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SARS CoV 2 Nsp5 Protein Causes Acute Lung Inflammation, a Dynamical Mathematical Model

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29 **Abstract**

30 In the present work we propose a dynamical mathematical model of the lung cells inflammation
31 process in response to SARS-CoV-2 infection. In this scenario, our model suggests that the main
32 protease Nsp5 enhances the inflammatory process by increasing the levels of NF κ B, IL-6, Cox2,
33 and PGE2 with respect to a reference state without the virus. This overstimulated immune state
34 becomes autonomous of the signals from other immune cells, and does not shut down by itself
35 neither when the external signals are turned off. Our model suggests that Nsp5 is effectively the
36 switch to start inflammation, the consequent overproduction of the ACE2 receptor, and an
37 important underlying cause of the most severe cases of COVID19.

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1 Introduction

Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) virus is an intracellular infectious agent whose replication cycle depends on the host's cell structures and functions. In particular, it uses the translational apparatus of different types of infected cells to express its proteins (Nakagawa et al., 2016). SARS-CoV-2 causes the Coronavirus Disease 2019 (COVID19) that has infected over 117,000,000 persons and killed over 2,400,000 worldwide since the end of 2019. No specific and effective therapeutic drugs to defeat SARS-CoV-2 infection have been developed yet¹, although several effective vaccines have been developed and are being applied.

SARS-CoV-2 virion is formed by four proteins: spike (S), envelope (E), membrane (M) and nucleocapside (N) that enclose the virus genome², which consists of a positive-sense nonsegmented single stranded mRNA ((+)ssRNA) of 30 kb. The open reading frames 1a (orf1a) and 1b (orf1b) are located near the untranslated 5' region (5'UTR) of the positive single stranded RNA ((+)ssRNA) and they encode for the polyproteins pp1a and pp1ab.

When SARS-CoV-2 virion infects the organism, S protein binds with high affinity to the surface receptor Angiotensin-Converting Enzyme 2 (ACE2), highly abundant in the lung alveolar type II cells and other cell types^{3,4}, and forms a molecular complex that begins the process of fusion of the virion envelope with the host cell membrane. Finally, viral (+)ssRNA is released into the host cytoplasm⁵ (Fig. 1).

The process of translation occurs in the cytoplasm and produces a set of 11 nonstructural proteins (Nsp) from the orf1a segment (Nsp1 to Nsp11), and 5 nonstructural proteins from the orf1b segment (Nsp12 to Nsp16). The set of 16 Nsp proteins form a replication-transcription complex (RTC) where a set of nested subgenomic negative-strands of RNA ((-)sgRNA) are synthesized by discontinuous transcription, and serve as the templates for the production of subgenomic mRNAs. The structural proteins E (Envelope), M (Membrane), N (Nucleocapside) and S (Spike), and the accessory proteins orf 3a, orf6, orf7a, orf7b, orf8, orf9b, orf9c and orf10 are synthesized from the subgenomic mRNAs^{1,6,7}. SARS-CoV-2 uses the host translational machinery, and redirects it to viral protein synthesis and replication while translation of the host mRNA is inhibited⁸. SARS-CoV and SARS-CoV-2 genomes have ~ 79% of homology⁹, and most of the set of structural and nonstructural proteins are practically the same. However, the virus species differ in the accessory proteins orf8, orf8a, orf 8b, orf9c and orf10^{3,10}.

Once the viral proteins are inserted into the host molecular machinery redirect a great number of host cell functions towards virus replication^{11,12}. Gordon and collaborators experimental work on the interaction of viral and host proteins, or *interactome*³, has been of help to clarify how the SARS-CoV-2 controls the host molecular network to replicate and propagate the infection¹³. From these results, is possible the construction of the undirected network representation of the interactome, and its statistical analysis indicates a modular free-scale hierarchical type of structure in which the proteins orf8, N and Nsp7 are the main hubs¹.

Despite all the molecular knowledge rapidly accumulated on SARS-CoV-2 itself and the infection or inflammation processes in which it is involved in human cells, we still lack dynamical models to understand which are the key nodes underlying severe inflammation processes in some cases. Mathematical and computational dynamic network models are useful tools to integrate data, find experimental holes and also propose or understand the underlying dynamics mechanisms involved in the complex viral/human networks involved during SARS-CoV-2 infection^{1,14,15,16}. The number

of nodes, the number of connections of each node to its neighbours, and the distribution of these connections in the network determine its complexity, structure and dynamical properties¹⁷.

In this network, the viral main protease Nsp5 has a special role in lung inflammation process through its link with Histone Deacetylase 2 (HDAC2)^{1,3,13} (Figs. 1 and 2); suggesting that Nsp5 represses HDAC2 inhibitory action on NF κ B (Figure 1), and thus enhances the transcription of the pro-inflammatory genes targeted by the Nuclear Factor κ B (NF κ B)¹⁸ (Figs. 1 and 2). Such role distinguishes SARS-CoV and SARS-CoV-2. The SARS-CoV virus uses N protein to promote the sustained transcription of the Cyclooxygenase 2 (Cox2) enzyme by binding directly to the NF- κ B transcriptional regulatory elements, and to the CCAAT/enhancer binding proteins of the gene *Cox2*¹⁹. Hence, SARS-CoV and SARS-CoV-2 produce severe acute respiratory syndrome through different molecular mechanisms.

The change from a hub (N protein in SARS-CoV) to a poor connected protein (Nsp5 in SARS-CoV-2) as the underlying cause of severe acute respiratory syndrome could be an adaptation that made SARS-CoV-2 more pathogenic. Nsp5 is also the main protease that cleavages the SARS-CoV-2 polyprotein at 11 conserved sites, and its malfunction or deletion could stop the viral replication cycle and lung inflammation^{20,21}. Thus, Nsp5 is a molecular switch that shows a low rate of mutation²², and its malfunctioning can decrease the strength of inflammation and the persistence of ACE2 in the lung epithelial cells giving rise to a less intense inflammatory response²³.

Activation of the NF κ B family of proteins (p65/RelA, p105/p50, p100/p52, RelB, and c-Rel) is mediated by the phosphorylation of the inhibitor of NF κ B proteins (I κ Bs) by I κ B kinases (IKKs) and the posterior ubiquitinylation and degradation of the phosphorylated I κ Bs (Fig. 1). The released NF κ B proteins enter the nucleus to activate specific genes¹⁸. In the nucleus, HDAC2 blocks the transcriptional activity of NF κ B, inhibiting the production and release of the cytokine Interleukin 6 (IL-6) and I κ B, and interfering with the functioning of the positive feedback circuit between IL-6 and NF κ B²⁴ (Fig. 3A). As we mentioned before, the sustained inactivation of HDAC2 by Nsp5 produces a sustained increase in the transcriptional activity of NF κ B (Figs. 1 and 2) that leads to a high production of IL-6 in the lung epithelium cells. Release of this excess of IL-6 to the vascular system contributes to the “cytokines storm” observed in critical patients^{25,26}, although the average serum concentration of IL-6 in critical COVID19 patients are lower with respect to other respiratory syndromes²⁶. However, the uninterrupted production of IL-6 is closely associated to a persistent presence of the ACE2 receptor in the alveolar type 2 cells due to enhanced *ACE2* gene expression mediated by the JAK-STAT pathway²⁷ (Fig. 2).

NF κ B also promotes transcription of the *Cox2* gene leading to the expression and increased enzymatic activity of Cox2 (Figs. 2 and 3B). However, NF κ B, Cox2 and IL-6 also form a positive feedback circuit in which NF κ B promotes Cox2 enzymatic activity that catalyses the conversion of Arachidonic acid (AA) into Prostaglandin G2 (PGG2), which is in turn modified by the peroxidase moiety of the Cox2 enzyme to produce prostaglandin H2 (PGH2) that is converted to prostaglandin E2 (PGE2)²⁸. PGE2 is then released to the vascular system, binds to its membrane EP2 and EP4 prostanoid receptors and promotes the cleavage of the I κ B-NF κ B complex increasing the amount of free NF κ B, which increases the production of IL-6^{29,30} (Fig. 3B). IL-6, in turn, increases NF κ B activity³¹ (Fig. 2). These regulatory circuits has three therapeutic targets: IL-6 production (Tocilizumab)²⁵, Cox2 (Nimesulide)²⁸, and NF κ B (Dexamethasone)^{32,33}. However, none of these have shown to be really effective to prevent or revert acute inflammatory cases after SARS-CoV-2 infection.

In this work, we propose a dynamical mathematical model that suggests that the three feedback loops mentioned above constitute the minimal circuit for the inflammatory process in the epithelial lung cells during SARS-CoV-2 infection (Fig. 2), and we use an ordinary differential equations (ODEs) continuous model to explore the effect of Nsp5 in the qualitative dynamics of this circuit in which this viral protein acts as an enhancing perturbation in the phase space of this dynamical system³⁴ (Fig. 2). For the model of the circuit, we chose as main molecular components the proteins NF κ B, I κ B, IL-6, Cox2, and PGE2. In this model, Nsp5 prevents the movement of HDAC2 into the nucleus allowing an increase in NF κ B transcriptional activity. The input that turns *on* the circuit in lung cells is a signal from monocytes, lymphocytes, and other immune cells that activates IL-6 production. We propose that this model represents the basic set of interactions that settles *on* the inflammation reaction to the invasion of the lung cells by SARS-CoV-2, and that it is a first approximation to understand the main dynamical features of this response.

Our hypothesis is that Nsp5 leads to the sustained overproduction of IL-6, Cox2, PGE2, and NF κ B, in the epithelial lung cells, which is *a necessary but not a sufficient condition* for a cytokine storm. In the particular case of epithelial lung cells, we found that the computational and mathematical analysis of the model supports our hypothesis, and that Nsp5 effectively enhances NF κ B, IL-6, Cox2 and PGE2 production during the time that remains bounded to HDAC2. Furthermore, the model predicts the existence of an over stimulated immune state (OSIS) in which the amount of IL-6 produced by the infected lung cells is high, and such state becomes autonomous and robust, independent of any stimulus from external sources like monocytes, and lymphocytes. Hence this OSIS cannot be completely shut down by classical anti-inflammatory drugs like Nimesulide and Dexamethasone, and can be possibly beaten only with specific inhibitors of Nsp5 like Saquinavir²¹.

2 Results

In this section we present in detail and focus on the results that are relevant to understand the role of Nsp5 in the process of acute lung inflammation. The rest of the analyses and results related to the mathematical aspects of the model are reported in the [Supplementary Stability Analysis of the Model](#).

We solved the model using the Euler predictor-corrector method with time step of 0.05 s and 720,000 integration steps to simulate a period of 10 hrs post infection, which is the estimated time for the first bursting of new virions into the extracellular medium^{17,35}. We used the parameter values shown in Table 1 to generate an *arbitrary reference state* (ARS) of the system for the initial conditions $ikbnf = 5 \text{ pg mL}^{-1}$, and the rest of the variables equal to 0 at time $t = 0$. In this case, the input $\alpha(t) = 10 \text{ pg mL}^{-1} \text{ s}^{-1}$ and $nsp5 = 0$ for $t \in [0, \infty)$. In this ARS, the translation velocities³⁶ of all genes are set to $2 \text{ pg mL}^{-1} \text{ s}^{-1}$. Fig. 4A shows that the output of the model is the transitory activation of NF κ B, Cox2 and PGE2, in contrast with a sustained high activation of IL-6. In order to clarify the source of the steady activation of IL-6, we solved the model with the same initial conditions and $\alpha(t) = 0$. In this case, the system does not *turn on* during all the time of simulation (data not shown), indicating that the signal from monocytes and other cells is a *necessary condition* to start the IL-6 sustained production in lung cells (Fig. 4A). Fig. 4B shows that when $nsp5 \neq 0$, $\alpha(t) = 0$ and equal gene translation rates, the response of the system is a high sustained level of activation of NF κ B, Cox2, and PGE2 with respect to the ARS. In this case, the concentration of IL-6 in the cell was only slightly affected by the virus. Figs. 4A - 4D show the first five minutes of simulation during which the circuit of Fig. 2 reaches its steady state. This result indicates that *the presence of the virus induce a sustained and persistent production of NF κ B, Cox2, and PGE2* in the circuit of Fig. 2.

Translation rates of the set of genes of the model rarely have the same value, even when their activation is promoted by the same transcription factor (NF κ B in this case). As shown in Table 1, the results of Figs. 4A and 4B are obtained when the rates of translation are the same for all genes. However, in real cells this is not the case, and *Cox2*, *Il-6*, *NF κ B* and *I κ B* are translated at different rates. In *Homo sapiens* the estimated average rate of translation ($V^{\max}$) of a gene is about 10^4 proteins per hour³⁶. However, this average value can be increased according to the state of activity of the cell. In the present work, we arbitrary chose the value of $V^{\max} = 2 \text{ pg mL}^{-1}\text{s}^{-1}$ for all genes in the ARF (Table 1). However, we made parameter variation to know how different values of this set of parameters affect the qualitative behavior of the model (see **Supplementary Stability Analysis of the Model**). Fig. 4C shows that when in the system $V_{nf}^{\max} = 10 \text{ pg mL}^{-1}\text{s}^{-1}$, $V_{I\kappa B}^{\max} = 15 \text{ pg mL}^{-1}\text{s}^{-1}$, $V_{Cox2}^{\max} = 2 \text{ pg mL}^{-1}\text{s}^{-1}$, and $V_{IL6}^{\max} = 2 \text{ pg mL}^{-1}\text{s}^{-1}$, $\alpha(t) = 10 \text{ pg mL}^{-1}\text{s}^{-1}$ and $nsp5 = 0$, the circuit tends to a steady state in which NF κ B, Cox2, and PGE2 exhibit a constant low concentration during the time of simulation, while IL-6 steady concentration is unaffected with respect to the value shown in Fig. 4A. When $nsp5 \neq 0$, the steady concentration of NF κ B, and Cox2, are increased ~ 10 fold with respect to their maximum concentration in the ARS, while PGE2 concentration is increased ~ 4 fold; Nsp5 produces a ~ 4 fold decrease in IL-6 steady concentration with respect to the reference concentration shown in Fig. 4A. This result indicates that a the difference in translation rates between *NF κ B* and *I κ B* (the intensity of the inhibition of NF κ B by I κ B, Fig. 3A) in absence of the virus with respect to their values in ARS induces a low sustained production of NF κ B, Cox2, and PGE2 without affecting IL-6 production. However the presence of Nsp5 induces a high production of NF κ B and Cox2, and a moderate increase in PGE2 concentration that decreases IL-6 concentration, indicating that a higher rate of *I κ B* translation with respect to *NF κ B* translation can decrease the concentration of the cytokine IL-6 in presence of the virus probably due to a lower concentration of PGE2 in the circuit (Figs. 2 and 3A).

Fig. 4D shows that when $\alpha(t) = 10 \text{ pg mL}^{-1}\text{s}^{-1}$, $nsp5 = 0$, $V_{nf}^{\max} = 10 \text{ pg mL}^{-1}\text{s}^{-1}$, $V_{I\kappa B}^{\max} = 15 \text{ pg mL}^{-1}\text{s}^{-1}$, $V_{Cox2}^{\max} = 2 \text{ pg mL}^{-1}\text{s}^{-1}$, and $V_{IL6}^{\max} = 20 \text{ pg mL}^{-1}\text{s}^{-1}$ the circuit tends to a steady state in which NF κ B, Cox2 and PGE2 exhibit a constant low concentration during the time of simulation, while IL-6 is unaffected with respect to its steady concentration in Fig. 4A. However, when $nsp5 \neq 0$ the dynamical behavior of the circuit changes because NF κ B and Cox2 show the same steady concentration that in Fig. 4C, but PGE2 has a higher steady concentration. In this case, IL-6 shows an increase of ~ 2 fold with respect to the level shown in Fig. 4C. Thus, a high rate of IL-6 translation can overcome the effect of an increased rate of *I κ B* translation in presence of the virus, allowing an overproduction of the cytokine.

All these results suggest an important dynamical property of the circuit shown in Fig. 2: a high value of the parameter $V_{IL6}^{\max}$ produces an over production of NF κ B, Cox2, PGE2, and IL-6 in the circuit in presence of Nsp5 with respect to the ARS (Figs. 4A and 4D). We name *over stimulated immune state* (OSIS) to the state of the system shown in Fig. 4D, for which: $V_{nf}^{\max} = 10 \text{ pg mL}^{-1}\text{s}^{-1}$, $V_{I\kappa B}^{\max} = 15 \text{ pg mL}^{-1}\text{s}^{-1}$, $V_{Cox2}^{\max} = 2 \text{ pg mL}^{-1}\text{s}^{-1}$, $V_{IL6}^{\max} = 20 \text{ pg mL}^{-1}\text{s}^{-1}$, $nsp5 \neq 0$ and $\alpha(t) = 10 \text{ pg mL}^{-1}\text{s}^{-1}$.

Fig. 5 shows the response of the circuit to a step function defined as:

$$\alpha(t) = \begin{cases} 10 & 0 \leq t < 10 \\ 0 & t > 10 \end{cases} \quad (16)$$

The latter simulates an external signal of IL-6 initially generated at a rate of $10 \text{ pg mL}^{-1} \text{ s}^{-1}$ during 10 minutes, and switched *off* after the 10 minutes. Fig. 5A is the response of the system in ARS to the step signal, in absence and presence of Nsp5. Interestingly, in the absence of Nsp5, the circuit exhibits a low concentration of NF κ B, Cox2 and PGE2, and a high concentration of IL-6, all which are zero when the external signal is switched *off*. In presence of Nsp5 the ARS is perturbed by $\alpha(t)$, and reaches a new steady state with high concentrations of NF κ B, Cox2, and PGE2 that remain even when the external signal is switched *off*. Furthermore, the high IL-6 concentration reached when the circuit is turned *on* falls to a lower concentration different from zero when $\alpha(t) = 0$.

Fig. 5B is the response of the circuit in OSIS to the step function. In absence of Nsp5, the circuit exhibits a low concentration of NF κ B, Cox2 and PGE2, and a high concentration of IL-6, all which are zero when the external signal is switched *off*. However, in presence of Nsp5 the OSIS is perturbed by $\alpha(t)$, and reaches a new steady state with high concentrations of NF κ B, Cox2, and PGE2 that remain even when the external signal is switched *off*. Furthermore, the higher IL-6 concentration reached when the circuit is turned *on* only falls $\sim 50\%$ of its maximum concentration when $\alpha(t) = 0$. These results suggest that the OSIS becomes *autonomous* of the external signal generated by monocytes and other immune cells in presence of Nsp5, i.e., *the OSIS becomes a persistent state in presence of SARS-CoV-2, and never shut down by itself or by removing external signaling during the 10 hrs of simulation.*

In order to reduce the effect of Nsp5 on the circuit of Fig. 2, we tested the effect of Nimesulide on NF κ B production. Nimesulide, a Cox2 inhibitor, acts inhibiting the synthesis of Cox2 and lipooxygenase enzyme and their products³⁷. We modified Eq. (14) to introduce an inhibition term that decreases the rate of production of Cox2 in presence of Nimesulide:

$$\frac{dcox2}{dt} = \frac{V_{Cox2}^{\max} p_{Cox2}^{on}(t)}{nim + 1} - k_{10} ccox2 \quad (17)$$

where nim is the amount of Nimesulide at time t , whose rate of variation of its concentration in the cell is:

$$\frac{d(nim)}{dt} = r_2 - k_{19} nim \quad (18)$$

where $r_2 = 40 \text{ pg mL}^{-1} \text{ s}^{-1}$ and $k_{19} = 3 \text{ s}^{-1}$. Solution of the modified model (Eq. 17) is presented in Fig. 6. In Fig. 6A, Nimesulide is applied to the circuit in presence of Nsp5, and with all gene translation rates set to $2 \text{ pg mL}^{-1} \text{ s}^{-1}$. In this case, Nimesulide has no effect on NF κ B, a slightly effect on IL-6, and a significant decrease in Cox2 and PGE2 concentrations to values near to zero in presence of 14 pg mL^{-1} of the drug. The same effect is observed when the circuit is in OSIS (Fig. 6B). Thus, Nimesulide is not an effective anti-inflammatory agent in the context of the dynamic circuit modeled here, and does not impact the concentration of IL-6 in the presence of Nsp5, even at high concentrations.

Dexamethasone (DX), is a potent synthetic glucocorticoid that inhibits NF κ B transcription, and is another anti-inflammatory drug that is being used to ameliorate acute inflammatory states due to SARS-CoV-2 infection. In order to test the effect of DX on the inflammatory process in the circuit of Fig. 2, we modified Eq. (8) to introduce an inhibition term that decreases the probability of activation of NF κ B:

$$\frac{dp_{NF\kappa B}^{on}(t)}{dt} = k_{15} \left(\frac{nf^*}{(hd^*+1)(DX+1)} \right) (1 - p_{NF\kappa B}^{on}(t)) - k_{-15} p_{NF\kappa B}^{on}(t) \quad (19)$$

where DX is the amount of DX at time t , whose rate of variation of its concentration in the cell is:

$$\frac{d(DX)}{dt} = r_3 - k_{20} DX \quad (20)$$

where $r_3 = 30 \text{ pg mL}^{-1} \text{ s}^{-1}$ and $k_{20} = 3 \text{ s}^{-1}$.

Fig. 7 shows the effect of DX on the circuit. Fig. 7A shows that when DX is applied to the circuit in presence of $Nsp5$, and when all genes have the same translation rate of $2 \text{ pg mL}^{-1} \text{ s}^{-1}$, the drug eliminates $NF \kappa B$, $Cox2$ and $PGE2$ from the circuit, but has a null effect on the concentration of $IL-6$. However, when the circuit is in OSIS (Fig. 7B), DX has little effect on the immune response in presence of $Nsp5$. Fig. 7C shows that when all translation rates are the same ($2 \text{ pg mL}^{-1} \text{ s}^{-1}$), DX can shut down the immune response in presence of the virus, when the circuit input $\alpha(t)$ is the function of Eq. (16). However, DX cannot shut down the effects of $Nsp5$ when the circuit is in OSIS, having a little anti-inflammatory effect (Fig. 7D). Hence, our results suggest that *DX is an effective anti-inflammatory drug during mild inflammatory states, but is less effective during critical acute lung inflammation due to SARS-CoV-2³⁸*. This dynamical behavior of the circuit of Fig. 2 is unaffected by higher doses of DX . When DX and Nimesulide are applied together (with 35 pg mL^{-1} of DX and 40 pg mL^{-1} of Nimesulide) to the circuit in OSIS, in which $\alpha(t)$ is the function of Eq. (16), there is also a limited anti-inflammatory effect (Fig. 8A).

In contrast, the predicted limited effect of Nimesulide and DX on reverting the OSIS, the molecular docking of $Nsp5$ with a hypothetical drug like Saquinavir²¹, is able to completely eliminate inflammation. We tested this hypothesis modifying $Nsp5$ production in Eq. (11) with an inhibitory term:

$$\frac{d(nsp5)}{dt} = \frac{\phi}{drug+1} - k_{18} nsp5 \quad (20)$$

where $drug$ is the amount of the hypothetical drug that can remove $Nsp5$ from the circuit, whose concentration at time t is given by:

$$\frac{d(drug)}{dt} = r_4 - k_{21} drug \quad (21)$$

with $r_4 = 30 \text{ pg mL}^{-1} \text{ s}^{-1}$ and $k_{21} = 3 \text{ s}^{-1}$.

Fig. 8B shows that when the hypothetical drug is applied to the circuit in OSIS, with $\alpha(t)$ given by Eq. (16), the inflammatory process is completely shut down, suggesting that *Nsp5 is effectively the key cause of the acute lung inflammation during SARS-CoV-2 infection, and a fundamental potential target for its treatment.*

3 Discussion

The complexity of the immune response to infections implies a complex set of different kinds of cells that involve complex regulatory networks and molecules. Together, they underlie specific responses that may lead to resolution or to inflammatory responses whose dynamical properties depend on the networks structure (i.e., the number of nodes, links, and the distribution of links between the nodes) occurring at different scales and levels^{39,40}. The structure of such complex and dynamical network is not random due to the specific nature of its nodes and links, so that if the network has a hierarchical structure its hubs determine the overall network dynamics and the structure of its phase space^{34,39,41}. In high dimensional phase spaces, steady points of diverse kinds can coexist with isolated closed curves (limit cycles), strange attractors of fractal structure, and other limit sets giving rise to a variety of biological dynamical behaviors like molecular switches, periodic and quasi-periodic oscillations, and bursting (among others). Two central problems in the analysis of this kind of dynamical systems are: a) the structure of the phase space is strongly dependent of the value of the set of parameters of the dynamical system, and b) the nonlinearity characteristic of biological systems such as the one treated here⁴² (see [Supplementary Introduction to Nonlinear Dynamics](#)).

In most of the ODEs-based biological models so far studied, there is limited knowledge of the real values of the parameters, and this leads to uncertainty concerning the extent to which the model explains the real process under study. In this case, the validation of the model relied on experimental data. Nonetheless, with the exception of some particular cases⁴³, qualitative models have to be proposed and quantitative validations remain ahead³⁹. This challenge increases with the dimensionality of the model. Thus, is necessary to choose some particular properties of the network under study (inflammation network in this case) that incorporate key dynamical features of the whole system under study, and that can also aid to reduce the number of parameters of the model to the minimum possible.

In this sense, we limited the scope of this work to the study of the inflammation process of epithelial lung cells during SARS-CoV-2 infection. The epithelial lung cells form a sub-network that have two highly connected nodes: the internal signal integrator NF κ B, and the external signal integrator IL-6 Receptor (IL6R)²⁵, which is activated by external IL-6 (Figs. 2 and 3). Both integrators trigger the activation of two important intermediate processes: Cox2 and PGE2 synthesis. Thus, we propose, as a first approximation, that the dynamical behavior of this these four nodes of the circuit of Fig. 2 reflect with certain accuracy the dynamical properties of the complex inflammation sub-network of lung cells. Thus, the dynamics of IL-6, NF κ B, Cox2 and PGE2 nodes is described by a model that consist of 12 nonlinear coupled ordinary differential equations (Table 1), that we assume represents the central dynamical core involved in the lung cells inflammation process in response to SARS-CoV-2 infection. Such core includes three coupled feedback loops shown in Fig. 3. We used this low-dimensionality model to explore the effect of Nsp5 in the qualitative dynamics of the inflammation circuit of Fig. 2^{17,34}.

SARS-CoV-2 main protease Nsp5 has a role as an epigenetic regulator of host DNA expression in lung cells; and thus its physical association with HADC2^{3,17} avoids the movement of the deacetylase from the cytoplasm to the nucleus, increasing the probability of the binding of NF κ B to the promoter site of its target genes⁴⁴ (Eqs. 5-8). NF κ B is a central integrator of the signals that initiate the inflammation process (Fig. 1), which include IL-6 secreted by monocytes, leucocytes, macrophages, among other cells, together with signals from TLR4 and TLR7 that suppress the I κ B-NF κ B complex in a MyD88 independent manner⁴⁵. NF κ B enhanced transcriptional activity

includes the transcription of *IL-6*, *Cox2*, *IκB* and *NF κB* itself that are key genes in the process of inflammation (Fig. 3).

Results obtained from our model suggest that Nsp5 can effectively transform a weak inflammation response into a persistent sustained one that has a relative high concentration of the cytokine IL-6 (Fig. 4B). In the example of Fig. 4B, all genes have the same translation rate, a condition that have low probably of occurrence in the real human immune system in which every gene is subject to different processes of post transcriptional regulation³⁶. When this restriction is removed, and IL-6 has a high translation rate, Nsp5 boosts the concentration of the four main proteins of the circuit with respect to their respective values in ARS, and the inflammation response becomes strongly persistent with a high IL-6 concentration. Furthermore, in this state that we named OSIS, the dynamics of the circuit of Fig. 2 becomes independent of any kind of external signal and never turns *off*, leading to the deregulation of the production of the cytokine IL-6 (Fig. 5B). In this form, Nsp5 changes the qualitative dynamical behavior of the system, in which the trajectories that initially span around the ARS stable node now span around the OSIS stable node³⁴ (see [Supplementary Stability Analysis of the Model](#)).

An important biological consequence of this change in the qualitative dynamics of the circuit is that ACE2 receptor synthesis also becomes persistent and autonomous of any external signal to the lung cells (Fig. 2). Furthermore, neither Nimesulide nor Dexamethasone or both can eliminate the OSIS suggesting that this dynamical behavior could be the form in which the virus assures the persistence of its reproductive cycle without any perturbation from the natural defenses of the body. This can be also a possible clue about the possible adaptive substitution of N protein used in SARS-CoV for the use of the main protease Nsp5 in SARS-CoV-2 as the switch for the inflammation process, which is necessary for ACE2 sustained production, taking into consideration that SARS-CoV N protein targets Cox2 while SARS-CoV-2 Nsp5 protein targets the master integrator NF κB. If this hypothesis is true, *this is the probable cause of the increased pathogenicity of SARS-CoV-2 with respect to SARS-CoV*.

It is of interest that the persistence of the OSIS is supported by a high translational rate of *IL-6*, which in the model must have a value of $V_{IL6}^{max} \geq 7 \text{ pg mL}^{-1} \text{ s}^{-1}$. This result pictures a probable hypothetical scenario where the external signal $\alpha(t)$ triggers the initial inflammation response with a low translation rate of IL-6 that can be shut down with DX. As viral infection continues, IL-6, NF κB, and IκB increase their rate of translation (V_{Cox2}^{max} value has a little weight in this process) and the dynamical behavior of the circuit of Fig. 2 becomes independent of the value of $\alpha(t)$, and *self-sustained*, i.e., the control of the production of the cytokine IL-6 is translated from the external immune cells to the lung cells, and cannot be stopped even with high doses of DX. In this form, in the model, V_{IL6}^{max} is a *bifurcation parameter* that changes the qualitative dynamical behavior of the circuit of Fig. 2 when $V_{nf}^{max} = 10 \text{ pg mL}^{-1} \text{ s}^{-1}$ and $V_{IκB}^{max} = 15 \text{ pg mL}^{-1} \text{ s}^{-1}$ are constant values (see [Supplementary Stability Analysis of the Model](#)).

In this extreme situation (OSIS), the unique treatment possible suggested by our model is an inhibitor of Nsp5 like *Saquinavir*²¹. In this case, the OSIS is completely shut down, which indirectly implies the down production of ACE2. Saquinavir is an anti-retroviral drug used against the main protease of HIV-1^{21,46}, with some undesirable effects like diarrhea, abdominal pain, and nausea. We suggest that this drug could be a plausible treatment against the effects of SARS-CoV-2 in acute lung inflammation, and as a complement to the new vaccine against this coronavirus.

In a previous work¹ the role of the viral protein Orf8 as a hub of the SARS-CoV-2 infection was suggested. This highly connected protein has as main targets those processes related to the vesicular trafficking required for the ensemble of new virions³ (Fig. 1). Thus, the inhibition of the effects of Orf8 in the host cells could block the reproduction cycle of the virus but not the inflammation process because Nsp5 is not directly linked to Orf8³. Rapamycin has been suggested as a drug against the effects of Orf8 but produces severe immunosuppressant effects, which, according to the results of our model, will not be of care in cases of severe lung inflammation because Nsp5 uncouple the circuit of Fig. 2 from the process of viral replication (Figs.1 and 5B). In this case, a treatment with Rapamycin and Saquinavir can be an alternative for patients with severe lung inflammation. This suggestion needs experimental verification and clinical evaluation.

4 Conclusions

In this work we propose a dynamical model that puts forward a novel hypothetical mechanisms underlying lung cells inflammation process in response to SARS-CoV-2. In this scenario the main protease Nsp5 effectively enhances the inflammatory process, increasing the levels of NF κ B, IL-6, Cox2, and PGE2 with respect to the reference state. When the translation rates of NF κ B and I κ B are increased to a high constant value, and the translation rate of IL-6 is increased above the threshold value of $7 \text{ pg mL}^{-1} \text{ s}^{-1}$ the circuit enters in a persistent over stimulated immune state (OSIS) with high levels of the cytokine IL-6. The OSIS never shuts down by itself, and becomes autonomous of the signals from other immune cells like macrophages and lymphocytes. Our model explains why DX or Nimesulide have little effect on the OSIS, and the only means to suppress such acute and sustained inflammation state is by inhibiting the protein Nsp5 with drugs such as Saquinavir.

In this form, the model suggests, in accordance to our hypothesis, that Nsp5 is effectively a key node underlying severe acute lung inflammation during SARS-CoV-2 infection. The persistent production of IL-6 by lung cells during infection can be one of the causes of the cytokine storm observed in critical patients with COVID19. From an evolutionary point of view, the use of Nsp5 as the switch to start inflammation, and the consequent overproduction of the ACE2 receptor, could have driven the increased pathogenicity of SARS-CoV-2 with respect to SARS-CoV.

The present theoretical work is part of the project “Dynamics of the SARS-CoV-2 network” and is based on the experimental data reported in the literature at the moment.

5 Model

Immune response to infections involves complex spatio-temporal processes involving multiple molecular components, non-linear interactions and cell-cell interactions. In consequence, the immune response to infections is a multidimensional, multiscale and multilevel dynamical process in which innate and acquired immunity mechanisms are involved to protect the organism against pathogens³⁹. In this scenario, the number of variables that become both sufficient and necessary for a detailed enough description of the spatio-temporal dynamics of the immune system is tremendously high. Some computational integrative efforts have been made in this direction⁴⁷, but imply very large systems that cannot be modeled with continuous quantitative models that are, in turn, necessary if we want to understand which are key nodes driving the system towards the OSIS. The lack of quantitative experimental data and knowledge of the values of most of the model parameters are additional challenges to achieve such dynamic models of the immune system in

response to specific infectious diseases. Thus, most models of the immune system have been qualitative^{39,48}.

A possible approach implies the use of ordinary differential equations (ODEs) qualitative modeling that is oriented to determine all the possible trajectories of a dynamical system along its n -dimensional phase space (with $n > 0$), when the system is subject to a set of initial conditions and parameters values. High dimensional nonlinear dynamical systems can exhibit a variety of coexisting dynamical behaviors that determine the structure of their phase space⁴⁹. However, the structure of the phase space of biological nonlinear dynamical systems is highly dependent on the set of parameters values. Variations in the value of one or more parameters can drive a drastic change in phase space generating a different qualitative dynamics of the system. This process is known as bifurcation, and it is important to find the possible bifurcations in the system and to identify which parameter or parameters are the responsible of these changes⁴⁹ (See [Supplementary Introduction to Nonlinear Dynamics](#)).

In order to avoid the problem due to the high dimensionality and complexity of the immune response to SARS-CoV-2 infection, and to use the minimum number of parameters in the model that we are proposing in this work, we assume that the interaction between nodes IL-6, NF κ B, I κ B, Cox2 and PGE2 settles on the main dynamical features of the inflammation process in lung epithelial cells due to the presence of the Nsp5 viral protein. In the proposed circuit shown in Fig. 2, IL-6, NF κ B, Cox2 and PGE2 define three feedback loops (Figs. 3A and 3B): a positive feedback loop between NF κ B and IL-6 (Fig. 3A); a positive feedback loop between NF κ B and I κ B (Fig. 3A); and a negative feedback loop between PGE2 and I κ B (Fig. 3B).

These feedback loops are distributed between three compartments:

a) IL-6 is secreted into the extracellular medium where it acts in paracrine and exocrine form. IL-6 binds to its receptor IL6R at the cell surface. Once the complex IL6-IL6R is formed, it activates IKK in the cytoplasm³¹. PGE2 is also secreted into this compartment where it binds to EP2 and EP4 receptors at the cell surface, activating Protein Kinase B (Akt) in the cytoplasm²⁹.

b) IKK, free NF κ B, and the complex I κ B-NF κ B are located in the cytoplasm, where I κ B is degraded by ubiquitination. Protein Cox2 is located in the endoplasmic reticulum (ER).

c) Free NF κ B enters the nucleus where induce the transcription of *IL6*, *Cox2*, *IKB* and *NF κ B* genes. Transcriptional activity of nuclear NF κ B is inhibited by HDAC2, and enhanced by Nsp5.

In this form, the model cell consists of three compartments: extracellular medium, cytoplasm and nucleus. We assume that the ratio of the cytoplasmatic volume to external volume (V_{cyt} / V_{out}) is one, and that the ratio of the cytoplasmatic volume to the nuclear volume (V_{cyt} / V_{nuc}) is 2. In each compartment the concentration of the molecules are measured in pg mL^{-1} .

In the circuit of Fig. 2, the rate of production of free cytoplasmic NF κ B depends on the rate of its release from the I κ B-NF κ B complex, which is proportional to the product of the concentrations of I κ B and IKK (Fig. 3A), its own rate of production due to nuclear NF κ B transcriptional activity inhibited by HDAC2 (Figs. 1 and 3A), the rate at which is recaptured by I κ B (Fig. 3A), and the rate of transport into the nucleus:

$$\frac{d(nf)}{dt} = V_{nf}^{\max} p_{nf}^{on}(t) + k_a ikk \cdot ikbnf - k_4 nf \cdot ikb - D \cdot nf \quad (1)$$

where nf is the concentration of free cytoplasmic NF κ B, $V_{nf}^{\max}$ is the maximum rate of translation of the NF κ B gene, $p_{nf}^{on}(t)$ is the probability that the NF κ B gene is in its active state (on), ikk is the concentration of IKK, $ikbnf$ is the concentration of I κ B-NF κ B complex, k_a and k_2 are rate constants and D is the transport constant (Table 1).

We assume that the concentration of IKK activated by IL-6 is proportional to the external concentration of the cytokine: $ikk = \gamma_1 \cdot il6$, where γ_1 is a constant of proportionality. In a similar form, PGE2 increases the rate of cleavage of the complex I κ B-NF κ B by phosphorylation of the IKK α subunit by Akt⁵⁰ (Fig. 3B). Thus, we assume that the concentration of Akt is proportional to the external concentration of PGE2: $akt = \gamma_2 \cdot pge2$. Substituting both expressions in Eq. (1) we finally obtain:

$$\frac{d(nf)}{dt} = V_{\max nf} p_{nf}^{on}(t) + k_1 il6 \cdot ikbnf + k_{11} pge2 \cdot ikbnf - k_4 nf \cdot ikb - D \cdot nf \quad (2)$$

where k_1 and k_{11} are the new rate constants (Table 1).

The rate at which the complex I κ B-NF κ B is broken in presence of IKK depends on the rate at which IKK and Akt remove I κ B from the complex under the action of IL-6 and PGE2, and the rate at which the complex is formed again.:

$$\frac{d(ikbnf)}{dt} = k_4 nf \cdot ikb - k_5 il6 \cdot ikbnf - k_{11} pge2 \cdot ikbnf \quad (3)$$

where $ikbnf$ is the concentration of the complex I κ B-NF κ B, and k_4 and k_5 are rate constants (Table 1).

The rate at which NF κ B enter the nucleus depends on the rate of transport of the molecule into de nucleus, adjusted for the change in volume between the cytoplasmic and nuclear compartments, and on the rate of inhibition of nuclear NF κ B by its inhibitors:

$$\frac{dnf^*}{dt} = D \left(\frac{V_{cyt}}{V_{nuc}} \right) nf - k_{deg} n f^* \quad (4)$$

where nf^* is the nuclear concentration of NF κ B and k_{deg} is a rate constant.

The probabilities of activation of genes $IL6$, $Cox2$, $I\kappa B$ and $NF \kappa B$ (Figs. 3A and 3B) are given by:

$$\frac{dp_{IL6}^{on}(t)}{dt} = k_2 \left(\frac{nf^*}{hd^* + 1} \right) (1 - p_{IL6}^{on}(t)) - k_{-2} p_{IL6}^{on}(t) \quad (5)$$

$$\frac{dp_{I\kappa B}^{on}(t)}{dt} = k_3 \left(\frac{nf^*}{hd^* + 1} \right) (1 - p_{I\kappa B}^{on}(t)) - k_{-3} p_{I\kappa B}^{on}(t) \quad (6)$$

$$\frac{dp_{Cox2}^{on}(t)}{dt} = k_7 \left(\frac{nf^*}{hd^* + 1} \right) (1 - p_{Cox2}^{on}(t)) - k_{-7} p_{Cox2}^{on}(t) \quad (7)$$

$$\frac{dp_{NF\kappa B}^{on}(t)}{dt} = k_{15} \left(\frac{nf^*}{hd^* + 1} \right) (1 - p_{NF\kappa B}^{on}(t)) - k_{-15} p_{NF\kappa B}^{on}(t) \quad (8)$$

see **Supplementary Master Equation for Gene Activation** for the mathematical deduction of these equations. For Eq. 5-7, the variable $p_i^{on}(t)$, with $i \in \{\text{IL-6, I}\kappa\text{B, Cox2, NF } \kappa\text{B}\}$, is the probability that the gene i is expressed at time t , hd^* is the amount of HDCA2 in nucleus, and $k_2, k_{-2}, k_3, k_{-3}, k_7, k_{-7}, k_{15}, k_{-15}$ are rate constants (Table 1).

We assume that free cytoplasmic HDAC2 is produced according to the rate equation:

$$\frac{d(hd)}{dt} = \frac{r}{\eta(nsp5 + 1)} - D_2 \cdot hd \quad (9)$$

where hd is the amount of free cytoplasmic HDAC2 that is produced at a constant rate r , D_2 is the rate of transport of free HDAC2 into the nucleus. In this equation, $nsp5$ is the amount of Nsp5 in the infected cell that sequester HDAC2 in cytoplasm⁴⁴, and η is a constant (Table 1). Free cytoplasmic HDCA2 enters the nucleus according to the equation:

$$\frac{d(hd^*)}{dt} = D_2 \left(\frac{V_{cyt}}{V_n} \right) hd - k_{deg2} hd^* \quad (10)$$

where hd^* is the amount of HDAC2 in the nucleus, and k_{deg2} is a rate constant (Table 1).

Nsp5 is a protein with only one link in the virus interactome^{1,3,13}, which is an input to the circuit of Fig. 2. However, the real kinetic mechanism of production of this viral protein is unknown, thus we assume a plausible simple kinetic mechanism that leads Nsp5 to have a maximum constant concentration of 20 nM. We set the parameters ϕ and k_{18} to the values shown in Table 1:

$$\frac{d(nsp5)}{dt} = \phi - k_{18} nsp5 \quad (11)$$

where ϕ is the constant rate of production of Nsp5, and k_{18} is a rate constant (Table 1). The effect of the variation of parameter ϕ on the dynamics of the circuit of Fig. 2 is analyzed in the **Supplementary Stability Analysis of the Model**.

During the early stage of infection by SARS-CoV-2, the rate of variation of the concentration of IL-6 outside the lung cell depends on the rate of production of IL-6 by monocytes, leucocytes and other cells under stimulation by Toll-like Receptors (TLRs) (denoted by $\alpha(t)$ ^{25,51}, on the rate of production by lung cells due to nuclear NF κ B transcriptional activity enhanced by Nsp5 (Fig. 3A),

and on the rate of degradation of IL-6 by different mechanisms. Finally, we assume that the concentration of IKK activated by IL-6 is proportional to the external concentration of the cytokine (see Eq. 2):

$$\frac{d(il6)}{dt} = \alpha(t) + V_{IL6}^{\max} p_{IL6}^{on}(t) - k_5 il6 \cdot ikbnf - k_6 il6 \quad (12)$$

where $V_{IL6}^{\max}$ is the maximum rate of translation of *IL6*, $p_{IL6}^{on}(t)$ is the probability that the *IL6* gene is activated at time t , and k_5 and k_6 are rate constants (Table 1).

The rate of variation of the amount of free IκB in the system is the balance between the rate of translation of gene *IκB* and the rate of formation of new IκB-NF κB complexes:

$$\frac{d(ikb)}{dt} = V_{IκB}^{\max} p_{IκB}^{on}(t) - k_4 ikb \cdot nf \quad (13)$$

where $V_{IκB}^{\max}$ is the maximum rate of translation of *IκB*, and $p_{IκB}^{on}(t)$ is the probability that the gene *IκB* is activated at time t (Table 1).

The rate of variation of the amount of the enzyme Cox2 in the system depends on the rate of translation of the *Cox2* gene, and its rate of inhibition by different mechanisms:

$$\frac{dcox2}{dt} = V_{Cox2}^{\max} p_{Cox2}^{on}(t) - k_{10} cox2 \quad (14)$$

where *cox2* is the amount of the enzyme in the system, $V_{Cox2}^{\max}$ is the maximum rate of variation, $p_{Cox2}^{on}(t)$ is the probability that the gene *Cox2* is activated at time t , k_8 , k_9 and k_{10} are rate constants (Table 1).

The rate of variation of the amount of PGE2 in the system depends on the amount of Cox2, on the amount of IL-6, and on its rate of degradation:

$$\frac{dpge2}{dt} = k_{17} cox2 - k_{11} pge2 \cdot ikbnf - k_{12} pge2 \quad (15)$$

where k_{11} , k_{12} and k_{17} are rate constants (Table 1).

6 Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

7 Author Contributions

Antonio Bensussen made the stability and sensitivity analysis of the model. Elena R. Álvarez-Buylla and José Díaz conceived the study, discussed the data, and wrote the paper. José Díaz coordinated the study.

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10 Supplementary Material

[Supplementary Stability Analysis of the Model](#)

[Supplementary Master Equation for Gene Activation](#)

[Supplementary Introduction to Nonlinear Dynamics](#)

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12 Tables

Table 1.- Equations and parameters of the basic model of inflammation

Basic Inflammation Circuit	Parameters
$\frac{dNF\kappa B}{dt} = V_{nf}^{\max} p_{nf}^{on}(t) + k_1 \cdot il6 \cdot i\kappa bnf + k_{11} pge2 \cdot i\kappa bnf - k_4 i\kappa b \cdot nf - D \cdot nf$ $i\kappa bn = I\kappa B - NF\kappa B \text{ molecular complex}$	$k_1 = 0.5 \text{ pg mL}^{-1} \text{ s}^{-1}$ $k_4 = 5 \text{ pg mL}^{-1} \text{ s}^{-1}$ $D = 1 \text{ s}^{-1}$
$\frac{d(nf^*)}{dt} = D \left(\frac{V_{cyt}}{V_{nuc}} \right) nf - k_{deg} nf^*$ $nf^* = \text{nuclear concentration of } NF\kappa B$	$k_{deg} = 2 \text{ s}^{-1}$ $\left(\frac{V_{cyt}}{V_{nuc}} \right) = 2$
$\frac{d(i\kappa bnf)}{dt} = k_4 nf \cdot i\kappa b - k_5 il6 \cdot i\kappa bnf - k_{11} pge2 \cdot i\kappa bnf$ $i\kappa bnf = I\kappa B - NF\kappa B \text{ molecular complex}$ $pge2 = \text{amount of PGE2}$	$k_4 = 5 \text{ pg mL}^{-1} \text{ s}^{-1}$ $k_5 = 0.5 \text{ pg mL}^{-1} \text{ s}^{-1}$
$\frac{dp_{IL6}^{on}(t)}{dt} = k_2 \frac{nf^*}{hd^* + 1} p_{IL6}^{off}(t) - k_{-2} p_{IL6}^{on}(t)$ $p_{IL6}^{on}(t) = \text{probability of } IL6 \text{ activation at time } t$ $hd^* = \text{amount of HDAC2 in nucleus}$	$k_2 = 0.1 \text{ (Number of Molecules)}^{-1} \text{ s}^{-1}$ $k_{-2} = 0.035 \text{ s}^{-1}$
$\frac{dp_{I\kappa B}^{on}(t)}{dt} = k_3 \frac{nf^*}{hd^* + 1} (1 - p_{I\kappa B}^{on}(t)) - k_{-3} p_{I\kappa B}^{on}(t)$ $p_{I\kappa B}^{on}(t) = \text{probability of } I\kappa B \text{ activation at time } t$	$k_3 = 0.1 \text{ (Number of Molecules)}^{-1} \text{ s}^{-1}$ $k_{-3} = 0.035 \text{ s}^{-1}$
$\frac{dp_{Cox2}^{on}(t)}{dt} = k_7 \frac{nf^*}{hd^* + 1} (1 - p_{Cox2}^{on}(t)) - k_{-7} p_{Cox2}^{on}(t)$ $p_{Cox2}^{on}(t) = \text{probability of } Cox2 \text{ activation at time } t$	$k_7 = 0.1 \text{ (Number of Molecules)}^{-1} \text{ s}^{-1}$ $k_{-7} = 0.035 \text{ s}^{-1}$
$\frac{dp_{NF\kappa B}^{on}(t)}{dt} = k_{15} \frac{nf^*}{hd^* + 1} (1 - p_{NF\kappa B}^{on}(t)) - k_{-15} p_{NF\kappa B}^{on}(t)$ $p_{NF\kappa B}^{on}(t) = \text{probability of } NF\kappa B \text{ activation at time } t$	$k_{15} = 0.1 \text{ (Num of Molecules)}^{-1} \text{ s}^{-1}$ $k_{-15} = 0.035 \text{ s}^{-1}$
$\frac{d(nsp5)}{dt} = \phi - k_{18} nsp5$ $nsp5 = \text{amount of viral Nsp5 in cell}$ $\phi = \text{rate of Nsp5 production}$	$k_{18} = 0.7 \text{ s}^{-1}$ $\phi = 14 \text{ pg mL}^{-1} \text{ s}^{-1}$

$\frac{d(hd)}{dt} = \frac{r}{nsp5+1} - D_2 \cdot hd$ <p>hd = amount of HDAC2 in cytoplasm D_2 = transport constant of HDAC2</p>	$r = 30 \text{ pg mL}^{-1} \text{ s}^{-1}$ $D_2 = 2 \text{ s}^{-1}$ $\eta = 1 \text{ pg mL}^{-1}$
$\frac{d(hd^*)}{dt} = D_2 \left(\frac{V_{cyt}}{V_n} \right) hd - k_{deg2} hd^*$	$D_2 = 2 \text{ s}^{-1}$ $k_{deg2} = 2 \text{ s}^{-1}$ $\left(\frac{V_{cyt}}{V_{nuc}} \right) = 2$
$\frac{d(il6)}{dt} = \alpha(t) + V_{IL6}^{\max} p_{IL6}^{on}(t) - k_5 \cdot il6 \cdot ikbnf - k_6 il6$	$k_5 = 0.5 \text{ pg mL}^{-1} \text{ s}^{-1}$ $k_6 = 0.5 \text{ s}^{-1}$ $\alpha(t) = 10 \text{ pg mL}^{-1} \text{ s}^{-1}$
$\frac{d(ikb)}{dt} = V_{IKB}^{\max} p_{IKB}^{on}(t) - k_4 ikb \cdot nf$	$k_4 = 5 \text{ pg mL}^{-1} \text{ s}^{-1}$
$\frac{d(cox2)}{dt} = V_{Cox2}^{\max} p_{Cox2}^{on}(t) - k_{10} cox2$	$k_{10} = 0.4 \text{ s}^{-1}$
$\frac{d(pge2)}{dt} = k_{17} cox2 - k_{11} pge2 \cdot ikbnf - k_{12} pge2$	$k_{17} = 0.8 \text{ s}^{-1}$ $k_{11} = 0.8 \text{ pg mL}^{-1} \text{ s}^{-1}$ $k_{12} = 3 \text{ s}^{-1}$

In the reference state the translation velocities are: $V_{nf}^{\max} = V_{Cox2}^{\max} = V_{IL6}^{\max} = V_{IKB}^{\max} = 2 \text{ pg mL}^{-1} \text{ s}^{-1}$

13 Figure Captions

Figure 1.- Process of infection of lung cells by SARS-CoV-2. Infection begins when the protein S of the virion binds with high affinity to the cells surface receptor ACE2 (Angiotensin-Converting Enzyme 2) that is highly abundant in the lung alveolar type II cells. The formation of the complex S-ACE2 initiates the process of fusion between the virion envelope and the cells membrane leading to the liberation of the nucleocapsid with the viral genome into the cytoplasm. SARS-CoV-2 genome consists of a positive-sense nonsegmented single stranded mRNA ((+)ssRNA) of ~ 30 kb. The open reading frames 1a (orf1a) and 1b (orf1b) are located near the 5'UTR of the (+)ssRNA and they code for the polyproteins pp1a and pp1ab. Maturation of these polyproteins results in 11 nonstructural proteins (Nsp) from the orf1a segment (Nsp1 to Nsp11) and 5 nonstructural proteins from the orf1b segment (Nsp12 to Nsp16). Nsp proteins form the replication-transcription complex (RTC) a set of nested subgenomic minus-strands of RNA ((-)sgRNA) are synthesized in a process of discontinuous transcription. These (-) sgRNAs serve as the templates for the production of subgenomic mRNAs from which the structural proteins E, M, N and S, together with the accessory proteins orf 3a, orf6, orf7a, orf7b, orf8, orf9b, orf9c and orf10 are synthesized. The presence of viral (+)ssRNA initiates the TLR3 and TLR4 mediated immune response, which increases the concentration of free NF κB in cytoplasm. Cytoplasmic NF κB enters the nucleus where promotes the transcription of a series of target genes. Viral protein Nsp5 inhibits the inhibition exerted by HDAC2 on nuclear NF κB transcription factor, enhancing the synthesis of inflammatory cytokines like IL-6. (Figure drawn with Biorender template <https://app.biorender.com/>).

Figure 2.- Circuit representation of inflammation. IL-6, NF κ B, Cox2, and PGE2 are the basic nodes of the circuit that represents the basic features of inflammation of lung cells during SARS-CoV-2 infection. In this circuit the arrows represent activation and the bars inhibition. In the figure are shown two positive feedback loops between IL-6 and NF κ B, and between NF κ B and Cox2. IL-6 also induces the expression of the ACE2 receptor in the lung cell membrane. NF κ B is inhibited by HDAC2, which in turn is inhibited by the viral protein Nsp5. The circuit has two targets for the anti-inflammatory drugs Nimesulide and Dexamethasone.

Figure 3.- Feedback loops of the model of inflammation. The mathematical model proposed in this work is based on the dynamical features of three feedback loops: A) two positive feedback loops between IL-6 and NF κ B, and NF κ B and I κ B; B) a negative feedback loop between PGE2 and I κ B. In Figure A, $\alpha(t)$ is an external signal that represents the amount of IL-6 produced by immune cells like monocytes and lymphocytes, among others.

Figure 4.- Effect of Nsp5 on IL-6 production. A) Arbitrary reference state (ARS) of the circuit of Figure 2. In absence of the viral protein Nsp5, the response to an external input $a(t) = 10 \text{ pg mL}^{-1}\text{s}^{-1}$ is a transitory activation of NF κ B, Cox2, and PGE2. In this case, IL-6 shows a steady high concentration. In this case, all the gene translation rates are set to $2 \text{ pg mL}^{-1}\text{s}^{-1}$. B) In presence of Nsp5 the circuit shows steady higher concentrations of NF κ B, Cox2, and PGE2 (red line) with respect to ARS (black line). IL-6 steady concentration (black line) is unaffected by the presence of the viral protein (red line). In this panel, $a(t) = 10 \text{ pg mL}^{-1}\text{s}^{-1}$, and all the gene translation rates are set to $2 \text{ pg mL}^{-1}\text{s}^{-1}$. C) When NF κ B and I κ B translation rates are $V_{nf}^{\max} = 10 \text{ pg mL}^{-1}\text{s}^{-1}$ and $V_{I\kappa B}^{\max} = 15 \text{ pg mL}^{-1}\text{s}^{-1}$, respectively, and Cox2 and IL-6 translation rates are 2 nM s^{-1} the circuit shows a weak sustained activation of NF κ B, Cox2, and PGE2 (black line) in response to the external input $a(t) = 10 \text{ pg mL}^{-1}\text{s}^{-1}$ and in absence of Nsp5. In this case, IL-6 steady concentration does not change (black line). In presence of Nsp5 the circuit shows steady higher concentrations of NF κ B, Cox2, and PGE2 (red line), although the concentration of IL-6 is lower than is absence of the virus. D) In the over stimulated immune state (OSIS), the presence of Nsp5 produces an enhanced production of NF κ B, Cox 2, PGE2 and IL-5 (red lines). This particular state is obtained when $a(t) = 10 \text{ pg mL}^{-1}\text{s}^{-1}$, $V_{nf}^{\max} = 10 \text{ pg mL}^{-1}\text{s}^{-1}$, $V_{I\kappa B}^{\max} = 15 \text{ pg mL}^{-1}\text{s}^{-1}$, $V_{Cox2}^{\max} = 2 \text{ pg mL}^{-1}\text{s}^{-1}$, and $V_{IL6}^{\max} = 20 \text{ pg mL}^{-1}\text{s}^{-1}$.

Figure 5.- Response of the circuit to a step function. A) When the external signal $\alpha(t)$ is the step function of Eq. (16) of main text and is applied to the ARS in absence of Nsp5 (black line) the level of cytokine IL-6 initially increases to a maximum value of 20 pg mL^{-1} , and becomes zero when the external signal is switched *off*. There is not any effect on NF κ B, Cox2 and PGE2 concentrations in the circuit. However, in presence of Nsp5 (red line) the levels of NF κ B, Cox2, PGE2 and IL-6 are increased in response to $\alpha(t)$ and never become zero when the external signal is switched *off*. B) When the step function of Eq. (16) is applied to the circuit in absence of Nsp5 (black line) with $V_{nf}^{\max} = 10 \text{ pg mL}^{-1} \text{ s}^{-1}$, $V_{IL6}^{\max} = 15 \text{ pg mL}^{-1} \text{ s}^{-1}$, $V_{Cox2}^{\max} = 2 \text{ pgmL}^{-1} \text{ s}^{-1}$ and $V_{IL6}^{\max} = 20 \text{ pg mL}^{-1} \text{ s}^{-1}$, there is a low increase in NF κ B, Cox2 and PGE2 concentrations that becomes zero when the external signal is switched *off*. IL-6 initially increases to a maximum value of 20 pg mL^{-1} , and becomes zero when the external signal is also zero. In contrast, when the circuit is in OSIS in presence of Nsp5 (red line) the concentrations of NF κ B, Cox2, PGE2 and IL-6 are increased in response to $\alpha(t)$ and remain at a high value even the external signal is switched *off*.

Figure 6.- Effect of Nimesulide on lung cell inflammation. A) Effect of 14 pg mL^{-1} of Nimesulide on the levels of NF κ B, Cox2, PGE2 and IL-6 when $\alpha(t) = 10 \text{ pg mL}^{-1}\text{s}^{-1}$, and all gene translation rates are $2 \text{ pg mL}^{-1}\text{s}^{-1}$ in presence of Nsp5 (red line). Nimesulide decreases the concentration of Cox2 and PGE2, but has not effect on NF κ B and IL-6 (blue line). B) Effect of 14 pg mL^{-1} of Nimesulide on the OSIS (red line). Nimesulide significantly decreases the concentration of Cox2 and PGE2, but has not effect on NF κ B, The drug has a weak effect on IL-6 concentration (blue line).

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Figure 7.- Effect of Dexamethasone on lung cell inflammation. A) Effect of 14 pg mL^{-1} of Dexamethasone (DX) on the levels of NF κ B, Cox2, PGE2 and IL-6 when $a(t) = 10 \text{ pg mL}^{-1} \text{ s}^{-1}$, and all gene translation rates are $2 \text{ pg mL}^{-1} \text{ s}^{-1}$ in presence of Nsp5 (red line). DX effectively decreases the concentration of NF κ B, Cox2 and PGE2 to zero, but has not effect on IL-6 (black line). B) Effect of 14 pg mL^{-1} of DX on the OSIS (red line). DX has a weak effect on the concentration of NF κ B, Cox2, PGE2, and IL-6 (red line). C) When the external signal $a(t)$ is the step function of Eq. (16) of main text and is applied to the circuit in presence of Nsp5 (red line), with all gene translation rates equal to $2 \text{ pg mL}^{-1} \text{ s}^{-1}$, 14 pg mL^{-1} of DX decreases the concentration of NF κ B, Cox2, PGE2 and IL-6 to zero (black line). D) When the external signal $a(t)$ is the step function of Eq. (16) of main text and 14 pg mL^{-1} of DX are applied to the circuit in the OSIS, the drug has a practically null effect on the inflammation process.

Figure 8.- Effect of the molecular docking of Nsp5 on lung inflammation. A) When DX and Nimesulide are applied together (with 35 pg mL^{-1} of DX and 40 pg mL^{-1} of Nimesulide) to the circuit in OSIS (red line), in which $\alpha(t)$ is the function of Eq. (16), there is also a little anti-inflammatory effect. B) The molecular docking of Nsp5 completely shuts down the inflammation process when the circuit is in OSIS, with $\alpha(t)$ given by Eq. (16), suggesting that Nsp5 is effectively the cause of the acute lung inflammation during SARS-CoV-2 infection.

Supplementary Files

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