

Bladder malignant granular cell tumor with EP300 gene mutation: a case report and literature review

Jian Huang (✉ 18665763598@163.com)

Guangdong Medical University <https://orcid.org/0000-0001-8117-1028>

Di Zhu

Guangdong Medical University

Xinle Ren

Guangdong Medical University

Bing Huang

Guangdong Medical University

Case report

Keywords: Malignant granular cell tumor, Bladder, Histopathology, EP300, Next-generation high throughput sequencing

Posted Date: May 3rd, 2021

DOI: <https://doi.org/10.21203/rs.3.rs-337088/v2>

License: © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Abstract

Background Malignant granular cell tumor (GCT) is extremely rare. Malignant GCT with EP300 gene mutation in the bladder has not been reported in the literature.

Case presentation We report a special case of 45 years old female with malignant GCT of the bladder. Physical examination found a pelvic mass in the patient. Magnetic resonance imaging showed a huge mass between the posterior wall of the bladder, the cervix, and the anterior wall of the vagina. Pathological examination showed that the mass was 11×11×4.5cm in size, involved in the bladder's posterior wall. Under the microscope, the tumor cells were arranged in the shape of a nest or cord to infiltrate the bladder's wall. The tumor cells were pleomorphic, red-stained granular within the cytoplasm, with increased nuclear/cytoplasmic ratio, vacuolar nuclei, and obvious nucleoli. The tumor cells were showed obvious nuclear atypia, and the mitosis was more than 5/50HPF. Coagulative necrosis was widely showed within the tumor. Immunohistochemistry(IHC) showed that S-100, NSE, CD68, CR, α-AT, and TFE-3 were strongly positive, and the Ki-67 proliferation index was around 15%. The next-generation high throughput sequencing indicated that EP300 gene was missense mutated (c.457A>G) with 33% mutation abundance, and genes of DPYD(c.1627A>G),ERCC1(c.354T>C),NQO1(c.559C>T),TPMT(c.719A>G) and XRCC1(c.1196A>G) were polymorphic mutated. The patient died after three months of the second surgical treatment.

Conclusions We report for the first time a primary bladder malignant GCM accompanied by mutations in special driving genes such as EP300. We also conducted a comprehensive literature review and an in-depth discussion.

Introduction

Granular cell tumor (GCT) is a benign tumor thought to originate from nerve sheath cells. It accounts for about 2% of soft tissue tumors and could occur in any part of the body[1-5]. The most common sites are the mouth, skin, soft tissue, breast, and female reproductive organs. GCT occurred in the bladder is rare, and only a few cases have been reported[6-9]. Malignant GCT is an extremely rare soft tissue malignant tumor, accounting for less than 2% of all soft tissue GCTs. Malignant GCT occurs in a wide range of ages, mostly in adults aged 30-70, with an average age of 50. Women are more common. Most of the clinical manifestations are painless masses in subcutaneous or deep soft tissue. Most of the cases occurred in the extremities, torso, head, neck, and breast[10-16]. To date, few cases of bladder malignant GCT have been reported[7, 17]. Few studies on the driving genes of malignant GCT, especially the specific driving genes of malignant GCT, are not clear[18-20]. Here, we reported a case of malignant bladder GCT with vulvar, vaginal, and pelvic soft tissue invasion. Histopathology and IHC confirmed that it was a malignant GCT originating from the bladder. The next-generation high throughput sequencing detected several gene mutations, including a tumor-specific driving gene EP300 and another five genes associated with chemotherapy. We also conducted a comprehensive literature review and in-depth discussion.

Case Presentation

The patient was a 45-year-old Chinese female, housewife and Han nationality. The patient found pelvic mass during physical examination a year ago, the clinical manifestations were intermittent lower abdominal discomfort, frequent urination, dysuria, no hematuria, no vaginal bleeding and so on. The outpatient clinic was admitted to the hospital with "pelvic mass". The patient was without any treatment before. Physical examination found a pelvic mass, did not care, untreated. Then intermittent lower abdominal discomfort, frequent urination, dysuria, no hematuria, no vaginal bleeding, and so on. B-ultrasound examination showed that the left side of the uterus was abnormally hypoechoic (10.4×9.6cm), the right ovary had no obvious abnormal echo, and the left ovary showed unclearly. Then she comes to our hospital, and the outpatient clinic is admitted to the hospital as a "pelvic mass". The patient was treated with abdominal hysterectomy and double salpingectomy, pelvic tumor resection, cystectomy, and ureterostomy under general anesthesia.

Magnetic resonance imaging (MRI) pelvic plain scanning showed that a large lobulated mass could be seen among the cervix, the anterior wall of the vagina, and below the bladder. The boundary of the mass was still clear. The range of the mass was about 12×11×10cm in size. T1W1 showed a slightly higher signal, T2W1 showed a high-low mixed-signal, and DWI showed a mixed high signal. An enhanced scan showed obvious uneven enhancement, uterine body, and cervical lifting. The bladder was connected tightly to the mass and compressed forward and upward (Figure 1). A mass in the pelvis that lies between the uterus and the bladder and is closely connected to the base of the bladder and the wall of the uterus. After a complete hysterectomy, a huge mass was found between the anterior wall of the vagina and the posterior wall of the bladder, which was hard and fixed at the pelvic. The anterior boundary of the tumor was not separated from the posterior wall of the bladder and protruded into the bladder. When the bladder was completely dissociated from the extraperitoneal cavity, it was seen that the tumor originated from the posterior wall of the bladder and continued down to the anterior wall of the urethra to the pelvic floor.

The postoperative specimens were carefully observed to describe the size, color, texture, and other mass characteristics. The sampling processes, dehydration, embedded in paraffin, cut slides, and hematoxylin and eosin (H&E) staining were performed according to the requirements. A total cystectomy was performed with a volume of 15×12×5cm in whole tissues. A volume of 11×11×4.5cm in tumor size was seen at the posterior wall of the bladder. The section of the tumor was grayish-yellow, tough, and lobulated. The tumor involved the whole layer of the bladder wall, did not break through the bladder adventitia, and the boundary between the tumor and the surrounding tissue was still clear (Figure 2).

The tumor tissue was separated by fibrous connective tissue, arranged in flake, nest and cord shape, showing infiltrative growth. The tumor cells were pleomorphic, round, polygonal, and fusiform. The ratio of the nucleus to the cytoplasm was increased. The cytoplasm was rich in red staining with a large number of eosinophilic bodies. The nuclei were heteromorphic with different sizes. Some of the nuclei

were vacuolated and had obvious nucleoli. Multinucleated tumor cells and scattered coagulative necrotic foci were seen locally, and the mitotic figure was more than 5/50HPF(Figure 3).

DAKO company's latest Omnis automatic immunohistochemical instrument was used for staining, and the process was carried out under the operation manual provided by the manufacturer. The sources, clones, and manufacturer of the antibodies used are shown in Table 1. Negative and positive controls were all set up for each antibody. The IHC results showed that S-100, NSE, CD68, CR, α -AT, and TFE-3 in the tumor cells were strongly positive, and the Ki-67 proliferation index in most of the tumor areas was around 15% (Figure 4). The IHC results showed that SMA, Desmin, α -inhibin, GFAP, MyoD1, HMB45, and CK \times pan \times were all negative(not shown).

Table 1 The sources, clones and manufacturer of the antibodies used in this study

Antibodies	Clones	Manufacturers
CD68	KP1	MXB
NSE	3-3-C	MXB
S-100	4C4.9	MXB
CR (Calretinin)	MX027	MXB
α -AT (AAT)	Polyclone	MXB
TFE-3	MRQ-37	MXB
SMA	1A4	MXB
Desmin	MX046	MXB
α -inhibin	AMY82	MXB
GFAP	MX047	MXB
Ki-67	MIB-1	Dako
MyoD1	5.8A	MXB
HMB45	HMB45	MXB
CK (pan)	AE1/AE3	Dako

Next-generation high-throughput sequencing (NGS \times covered 425 gene exons, fusion-related introns, variable splicing regions, and specific microsatellite (MS) sites, and other common 1.46Mb base sites was used in this study. The detection results included point mutation, small fragment insertion and deletion mutation, gene fusion and copy number variation, microsatellite (MS) analysis, and tumor mutational burden (TMB). The whole sequencing process was completed on the Illumina-based Hiseq 4000 \times performed at Nanjing Geneseeq Technology Inc., Nanjing, Jiangsu, China. The 425 genes

sequenced include five functional partitions, which are key genes for targeted treatments, key genes for chemotherapy, genetic risk prediction genes, important tumor-driving genes, but there are no targeted drugs at present and key genes in the tumor-related signaling pathways. In the sequencing of the 425 genes, the results of NGS indicated that the overall mutation rate is very low. The results revealed that the EP300 gene was missense mutated (c.457A>G) with 33% mutation abundance. The gene of EP300 is in the classification of "key genes in the tumor-related signaling pathways". The NGS results also showed that genes of DPYD(c.1627A>G), ERCC1 (c.354T>C), NQO1(c.559C>T), TPMT(c.719A>G), and XRCC1(c.1196A>G) were polymorphic mutated, which these genes are all in the classification of "key genes for chemotherapy". The detailed locations and functions as shown in Table 2. NGS results also showed that the tumor mutational burden (TMB) was very lower (2.1 mutations/Mb) and the microsatellite (MS) analysis showed that no MIS-H related genes were detected. The list of the 425 genes and functional partition was shown in supplement data (Table 3).

Table 2 The mutations of genes detected by NGS

Genes	Variations	Mutant types	Ways of mutation	Possible functions
EP300	p.M153V	c.457A>G (p.M153V)	Missense mutation in exon 2 (33% in MF*)	Possible tumor driving genes
DPYD	p.I543V	c.1627A>G(p.I543V)	Heterozygous polymorphism	Side effects may increase for drugs of 5-FU, capecitabine, etc.
ERCC1	p.N118N	c.354T>C(p.N118=)	Homozygous polymorphism	The curative effect by using drugs of platinum may be better than that of the wild type.
NQO1	p.P187S	c.559C>T(p.P187S)	Heterozygous polymorphism	The curative effect by using mitomycin C may be worse than that of the wild type.
TPMT	p.Y240C	c.719A>G(p.Y240C)	Heterozygous polymorphism	Toxic side effects by using mercaptopurine may increase
XRCC1	p.Q399R	c.1196A>G(p.Q399R)	Homozygous polymorphism	The curative effect by using drugs of platinum may be better than that of the wild type.

*** Mutation Frequency**

Three months after the first operation, the patient was re-admitted to the hospital because of "vaginal mass found for more than one week". Specialist examination: the right vaginal wall can reach a mass with a diameter of about 3cm, lobulated, moveable, tenderness, clear boundary, close to the orifice of the vulva. What was seen during the operation: the vulvar mass was located in the lower part of the vagina behind the right greater labia, lobulated, with a volume of about 4×3×1 cm, with a clear boundary and hard nature. MRI indicates local recurrence of the tumor. The pathological diagnosis after the second operation was consistent with malignant GCT with vulvar and cervical invasion. The patient died three months after the second operation.

Discussion And Conclusions

The diagnostic criteria for malignant GCT were proposed by Fanburg-Smith and his colleagues as early as 1998. Malignant histological criteria included coagulative necrosis, fusiform, vesicular nucleus of large nucleolus, increased mitotic activity (>2 mitoses/high magnification ×200), increased nuclear/cytoplasmic ratio, and pleomorphism[21]. According to the pathological criteria for the diagnosis of malignant GCT, based on both histopathology and immunohistochemical expression of this presented case, it was no doubt a malignant GCT[22]. In addition to the criteria of histopathology, in this case, we also identified biological changes associated with malignant characteristics, such as invasive growth and vascular invasion. The differential diagnosis of benign and malignant GCT is often tricky in both pathology and clinic. However, this case is not difficult, based on histopathology and biological behavior, and other aspects. The tumor originated from the bladder wall and invaded the muscular layer of the bladder. In addition, the tumor also widely invades the vagina, cervix, and soft tissues in the pelvic, showing extremely malignant biological behavior. The patient did not respond to chemotherapy and died three months after the second operation.

Malignant GCM is extremely rare, and few cases have been reported [7, 11, 17, 23, 24]. So far, we know little about the molecular biology and molecular genetics of malignant GCT. Recently, Richard Davis et al. reported a case of malignant GCT of the lung, the NGS indicated that there was no mutation associated with lung adenocarcinoma, but the ATM gene mutation was found. ATM gene encodes DNA repair protein involved in cell cycle regulation[18]. Xu and colleagues apply the NGS to identify mutations in the genes of ASXL, NOTCH, and PARP pathways as potential driver mutations in pulmonary malignant GCT[25].

In our case, based on the high-throughput second-generation sequencing of 425 genes, it was found that the overall mutation rate of the tumor was very low. In the classification of "the key genes in tumor-related signaling pathway", the EP300 gene was missense mutated (C.457A > G) with 33% mutation abundance. As we know, the EP300 gene is located on chromosome 22, and encodes adenovirus E1A-related intracellular p300 protein, which has acetyltransferase activity, regulates transcriptional activity through chromosome remodeling. The EP300 and its homolog CREB binding protein (CBP) are two lysine acetyltransferases, which act as transcriptional factors and play an essential role in developing, progression, and treating many diseases, especially in tumors. Some cancer-related point mutations are gain-of-function alterations in EP300/CBP that might also contribute to cancer development[26-28]. It has been found that the inactivation mutations of histone acetyltransferase CBP and EP300 are prevalent in diffuse large B cell lymphoma (DLBCL). The functional inactivation mutations of CBP and EP300 will down-regulate the expression of CD20, which leads to the failure of anti-CD20 antibody therapy[29]. Other studies have also shown that targeting EP300 may effectively treat fatal leukemia[30]. Therefore. The use of synthetic small-molecular compounds to inhibit the function of mutant EP300 brings hope for targeted therapy of malignant tumors[31]. A recent study also shows that the EP300 protein is related to the grade, pathological T stage, lymph node metastasis, and survival rate of esophageal squamous cell carcinoma[32].

Gongmin Zhu and colleagues found that EP300 mutation in bladder cancer is associated with a higher tumor mutation burden (TMB) and up-regulates the immune system's signal pathway, which can be used

as a biomarker to predict the effect of immunotherapy[33]. In our case, NGS results showed that the TMB was significantly lower (2.1 mutations/Mb). The relationship between EP300 mutation and TMB may be related to different tumor types and mutation sites. How genetic or epigenetic changes in EP300 will affect the phenotype of various cancers is far from clear.

In our case, in addition to the EP300 missense mutation, it was also detected that DPYD Δ NQO1 and TPMT had heterozygous polymorphism mutations, that ERCC1 and XRCC1 had homozygous polymorphism mutations, which these genes are all in the classification of “ the key genes for chemotherapy” from total 425 genes. Heterozygous mutation of the DPYD gene in p. I543V is a missense mutation caused by single nucleotide polymorphism rs1801159, which can decrease the activity of DPYD, increase the toxicity and side effects of 5-fluorouracil, and increase the risk of vomiting and nausea in patients[34]. ERCC1 gene p.N118N homozygous mutation is a synonymous mutation caused by single nucleotide polymorphism rs11615. This homozygous genotype can increase the risk of lung cancer and breast cancer. Besides, this mutation can reduce mRNA level by affecting the stability of ERCC1 transcripts and increase cell response to platinum drugs by reducing DNA repair function[35]. TPMT gene p.Y240C heterozygous mutation (TPMT*3C) is a missense mutation caused by single nucleotide polymorphism rs1142345. This heterozygous polymorphism mutation can reduce mercaptopurine methyltransferase activity by about 50%, which leads to serious poor metabolism of mercaptopurine and side effects[36]. XRCC1 gene p.Q399R homozygous mutation is a missense mutation caused by single nucleotide polymorphism rs25487, which is associated with the risk of non-small cell lung cancer, breast cancer, colorectal cancer, gastric cancer, and other tumors. This homozygous mutation is related to platinum chemotherapeutic drugs' efficacy and can increase the response rate of cells to platinum drugs[37].

There is no targeted drug for malignant GCT, and whether it is sensitive to chemotherapy may be related to its particular gene mutation. Wei, L, and colleagues performed whole-genome sequencing of a malignant GCT that was metabolically responsive to pazopanib. The results showed that the mutation rate was shallow, and the whole genome was stable[20]. Due to the small number of cases, there are not many reports on the survival of malignant GCM. However, it is generally believed that it is closely related to the tumor's size, local invasion, and distant metastasis[24, 38]. It took less than a year from discovery to death in our case.

In summary, few cases of malignant GCM of the bladder have been reported, but none of them have been studied by molecular genetics. We performed molecular genetic analysis of malignant GCM of the bladder for the first time, and in particular, detected a missense mutation in exon 2 of EP300 gene p.M153V with a high mutation abundance. The clinical significance of EP300 gene mutation in malignant GCM of the bladder is not clear. Whether it leads to the EP300 protein dysfunction to activate downstream signal pathways and participating in tumorigenesis and development, more clinical cases and experimental studies are needed to confirm them.

Abbreviations

GCT: Granular cell tumor

NGS: Next-generation high throughput sequencing

MRI: Magnetic resonance imaging

IHC: Immunohistochemistry

H&E: Hematoxylin and Eosin

MS: Microsatellite

TMB: Tumor Mutational Burden

CBP: CREB binding protein

DLBCL: Diffuse large B cell lymphoma

Declarations

Ethics approval and consent to participate

This case was reviewed and approved by the Ethics Committee of Affiliated Hospital, Guangdong Medical University, China.

Consent for Publication

Written informed consent was obtained from the patient's next of kin for publication of this case report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

Availability of data and materials

All data generated and described in this article are freely available to any scientist wishing to use them for noncommercial purposes, without breaching participant confidentiality.

Competing interests

The author(s) declared no potential conflicts of interest for the research, authorship, and/or publication of this article.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

Authors' contributions

J.H. found the specific case ; J.H. and B.H., two senior diagnostic pathologist made the final pathological diagnosis for the case; J.H. and D.Z. wrote the main manuscript text; D.Z., XL.R. and J.H. prepared figures 1-4; All authors reviewed the manuscript.

Acknowledgements

Not Applicable

References

- [1]. Davis, R., K. Deak and C.H. Glass, Pulmonary Granular Cell Tumors: A Study of 4 Cases Including a Malignant Phenotype. *Am J Surg Pathol*, 2019. 43(10): p. 1397-1402.
- [2]. Stemm, M., et al., Typical and Atypical Granular Cell Tumors of Soft Tissue: A Clinicopathologic Study of 50 Patients. *Am J Clin Pathol*, 2017. 148(2): p. 161-166.
- [3]. Rubino, F., et al., Granular cell tumors of the sellar region: what should be done after subtotal resection? A systematic review. *Pituitary*, 2020. 23(6): p. 721-732.
- [4]. Aoyama, K., et al., Granular cell tumors: a report of six cases. *World J Surg Oncol*, 2012. 10: p. 204.
- [5]. Shi, Y., et al., Experience with Esophageal Granular Cell Tumors: Clinical and Endoscopic Analysis of 22 Cases. *Dig Dis Sci*, 2020.
- [6]. Lee, C., et al., Clinicopathologic Features of Benign Neurogenic Tumor of Urinary Bladder. *Int J Surg Pathol*, 2018. 26(3): p. 221-228.
- [7]. Sun, Y., et al., Granular Cell Tumor of the Bladder: A Report of Six Cases. *Urology*, 2018. 121: p. 203.e1-203.e5.
- [8]. Yoshida, T., et al., Granular cell tumor of the urinary bladder. *Int J Urol*, 2001. 8(1): p. 29-31.
- [9]. Eandi, J.A., et al., Granular cell tumor of the urinary bladder with pseudoepitheliomatous hyperplasia and colocalization with adenocarcinoma. *Int J Urol*, 2007. 14(9): p. 862-4.
- [10]. Shah, K., et al., Malignant Granular Cell Tumor of the Lower Extremity in an Adolescent Male. *J Pediatr Hematol Oncol*, 2020.
- [11]. Stasek, M., et al., Malignant Granular Cell Tumor of the Esophagus: A Case Report. *Ann Thorac Cardiovasc Surg*, 2020. 26(6): p. 359-364.
- [12]. Bradford, B.E., et al., Benign and Malignant Granular Cell Tumor of the Hypopharynx: Two Faces of a Rare Entity. *Head Neck Pathol*, 2020.

- [13]. Tsukamoto, S., et al., Malignant granular cell tumor of the median nerve: a case report with a literature review of 157 cases. *Skeletal Radiol*, 2019. 48(2): p. 307-316.
- [14]. Jia, X., et al., Large malignant granular cell tumor with suprascapular nerve and brachial plexus invasion: A case report and literature review. *Medicine (Baltimore)*, 2017. 96(44): p. e8531.
- [15]. Akahane, K., et al., Malignant granular cell tumor of the breast: case report and literature review. *Breast Cancer*, 2015. 22(3): p. 317-23.
- [16]. Liu, T.T., et al., Primary cutaneous malignant granular cell tumor: a case report in China and review of the literature. *Diagn Pathol*, 2015. 10: p. 113.
- [17]. Zhuang, Y.Z., X.Y. Jiang and P.Q. Chen, [Malignant granular cell tumor of the urinary bladder]. *Zhonghua Bing Li Xue Za Zhi*, 2006. 35(3): p. 188.
- [18]. Davis, R., K. Deak and C.H. Glass, Pulmonary Granular Cell Tumors: A Study of 4 Cases Including a Malignant Phenotype. *Am J Surg Pathol*, 2019. 43(10): p. 1397-1402.
- [19]. Pareja, F., et al., Loss-of-function mutations in ATP6AP1 and ATP6AP2 in granular cell tumors. *Nat Commun*, 2018. 9(1): p. 3533.
- [20]. Wei, L., et al., Whole-genome sequencing of a malignant granular cell tumor with metabolic response to pazopanib. *Cold Spring Harb Mol Case Stud*, 2015. 1(1): p. a000380.
- [21]. Fanburg-Smith, J.C., et al., Malignant granular cell tumor of soft tissue: diagnostic criteria and clinicopathologic correlation. *Am J Surg Pathol*, 1998. 22(7): p. 779-94.
- [22]. Fanburg-Smith, J.C., et al., Malignant granular cell tumor of soft tissue: diagnostic criteria and clinicopathologic correlation. *Am J Surg Pathol*, 1998. 22(7): p. 779-94.
- [23]. Quinn, P.L., et al., Malignant Granular Cell Tumor of the Bile Duct. *ACG Case Rep J*, 2019. 6(8): p. e00193.
- [24]. Moten, A.S., et al., Malignant granular cell tumor: Clinical features and long-term survival. *J Surg Oncol*, 2018. 118(6): p. 891-897.
- [25]. Xu, S., et al., Next Generation Sequencing Uncovers Potential Genetic Driver Mutations of Malignant Pulmonary Granular Cell Tumor. *J Thorac Oncol*, 2015. 10(10): p. e106-9.
- [26]. Attar, N. and S.K. Kurdistani, Exploitation of EP300 and CREBBP Lysine Acetyltransferases by Cancer. *Cold Spring Harb Perspect Med*, 2017. 7(3).
- [27]. Breen, M.E. and A.K. Mapp, Modulating the masters: chemical tools to dissect CBP and p300 function. *Curr Opin Chem Biol*, 2018. 45: p. 195-203.

- [28]. Blobel, G.A., CREB-binding protein and p300: molecular integrators of hematopoietic transcription. *Blood*, 2000. 95(3): p. 745-55.
- [29]. Scialdone, A., et al., Depletion of the transcriptional coactivators CREB-binding protein or EP300 downregulates CD20 in diffuse large B-cell lymphoma cells and impairs the cytotoxic effects of anti-CD20 antibodies. *Exp Hematol*, 2019. 79: p. 35-46.e1.
- [30]. Huang, Y., et al., The Leukemogenic TCF3-HLF Complex Rewires Enhancers Driving Cellular Identity and Self-Renewal Conferring EP300 Vulnerability. *Cancer Cell*, 2019. 36(6): p. 630-644.e9.
- [31]. Yang, Y., et al., Discovery of Highly Potent, Selective, and Orally Efficacious p300/CBP Histone Acetyltransferases Inhibitors. *J Med Chem*, 2020. 63(3): p. 1337-1360.
- [32]. Bi, Y., et al., EP300 as an oncogene correlates with poor prognosis in esophageal squamous carcinoma. *J Cancer*, 2019. 10(22): p. 5413-5426.
- [33]. Zhu, G., et al., EP300 mutation is associated with tumor mutation burden and promotes antitumor immunity in bladder cancer patients. *Aging (Albany NY)*, 2020. 12(3): p. 2132-2141.
- [34]. Zhang, H., et al., DPYD*5 gene mutation contributes to the reduced DPYD enzyme activity and chemotherapeutic toxicity of 5-FU: results from genotyping study on 75 gastric carcinoma and colon carcinoma patients. *Med Oncol*, 2007. 24(2): p. 251-8.
- [35]. Nebert, D.W., et al., NAD(P)H: quinone oxidoreductase (NQO1) polymorphism, exposure to benzene, and predisposition to disease: a HuGE review. *Genet Med*, 2002. 4(2): p. 62-70.
- [36]. Almoguera, B., et al., Imputation of TPMT defective alleles for the identification of patients with high-risk phenotypes. *Front Genet*, 2014. 5: p. 96.
- [37]. Li, D., et al., Effects of base excision repair gene polymorphisms on pancreatic cancer survival. *Int J Cancer*, 2007. 120(8): p. 1748-54.
- [38]. Alnashwan, Y.A., K. Ali and S.S. Amr, Metastasizing Malignant Granular Cell Tumor (Abrikossoff Tumor) of the Anterior Abdominal Wall, with Prolonged Survival. *Case Rep Pathol*, 2019. 2019: p. 9576487.

Figures



Figure 1

MRI showed a large mass between the cervix, the anterior wall of the vagina, and the bottom of the bladder, and the boundary was still clear. A. A coronal section (The red arrow showed the extent of the tumor); B. A sagittal section (the red arrow showed the extent of the tumor, the yellow arrow showed the connection between the tumor and the bottom of the bladder.).

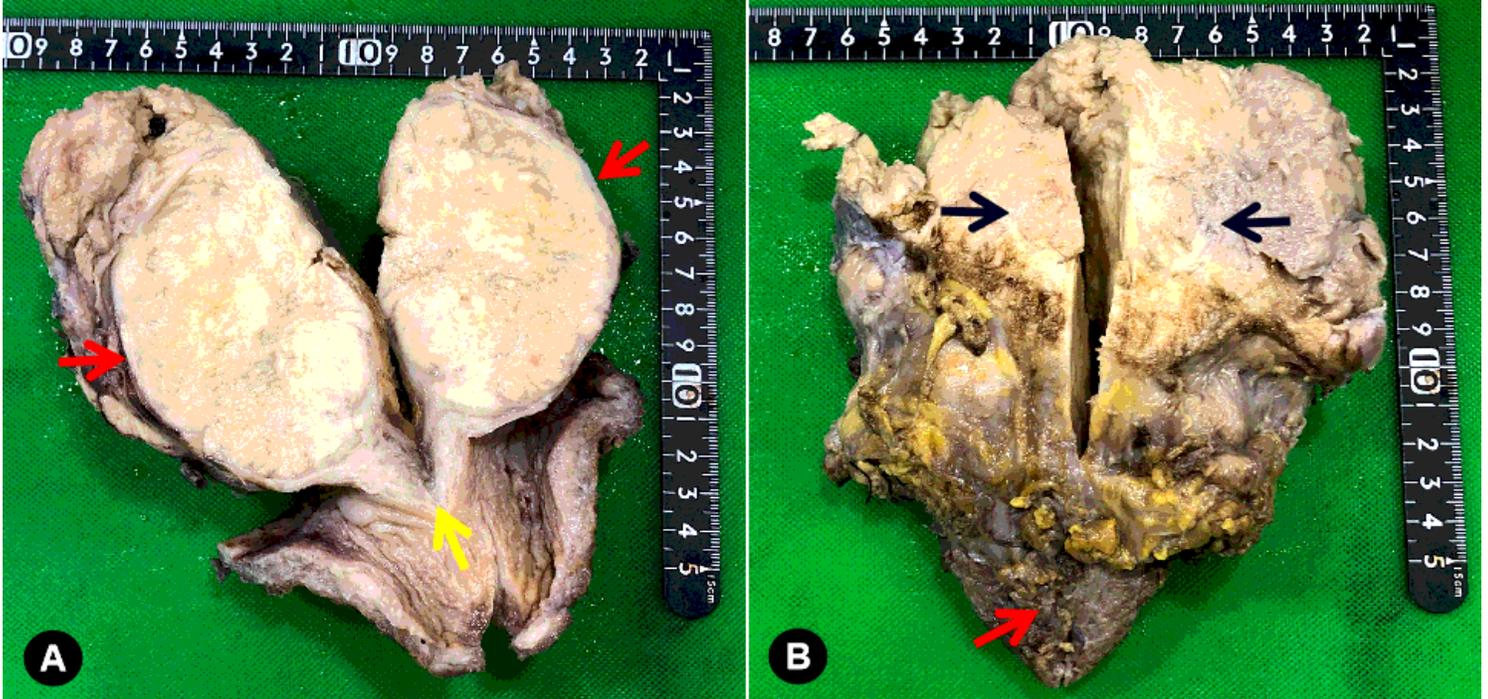


Figure 2

A volume of 11×11×4.5cm in tumor size was collected to the posterior wall of the bladder. A. The section of the tumor was grayish-yellow, tough, and lobulated. The red arrow showed the extent of the tumor, the

yellow arrow showed the connection between the tumor and the bottom of the bladder. B. Back view: The black arrow shows the adhesion between the tumor and the pelvic soft tissue. The red arrow showed the bottom of the bladder.

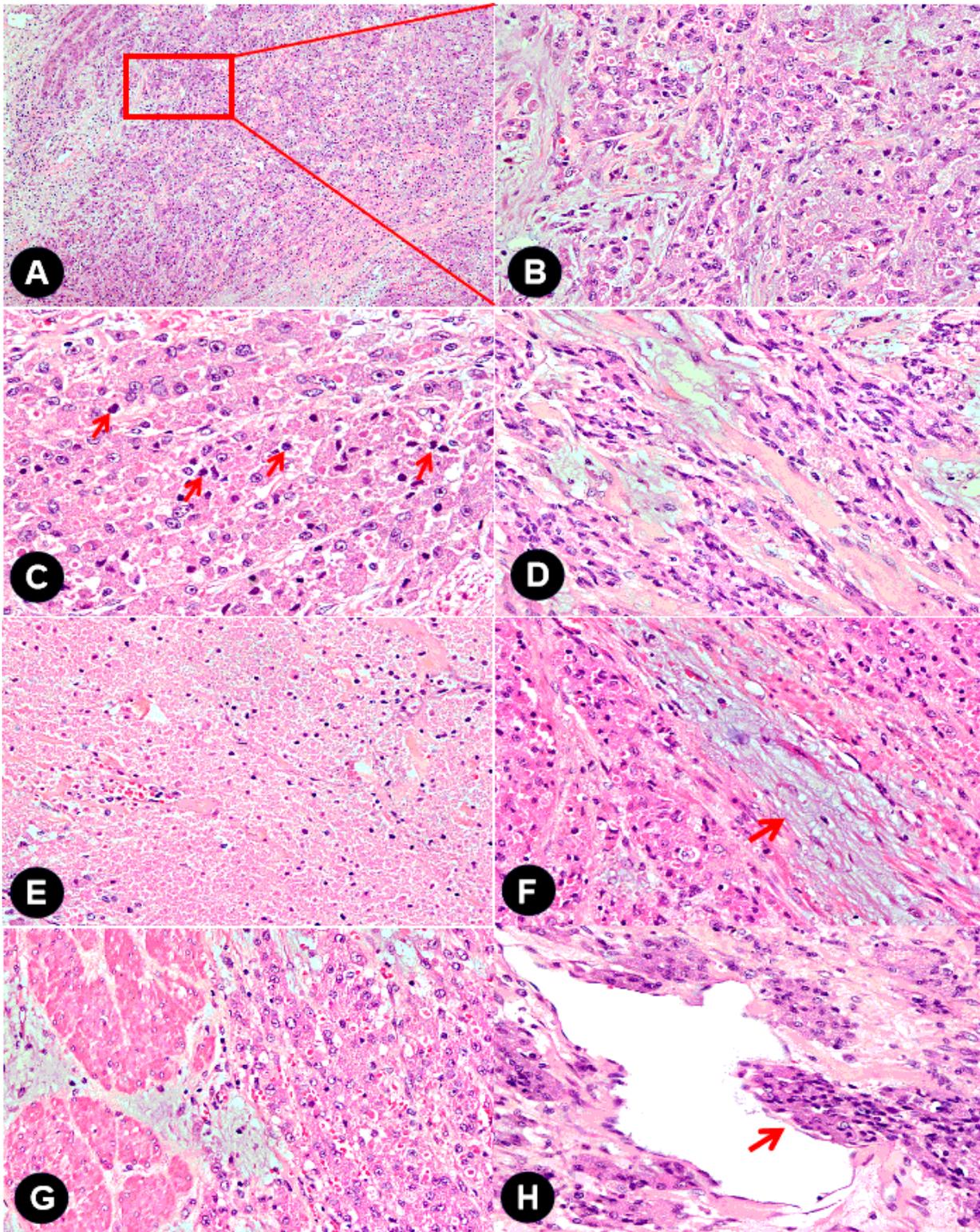


Figure 3

Under a microscope: A. Overall the histopathology. B. The tumor cells were pleomorphic, round, polygonal, and fusiform. The ratio of the nucleus to the cytoplasm was increased. The cytoplasm was rich in red

staining with a large number of eosinophilic bodies. C. The mitotic figure was more than 5/50HPF (shown by the red arrow). D. The tumor tissue was separated by fibrous connective tissue, arranged in flake, nest and cord shape. E. Coagulative necrosis. F. Interstitial myxoid degeneration shown by the red arrow. G. The tumor cells invaded the bladder muscle wall. H. The tumor cells invade the lumen of blood vessels (shown by the red arrow).

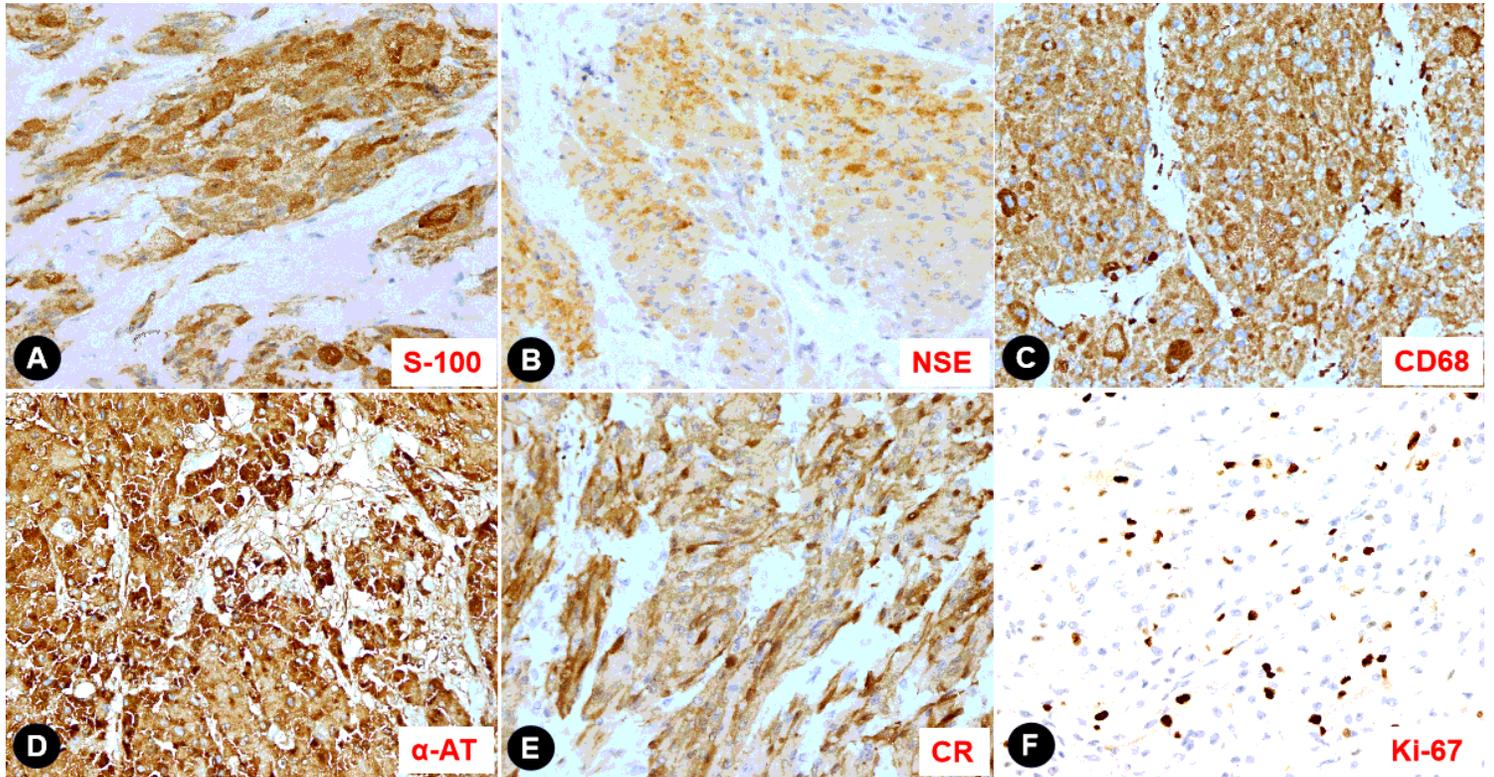


Figure 4

The IHC results showed that S-100 (A), NSE (B), CD68 (C), α-AT (D) (CR (E)), and TFE-3 in the tumor cells were strongly positive, and Ki-67 proliferation index in most of the tumor areas was around 15% (F).

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [OnlineSupplementdatatable31.png](#)
- [OnlineSupplementdatatable32.png](#)
- [CAREchecklist.pdf](#)