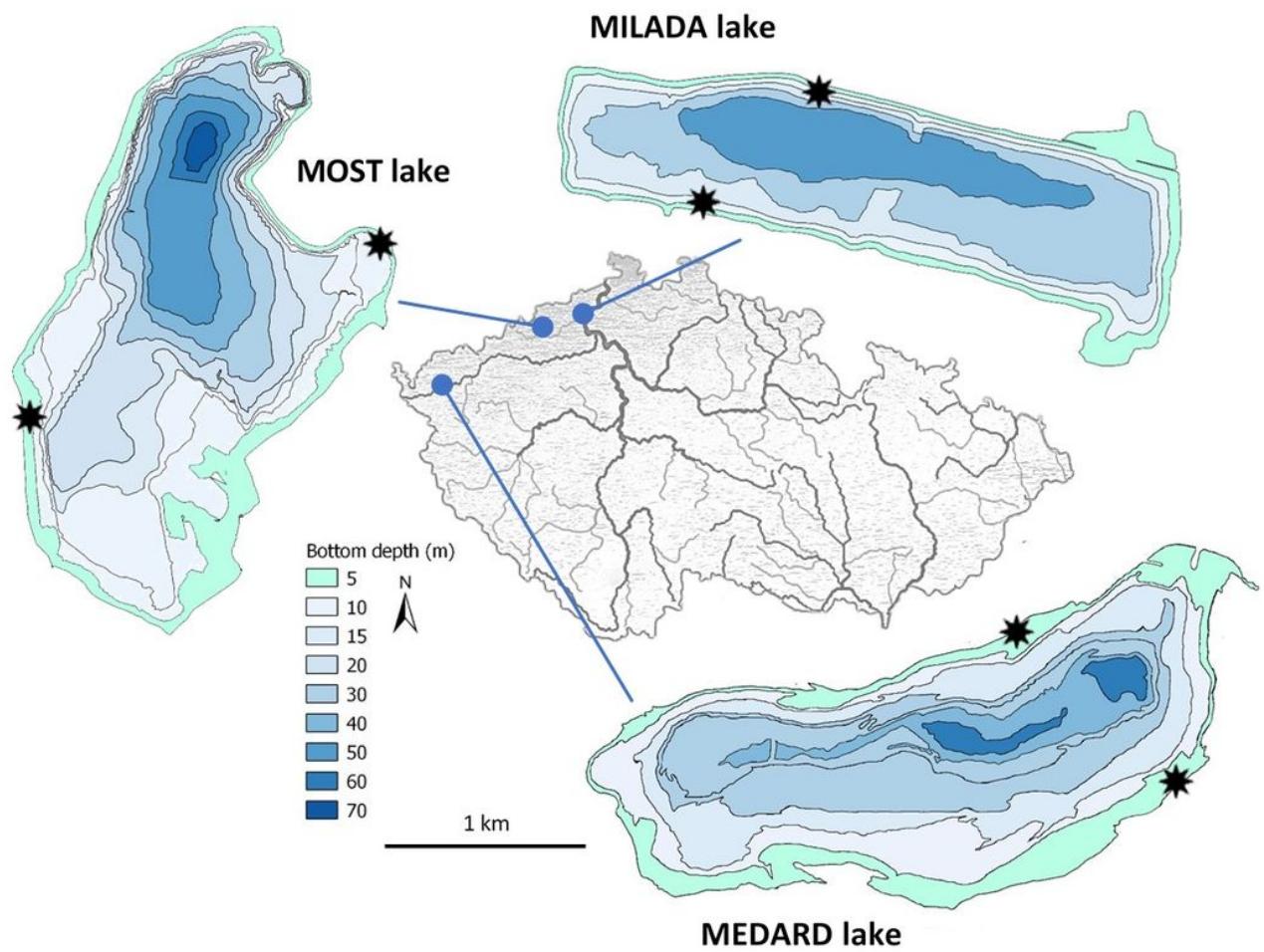
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**Table 3.** (cont.)

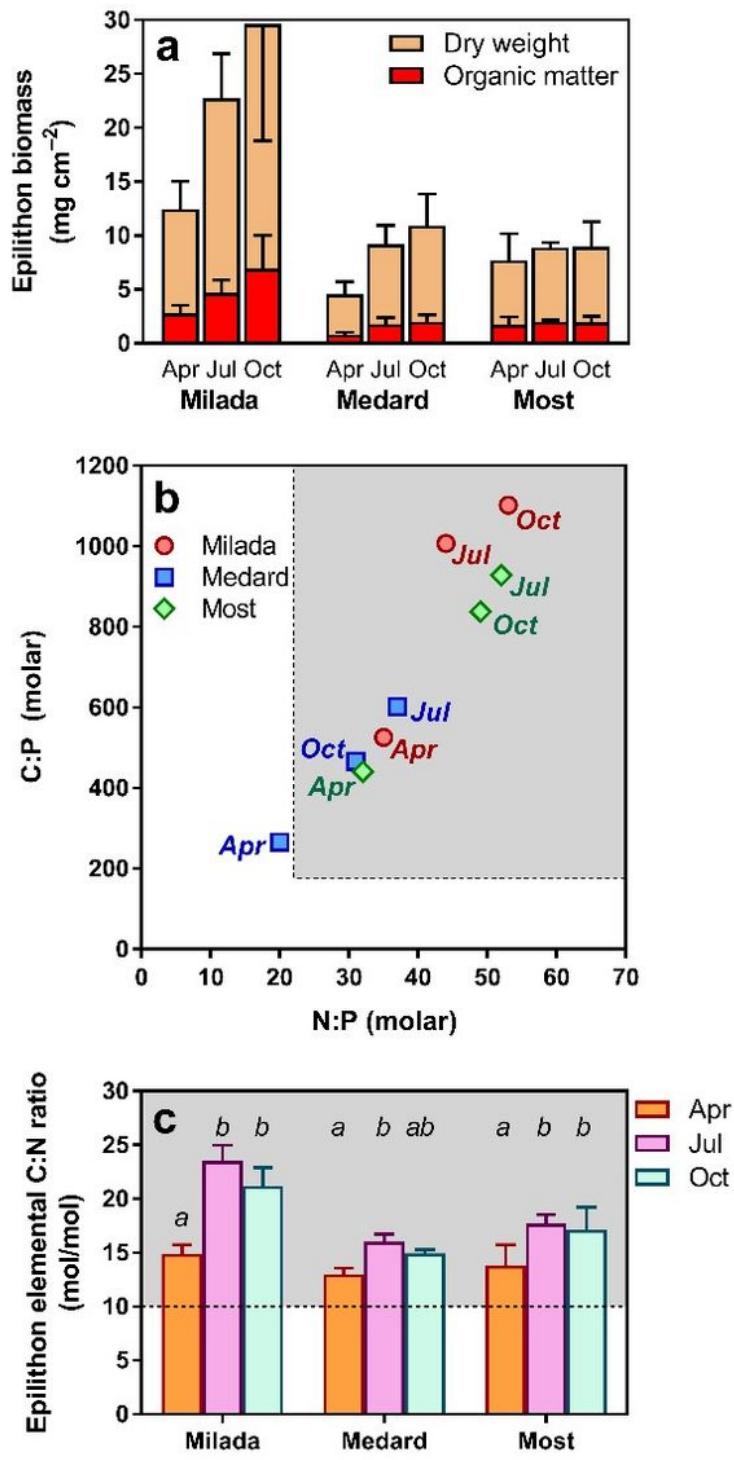
Date	Lake	Site <sup>1</sup>	Depth (m)	Best model <sup>2</sup>	F-test p	R <sup>2</sup> <sub>adj</sub>	V <sub>max</sub> (mg P g OM <sup>-1</sup> h <sup>-1</sup> )	K <sub>s</sub> (mg P L <sup>-1</sup> )	SPUA <sub>E</sub> (L g OM <sup>-1</sup> h <sup>-1</sup> )
21 Oct 2019	Medard	N	0.5	MM	0.0099	0.4554	2.32 ± 0.95	8.87 ± 6.6	0.261 ± 0.122
21 Oct 2019	Medard	N	1.5	MM	0.016	0.4669	1.51 ± 0.55	3.3 ± 2.62	0.456 ± 0.249
21 Oct 2019	Medard	S	0.5	MM	0.0145	0.7797	12.3 ± 4.4	29.1 ± 15.1	0.422 ± 0.099
21 Oct 2019	Medard	S	1.5	MM	0.042	0.6734	3.35 ± 1.57	25.9 ± 17.5	0.129 ± 0.04
29 Apr 2019	Most	N	0.5	MM	0.0217	0.8198	47.7 ± 19.1	46.7 ± 25	1.02 ± 0.21
29 Apr 2019	Most	N	1.5	MM	0.0116	0.9177	101 ± 33	103 ± 41	0.987 ± 0.11
29 Apr 2019	Most	S	0.5	L	0.1388	0.9414	–	–	0.897 ± 0.063
29 Apr 2019	Most	S	1.5	MM	0.0024	0.9074	73.9 ± 18.7	47 ± 15.4	1.57 ± 0.19
29 Jul 2019	Most	N	0.5	MM	0.0017	0.6322	6.04 ± 1.69	6.92 ± 3.59	0.872 ± 0.285
29 Jul 2019	Most	N	1.5	MM	0.0002	0.6232	7.73 ± 1.40	5.87 ± 2.73	1.32 ± 0.44
29 Jul 2019	Most	S	0.5	MM	<0.0001	0.814	4.83 ± 0.70	3.17 ± 1.07	1.52 ± 0.37
29 Jul 2019	Most	S	1.5	MM	<0.0001	0.9743	7.55 ± 0.60	9.81 ± 1.53	0.77 ± 0.08
30 Sep 2019	Most	N	0.5	MM	0.0016	0.7205	4.14 ± 1.09	8.76 ± 4.04	0.473 ± 0.131
30 Sep 2019	Most	N	1.5	MM	0.0007	0.7974	3.68 ± 0.70	4.49 ± 1.7	0.821 ± 0.201
30 Sep 2019	Most	S	0.5	MM	0.0124	0.5693	3.01 ± 1.00	6.17 ± 4.1	0.488 ± 0.207
30 Sep 2019	Most	S	1.5	L	0.1015	-0.3157	–	–	0.2 ± 0.046

# Figures



**Figure 1**

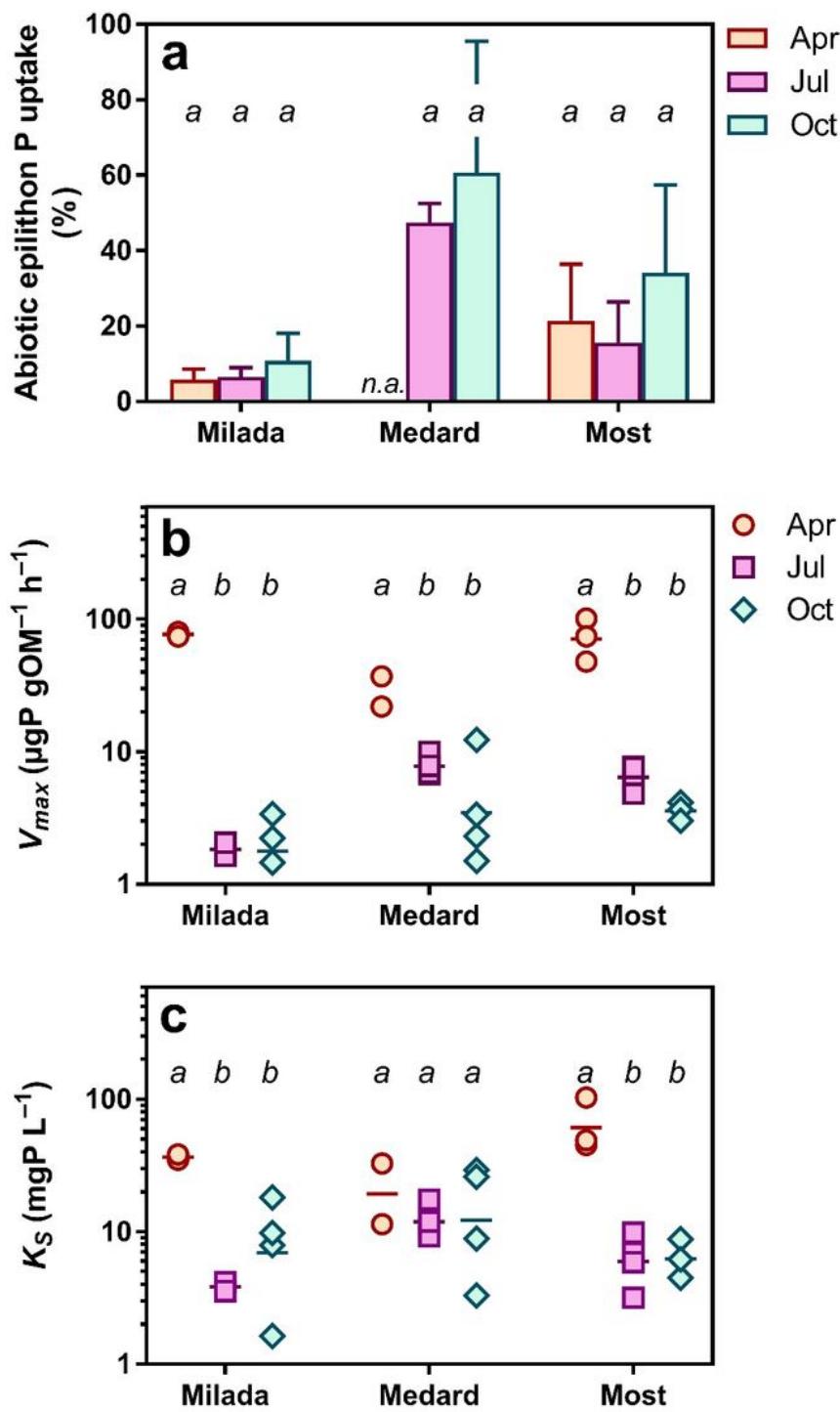
Maps of the investigated post-mining lakes and location of the sampling sites (asterisks).



**Figure 2**

Biomass and elemental composition of epilithon in three post-mining lakes sampled in the Czech Republic in 2019. (a) epilithon dry weight and organic matter content expressed per overgrown area, (b) relationship between epilithon molar N:P and C:P molar ratios (error bars omitted for the sake of clarity), (c) epilithon C:N ratios. Columns show averages of 4 ( $2 \times 2$ ) values from two sampling sites and two depths  $\pm$  SD. Different lowercase letters indicate significant seasonal differences within lakes ( $p < 0.05$ ,

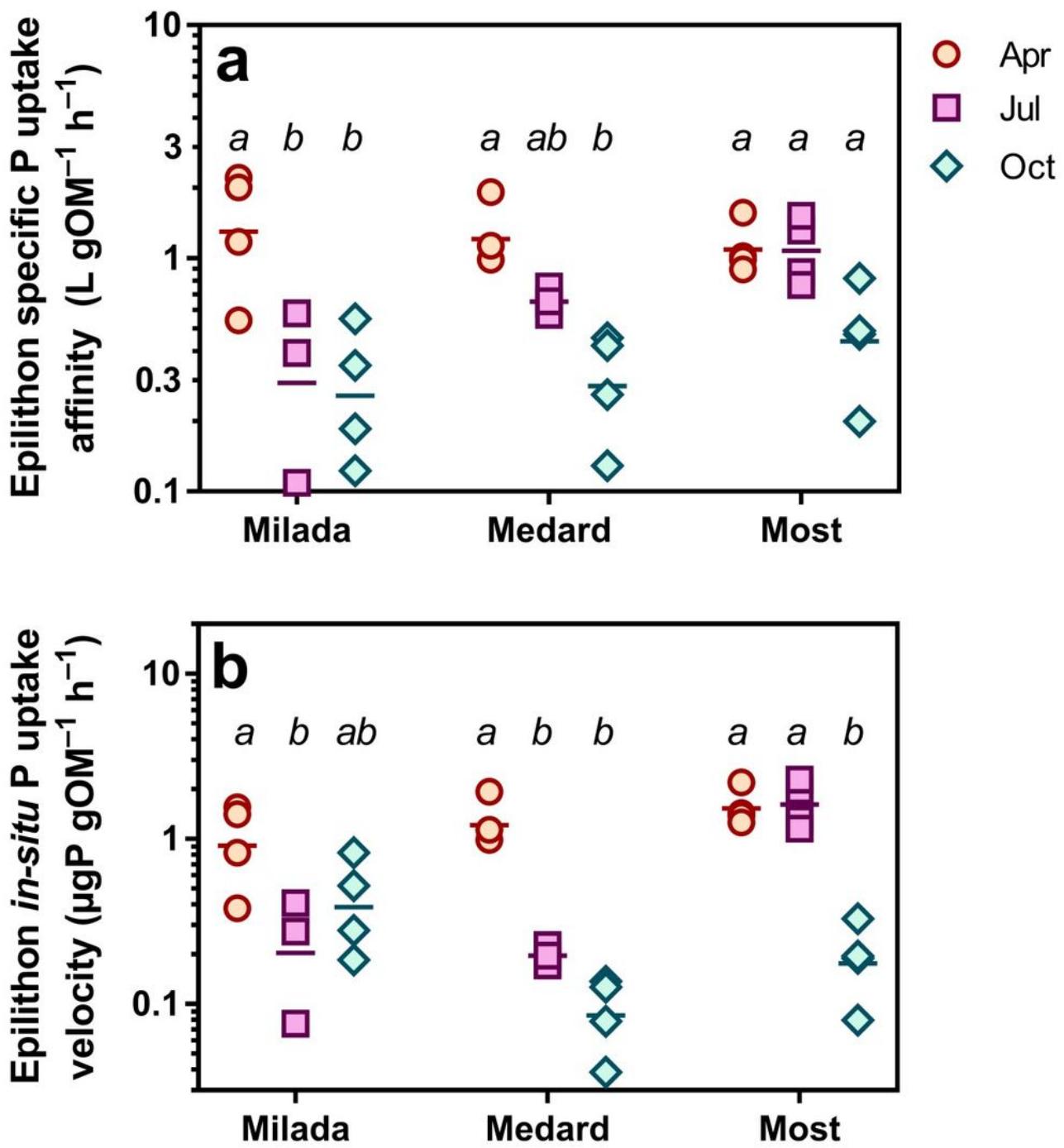
Two-way ANOVA with Tukey's multiple comparisons post-test). Shaded areas (b, c) show values indicating P and N limitation, respectively, given by Hillebrand and Sommer26.



**Figure 3**

Epilithon P uptake in three post-mining lakes sampled in the Czech Republic in 2019. (a) Abiotic P uptake (adsorption). Columns show averages of 4 values from two sampling sites and two depths  $\pm$  SD. (b) Maximum P uptake capacity,  $V_{max}$ . (c) Half-saturation constant,  $K_s$ . Symbols in two lower panels show

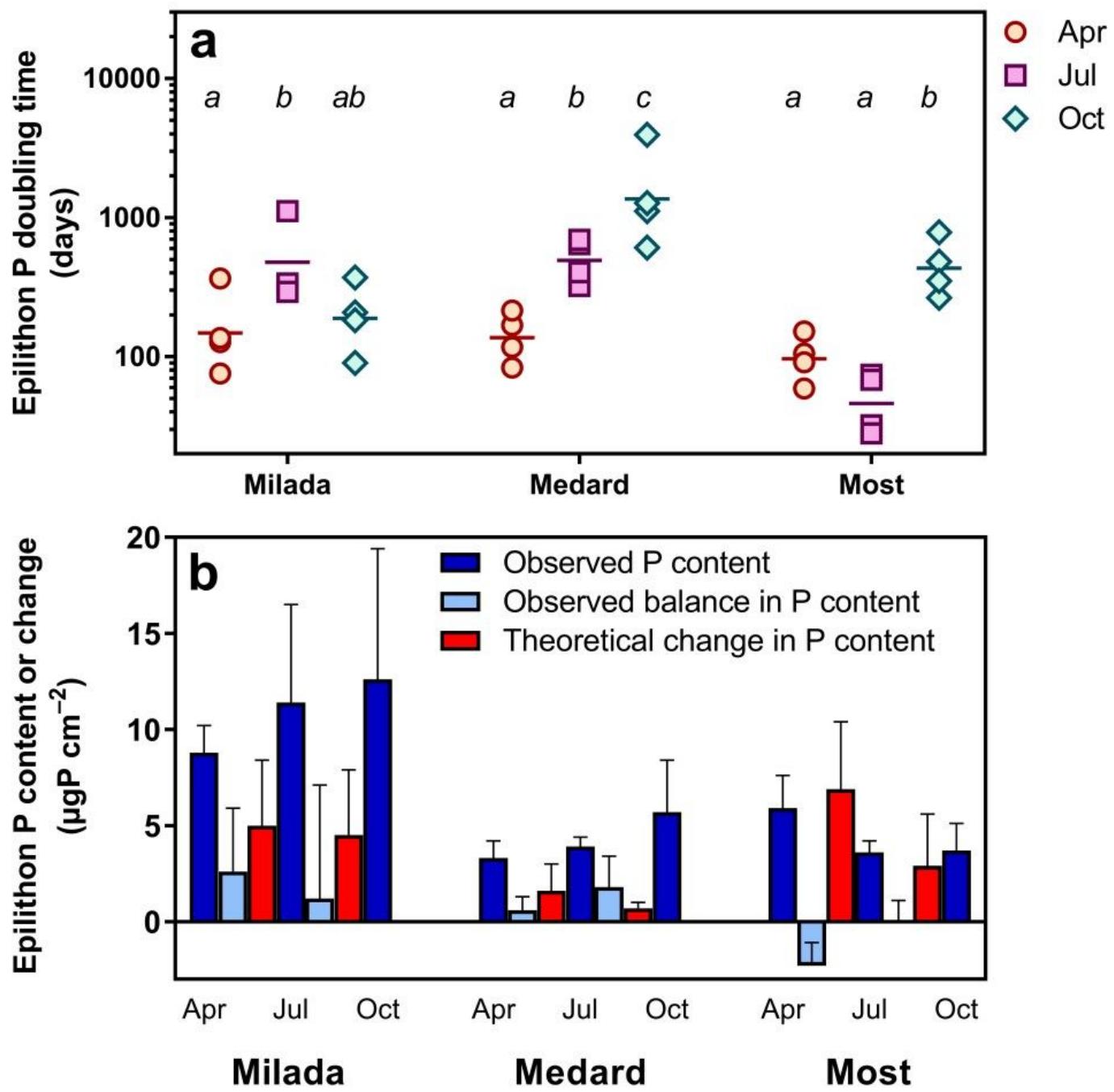
individual values. Different lowercase letters indicate significant seasonal differences within lakes ( $p < 0.05$ , Two-way ANOVA and Tukey's multiple comparisons post-test).



**Figure 4**

uptake parameters of epilithon in three post-mining lakes sampled in the Czech Republic in 2019. (a) Epilithon specific phosphorus uptake affinity (SPUAE), (b) calculated in-situ P uptake by epilithon.

Symbols show individual values. Different lowercase letters indicate significant seasonal differences within lakes ( $p < 0.05$ , Two-way ANOVA and Tukey's multiple comparisons post-test).



**Figure 5**

Calculated epilithon internal P dynamics based on measured epilithon P uptake properties in three post-mining lakes sampled in the Czech Republic in 2019. (a) Epilithon phosphorus doubling time. Symbols show individual values; different lowercase letters indicate significant seasonal differences within lakes ( $p < 0.05$ ; Two-way ANOVA and Tukey's multiple comparisons post-test), (b) comparison of observed

(blue columns) and theoretical (red columns) changes in epilithon P content. Columns show averages of 4 (2×2) values from two sampling sites and two depths ± SD.

## Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [EpilithonPuptakeKonopacovaSupplementaryTablesandFigures.pdf](#)