

HDL-proteome enrichment with apolipoprotein A-IV compensates for HDL loss of function in diabetes mellitus kidney disease

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Keywords: diabetes mellitus, chronic kidney disease, HDL, apolipoprotein A-IV, proteomics, atherosclerosis

Posted Date: June 9th, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-33407/v1>

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Version of Record: A version of this preprint was published on September 14th, 2020. See the published version at <https://doi.org/10.1186/s12944-020-01381-w>.

Abstract

Background and aims:

Diabetes mellitus kidney disease (DKD) is associated with lipid derangements worsening kidney function and enhancing cardiovascular (CV) risk. The management of dyslipidemia, hypertension and other traditional risk factors does not completely prevent CV complications bringing up the participation of untraditional risk factors such as advanced glycation end products (AGEs), carbamoylation and changes in HDL proteome and functionality. We analyzed HDL composition, proteome, chemical modification and functionality in non-dialytic DKD subjects categorized according to glomerular filtration rate (GFR) and urinary albumin excretion rate (AER).

Methods:

DKD individuals were divided in GFR > 60 mL/min/1.73 m² plus AER stages A1 and A2 (n = 10) and GFR < 60 plus A3 (n = 25) and matched by age with control subjects (GFR > 60; n = 8).

Results:

Targeted proteomic analyses quantified 29 proteins associated with HDL in all groups, although only 2 were more expressed in GFR < 60 + A3 group in comparison to controls: apolipoprotein D (apo D) and apo A-IV. HDL from GFR < 60 + A3 presented higher levels of total AGEs, pentosidine and carbamoylation (1.2, 1.1 and 4.2 times, respectively) and a reduced ability in removing ¹⁴C-cholesterol from macrophages (33%) in comparison to controls. The antioxidant role of HDL (lag time for LDL oxidation) was similar among groups but HDL from GFR < 60 + A3 presented a higher ability in inhibiting the secretion of IL6 and TNF alpha (95%) in LPS-elicited macrophages in comparison to control group.

Conclusion:

The increment in apo A-IV that presents many antiatherogenic actions seems to counteract the HDL chemical modification by AGE and carbamoylation and its increment in apo D that occurred in well-established DKD.

Background

In diabetes mellitus (DM), abnormal kidney function is one of the most frequent complications being the leading cause of end-stage kidney disease. Besides, kidney function impairment increases the risk of cardiovascular disease (CVD) that is the major cause of mortality in both type 1 (DM1) and type 2 DM (DM2) [1]. Apart from traditional risk factors that are commonly observed in DM, including dyslipidemia, hypertension and other components of the metabolic syndrome, untraditional risk factors - advanced glycation, carbamoylation and oxidation - contribute to macrovascular disease in DM kidney disease (DKD) [2-4].

In DKD, the reduction in kidney function represented by the diminished glomerular filtration rate (GFR) is not invariably accompanied in the same extension by elevation in the urinary albumin excretion rate (AER) [5]. In fact, many individuals with DKD with a marked reduction in GFR can still present normal (A1 stage) or slightly reduced AER (A2 stage) and in some cases people in the A2 stage can revert to A1 [6]. The incidence of CVD is positively related to the reduction in GFR as well as AER, and both parameters have additive effects on CV risk in any stage of abnormal kidney function [7].

Advanced glycation end products (AGEs) are prevalent in DM and in chronic kidney disease (CKD) due to hyperglycemia, oxidative stress and detoxification failure of intermediate compounds of the glycation reaction. AGEs are independent predictors of CV risk by impairing the reverse cholesterol transport (RCT) and lipid metabolism, inducing inflammation and altering vasodilation [8-10]. In addition, the reaction of isocyanic acid derived from urea with proteins leads to protein carbamylation that is also related to atherogenesis [4].

The reduction of high-density lipoprotein (HDL) cholesterol in plasma is a hallmark in DM although HDL dysfunction is also considered as having an important role in CV morbidity and mortality. This is especially reputable when analyzing clinical trials where the increment in HDL cholesterol did not contribute for CV risk improvement. HDL are antiatherogenic particles that mediate the removal of excess cholesterol from the arterial wall macrophages allowing its traffic to liver and excretion in feces by the RCT. Besides, HDL exert several other atheroprotective actions including antioxidant, antiinflammatory, vasodilation and anti-aggregant and improving glucose tolerance and insulin sensitivity. HDL is a cargo lipoprotein transporting many proteins and microRNAs that are able to control metabolism in different tissues and in the arterial wall [11].

The HDL proteome that follows its complexity and functionality has been analyzed in CKD associated or not with DM, but in most of the studies, the individuals were on dialysis that may have influenced the results. In addition, the stratification of CKD subjects by GFR or AER alone may not reflect changes in HDL proteome and function that takes place in an unusual evolution of chronic disease typically in DKD. We then analyzed the HDL proteome, composition and chemical modification by advanced glycation and carbamylation, together with its functionality in non-dialysis DKD subjects categorized according to the GFR and AER in comparison to age-matched control subjects. We found that the HDL proteome was enriched in apolipoprotein D (apo D) and apo A-IV and that HDL was modified by advanced glycation and carbamylation, according to the reduction in GFR and increased AER. The ability of HDL in removing cellular cholesterol was reduced in DKD with GFR < 60 mL/min/1.73 m² plus A3 although its antioxidant activity was preserved and its capacity to prevent inflammation in macrophages was even increased. This may be ascribed to the increment of HDL in apo A-IV that exerts antiatherogenic actions counteracting HDL loss of function that is reported in CKD.

Material And Methods

DM2 subjects with DKD were recruited at the Hospital das Clínicas da Faculdade de Medicina da Universidade de São Paulo. Healthy control individuals matched by age and gender were selected at Faculdade de Medicina da Universidade de São Paulo. All participants were properly informed about the procedures and the study and signed an informed written consent form that was previously approved by The Ethical Committee for Human Research Protocols of the Clinical Hospital (#15024), in accordance with the Declaration of Helsinki.

Subjects with other chronic diseases, rapid loss in GFR ($> 3 \text{ mL} / \text{min} / \text{year}$), refractory hypertension, BMI $< 18.5 \text{ Kg} / \text{m}^2$, current smoke or alcohol abuse were not included. Blood was drawn after overnight fasting and plasma was immediately isolated in refrigerated centrifuge (4°C). Glycemia, triglycerides (TG), total cholesterol (TC), HDL cholesterol (HDLc), fructosamine, TSH, creatinine and urea were determined in plasma by enzymatic techniques after overnight fasting and albumin in 24h-urine. HbA1c was determined by high performance liquid chromatography (HPLC). DKD individuals were categorized according to the GFR above $60 \text{ mL}/\text{min}/1.73 \text{ m}^2$ plus AER stages A1 ($< 30 \text{ mg}/\text{g}$ creatinine) and A2 ($30 - 300 \text{ mg}/\text{g}$ creatinine) and GFR below $60 \text{ mL}/\text{min}/1.73 \text{ m}^2$ plus stage A3 ($> 300 \text{ mg}/\text{g}$ creatinine). Control subjects presented GFR above $60 \text{ mL}/\text{min}/1.73 \text{ m}^2$ plus A1.

Isolation of lipoproteins

Venous blood samples were drawn after overnight fasting and plasma immediately isolated in a refrigerated centrifuge. Preservatives were added to the plasma and density adjusted with bromide potassium to $1.21 \text{ g}/\text{mL}$. Low-density (LDL; $d = 1.019-1.063 \text{ g}/\text{mL}$) and high-density lipoprotein (HDL; $d = 1.063-1.21 \text{ g}/\text{mL}$) were isolated from plasma by discontinuous density gradient ultracentrifugation ($100\,000\text{g}$, 24 h , 4°C , Sw40 rotor; Beckman ultracentrifuge). Samples were dialyzed against phosphate buffer saline containing EDTA (PBS).

HDL composition in lipids and apoA-I

The amount of lipids and apo A-I in HDL were determined, respectively, by enzymatic techniques [TC and TG; Labtest diagnóstica S. A., Minas Gerais, Brazil; phospholipids (PL); Randox Laboratorier LTD. Crumlin, Co. Antrem, United Kingdom] and immunoturbidimetry.

Determination of total AGE and pentosidine in HDL

The contents of total AGE and pentosidine were determined by fluorescence measurement (Synergy HT Multi-Mode Microplate Reader, SpectraMax M5). Samples were excited at a wavelength of 370nm and the fluorescence emitted at 440 nm and 378 nm , respectively, for total AGE and pentosidine.

Proteolytic digestion of HDL

The HDL protein concentration was determined by the Bradford assay (Bio-Rad, Hercules, CA, USA). Ten micrograms of HDL-protein was solubilized in 100 mM ammonium bicarbonate, dithiothreitol, and iodoacetamide, following digestion with trypsin ($1:40$, w/w Promega, Madison, WI, USA) for 4 h at 37°C .

Trypsin was further added to samples (1:50, w/w HDL) and the incubation was done overnight at 37 °C. Samples were desalted using solid phase extraction (Oasis PRIME HLB SPE column; Waters) after acidic hydrolysis with 2% trifluoroacetic acid and kept frozen at - 80°C until MS analyses. Prior to MS analysis, samples were resuspended in 0.1% formic acid (final protein concentration of 25 ng/μL).

Targeted proteomic analyses

Digested HDL proteins (50 ng protein) were quantified by parallel reaction monitoring (PRM), as previously described ^[12]. Briefly, an Easy-nLC 1200 UHPLC (Thermo Scientific, Bremen, Germany) was used for peptide separation. Acquisition of the data was performed in an Orbitrap Fusion Lumos mass spectrometer (Thermo Scientific, Bremen, Germany) using a nanospray Flex NG ion source (Thermo Scientific, Bremen, Germany). A scheduled (3-min window) inclusion list containing m/z of precursor peptides of interest and corresponding retention times was generated using Skyline software ^[13].

Selection of HDL peptides for targeted quantification

PRM methodology was assembled from shotgun proteomics analyses as previously described ^[12]. Ninety-one proteins were identified, but this number was reduced to 47 proteins eliminating potentials contaminant proteins (keratin, proteins with <2 unique peptides and peptides with high interfering signal and mass error >10 ppm). Peptides susceptible to ex-vivo modification (e.g., methionine-containing peptides) were also avoided, and only peptides satisfactorily detected were included in the final analysis. After our exclusion criteria, 31 proteins remained. For statistical analysis, we considered the best peptide that represents each protein of interest. In order to find these surrogate peptides, firstly the peptide pair with best Pearson's correlation coefficient was determined. From these 2 peptides, the peptide with the lowest CV was finally selected. The 31 surrogate peptides chosen for HDL proteins are highlighted in Supplementary Table 1.

Acetylation of LDL

LDL was acetylated as previously described by Basu et al ^[14]. Samples were extensively dialyzed before incubation with macrophages.

Measurement of ¹⁴C-cholesterol efflux

Bone marrow-derived cells were isolated from mice and macrophages were differentiated as previously described ^[15]. Cells were overloaded with acetylated LDL (50 μg/mL DMEM) and ¹⁴C-cholesterol (0.3 μCi/mL) for 48 h. HDL from controls and DKD subjects (50 μg/mL) were utilized as cholesterol acceptors in 6- h incubations and cholesterol efflux calculated as previous described ^[8].

Measurement of the HDL antioxidant activity

The ability of HDL from controls and DKD individuals in inhibiting LDL oxidation was determined by incubation of LDL (40 μg/mL) isolated from a unique healthy plasma donor with CuSO₄ solution (1 mL)

in the presence of HDL (80 µg/mL). Lipoproteins were dialyzed against PBS without EDTA prior incubations. The absorbance at 234 nm was continuously monitored every 3 min during 4 h and the lag time phase for LDL oxidation (min) and the maximum ratio of conjugated dienes formation calculated [15].

Measurement of the HDL antiinflammatory activity

BMDM were isolated and cultured as previously described; then overloaded with acetylated LDL (50 µg/mL DMEM) and treated for 24 h with HDL from controls and DKD subjects. After washing, macrophages were incubated with lipopolysaccharide (LPS; 1 µg/mL DMEM) for 24 h. Medium was collected and the amount of TNF alpha and interleukin 6 (IL-6) determined by ELISA (R&D System-Duo Set, Minneapolis, EUA) [9].

Statistical analysis

Statistical analysis was performed using the GraphPad Prism 5 program (GraphPad Software, Inc. 2007). Comparisons were made by one-way analysis of variance (ANOVA) with Dunnett post-test, Student t test and Spearman linear correlation as appropriated. The value of $p < 0.05$ was considered statistically significant. Proteome data were compared by multiple comparisons.

Results

Anthropometric and biochemical data of control and DKD subjects are depicted in table 1. In the group GFR <60 + A3 there was a greater predominance of male individuals (72%) compared to the control (50%) and GFR > 60 + A1 and A2 (30%) groups. Age was similar among groups as well as the time of disease comparing both DKD groups. BMI and CVD history were higher in the group GFR > 60 + A1 and A2. Fructosamine and HbA1c levels were higher in DM groups as compared to controls but CML levels did not reached statistically difference among groups. Total cholesterol (TC) was lower in the group GFR > 60 + A1 and A2 as compared to controls.

Eighty percent of DKD subjects were in use of insulin and 70 % were on statins and beta-blockers. Angiotensin-converting-enzyme inhibitors (ACEi) and metformin were used for A1 and A2 subjects only and angiotensin II receptor blockers (ARBs) by groups A1 (39 %), A2 (50 %) and A3 (36%). Eritropoietin was utilized by A2 (7 %) and A3 (12 %) groups.

Twenty-nine HDL-associated proteins were quantified by PRM using nanoscale liquid chromatography coupled with mass spectrometry (Nano-LC / MS / MS). Of these, 2 were more expressed in the GFR < 60 + A3 group as compared to the control group: apo A-IV (**figure 1 panel A**) and apoD (**figure 1 panel B**).

A positive correlation was observed between apoA-IV (**figure 2 panel A**) and cystatin C (**Figure 2 panel B**) with the AER. On the other hand, a negative correlation was observed between apoA-I (**figure 2 panel C**), serum amyloid protein A-4 (SAA4, **figure 2 panel D**) and apoC-IV (**figure 2 panel E**) with the AER.

The HDL composition in lipids and apoA-I is shown in **figure 3**. The HDL content in TC (**figure 3 panel A**) and TG (**figure 3 panel B**) was similar between all groups. There was a decrease in PL (**figure 3 panel C**) and apoA-I (**figure 3 panel D**) in the HDL of the TFG group <60 + A3 compared to the control group. A positive correlation was observed between HDL-PL (**Figure 3 panel E**), and apoA-I (**Figure 3 panel F**) with the GFR.

Total AGE (**figure 4 panel A**) and pentosidine (**figure 4 panel B**) were determined in HDL, being, for both cases, higher in HDL isolated from individuals with GFR < 60 + A3 (80% and 93.7 %, respectively), as compared to the control group. Although the values of HbA1c and fructosamine were similar between groups, carbonyl stress as a function of albuminuria and GFR < 60 can be attributed to the renal changes that accompany macroalbuminuria and are reflected in the lower detoxification of glycation reaction precursors and greater oxidative stress. The modification of HDL by isocyanic acid that reflects uremic stress was greater in the group with GFR < 60 + A1 compared to the control group (**figure 4 panel C**). A positive correlation was observed between total AGE (**Figure 4, panel D**) and pentosidine in HDL (**Figure 4 panel E**) with AER.

HDL was utilized as acceptors of ¹⁴C-cholesterol from BMDM. As shown in the **figure 5 (panel A)**, ¹⁴C-cholesterol efflux mediated by HDL isolated from the TFG < 60 + A3 group was lower as compared to that mediated by the HDL from the control group. The antioxidant activity of HDL was determined by measuring LDL oxidation with CuSO₄ along time. The lag phase for LDL oxidation determined by the presence of HDL was similar among all groups (**figure 5 panel B**) as well as the maximum ratio of conjugated dienes formation in LDL (**figure 5 panel C**). In the **figure 5 (panel D and E)**, it is demonstrated the secretion of inflammatory cytokines, IL6 (panel D) and TNF alpha (panel E) in macrophages treated with HDL from controls and DKD patients with GFR < 60 + A3 groups and after challenged with LPS. For both cytokines, it was observed a very lower secretion when macrophages were exposed to DKD HDL in comparison to C-HDL.

Discussion

The prevalence of kidney complications in DM is high and the AER together with the reduced GFR independently and synergistically predict CV morbidity and mortality^[16]. Changes in the HDL proteome and functionality may modulate the antiatherogenic actions of this lipoprotein and consequently the development of atherosclerosis^[17]. In this study, we evaluated in DM2 patients with DKD, the composition, chemical modification and proteomics of the HDL particles and their ability to remove cholesterol from macrophages, inhibit LDL oxidation and macrophage inflammation.

Targeted proteomics quantified 29 proteins in HDL of all experimental groups, although only 2 were differentially expressed in DKD with GFR < 60 plus A3. Apo D is an atypical HDL apolipoprotein closely related to retinol-binding protein. It contributes for the HDL hydrophobic nucleus remodeling by facilitating the lecithin-cholesterol acyltransferase (LCAT) anchoring to the lipoprotein structure and carrying lysophosphatidylcholine, although it is not clear whether it has the potential to activate or inhibit

the enzyme ^[18]. Some studies show an increase of apoD in the HDL proteome of individuals with coronary artery disease and in areas of human atherosclerotic lesion as well as in apolipoprotein E knockout mice plasma ^[18, 19, 20]. However, it is not clear yet whether this increase in apoD in these conditions refers to its role in inducing atherosclerosis or whether it represents a compensatory adaptive mechanism to changes observed in cardiovascular disease ^[18].

Several studies have demonstrated that apoA-IV can be used as an early marker of kidney failure in individuals with CKD and in the general population, although further studies are needed to understand the pathophysiological basis of this association ^[21]. Besides, in individuals on hemodialysis, the increment in apoA-IV was associated with an increased risk of all causes mortality ^[22]. Our data agree with these findings and an increased apoA-IV expression was observed in the HDL proteome of individuals with higher AER and reduced GFR as compared to controls. Also, apoA-IV was positively correlated to AER. This apolipoprotein is synthesized in the intestine and secreted in the mesenteric lymph, being transported by chylomicrons but mainly by HDL, being the third most abundant apolipoprotein in this lipoprotein ^[21, 22]. ApoA-IV presents many anti-atherogenic functions, acting in the removal of cellular cholesterol ^[23] and promoting activation of lipoprotein lipase ^[24], LCAT and cholesteryl ester transfer protein (CETP) ^[25]. In addition, its anti-atherogenic activity is complemented by its anti-inflammatory and antioxidant properties ^[26, 27].

Similarly to other HDL-apolipoproteins, apoA-IV is modified by advanced glycation in DM and CKD. Recently, our group demonstrated that E. coli recombinant apo A-IV submitted to advanced glycation in vitro maintains its ability in removing macrophage excess cholesterol, despite its large amount pyrroline, CML and argpyrimidine ^[28]. This may explain the fact that no major reductions in cholesterol efflux were observed in the present study, even with a significant increase in total AGE and pentosidine in the HDL of the group GFR < 60 + A3 as compared to controls. In addition, the in vitro glycation of apo A-IV only partially impaired its ability to inhibit the inflammatory response promoted by LPS in macrophages ^[280]. On the contrary, apoA-I has its ability in removing cell cholesterol and antioxidant and anti-inflammatory properties severely impaired by advanced glycation ^[29].

The HDL proteome has been analyzed in CKD, but the vast majority of studies were performed in individuals undergoing hemodialysis, which can interfere with the observed results. Also, many studies included CKD together with DKD indistinctively. Mangé et al ^[30] found 40 differently expressed proteins in HDL from CKD subjects. ApoC-II and apoC-III were higher while transthyretin and haptoglobin-related protein (HPTR) were lower in HDL from CKD subjects on hemodialysis compared to healthy subjects. In another study, individuals on hemodialysis, with or without DM, an increase in SAA1, albumin, phospholipase A2 and apoCIII was observed in HDL proteome. Along with the reduction in the content of PL and an increase in TG and lysophospholipids, these modifications were linked to the reduction in the HDL-mediated cholesterol efflux ^[31]. In agreement HDL dysfunction was also related to changes in HDL proteome by Florens et al ^[32]. Shao et al ^[33] found proteins related to renal injury (beta 2 microglobulin, complement factor D, cystatin C, prostaglandin-H2 D-isomerase, retinol-binding protein 4 and AMBP)

increased in HDL of individuals with CKD. Others were more present in the control group, among them: apoA-I, apoA-II, apoL-I, apoM and paraoxonase 1, conferring greater damage to the anti-atherogenic activities of HDL in CKD.

Recently, the HDL proteome was analyzed in 191 individuals with DM1, in the Diabetes Control and Complications Trial (DCCT) study. Eight proteins were associated with proteinuria, although only 1 was simultaneously associated with AER and coronary calcium content ^[34]. Wang et al ^[35] demonstrated enrichment in 8 proteins related to inflammation and lipid metabolism (serum amyloid A1, A2, and A4; hemoglobin beta, HPTR, CETP, PLTP and apo E) in HDL isolated from individuals in short-term as compared to long-term dialysis therapy.

Although not quantitatively changed in our proteomics analysis, cystatin C levels in HDL were positively correlated with the AER. Cystatin is directly linked to impaired renal function and is also considered by some investigators as a marker of coronary atherosclerosis and cerebrovascular events ^[36]. On the other hand, SAA4 and apoC-IV were inversely correlated with AER. Higher levels of SAA associated to HDL have been shown as correlated with a reduced ability of HDL in removing cell cholesterol ^[31] and are associated with CV mortality ^[37].

ApoA-I determined by immunoturbidimetry was reduced in HDL of the TFG group < 60 + A3 with a negative correlation with AER. Therefore, it is likely that in vivo, cellular cholesterol removal may be even more compromised than that observed in the present study due to the lower content of apo A-I in individuals with established DKD. In our efflux assay HDL concentration was matched by the protein component that is mainly represented by apoA-I, making more difficult to observed differences.

A decrease in the HDL-PL was also observed in the GFR group < 60 + A3 with a positive correlation with GFR. Notably, PLs are positive modulators of cell cholesterol removal, as they facilitate the interaction of HDL with plasma membrane domains that guarantee cholesterol exportation by diffusion and / or mediated by specific receptors.

Reduced ability in removing cell cholesterol was previously reported in CKD individuals stage 3 and 4 as compared to healthy controls although not independently of age ^[380]. In this sense, there are studies showing the role of aging on the ability of HDL in mediating cholesterol efflux that may be a bias for many studies dealing with controls and CKD subjects at different ages ^[39, 40]. In dialytic individuals, Yamamoto et al ^[41] reported a reduced cholesterol efflux mediated by HDL from DM with or without DKD in comparison to controls.

Differently from our work, there are many studies that measured the cholesterol efflux mediated by the apoB depleted serum. Although, HDL is the only lipoprotein in that serum, there are many other components including albumin, cytokines, haptoglobin and insulin that could affect cholesterol removal. Besides, the measurement of cholesterol removal mediated by apo B-depleted serum mitigates variations in HDL cholesterol level that are frequently altered in DM and DKD ^[42, 43].

Ganda et al ^[44] did not find any alteration in the cholesterol efflux mediated by the apo B-serum isolated from CKD subjects (stages 4 and 5), although the expression of *Abca1* in monocytes isolated from those subjects was reduced compromising the cholesterol efflux to apoA-I. Chemical modification of HDL by advanced glycation and carbamoylation that was observed in the present study was related to the stage of AER and GFR. In individuals in hemodialysis or peritoneal dialysis a greater amount of pentosidine was found in plasma that correlated to the progression of kidney disease. Besides, pentosidine levels were higher in CKD stage 5 subjects as compared to stage 1 ^[45]. In CKD subjects undergoing hemodialysis it was demonstrated elevated concentration of carbamoylated HDL, which is dysfunctional impairing the RCT ^[46, 47].

Conclusion

The antioxidant capacity of HDL assessed by the lag time for LDL oxidation was similar among groups, while the HDL anti-inflammatory ability was greatly increased. This was demonstrated by the reduced secretion of inflammatory cytokines by macrophages challenged by LPS. These results may be ascribed to the enhanced amount of apo A-IV in HDL that seems to compensate for chemical modifications of HDL that take place in established DKD ^[48]. The exact role of apoA-IV enhancement in HDL proteome should be investigated in detail in order to clarify its contribution for CVD prevention in DKD.

Abbreviations

apo A-I
apolipoprotein A-I
apo A-IV
apolipoprotein A-IV
apo C-IV
apolipoprotein C-IV
apo D
apolipoprotein D
AER
albumin excretion rate
AGEs
advanced glycation end products
BMDM
bone marrow derived-macrophages
CKD
chronic kidney disease
CVD
cardiovascular disease
DKD

diabetes mellitus kidney disease
GFR
glomerular filtration rate
HDL
high density lipoprotein
LDL
low density lipoprotein
RCT
reverse cholesterol transport
SAA4
serum amyloid protein A-4

Declarations

Ethics approval and consent to participate: All participants were properly informed about the procedures and the study and signed an informed written consent form that was previously approved by The Ethical Committee for Human Research Protocols of the Clinical Hospital (#15024), in accordance with the Declaration of Helsinki.

Consent for publication: Not applicable

Availability of data and material: The dataset used and/or analyzed during the current study are available from the corresponding author on reasonable request. The authors declare that all data generated or analyzed during this study are included in this published article [and its supplementary information files].

Competing interests: there are no conflicts of interest. All authors have read and approved the submission of the manuscript; the manuscript has not been published and is not being considered for publication elsewhere, in whole or in part, in any language.

Funding

The authors would like to thank the financial support from Fundação de Amparo à Pesquisa do Estado de São Paulo, FAPESP (grants #2018/18545-4 to MMS, 2018/00172-0 to RSP, 2015/21072-5 to MP and 2016/15603-0 to MP and MLCCG, 2016/00696-3 to GER, 2017/07725-1 to ARMS and GER) and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – Brasil (CAPES) – Finance Code 001. MP and MLCCG are recipients of a research award from Conselho Nacional de Desenvolvimento Científico e Tecnológico, CNPq, Brazil.

Autors contribution

MFMS: performed all experiments and helped in manuscript preparation; ALAL: selected patients and controls and helped in clinical data processing; RSP: helped in experiments performance; CAM: helped in patients recruitment and clinical data; ARMS: performed proteome analysis and data processing;

MIBACS: helped in statistics; ERN: helped in data interpretation; MLCCG: helped in data interpretation; GER: supervised the proteome analysis and data interpretation; MSQ: supervised patients recruitment and helped in data interpretation; MP: conceived the study and experiments and wrote the manuscript.

Acknowledgments: The authors would like to thank Fundação Faculdade de Medicina and Laboratórios de Investigação Médica (LIM) do Hospital das Clínicas da Faculdade de Medicina da Universidade de São Paulo.

References

1. Maqbool M, Cooper ME, Jandeleit-Dahm KAM. Cardiovascular disease and Diabetic Kidney Disease. *Semin Nephrol.* 2018;38(3):217–32. doi:10.1016/j.semnephrol.2018.02.003.
2. Rabbani N, Thornalley PJ. Advanced glycation end products in the pathogenesis of chronic kidney disease. *Kidney Int.* 2018;93(4):803–13. doi:10.1016/j.kint.2017.11.034.
3. Ravarotto V, Simioni F, Pagnin E, et al. Oxidative stress - chronic kidney disease - cardiovascular disease: A vicious circle. *Life Sci.* 2018;210:125–31. doi:10.1016/j.lfs.2018.08.067.
4. Delanghe S, Delanghe JR, Speeckaert R, et al. Mechanisms and consequences of carbamoylation. *Nat Rev Nephrol.* 2017;13(9):580–93. doi:10.1038/nrneph.2017.103.
5. Tsalamandris C, Allen TJ, Gilbert RE, et al. Progressive decline in renal function in diabetic patients with and without albuminuria. *Diabetes.* 1994;43(5):649–55. doi:10.2337/diab.43.5.649.
6. Halimi JM. The emerging concept of chronic kidney disease without clinical proteinuria in diabetic patients. *Diabetes Metabolism.* 2012;38:291–97. doi:10.1016/j.diabet.2012.04.001.
7. Chronic Kidney Disease Prognosis Consortium. Matsushita K, van der Velde M, et al. Association of estimated glomerular filtration rate and albuminuria with all-cause and cardiovascular mortality in general population cohorts: a collaborative meta-analysis. *Lancet.* 2010;375(9731):2073–81. doi:10.1016/S0140-6736(10)60674-5.
8. Machado-Lima A, Iborra RT, Pinto RS, et al. Advanced glycated albumin isolated from poorly controlled type 1 diabetes mellitus patients alters macrophage gene expression impairing ABCA1-mediated reverse cholesterol transport. *Diabetes Metab Res Rev.* 2013;29(1):66–76. doi:10.1002/dmrr.2362.
9. Okuda LS, Castilho G, Rocco DD, et al. Advanced glycated albumin impairs HDL anti-inflammatory activity and primes macrophages for inflammatory response that reduces reverse cholesterol transport. *Biochim Biophys Acta.* 2012;1821(12):1485–92. doi:10.1016/j.bbaliip.2012.08.011.
10. Jiang J, Chen P, Chen J, et al. Accumulation of tissue advanced glycation end products correlated with glucose exposure dose and associated with cardiovascular morbidity in patients on peritoneal dialysis. *Atherosclerosis.* 2014;224(1):187–94. doi:10.1016/j.atherosclerosis.2012.06.022.
11. Ben-Aicha S, Badimon L, Vilahur G. Advances in HDL: Much More than Lipid Transporters. *Int J Mol Sci.* 2020;21(3):732. doi:10.3390/ijms21030732.

12. Silva ARM, Toyoshima MTK, Passarelli M, Di Mascio P, Ronsein GE. Comparing data-independent acquisition and parallel reaction monitoring in their abilities to differentiate high-density lipoprotein subclasses. *J Proteome Res.* 2020;19(1):248–59. doi:10.1021/acs.jproteome.9b00511.
13. MacLean B, Tomazela DM, Shulman N, et al. Skyline: an open source document editor for creating and analyzing targeted proteomics experiments. *Bioinformatics.* 2019;26(7):966–8. doi:10.1093/bioinformatics/btq054.
14. Basu SK, Goldstein JL, Anderson, Brown RGWMS. Degradation of cationized low density lipoprotein and regulation of cholesterol metabolism in homozygous familial hypercholesterolemia fibroblasts. *Proc Natl Acad Sci U S A.* 1976;73(9):3178–82. doi:10.1073/pnas.73.9.3178.
15. Iborra RT, Ribeiro IC, Neves MQ, et al. Aerobic exercise training improves the role of high-density lipoprotein antioxidant and reduces plasma lipid peroxidation in type 2 diabetes mellitus. *Scand J Med Sci Sports.* 2008;18(6):742–50. doi:10.1111/j.1600-0838.2007.00748.x.
16. Gansevoort RT, Matsushita K, Van Der Velde M, et al. Lower estimated GFR and higher albuminuria are associated with adverse kidney outcomes. A collaborative meta-analysis of general and high-risk population cohorts. *Kidney Int.* 2011;80(1):93–104. doi:10.1038/ki.2010.531.
17. Kronenberg F. HDL in CKD – the devil is in the detail. *J AM Soc Nephrol.* 2018;29(5):1356–71. doi:10.1681/ASN.2017070798.
18. Perdomo G, Henry Dong H. Apolipoprotein D in lipid metabolism and its functional implication in atherosclerosis and aging. *Aging.* 2009;1(1):17–27. doi:10.18632/aging.100004.
19. Vaisar T, Pennathur S, verde PS, et al. Shotgun proteomics implicates protease inhibition and complement activation in the anti-inflammatory properties of HDL. *J Clin Invest.* 2007;117(3):746–56. doi:10.1172/JCI26206.
20. Sarjeant JM, Lawrie A, Kinnear C, et al. Apolipoprotein D inhibits platelet-derived growth factor-BB-induced vascular smooth muscle cell proliferation by preventing translocation of phosphorylated extracellular signal regulated kinase 1/2 to the nucleus. *Arterioscler Thromb Vasc Biol.* 2003;23(12):2172–7. doi:10.1161/01.ATV.0000100404.05459.39.
21. Kronenberg F. Apolipoprotein L1 and apolipoprotein A-IV and their association with kidney function. *Curr Opin Lipidol.* 2017;28(1):39–45. doi:10.1097/MOL.0000000000000371.
22. Kollerits B, Krane V, Drechsler C, et al. Apolipoprotein A-IV concentrations and clinical outcomes in haemodialysis patients with type 2 diabetes mellitus—a post hoc analysis of the 4D Study. *J Intern Med.* 2012;272(6):592–600. doi:10.1111/j.1365-2796.2012.02585.x.
23. Steinmetz A, Barbaras R, Ghalim N, et al. Human apolipoprotein A-IV binds to apolipoprotein A-I/A-II receptor sites and promotes cholesterol efflux from adipose cells. *J Biol Chem.* 1990;265(14):7859–63.
24. Goldberg IJ, Scheraldi CA, Yacoub LK, et al. Lipoprotein apoC-II activation of lipoprotein lipase. Modulation by apolipoprotein A-IV. *J Biol Chem.* 1990;265(8):4266–72.
25. Guyard-Dangremont V, Lagrost L, Gambert P. Comparative effects of purified apolipoproteins A-I, A-II, and A-IV on cholesteryl ester transfer protein activity. *J Lipid Res.* 1994;35(6):982–92.

26. Peng J, Li Xiang-Ping. Apolipoprotein A-IV: a potential therapeutic target for atherosclerosis. *Prostaglandins Other Lipid Mediators*. 2018;139:87–92. doi:10.1016/j.prostaglandins.2018.10.004.
27. Duverger N, Trempe G, Caillaud JM, et al. Protection against atherogenesis in mice mediated by human apolipoprotein A-IV. *Science*. 1996;273(5277):966–8. doi:10.1126/science.273.5277.966.
28. Okuda LS, Iborra RT, Pinto PR, et al. Advanced glycosylated apoA-IV loses its ability to prevent the LPS-Induced reduction in cholesterol efflux-related gene expression in macrophages. *Mediators of Inflammation*. Article ID 6515401. doi.org/10.1155/2020/6515401.
29. Nobécourt E, Tabet F, Lambert G, et al. Nonenzymatic glycation impairs the antiinflammatory properties of apolipoprotein A-I. *Arterioscler Thromb Vasc Biol*. 2010;30(4):766–72. doi:10.1161/ATVBAHA.109.201715.
30. Mangé A, Goux A, Badiou S, et al. HDL proteome in hemodialysis patients: A quantitative nanoflow liquid chromatography-tandem mass spectrometry approach. *PLoS One*. 2012;7(3):e34107. doi:10.1371/journal.pone.0034107.
31. Holzer M, Birner-Gruenberger R, Stojakovic T, et al. Uremia alters HDL composition and function. *J Am Soc Nephrol*. 2011;22(9):1631–41. doi:10.3390/toxins11110671.
32. Florens N, Calzada C, Delolme F, et al. Proteomic Characterization of High-Density Lipoprotein Particles from Non-Diabetic Hemodialysis Patients. *Toxins (Basel)*. 2019;11(11).pii.E671. doi:10.3390/toxins11110671.
33. Shao B, De Boer I, Tang C, et al. A Cluster of Proteins Implicated in Kidney Disease Is Increased in High-Density Lipoprotein Isolated from Hemodialysis Subjects. *J Proteome Res*. 2015;14(7):2792–806. doi:10.1021/acs.jproteome.5b00060.
34. Shao B, Zelnick LR, Wimberger J, et al. Albuminuria, the High-Density Lipoprotein Proteome, and Coronary Artery Calcification in Type 1 Diabetes Mellitus. *Arterioscler Thromb Vasc Biol*. 2019;39(7):1483–91. doi:10.1161/ATVBAHA.119.312556.
35. Wang K, Zelnick LR, Hoofnagle AN, et al. Alteration of HDL protein composition with hemodialysis initiation. *Clin J Am Soc Nephrol*. 2018;13(8):1225–33. doi:10.2215/CJN.11321017.
36. Zhu Z, Zhong C, Xu T, et al. Prognostic significance of serum cystatin C in acute ischemic stroke patients according to lipid component levels. *Atherosclerosis*. 2018;274:146–51. doi:10.1016/j.atherosclerosis.2018.05.015.
37. Kopecky C, Genser B, Drechsler C, et al. Quantification of HDL proteins, cardiac events, and mortality in patients with type 2 diabetes on hemodialysis. *Clin J Am Soc Nephrol*. 2015;10:224–31. doi:10.2215/CJN.06560714.
38. Kaseda R, Tsuchida Y, Yang HC, et al. Chronic kidney disease alters lipid trafficking and inflammatory responses in macrophages: Effects of liver X receptor agonism. *BMC Nephrol*. 2018;19(1):17. doi:10.1186/s12882-018-0814-8.
39. Cornoni-Huntley J, Ostfeld AM, Taylor JO, et al. Established populations for epidemiologic studies of the elderly: study design and methodology. *Aging (Milano)*. 1993;5(1):27–37. doi:10.1007/bf03324123.

40. Corti MC, Guralnik JM, Salive ME, et al. Clarifying the direct relation between total cholesterol levels and death from coronary heart disease in older persons. *Ann Intern Med.* 1997;126(10):753–60. doi:10.7326/0003-4819-126-10-199705150-00001.
41. Yamamoto S, Yancey PG, Ikizler TA, et al. Dysfunctional high-density lipoprotein in patients on chronic hemodialysis. *J Am Coll Cardiol.* 2012;60(23):2372–9. doi:10.1016/j.jacc.2012.09.013.
42. Meier SM, Wultsch A, Hollaus M, et al. Effect of chronic kidney disease on macrophage cholesterol efflux. *Life Sci.* 2015;136:1–6. doi:10.1016/j.lfs.2015.06.005.
43. Maeba R, Kojima K, ichiro, Nagura M, et al. Association of cholesterol efflux capacity with plasmalogen levels of high-density lipoprotein: A cross-sectional study in chronic kidney disease patients. *Atherosclerosis.* 2018;270:102–9. doi:10.1016/j.atherosclerosis.2018.01.037.
44. Ganda A, Yvan-Charvet L, Zhang Y, et al. Plasma metabolite profiles, cellular cholesterol efflux, and non-traditional cardiovascular risk in patients with CKD. *J Mol Cell Cardiol.* 2017;112:114–22. doi:10.1016/j.yjmcc.2017.05.001.
45. MacHowska A, Sun J, Qureshi AR, et al. Plasma pentosidine and its association with mortality in patients with chronic kidney disease. *PLoS One.* 2016;11(10):e0163826. doi:10.1371/journal.pone.0163826.
46. Holzer M, Gauster M, Pfeifer T, et al. Protein carbamylation renders high-density lipoprotein dysfunctional. *Antioxidants Redox Signal.* 2011;14(12):2337–46. doi:10.1089/ars.2010.3640.
47. Holzer M, Zangger K, El-Gamal D, et al. Myeloperoxidase-derived chlorinating species induce protein carbamylation through decomposition of thiocyanate and urea: Novel pathways generating dysfunctional high-density lipoprotein. *Antioxidants Redox Signal.* 2012;17(8):1043–52. doi:10.1089/ars.2011.4403.
48. Rysz J, Gluba-Brzózka A, Rysz-Górzyńska M, Franczyk B. The Role and Function of HDL in Patients with Chronic Kidney Disease and the Risk of Cardiovascular Disease. *Int J Mol Sci.* 2020;21(2):601. doi:10.3390/ijms21020601. Published 2020 Jan 17.

Tables

Table 1. Anthropometric and clinical data of control subjects and individuals with DKD categorized according to the glomerular filtration rate (GFR; mL/min/1.73 m²) and stage of urinary albumin excretion (A1 = normoalbuminuria, A2 = microalbuminuria; A3 = macroalbuminuria).

	n (F/M)	Age (years)	Time of DM (years)	CVD history (%)	IMC (Kg/m ²)	HbA1c (%)	Fructosamine (μmol/L)	TC (mg/dL)	TG (mg/dL)	HDLc (mg/dL)	CML (μg/mL)
Control		70.0 ±	-	-	24.8 ± 2.5	5.5 ±	227 ± 21	210 ±	137 ±	57 ± 21	
GFR > 60	8 (4/4)	8.6				0.2		47	95		0.5 (0.3 - 1.0)
DM		65.0 ±	13.8 ±		33.4 ± 6.0	8.1 ±	305 ± 68	143 ±	97 ± 28	52 ± 17	
GFR > 60 + A1 and A2	10 (7/3)	7.8	6.4	2 (20 %)	* 33.4 ± 6.0	0.6*		21*			1.7 (0.5 - 3.4)
DM		67.4 ±	17.8 ±		29.0 ± 5.0	8.3 ±	347 ± 71 *	174 ±	148 ±	47 ± 22	
GFR < 60 + A3	25 (7/18)	8.0	6.5	11 (44 %)		0.9		54	64		1.0 (0.7 - 7.3)

BMI = body index mass; HbA1c = glycated hemoglobin; TC = total cholesterol; TG = triglycerides; HDLc = HDL cholesterol (mean ± SD); CML = carboxymethyl-lysine (median - range). Results were compared by one-way ANOVA with Dunnett's post-test; * p < 0.05 in comparison to Control GFR > 60.

Figures

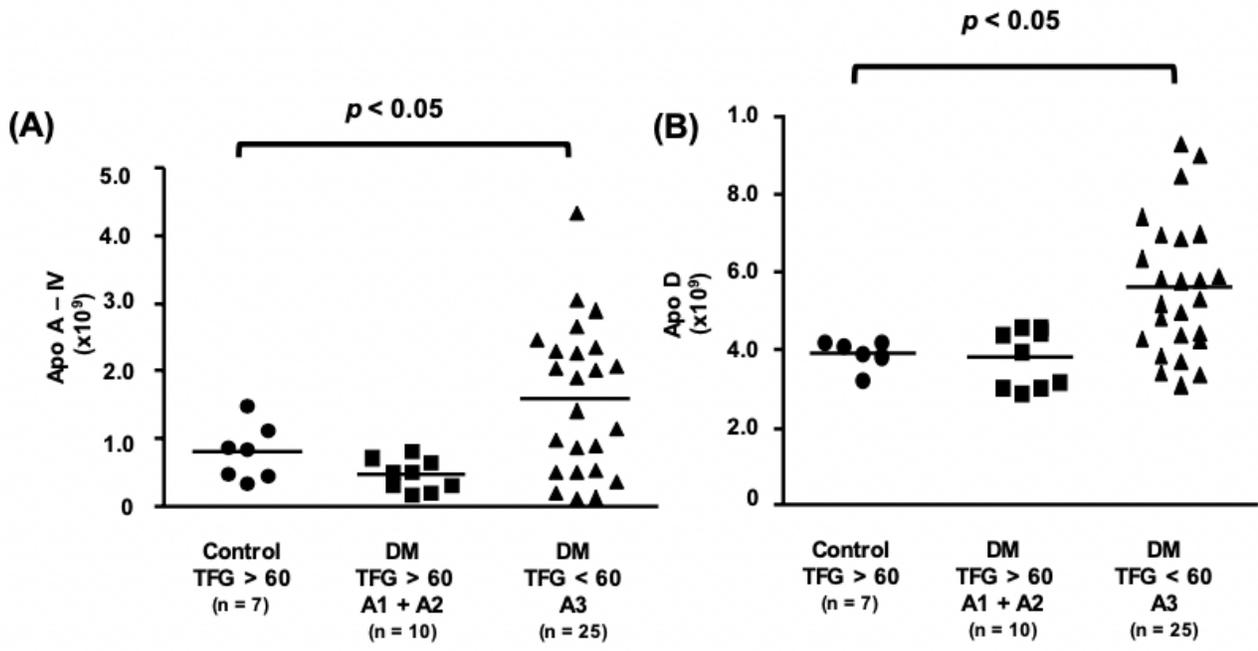


Figure 1

HDL proteome. HDL proteins were quantified by targeted proteomics (panels A to B). Comparisons were made with Prism software by one-way ANOVA, with Dunnett's post-test.

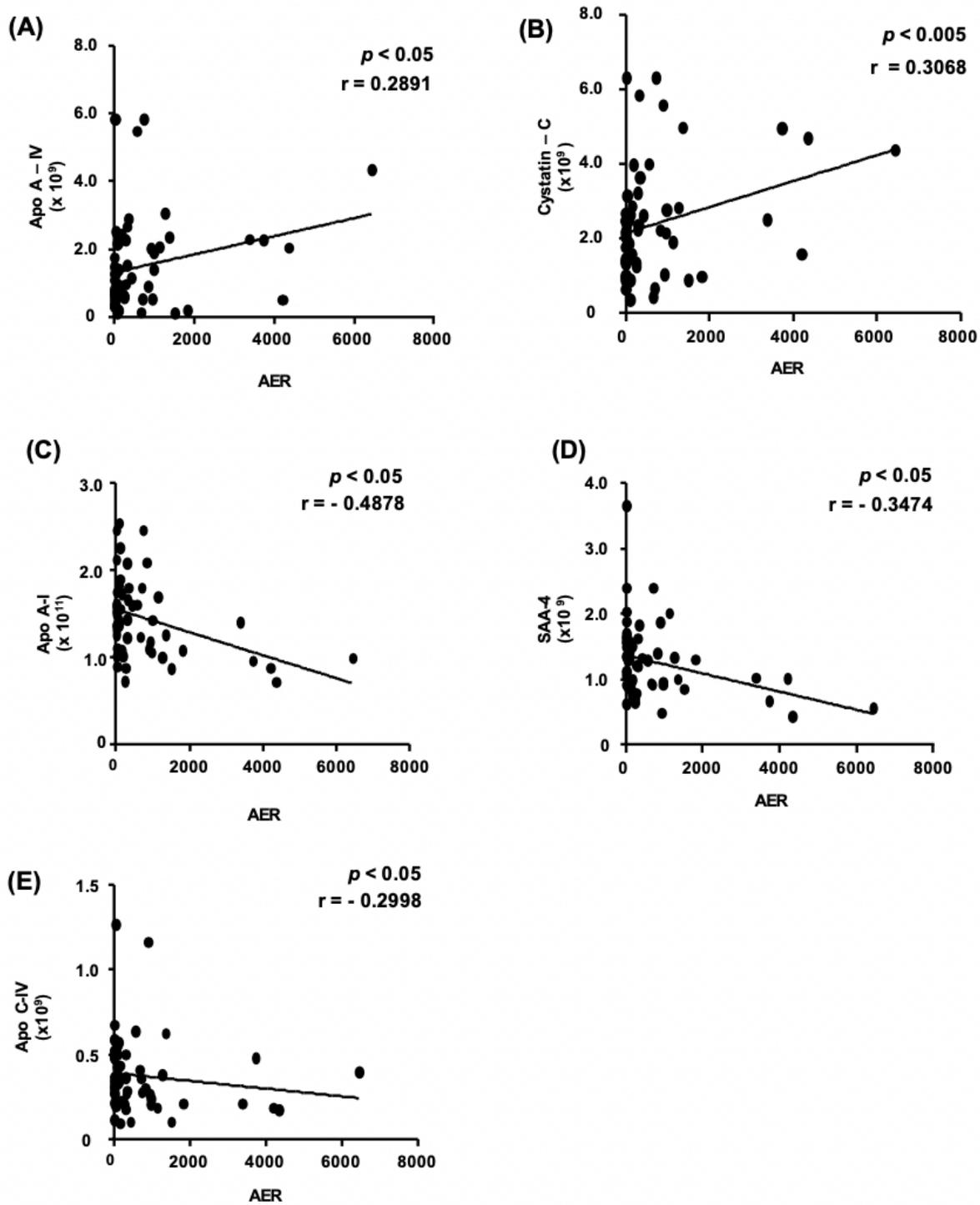


Figure 2

Correlation of HDL proteome with albumin excretion rate Correlations between HDL proteome with the AER (panels A to C) were performed using Spearman correlation.

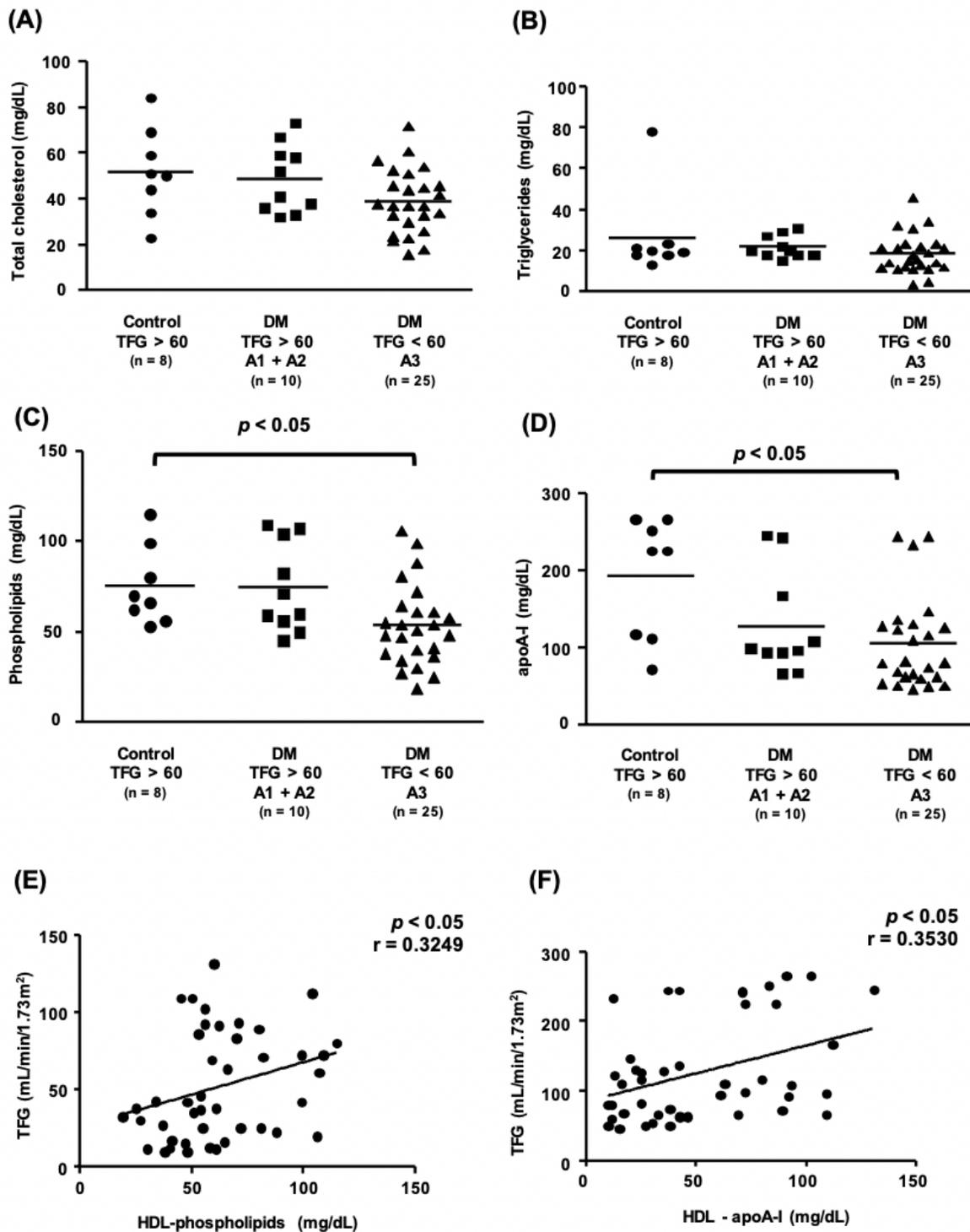


Figure 3

HDL composition. The HDL content of TC, TG and PL was determined by colorimetric enzymatic methods and apoA-I, by immunoturbidimetry (panel A to D). The results were compared by one-way ANOVA with Dunnett's post-test. Associations between HDL components with GFR (panels E to F) were performed by Spearman's correlation.

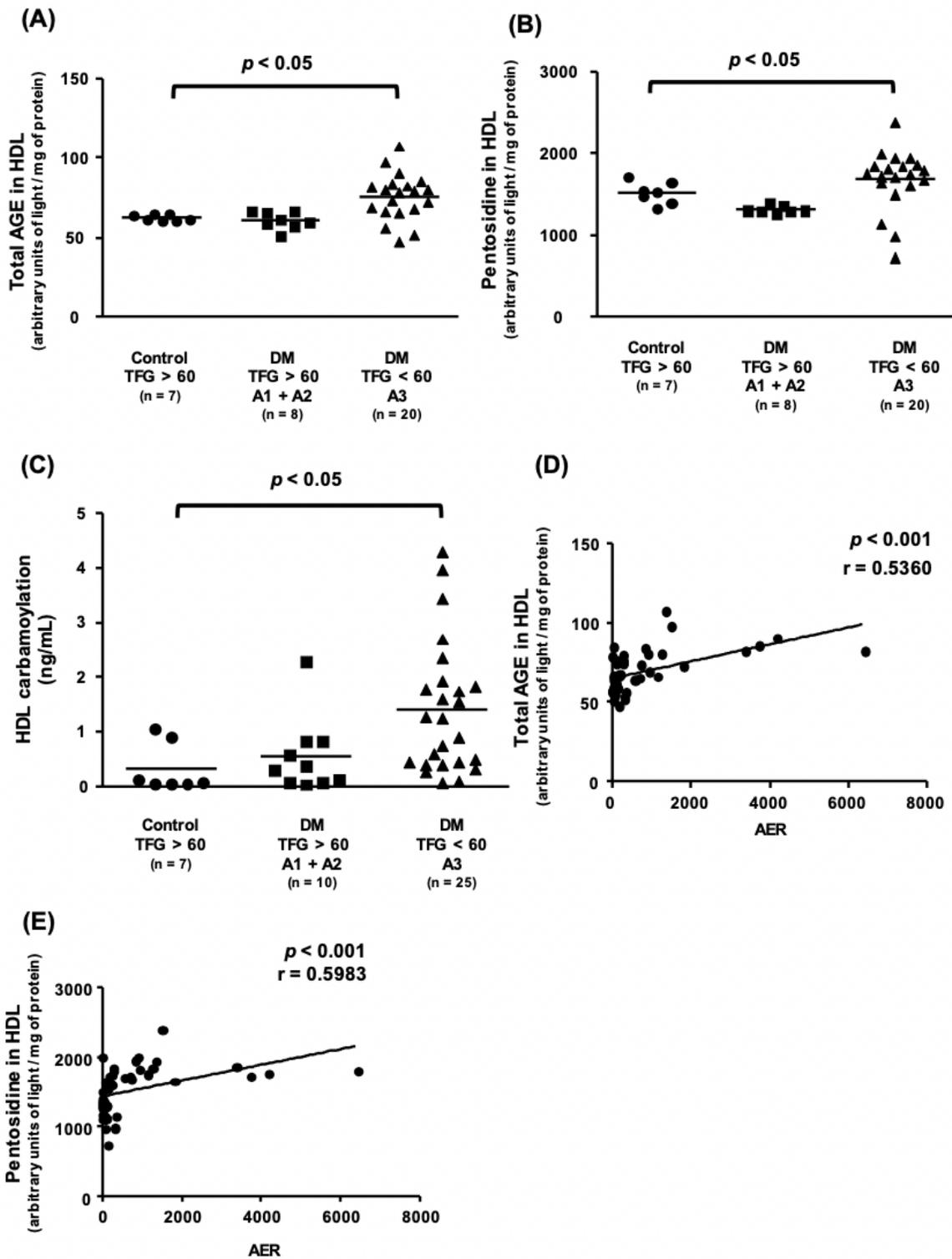


Figure 4

HDL modification by advanced glycation and carbamoylation. The amount of total AGE (panel A), and pentosidine (panel B) was determined in HDL by measuring the absorbance in the fluorescence range at 440 nm (total AGE) and 378 nm (pentosidine) and carbamoylation (panel C), by ELISA. The results were compared by one-way ANOVA with Dunnett's post-test. Associations between HDL chemical modification with AER (panels D and E) were performed by Spearman's correlation.

Figure 5

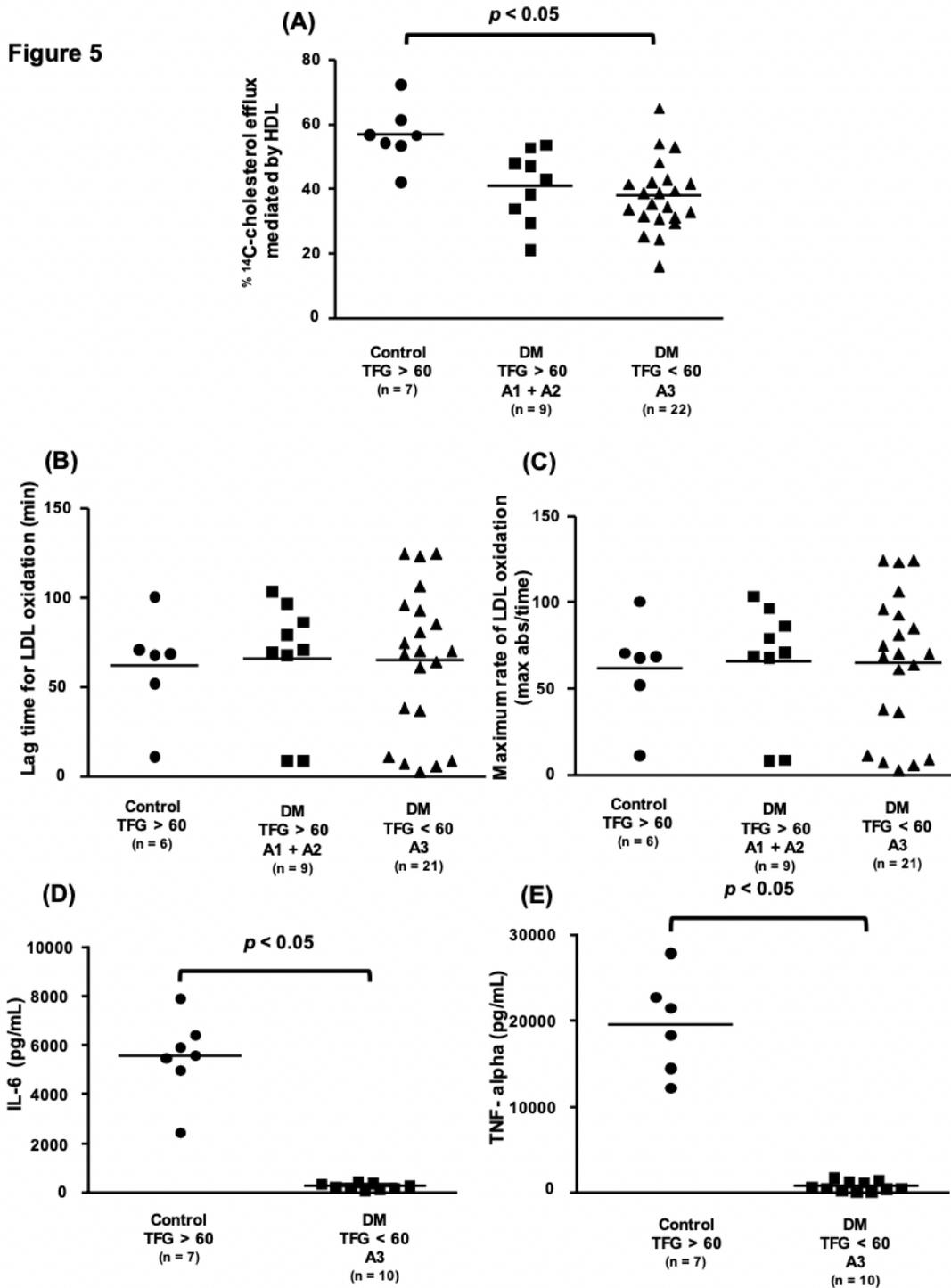


Figure 5

HDL functionality. (panel A) cholesterol efflux: HDL was isolated from DKD and control subjects and utilized as acceptor of cellular cholesterol. Bone marrow-derived macrophages (BMDM) overloaded with acetylated LDL and ¹⁴C-cholesterol were incubated with 50 μg of HDL / mL of medium. Cholesterol efflux was determined after measuring the radioactivity in the culture medium and that remaining in cells, being calculated as: ¹⁴C-cholesterol in the medium / ¹⁴C-cholesterol in the medium + ¹⁴C-cholesterol in cell x

100. Control incubations were performed in the presence of the DMEM/FAFA in the absence of HDL and the results subtracted from those obtained in the presence of HDL. (panels B and C) antioxidant activity: the lag time (panel B) and the maximum rate of LDL oxidation (panel C) was determined in incubations with LDL isolated from a healthy donor with CuSO₄ solution and HDL from DKD or controls. (panels D and E) antiinflammatory activity: BMDM overloaded with acetylated LDL were treated with HDL for 24 h. After washing, cells were treated with LPS for 24 h and interleukin 6 (IL-6, panel D) and TNF-alfa (panel E) were determined in the medium by ELISA. Results were compared by one-way ANOVA with Dunnett's post-test or Student t test.

Supplementary Files

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