

Identification of Potential Biomarkers for Abdominal Pain in IBS Patients by Bioinformatics Approach

Zhongyuan Lin

Sun Yat-sen University First Affiliated Hospital

Yimin Wang

Guangdong Second Provincial General Hospital

Shiqing Lin

Sun Yat-sen University First Affiliated Hospital

Decheng Liu

Sun Yat-sen University First Affiliated Hospital

Guohui Mo

Sun Yat-sen University First Affiliated Hospital

Hui Zhang

Guangdong Second Provincial General Hospital

Yunling dou (✉ sysdyl@163.com)

Sun Yat-sen University First Affiliated Hospital

Research article

Keywords: Bioinformatics analysis, Biomarker, Visceral hypersensitivity, Abdominal pain, Irritable bowel syndrome

Posted Date: November 16th, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-33139/v2>

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Version of Record: A version of this preprint was published on February 2nd, 2021. See the published version at <https://doi.org/10.1186/s12876-021-01626-7>.

Abstract

Background: Irritable bowel syndrome (IBS) is the most common functional gastrointestinal disease characterized by chronic abdominal discomfort and pain. The mechanisms of abdominal pain, as a relevant symptom, in IBS are still unclear. We aimed to explore the key genes and neurobiological changes specially involved in abdominal pain in IBS.

Methods: Gene expression data (GSE36701) was downloaded from Gene Expression Omnibus (GEO) database. Fifty-three rectal mucosa samples from Irritable Bowel Syndrome with diarrhea (IBS-D) patients and 40 samples from healthy volunteers (HV) as controls were included. Differentially expressed genes (DEGs) between two groups were identified using the GEO2R online tool. Functional enrichment analysis of DEGs was performed on the DAVID database. Then a protein-protein interaction (PPI) network was constructed and visualized using STRING database and Cytoscape.

Results: The microarray analysis demonstrated a subset of genes (*CCKBR*, *CCL13*

ACPP, *BDKRB2*, *GRPR*, *SLC1A2*, *NPFF*, *P2RX4*, *TRPA1*, *CCKBR*, *TLX2*, *MRGPRX3*, *PAX2*, *CXCR1*) specially involved in pain transmission. Among these genes, we identified *GRPR*, *NPFF* and *TRPA1* genes as potential biomarkers for irritating abdominal pain of IBS patients.

Conclusions: Overexpression of certain pain-related genes (*GRPR*, *NPFF* and *TRPA1*) may contribute to chronic visceral hypersensitivity, therefore be partly responsible for recurrent abdominal pain or discomfort in IBS patients. Several synapses modification and biological process of psychological distress may be risk factors of IBS.

Background

Chronic abdominal pain is an important symptom of irritable bowel syndrome (IBS), which affects 3.8%-9.2% of the global population[1]. Recurrent abdominal pain can substantially reduce quality of life in IBS patients and there is no effective and standard treatment currently[2]. In order to identify new therapies, it is important to understand the underlying mechanism of chronic abdominal pain.

Abdominal pain in IBS is thought to be secondary to visceral hypersensitivity (VH), which has been described as low thresholds of stimuli perception arising from the gut[3]. Visceral hypersensitivity is characterized by two components: allodynia, a painful response to stimuli that are normally not painful and hyperalgesia, an enhanced response to a painful stimulus[4]. The underlying pathogenesis that lead to VH in patients who have IBS is not fully understood. However, the prevailing viewpoint in the pathogenesis involves psychosocial factors, subtle inflammation and alteration in the neuronal excitability of gut sensory pathways, which is responsible for transmitting nociceptive information from the periphery to the central nervous system[5]. Specific changes of a variety of receptors and ion channels both in the expression and function have been documented in visceral pain conditions relevant to irritable bowel syndrome. Researches have shown genetic factors may play a role in those specific

changes in IBS[6]. Beyder and colleagues have reported that a mutation of the SCN5A-encoded voltage gated sodium channel, type V (alpha subunit) was associated with abdominal pain[7]. Other investigators have investigated genetic changes related to enteric infection, epithelial barrier function and immune regulation, serotonin signaling, cannabinoid receptors, and bile acid synthesis, with varying results[8].

Bioinformatics analysis provides tools to produce overviews of genetic networks and potential biological pathways based on large-scale information. This study aimed to explore the molecular mechanisms specially involved in pain transmission underlying IBS and to identify hub genes and potential pathways associated with the pathogenesis of IBS.

Methods

Microarray Data Search and Selection of Eligible Data Set

The microarray data of GSE36701 was downloaded from the GEO database (www.ncbi.nlm.nih.gov/geo/) with its microarray platform as GPL570 (Affymetrix Human Genome U133 Plus 2.0 Array). In this microarray, 53 samples of rectal mucosa from IBS-D patient with obvious abdominal pain were obtained and 40 samples from healthy volunteer as controls. Subjects completed a Talley IBS symptom questionnaire modified to include days with pain. IBS was diagnosed according to the Rome II criteria. Those with IBS also completed IBS quality of life questionnaire and the IBS Symptom Severity Score Questionnaire including severity of visceral pain. Those agreeing to take part underwent sigmoidoscopy without bowel preparation. Biopsies were immersed in RNALater to inactivate degrading enzymes and further extracted for mRNA. Details of clinical data collection, RNA extraction and microarray processing can be found in GSE36701 dataset citation (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE36701>).

DEGs' Screening

The expression profiles were expressed as FC (fold change) to produce a normally distributed variable. We screened out DEGs using the online tool GEO2R/R, which was based on limma (Linear Models for Microarray Analysis) R package 3.26.8 (<https://www.ncbi.nlm.nih.gov/geo/info/geo2r.html>). We applied Benjamini & Hochberg false discovery rate method for adjustment of the P-values. In our study, DEGs between healthy volunteers and IBS-D patients were screened and selected by the cut-off point of FC ≥ 1.2 and P-value < 0.05 .

Functional Enrichment Analysis

The selected DEGs were submitted to the Database for Annotation, Visualization, and Integrated Discovery (DAVID) for further analysis. The DAVID database, version 6.8 Beta (<https://david-d.ncifcrf.gov/>), offers biological function annotation for researchers[9]. In this study, Gene Ontology (GO) analysis and The Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways of DEGs were obtained from the DAVID database. The biological processes and pathways might contribute to verify the

significant DEGs and to IBS treatments. Functional annotation analysis was performed by two-sided hypergeometric test and Benjamini & Hochberg methods was applied for multiple testing correction. Min.Count ≥ 2 and P -value < 0.05 was chosen as the threshold.

Protein-Protein Interaction (PPI) Analysis

Then we constructed PPI network of DEGs from Search Tool for the Retrieval of Interacting Genes (STRING, <http://string-db.org/>)[10]. STRING is a public database of known and predicted protein-protein interactions. In this study, we selected confidence score > 0.4 as a threshold to construct the PPI network. The combined score is calculated by combining the probabilities from the different evidence channels, including high-throughput experimental data and literature, and corrected for the probability of randomly observing an association[11]. Then, the list of PPI pairs was downloaded for further analysis. Module clustering analysis for the network was then performed to identify the potential functional modules in the network, using the Molecular Complex Detection (MCODE 1.6.1) in Cytoscape 3.7.1 plugin[12, 13]. The degree cut-off value to 2 and the node score cut-off to 0.2 were set as criterions in the MCODE process.

Results

Screening and prioritization for DEGs

An available Expression profiling data set (GSE36701) in NCBI was selected in this study to identify differentially expressed genes in the comparisons between IBS-D patients and the healthy volunteers. Then the differentially expressed genes were further filtered for functional associations with neurotransmitters/mediators of pain. Each array was normalized by quantile, and then the DEGs analysis was performed (P -value < 0.05 , $FC \geq 1.2$ or ≤ 0.8). Compared with the HVs, we find out 9 upregulated genes (*CCKBR*, *CCL13*, *ACPP*, *BDKRB2*, *GRPR*, *SLC1A2*, *NPFF*, *P2RX4*, and *TRPA1*) and 4 downregulated genes (*TLX2*, *MRGPRX3*, *CXCR1* and *PAX2*) that may have relevance in the initiation and development of visceral pain. The most significant upregulated or downregulated genes specially linked to pain transmission are illustrated in Table 1 and Table 2.

Table 1 Upregulated genes specially involved in pain transmission in IBS-D patients in comparison with HVs.

ID		P Value	Gene symbol	Gene title	FC
206407	s at	1.98E-04	CCL13	C-C motif chemokine ligand 13	1.58
205870	at	1.51E-03	BDKRB2	bradykinin receptor B2	1.38
217590	_s_at	1.62E-02	TRPA1	transient receptor potential cation channel subfamily A member 1	1.23
204393	s at	4.26E-03	ACPP	acid phosphatase, prostate	1.55
208389	s at	1.11E-02	SLC1A2	solute carrier family 1 member 2	1.31
204088	at	1.41E-02	P2RX4	purinergic receptor P2X4	1.23
207929	at	2.62E-02	GRPR	gastrin releasing peptide receptor	1.37
206402	s at	4.65E-02	NPFF	neuropeptide FF-amide peptide precursor	1.25
234475	x at	1.70E-02	CCKBR	cholecystokinin B receptor	1.77

HVs: healthy volunteers; FC: fold change

Table 2 Downregulated genes specially involved in pain transmission in IBS-D patients in comparison with HV.

ID		P Value	Gene symbol	Gene title	FC
207410	s at	2.27E-02	TLX2	T-cell leukemia homeobox 2	0.65
1553293	at	1.24E-02	MRGPRX3	MAS related GPR family member X3	0.71
206228	at	2.58E-02	PAX2	paired box 2	0.76
207094	at	7.11E-03	CXCR1	C-X-C motif chemokine receptor 1	0.57

HVs: healthy volunteers; FC: fold change

Functional Enrichment Analysis

Functional analysis was carried out with GO biological process and KEGG pathways. Genes differentially were classified according to their putative Gene Ontology (GO) based on similarity with known genes recorded in public databases (Fig. 1). To further understand the functions of the DEGs, we represented significant pathways based on KEGG databases (Fig. 2). The upregulated DEGs were mainly associated with Glutamatergic synapse, Dilated cardiomyopathy, Arginine biosynthesis. The downregulated DEGs were linked to Circadian entrainment, Alcoholism, Aldosterone-regulated sodium reabsorption, Maturity onset diabetes of the young, cGMP-PKG signaling pathway, Dopaminergic synapse, Morphine addiction, Systemic lupus erythematosus, Glutamatergic synapse, Glycine, serine and threonine metabolism, Retrograde endocannabinoid signaling, Renin secretion.

PPI Network

A PPI network of the proteins encoded by identified DEGs was constructed by STRING. To identify the key modules of the PPI network, module clustering was then performed through the MCODE plugin of Cytoscape (Fig. 3). The modules showed that several pain or itch related genes like *GRPR*, *NPFF*, *CCKBR*, *BDKRB2*, *CXCR1* and *CCL13* genes (Red Rectangle, Fig. 3) were also clustered and most of these genes also occurred in GO terms or KEGG pathways enriched above. Finally, we identified that *GRPR*, *NPFF* and *TRPA1* as potential biomarkers for abdominal pain of IBS patients.

Discussion

Persistent hypersensitivity of sensory pathways innervating the colon is regarded as primary mechanism contributing to the initiation, development, and maintenance of chronic discomfort and abdominal pain in IBS patients (15, 16, 39). Therefore, determining the mechanisms contributing to these processes is crucial. Swan C et.al originally submitted the GSE36701 dataset, performed mRNA expression profiling study of rectal biopsies from donors with healthy volunteers and IBS-D patients[14]. They collected detailed clinical data (including abdominal pain frequency, anxiety and depression scale), using a combination of bioinformatics and experimental approaches to identify candidate genetic polymorphisms in the IBS[14]. While we further filtered for functional associations related to neurotransmitters/mediators of pain. Most notable among those genes were shown in Table 1 (upregulated genes) and Table 2 (downregulated genes). Interestingly, *GRPR*, *NPFF*, *TRPA1*, *BDKRB2*, *MRGPRX3*, that commonly regarded as factor also regulating itch signaling pathways were found[15-17]. Finally, three genes including *GRPR*, *NPFF* and *TRPA1* were considered to play an essential role in abdominal pain in IBS.

In the colon, afferent sensitization occurs via a variety of processes[18], including histamine-dependent mechanisms and histamine-independent mechanisms [19, 20]; Evidence have been accumulated that activation of receptors associated with the above two itch pathways on colon-innervating afferents induces visceral hypersensitivity[19, 21]. Gastrin-releasing peptide receptor expressing (GRPR)+ neurons have a central role in the spinal transmission of both histaminergic and non-histaminergic itch[22]. In our study, *GRPR* was significantly upregulated in IBS-D patients and identified as a hub gene by MCODE. GRPR is a G protein-coupled receptor and mediates itch sensation mainly via the PI3K/Akt pathway[23]. At the same time, GRP induces neutrophil chemotaxis through GRPR via p38, ERK1/2[24] and PI3K activation and p38, ERK and PI3K are important mediator in visceral pain[25-27]. Itch and pain signals are conveyed by distinct yet interacting neuronal pathways: pruritogens at higher doses produce pain to suppress itch[28, 29]; The frequency of abdominal pain in IBS was higher in patients with chronic pruritus than in healthy controls[30]. In addition, Neuropeptide FF (*NPFF*), the histamine-independent itch receptors agonist, evoking colonic afferent mechanical hypersensitivity[17, 19], was also clustered by Cytoscape module in our study. Therefore, we speculated that altered GRP-GRPR signaling and *NPFF* in spinal dorsal horn participate in visceral hypersensitivity through sensitization of itch transmission neurons, thereby contributing to abdominal pain or discomfort.

In the present study, another gene related to pain transmission deserve attention. Our results, together with previous studies have indicated significantly up-regulated *TRPA1*(Transient receptor potential ankyrin 1) mRNA expression in biopsies of IBS patients[31]. TRPA1 has been implicated in mechanical hypersensitivity of colonic afferents and both bradykinin and TNF- α induce visceral hypersensitivity through a TRPA1-dependent mechanism[32]. On the other hand, TRPA1 can also induce the inflammatory response via neurogenic inflammation: activation and sensitization of TRPA1 and release of substance P contribute to the initiation and development of colitis in mice, which correspondingly re-sensitises nociceptors[33]. TRPA1 expressed by intestinal enterochromaffin cells can serves as the primary detector of intestinal irritants prior to direct sub-mucosal damage[34]. These findings further highlight TRPA1 as an important integrator of sensory signals in colonic afferents by inducing mechanical visceral hypersensitivity.

As we known, IBS is a multifactorial disease. Abnormal stress response, psychological distress and infectious or inflammatory response in susceptible population may initiate visceral hypersensitivity that results in the development of IBS symptoms. According to the functional enrichment analyses, two pathways were associated with dysregulation of renin-angiotensin system (RAS), which is a potent target of stress-induced intestinal inflammation in a murine model of IBS [35]. Circadian entrainment and disorder of several synapses such as Glutamatergic synapse, Dopaminergic synapse may be risk factors of IBS. Besides, certain pathways directly involved in pain transmission were enriched, including Morphine addiction and Retrograde endocannabinoid signaling. Endocannabinoids are involved in controlling motility, secretion and intestinal inflammation[36]. The endocannabinoid system in DRG neurons mediate stress-induced visceral hypersensitivity in a mouse model of IBS[37].

Conclusions

Our results showed that *GRPR*, *NPFF* and *TRPA1* genes may be potential biomarkers for the diagnosis and new targets for treatment of abdominal pain in IBS. Several synapses modification and biological process of psychological distress may be risk factors of IBS. However, further studies are required to confirm the clinical significance of these findings.

Abbreviations

IBS: Irritable Bowel Syndrome ;HV: Healthy Volunteer; GRPR: Gastrin-Releasing Peptide Receptor; VH: Visceral hypersensitivity; TRPA1: Transient receptor potential ankyrin 1; NPFF: Neuropeptide FF; GEO: Gene Expression Omnibus; DEGs: Differentially expressed genes ; IBS-D: Irritable bowel syndrome with diarrhea; DAVID: Database for Annotation, Visualization, and Integrated Discovery ;STRING: Search Tool for the Retrieval of Interacting Genes ;GO: Gene Ontology; KEGG: Kyoto Encyclopedia of Genes and Genomes ; PPI: Protein-Protein Interaction ; MCODE: Molecular Complex Detection ;CXCL13: C-X-C motif chemokine ligand 13;CXCR1: C-X-C motif chemokine receptor 1

Declarations

Acknowledgements

Not applicable.

Authors' contributions

ZYL and YMW contributed to this work equally. ZYL: data collection and writing up of the first draft of the paper; YMW: data analysis. SQL, DCL and GHM did literature research and analysis of the data. Correspondence and requests for materials should be addressed to YLD and HZ: study design and modifying paper. All authors have read and approved the manuscript and ensure that this is the case.

Funding

No funding.

Availability of data and materials

The raw data of microarray experiment (its accession numbers are GSE36701) was downloaded from the Gene Expression omnibus (GEO) repository (<https://www.ncbi.nlm.nih.gov/geo/>). All data are publicly accessible. All data generated or analyzed during this study are included in this published article.

Ethics approval and consent to participate

Ethical approval is not required because this analysis was performed on public database. The original research ethics information can be accessed in GEO database (<https://www.ncbi.nlm.nih.gov/geo/>).

Consent for publication

Not applicable.

Competing interests

No competing interests.

Author details

¹Department of Anesthesiology, The First Affiliated Hospital of Sun Yat-sen University, Guangzhou, 510000, Guangdong, China. ²Department of Anesthesiology, Guangdong Second Provincial General Hospital, Guangzhou, 510000, Guangdong, China.

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Figures

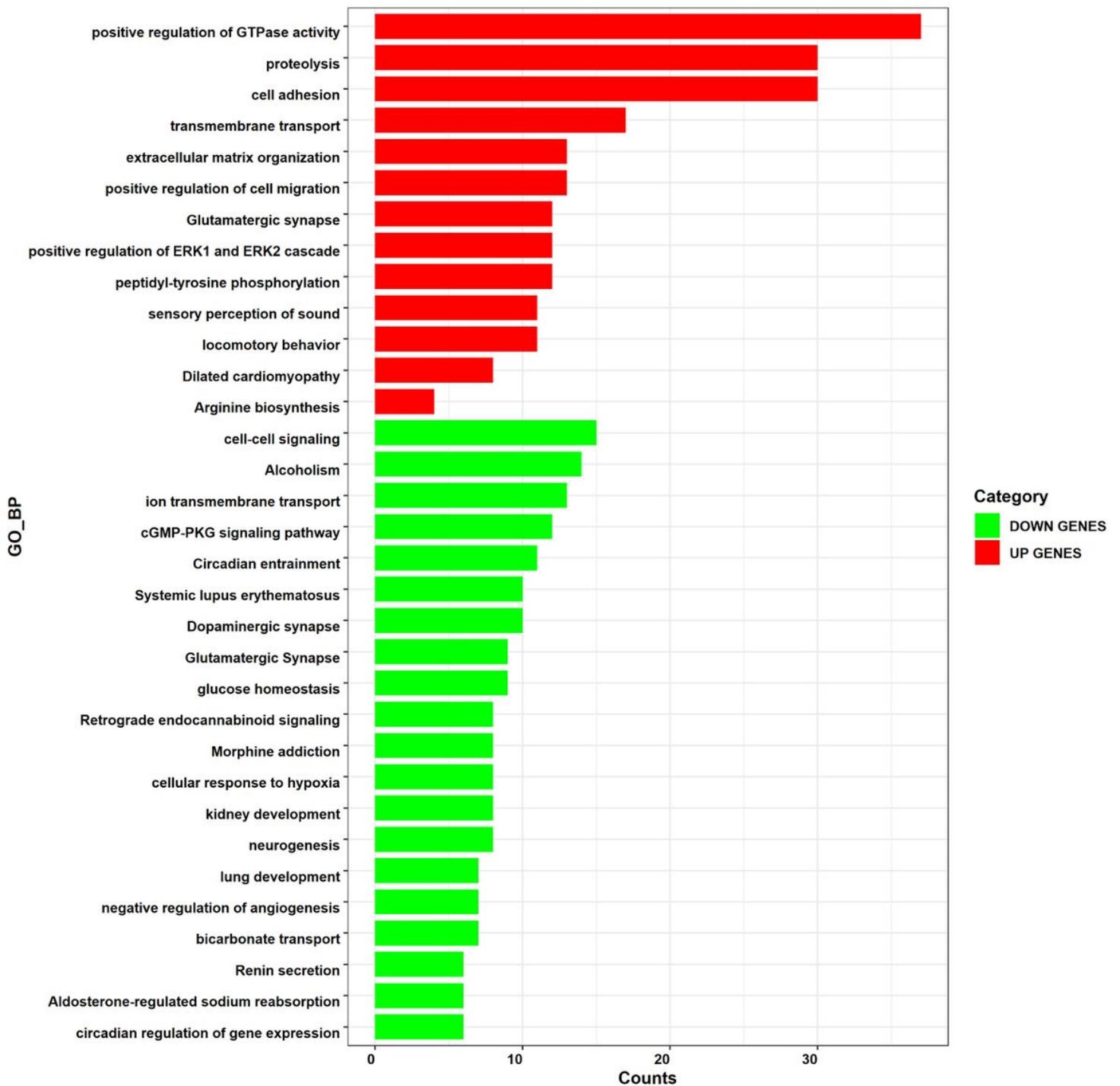


Figure 1

Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways enriched by the differentially expressed genes (DEGs). The x-axis of graph shows the counts, the number of genes clustered in each category, increasing with bar length. The threshold of P-value was set as 0.05.

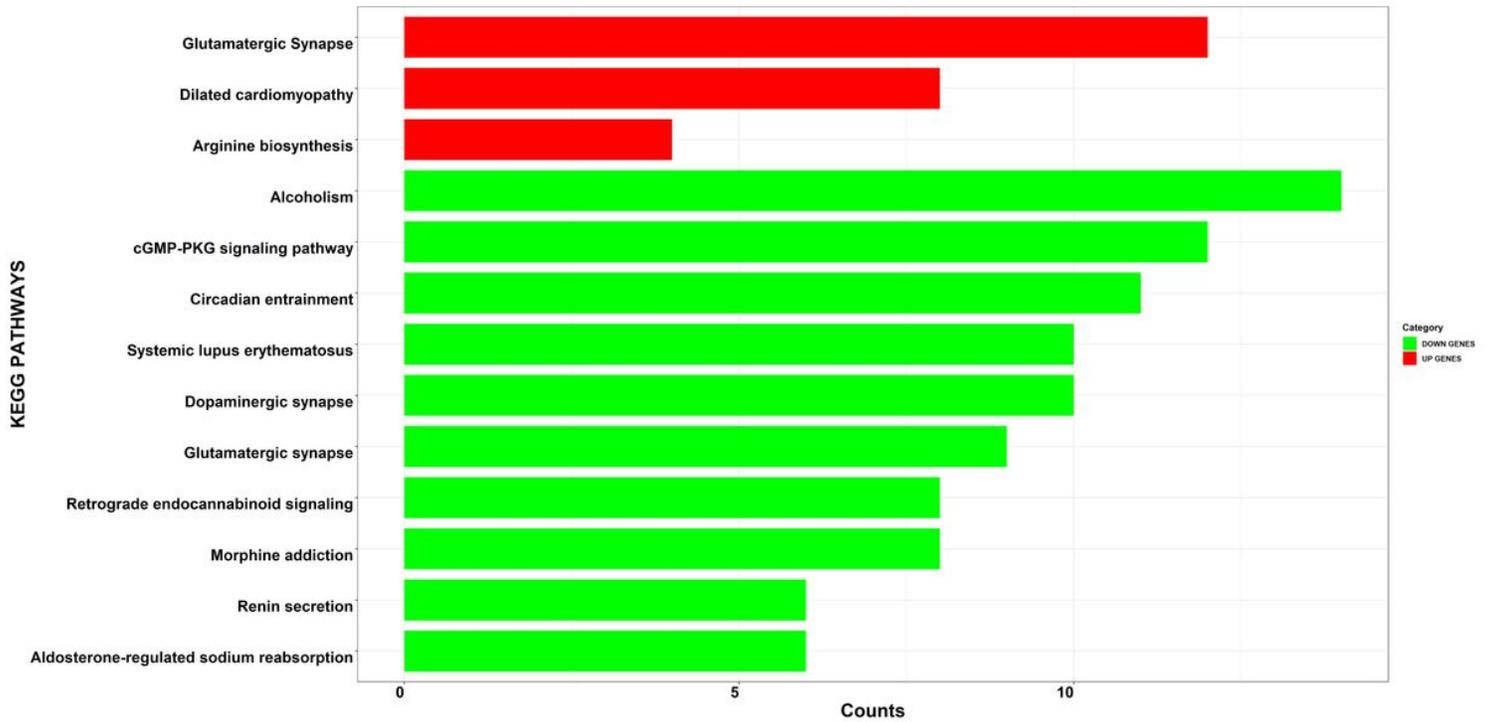


Figure 2

Biological process (BP) enriched by the differentially expressed genes (DEGs). The x-axis of graph shows the counts, the number of genes clustered in each category, increasing with bar length. The threshold of P-value was set as 0.05.

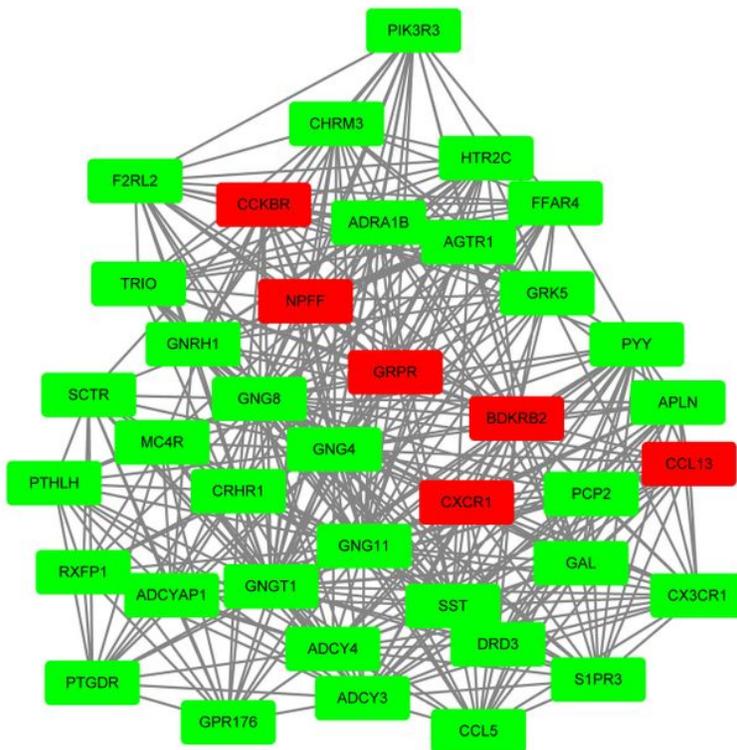


Figure 3

The Molecular Complex Detection (MCODE) components of the protein-protein interaction (PPI) network identified genes associated with pain transmission. Several outstanding hub genes such as CCKBR, NPFF, GRPR, BDKRB2, CXCR1 and CCL13 (red rectangle) was clustered. Degree >10 was set as the cut-off criterion. Rectangle represent DEGs and lines represent interaction and relationship between nodes.