

# Effects on serum protein levels from one bout of high intensity interval training in individuals with axial spondyloarthritis and controls

Åsa Andersson (✉ [asa.andersson@hh.se](mailto:asa.andersson@hh.se))

FIH, Halmstad University

M. Charlotte Olsson

FIH, Halmstad University

Anna Torell

Ängelholm Hospital

Elisabeth Mogard

Lund University, Skåne University Hospital

Emma Haglund

FIH, Halmstad University

---

## Research Article

**Keywords:** Axial spondyloarthritis, high-intensity interval training, exercise rehabilitation, IL-6, BMP-7, CRP

**Posted Date:** November 8th, 2023

**DOI:** <https://doi.org/10.21203/rs.3.rs-3564226/v1>

**License:** © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

**Additional Declarations:** No competing interests reported.

---

# Abstract

## Background

Axial spondyloarthritis (axSpA) is a chronic inflammatory disease primarily affecting the axial skeleton causing pain, inflammation, and stiffness. Individuals with axSpA are at greater risk of developing cardiovascular disease, which can be counteracted by physical activity. High-intensity interval training (HIIT) has been shown to improve cardiovascular health, but the effect on disease activity and the level of inflammation in axSpA has been less studied. With the aim of investigating how levels of inflammatory cytokines, myokines, and protein markers for bone metabolism are acutely affected by one bout of HIIT, we studied serum from individuals with axSpA and healthy controls (HC).

## Methods

Ten participants with axSpA and 11 age- and sex-matched HC performed a single HIIT bout on a cycle ergometer: 4x4 minutes intervals with three minutes active rest in between. Blood samples were taken before and one hour after the HIIT bout. Serum proteins (IL-6, IL-17, IL-18, TNF $\alpha$ , CXCL-10, VEGF-A, BDNF, DKK-1, osteoprotegerin, osteocalcin, osteopontin, BMP-7, CRP) were analyzed with a Luminex system or ELISA. Descriptive data are presented as mean with standard deviation. A two-way ANOVA was used for comparisons.

## Results

A main effect from baseline to one hour post HIIT showed that both groups had a significant increase in serum levels (pg/ml) of IL-6: axSpA 2.2 (3.0) to 3.2 (1.8) and HC 0.4 (0.4) to 1.9 (2.0),  $p = 0.03$ . VEGF-A (pg/ml) was significantly lower in the axSpA group: 159 (138) vs. HC 326 (184),  $p = 0.03$ , but was not affected by the HIIT bout. BMP-7 (ng/ml) increased in both groups after the HIIT: axSpA 61.6 (13.1) to 75.2 (20.0) and HC 64.6 (20.8 to 75.0 (17.8),  $p < 0.001$ . For the other proteins analyzed, there were no significant differences in serum concentrations between individuals with axSpA and HC, or within the two groups before and after one bout of HIIT.

## Conclusions

One acute bout of HIIT significantly increases the serum concentrations of IL-6 and BMP-7 after 1 hour in both individuals with axSpA and HC.

## Background

Spondyloarthritis is an umbrella term for a group of inflammatory rheumatic diseases, including axial spondyloarthritis (axSpA), which primarily affects the axial skeleton and the sacroiliac joint (1). The

clinical manifestations encompass stiffness, aching, and pain around the spine and pelvis and the disease process can lead to fusing of the vertebrae, but also of the sacroiliac joints (i.e., ankylosis) (2–4). axSpA includes patients with X-ray changes in the sacroiliac joint or spine, in addition to patients without X-ray verified changes (non-radiographic axSpA) (1).

Studies have shown that individuals with axSpA have an increased risk of cardiovascular disease, which influences the treatment of the disease and overall health (5). The elevated cardiovascular risk is believed to be the result of a chronic systemic inflammation and insufficient level of physical activity, but other comorbidities might play a role as well (6, 7). Recommended treatment of axSpA is a combination of pharmacological and non-pharmacological treatment including exercise interventions (8, 9), but to date, the most suitable form of exercise for individuals with axSpA is not known (8, 10). Exercise guidelines for individuals with axSpA include domains of cardiovascular-, flexibility-, and resistance training. Moreover, guidelines need to be individually tailored and be based on shared decision making (8, 10).

Traditionally, exercise interventions for individuals with axSpA involve moderate intensity exercise training, but lately some researchers have promoted the incorporation of high intensity interval training (HIIT) (11, 12). HIIT is described as high intensity exercise bouts separated by periods of active recovery phases. It has been demonstrated that regular HIIT bouts can lead to better cardiovascular health through increased  $VO_{2max}$  and improved muscle metabolism (13, 14). Moreover, studies in metabolic disorders show that HIIT has a positive effect on systemic inflammatory markers linked to the disease state (15, 16). The significance of HIIT for disease development in rheumatic disorders in general, including the response in levels of inflammatory markers, have not been widely studied. A three-month HIIT intervention improved disease symptoms and cardiovascular health, in addition to maintaining an unaltered C-reactive protein (CRP) level, in individuals with axSpA (11, 12). Levels of pro-inflammatory cytokines were not affected when measured 48 hours after the HIIT-bout (11). During extensive muscle activity, a large set of proteins, called myokines, are released which are suggested to influence many organ systems with impact on metabolism and inflammation among other functions (17, 18). One of the most studied myokines, IL-6, is immediately released in response to muscle activity (19). IL-6 is a pleiotropic cytokine with both pro-inflammatory and anti-inflammatory effects. The release of myokines, like IL-6, upon exercise is suggested to counteract inflammation and, thus, cardiovascular issues in rheumatic diseases, since systemic inflammation is an important factor in atherosclerosis (18, 20). Other examples of proteins produced by muscle and other tissues are brain derived neurotrophic factor (BDNF), a neurotrophin with effects on synapse activities in the brain (21), and vascular endothelial growth factor (VEGF) important for vascularization of skeletal muscle tissue in response to exercise training (22).

The ankylosis that features axSpA is part of bone metabolism, which extends from the formation of new bone (syndesmophytes) to osteoporosis and bone erosion (23). It is not yet fully understood whether bone metabolism with new bone formation is linked to the inflammatory process in axSpA. Moreover, changes in levels of proteins involved in bone metabolism in response to exercise training have not been studied in this disease.

To safely recommend HIIT training to individuals with axSpA, it is important to first study the acute effects after a single HIIT bout regarding disease symptoms and state of inflammation. Previous studies on the acute effect of a single bout of HIIT in healthy adults, overweight, and obese individuals have shown that changes in levels of serum inflammatory markers are dependent on exercise intensity and metabolic status (24), and that there is no severe inflammation due to muscle damage (25).

With an overall research aim to study the role for HIIT as an exercise intervention in individuals with axSpA regarding influence on systemic inflammation and bone metabolism, in addition to the effect on lowering of the risk for cardiovascular disease, we performed the herein presented study. This cross-sectional study was undertaken to provide a controlled evaluation of the acute effects of one HIIT bout on disease activity measured through serum levels of a set of inflammatory cytokines, myokines, and markers for bone metabolism in age- and sex-matched individuals with axSpA and HC.

## Methods

### Participants

The study included 21 individuals (10 females) between the ages of 18 and 50 (mean  $\pm$  SD age  $40 \pm 7$ ). Ten participants from two rheumatology clinics in southern Sweden were diagnosed with axSpA according to the Assessment in Spondylo-Arthritis international Society (ASAS) classification criteria (26) and eleven participants were age- and sex-matched HC. Exclusion criteria were cardiovascular disease, lung disease, or any other disease that reduced the physical capacity or the possibility of completing a HIIT bout. The individuals in the axSpA group, except for one participant, used anti-inflammatory drugs such as non-steroidal anti-inflammatory drugs (NSAIDs) and biologics. Participants were asked to avoid food and drinks two hours before arrival, tobacco 30 minutes before, and to avoid exercising at high intensity the evening before the testing procedures (see below) and the HIIT bout. The study was approved by the national ethical review board (Dnr: 2019–04155 and Dnr: 2022-03114-02) and followed the ethical principles of the declaration of Helsinki.

### Testing procedures

Before inclusion, electrocardiogram (ECG) and blood pressure were measured. In the event of deviating results regarding ECG and/or a blood pressure  $> 140/90$ , the person was excluded and referred to the health center for further investigation. Anthropometrics such as height was measured with a stadiometer and body composition with bioelectrical impedance analysis (InBody 770, Body Space South Korea) (Table 1). Thereafter, participants were fitted with a heart rate (HR) monitor (Polar, Finland) and the Åstrand's submaximal cycle ergometer (Monark, Vansbro, Sweden) test for prediction of maximal oxygen consumption ( $VO_{2max}$ ) was performed (27).

Table 1

Baseline data for participants with axial spondyloarthritis (axSpA) and age- and sex matched healthy controls (HC).

	axSpA (n = 10)	HC (n = 11)	p-value <sup>a</sup>
<b>Age</b>	41 (7)	40 (7)	0.705
<b>Sex</b> (men/women)	5/5	5/6	NA
<b>Skeletal muscle mass (kg)</b>	32.8 ± 5.8	33.5 ± 8.5	0.705
<b>Body fat (%)</b>	22.0 ± 5.8	19.8 ± 7.8	0.468
<b>Systolic blood pressure</b> (mm Hg)	127 (11)	128 (9)	0.710
<b>Diastolic blood pressure</b> (mm Hg)	81 (6)	80 (5)	0.824
<b>VO<sub>2</sub>max</b> (ml O <sub>2</sub> /kg/min)	39.9 ± 8.4	42.6 ± 10.7)	0.973
<b>Smoker or using tobacco in other forms</b>	3	2	NA
<b>EQ-5D, 0-1<sup>b</sup></b> health status	0.87 (0.11)	0.93 (0.10)	0.251
<b>BASDAI, 0-10<sup>c</sup></b> disease activity	1.6 (0.8)	NA	NA
<b>BASFI, 0-10<sup>c</sup></b> physical function	0.7 (0.8)	NA	NA
<b>BASG 0-10<sup>c</sup></b> wellbeing	1.5 (1.0)	NA	NA
<b>Anti-inflammatory drugs<sup>d</sup></b>	9/10	2/11	NA
<b>Drugs other than anti-inflammatory<sup>e</sup></b>	0/10	1/11	NA
Results presented as mean (± SD). <sup>a</sup> Difference between axSpA and HC analyzed with Mann-Whitney U-test. <sup>b</sup> Worst to best. <sup>c</sup> Best to worst. <sup>d</sup> Non-steroidal anti-inflammatory drugs (NSAID), TNFα-inhibitors, Corticosteroids (allergy/asthma). <sup>e</sup> Synthetic thyroid hormone. BASDAI, Bath Ankylosing Spondylitis Disease Activity Index; BASFI, Bath Ankylosing Spondylitis Functional Index; BASG, Bath Ankylosing Spondylitis Global.			

After a brief rest, participants began the HIIT-bout while wearing a HR monitor. HIIT was performed on a rpm-independent ergometer bike (LC-6 or 928E, Monark, Vansbro. Sweden) and included four high intensity intervals, each 4 minutes in duration, with 3 minutes active rest (about 70% of maximum HR) between intervals. During the high intensity intervals, the participants should reach > 17 on the rating of the perceived exertion (RPE) scale (28) and reach 90% of estimated maximum HR (HR<sub>max</sub>) any time during the interval (29). After the HIIT-bout, the participants filled out the disease-specific questionnaire Bath Ankylosing Spondylitis Disease Activity Index (BASDAI) on disease activity scoring 0–10 (best-worst) (30). For descriptive data, health status was measured with the generic questionnaire EuroQol-5

domain (EQ-5D) consisting of five questions covering mobility, self-care, usual activity, pain/discomfort, and anxiety/depression scoring 0–1 (no health to full health) (31).

Blood samples were collected before (baseline) and one hour after the HIIT-bout ended. Blood samples rested for 30 minutes followed by centrifugation for 10 minutes. The serum supernatant was aliquoted and stored at -80°C in biobank number 85 at FoU Spenshult, Halmstad. Baseline plasma levels of lipids (P-cholesterol, P-triglycerides, P-LDL, P-HDL) and glucose were analyzed at clinical chemistry departments at the nearest hospitals.

#### Serum analyses of cytokines, myokines and bone-associated proteins

Serum was analyzed with the Luminex MAGPIX® (Luminex corporation, Austin, US) and ELISA (Spectramax Plus 384, Molecular Devices, San Jose, US). Selected markers from the Human Cytokine/Chemokine/Growth Factor Panel A (HCYTA-60K), Human Myokine Panel (HMYOMAG-56K), and Human Bone Panel (HBNMAG-51K) (Merck KGaA, Darmstadt, Germany) were used for the analysis of IL-6, IL-17, IL-18, TNF $\alpha$ , CXCL-10, VEGF-A, BDNF, DKK-1, osteoprotegerin, osteocalcin, and osteopontin. The Belysa™ Immunoassay Curve Fitting Software (Merck KGaA, Darmstadt, Germany) was used for analysis of Luminex MAGPIX data. CRP and BMP-7 were analyzed with Human C-reactive protein and BMP-7 Quantikine ELISA (R&D systems INC Minneapolis, US), respectively, according to the manufacturers protocol. For ELISA data, the "Quest Graph™ Four Parameter Logistic (4PL) Curve Calculator." AAT Bioquest, Inc. <https://www.aatbio.com/tools/four-parameter-logistic-4pl-curve-regression-online-calculator> was used for the standard curve fitting. The lowest limit of detection for the different serum proteins was: IL-6 (0.18 pg/ml), IL-17 (0.89 pg/ml), IL-18 (0.42 pg/ml), CXCL-10 (2.47 pg/ml), TNF $\alpha$  (4.4 pg/ml), VEGF-A (0.91 pg/ml), BDNF (3 pg/ml), DKK-1 (1.2 pg/ml), osteoprotegerin (1.8 pg/ml), osteocalcin (37.5 pg/ml), osteopontin (15.6 pg/ml), BMP-7 (2.44 pg/ml), CRP (0.010 ng/ml). For a few samples some protein levels were below detection as indicated in Table 2.

Table 2

Serum protein levels in individuals with axial spondyloarthritis (axSpA) and age- and sex matched healthy controls (HC) at baseline and one hour after a 4 minutes x 4 times high intensity interval training (HIIT) session. All values are means  $\pm$  standard deviations.

Serum analyte	axSpA baseline (n = 10)	HC baseline (n = 11)	axSpA post-HIIT (n = 10)	HC post-HIIT (n = 11)	2*2 ANOVA p-value	Post-Hoc p-value
IL-6 (pg/ml)	2.2 $\pm$ 3.0 n = 8 <sup>a</sup>	0.4 $\pm$ 0.4 n = 10	3.2 $\pm$ 1.8 n = 8	1.9 $\pm$ 2.0 n = 10	MEpre-post 0.03	AxSpA 0.23 HC 0.04
IL-17 (pg/ml)	3.5 $\pm$ 5.1 n = 5	4.9 $\pm$ 3.1 n = 8	3.7 $\pm$ 4.5 n = 5	4.8 $\pm$ 3.2 n = 8	> 0.05	NA
IL-18 (pg/ml)	14.9 $\pm$ 7.1 n = 9	24.4 $\pm$ 14.4 n = 10	16.2 $\pm$ 6.8 n = 9	24.7 $\pm$ 14.6 n = 10	> 0.05	NA
TNF $\alpha$ (pg/ml)	51.6 $\pm$ 94.0	29.0 $\pm$ 47.9	48.2 $\pm$ 75.1	41.2 $\pm$ 88.2	> 0.05	NA
CXCL-10 (pg/ml)	82.7 $\pm$ 65.1	68.9 $\pm$ 35.4	75.7 $\pm$ 54.5	66.3 $\pm$ 31.4	> 0.05	NA
VEGF-A (pg/ml)	159 $\pm$ 138	326 $\pm$ 184	172 $\pm$ 122	325 $\pm$ 170	MEgroup 0.03	Baseline 0.031 Post 0.030
BDNF (ng/ml)	15.6 $\pm$ 5.2	15.8 $\pm$ 3.9	16.1 $\pm$ 3.9	16.3 $\pm$ 3.3	> 0.05	NA
DKK-1 (pg/ml)	1744 $\pm$ 379	1527 $\pm$ 212	1767 $\pm$ 364	1549 $\pm$ 207	> 0.05	NA
OPG (pg/ml)	280 $\pm$ 68	261 $\pm$ 39	278 $\pm$ 57	267 $\pm$ 47	> 0.05	NA
OC (ng/ml)	16.2 $\pm$ 9.1	14.5 $\pm$ 5.3 n = 10	16.1 $\pm$ 8.4	15.1 $\pm$ 6.0 n = 10	> 0.05	NA
OPN (ng/ml)	18.4 $\pm$ 14.3	17.8 $\pm$ 6.8	14.8 $\pm$ 11.1	16.7 $\pm$ 9.1	> 0.05	NA

<sup>a</sup>number of values included in the calculation if different from the total number; ME, main effect; IL-6,-17,-18, Interleukin-6,-17,-18; TNF $\alpha$ , Tumor Necrosis Factor alpha; CXCL-10, C-X-C motif chemokine ligand 10; VEGF-A, Vascular Endothelial Growth Factor-A; BDNF, Brain Derived Neurotrophic Factor; DKK-1, Dickkopf WNT signaling pathway inhibitor 1, OPG, Osteoprotegerin; OC, Osteocalcin; OPN, Osteopontin; BMP-7, Bone Morphogenetic Protein-7; CRP, C-reactive protein.

Serum analyte	axSpA baseline (n = 10)	HC baseline (n = 11)	axSpA post-HIIT (n = 10)	HC post-HIIT (n = 11)	2*2 ANOVA p-value	Post-Hoc p-value
<b>BMP-7 (pg/ml)</b>	61.6 ± 13.1	64.6 ± 20.8	75.2 ± 20.0	75.0 ± 17.8	MEpre-post < 0.001	AxSpA 0.004 HC 0.018
<b>CRP (µg/ml)</b>	1.12 ± 1.06	0.77 ± 0.85	1.11 ± 1.12	0.71 ± 0.75	> 0.05	NA
<sup>a</sup> number of values included in the calculation if different from the total number; ME, main effect; IL-6,-17,-18, Interleukin-6,-17,-18; TNFα, Tumor Necrosis Factor alpha; CXCL-10, C-X-C motif chemokine ligand 10; VEGF-A, Vascular Endothelial Growth Factor-A; BDNF, Brain Derived Neurotrophic Factor; DKK-1, Dickkopf WNT signaling pathway inhibitor 1, OPG, Osteoprotegerin; OC, Osteocalcin; OPN, Osteopontin; BMP-7, Bone Morphogenetic Protein-7; CRP, C-reactive protein.						

## Statistics

Statistical analyses were performed with SPSS (version 28.0, IBM SPSS Statistics, IBM corp. USA). Differences in baseline descriptive data was calculated with Mann-Whitney U test. A repeated measures analysis of variance (ANOVA) was used to detect differences between groups and before and after a single HIIT bout in a 2 (group) \* 2 (pre-post) design. For main effects or interactions significant at  $p \leq 0.05$ , Least Significant Difference (LSD) post-hoc analyses were performed. Data is reported as mean ± standard deviation (SD) and statistical significance was set to  $p \leq 0.05$ .

## Results

Twenty-two participants were recruited, of which one was excluded due to abnormal ECG in the clinical screening. Ten individuals diagnosed with axSpA (age  $41 \pm 7$  years), and 11 HC (mean age  $40 \pm 7$  years) completed the HIIT-bout. Physiological measurements, values of self-reported disease activity and health status, are shown in Table 1. Except for the self-reported disease parameters not applicable for HC, the participants in the two groups were well matched for body composition (skeletal muscle mass: axSpA  $32.8 \pm 5.8$ , HC  $33.5 \pm 8.5$  kg; body fat: axSpA  $22.0 \pm 5.8$ , HC  $19.8 \pm 7.8\%$ ), resting blood pressure (axSpA  $127/81 \pm 11/6$ , HC  $128/80 \pm 9/5$  mmHg) and maximal oxygen consumption ( $VO_{2max}$ ) (axSpA  $40 \pm 8$ , HC  $43 \pm 10$  mlO<sub>2</sub>/kg/min) (Table 1). Furthermore, no significant differences were observed for CRP (Table 2), plasma lipoproteins, triglycerides, and glucose (data not shown). All participants with axSpA, except one, were treated with anti-inflammatory drugs (Table 1).

## Baseline and post-HIIT levels of cytokines, myokines and bone-associated proteins



Sera from the two participating groups were analyzed for proteins involved in inflammation and bone metabolism, in addition to myokines (Table 2). A significant ANOVA pre-post main effect ( $p = 0.03$ ) was seen for IL-6 (pg/ml): axSpA 2.2 (3.0) to 3.2 (1.8) and HC 0.4 (0.4) to 1.9 (2.0). Post-hoc analyses showed increased levels of IL-6 in HC from baseline to post-HIIT ( $p = 0.04$ ) but not in axSpA ( $p = 0.23$ ). There was no difference in the level of IL-6 (pg/ml) between the axSpA and HC groups at baseline: axSpA 2.2 (3.0) and HC 0.4 (0.4) ( $p = 0.07$ ) or post-HIIT: axSpA 3.2 (1.8) and HC 1.9 (2.0) ( $p = 0.20$ ) (Table 2). We found a significant group main effect ( $p = 0.03$ ) for the level of serum VEGF-A (pg/ml), with lower levels in the axSpA group at both baseline: axSpA 159 (138) and HC 326 (184) ( $p = 0.03$ ), and post-HIIT: axSpA 172 (122) and HC 325 (170) ( $p = 0.03$ ) (Table 2). No differences in serum levels (ANOVA  $p > 0.05$ ) for any of the remaining myokines and cytokines analyzed were observed between or within the axSpA and HC groups at baseline and post-HIIT (Table 2). For the bone associated proteins a significant pre-post main effect ( $p < 0.001$ ) was found for BMP-7. The serum-concentration of BMP-7 (pg/ml) increased significantly from baseline: axSpA 61.6 (13.1) and HC 64.6 (20.8), to post-HIIT: axSpA 75.2 (20) ( $p = 0.004$ ) and HC 75.0 (17.8) ( $p = 0.02$ ) (Table 2).

## Discussion

HIIT is an exercise form that has become popular in recent years, not only in healthy populations but also in individuals with different chronic diseases. Briefly, the health benefits associated with HIIT are: increased cardiovascular capacity, lower blood pressure and improved arterial function (32–34); increased metabolic health with weight control and improved insulin sensitivity (35); increased muscle strength; and influence on mental health with reports on reduced depression and anxiety (36). HIIT has, however, not been extensively studied as an exercise intervention in individuals with various rheumatic disorders, and therefore it is important to understand the acute effects of a HIIT-bout on disease symptoms and the state of inflammation before starting a HIIT-intervention. In the present study, we investigated a set of serum markers and physiological measurements in age- and sex-matched individuals with axSpA and HC before and after one bout of HIIT with the aim of determining levels of specific serum markers in axSpA and how these are affected by one acute bout of HIIT. One goal with the present and other ongoing studies in our group, is to establish biomarkers that could be used for the evaluation of disease progression and the effect of regular HIIT bouts on the disease process to optimize and individualize HIIT-based interventions. Furthermore, due to the systemic inflammation, individuals with axSpA are at higher risk for developing cardiovascular disease and HIIT has been shown to be a possible intervention to manage the disease and comorbidities when appropriate (12).

In this study, we analyzed serum markers including pro-inflammatory cytokines, myokines, and proteins involved in bone metabolism, in addition to CRP (Table 2). Except for IL-6, no significant differences in cytokine levels were observed between the two groups at baseline or post-HIIT. The acute increase in IL-6 levels, where the levels from baseline to post-HIIT increased more in the HC than in axSpA, is consistent with previous studies on the effect of one acute bout of HIIT on serum IL-6 (37–39). We choose to measure the serum analytes one hour after the HIIT bout, since this time point may detect proteins released already during the HIIT bout, but also proteins that emerge in serum at different times after the

HIIT bout. In a preliminary study on 12 healthy young individuals (age 18–29), we showed that already at five minutes post-HIIT, the levels of IL-6 increased 2.4 times compared to the baseline level. After one hour this increase was only 1.6 times the baseline level (see Additional file 1). This is in line with previous studies (38, 39) and indicates that measuring the IL-6 serum concentration already at five minutes post HIIT may show a more exact increase in the individual production of IL-6.

The pathogenesis of axSpA is strongly dependent on the genetic linkage to HLA-B-27, but other associated genes have also been identified (1). Non-genetic factors suggested to be involved in the triggering of the disease are gut dysbiosis and mechanical stress contributing to enthesitis and pathology in the subchondral bone (2). It is believed that inflammation is primarily driving bone destruction, but there might also be an interaction between inflammation-driven factors and new bone formation ongoing in the inflamed tissue. We did not find any differences in the levels of analyzed proteins involved in bone metabolism (DKK-1, OPG, OC, OPN), except for BMP-7 where an increase in serum concentration after HIIT was found in both groups (Table 2). The bone morphogenetic proteins (BMPs) belong to the transforming growth factor (TGF)- $\beta$  family and have important roles in osteogenesis and bone formation (40). It has been suggested that new bone formation in axSpA is partly driven by BMPs (41). BMP-7 is a growth factor involved in many cellular processes, including those having anti-inflammatory effects (40, 42), but is considered most important for bone formation (43). Studies have shown an increase in levels of BMP-7 in axSpA patients (44) with a correlation to radiographic damage, which is linked to new bone formation (45, 46). To our knowledge, BMP-7 has not been studied in the context of HIIT, but it has been shown that BMP-7 levels decrease with increasing age and that exercise and subsequent release of myokines, including BMP-7, is important for bone health in the elderly (43). The fact that we observe an increase in serum concentrations of BMP-7 post HIIT, may indicate that HIIT can contribute to improvement of bone metabolism as well as inflammation through BMP-7. More research is however needed to understand the role for BMP-7 in axSpA with regards to pathological bone formation versus anti-inflammatory features.

VEGF-A is a main signaling factor in angiogenesis by regulating proliferation of endothelial cells and vascular permeability. It is released by many different cell types and can activate inflammatory cells and the production of pro-inflammatory mediators. The level of VEGF in healthy adults has been shown not to be influenced by HIIT in the short term, which is in line with the present results (47, 48). In individuals with axSpA VEGF-levels are elevated, but the levels are significantly decreased by anti-inflammatory treatment such as TNF $\alpha$  inhibitors (49, 50). The participants with axSpA in the present study had lower levels of VEGF-A compared to the HCs, which could be explained by the fact that all, but one, were on treatment with anti-inflammatory drugs. Similarly, there was no difference in the baseline levels of CRP between the two groups. Reports from studies on the effect of HIIT on CRP levels have shown conflicting results probably due to a variation in disease status and group size (36, 51). In the present study there was no significant difference in CRP levels between the axSpA and HC groups and we did not observe any changes in CRP levels from baseline to post-HIIT, which suggests that one bout of HIIT does not influence the general inflammatory status and can be safely performed.

The present study was performed to address the impact of one single bout of HIIT on serum levels of inflammatory cytokines, myokines, and markers for bone metabolism in individuals with axSpA and HC. One limitation of the study is the small group sizes, which however were well matched regarding sex, age, and physiological measures. Due to the small groups, there was not sufficient power in the calculations to analyze sex differences for the different serum markers. In addition, all participants with axSpA, except one, were treated with anti-inflammatory drugs, which had a considerable impact on certain serum markers.

## Conclusion

In this study we investigated how one bout of HIIT acutely affect serum levels of specific pro- and anti-inflammatory cytokines, myokines, and proteins involved in bone metabolism in age- and sex-matched individuals with axSpA compared to HC. From the herein presented results we conclude that one bout of HIIT significantly increases the serum concentrations of IL-6 and the bone associated protein BMP-7 in both groups, but that the concentrations of none of the other proteins investigated, including CRP and VEGF-A, were significantly changed.

## Abbreviations

axSpA	Axial spondyloarthritis
BDNF	Brain derived neurotrophic factor
BMP-7	Bone morphogenetic protein-7
CRP	C-reactive protein
DKK-1	Dickkopf-related protein 1
HC	Healthy controls
HIIT	High-intensity interval training
IL-6, -17, -18	Interleukin-6, -17, -18
NSAID	Non-steroidal anti-inflammatory drugs
OC	Osteocalcin
OPG	Osteoprotegerin
OPN	Osteopontin
TNF $\alpha$	Tumor Necrosis factor alpha

VEGF            Vascular endothelial growth factor

## Declarations

### Ethics approval and consent to participation

The study was approved by the Swedish Ethics Review Authority (Dnr: 2019-04155 and Dnr: 2022-03114-02) and performed in accordance with the Declaration of Helsinki. Each participant signed an informed consent for participation in the study.

### Consent for publication

Not applicable

### Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

### Competing interests

The authors declare that they have no competing interests.

### Funding

Open access funding was provided by Halmstad University. This work was supported by grants from The Swedish Rheumatism Association and The Norrbacka-Eugenia foundation.

### Authors contributions

ÅA, EH, MCO conceived and designed the study. All authors participated in the acquisition of data. ÅA performed the serum analyses and ÅA, MCO and EH drafted the manuscript. MCO and EH performed the statistical analyses. All authors contributed to the interpretation of the results. All authors read and approved the final manuscript.

### Acknowledgements

We would like to thank the participants in the study who provided their informed consent to use their data for research, the staff at the involved Hospitals, and Emma Berthold, Maria Imberg, Alexandra Köllerfors, Belinda Wu, and Midya Tasin for technical assistance.

## References

1. Sieper J, Poddubnyy D. Axial spondyloarthritis. Lancet. 2017;390(10089):73–84.

2. Navarro-Compán V, Sepriano A, El-Zorkany B, van der Heijde D. Axial spondyloarthritis. *Ann Rheum Dis.* 2021;80(12):1511–21.
3. Sieper J, van der Heijde D, Review. Nonradiographic axial spondyloarthritis: new definition of an old disease? *Arthritis Rheum.* 2013;65(3):543–51.
4. Walsh JA, Magrey M. Clinical Manifestations and Diagnosis of Axial Spondyloarthritis. *J Clin Rheumatol.* 2021;27(8):e547–e60.
5. Toussiot E. The Risk of Cardiovascular Diseases in Axial Spondyloarthritis. *Curr Insights Front Med (Lausanne).* 2021;8:782150.
6. Zhao SS, Robertson S, Reich T, Harrison NL, Moots RJ, Goodson NJ. Prevalence and impact of comorbidities in axial spondyloarthritis: systematic review and meta-analysis. *Rheumatology (Oxford).* 2020;59(Suppl4):iv47–iv57.
7. Rueda-Gotor J, Ferraz-Amaro I, Genre F, Gonzalez Mazon I, Corrales A, Portilla V, et al. Cardiovascular and disease-related features associated with extra-articular manifestations in axial spondyloarthritis. A multicenter study of 888 patients. *Semin Arthritis Rheum.* 2022;57:152096.
8. Rausch Osthoff AK, Niedermann K, Braun J, Adams J, Brodin N, Dagfinrud H, et al. 2018 EULAR recommendations for physical activity in people with inflammatory arthritis and osteoarthritis. *Ann Rheum Dis.* 2018;77(9):1251–60.
9. van der Heijde D, Ramiro S, Landewe R, Baraliakos X, Van den Bosch F, Sepriano A, et al. 2016 update of the ASAS-EULAR management recommendations for axial spondyloarthritis. *Ann Rheum Dis.* 2017;76(6):978–91.
10. Martey C, Sengupta R. Physical therapy in axial spondyloarthritis: guidelines, evidence and clinical practice. *Curr Opin Rheumatol.* 2020;32(4):365–70.
11. Sveaas SH, Berg IJ, Provan SA, Semb AG, Hagen KB, Vollestad N, et al. Efficacy of high intensity exercise on disease activity and cardiovascular risk in active axial spondyloarthritis: a randomized controlled pilot study. *PLoS ONE.* 2014;9(9):e108688.
12. Sveaas SH, Bilberg A, Berg IJ, Provan SA, Rollefstad S, Semb AG, et al. High intensity exercise for 3 months reduces disease activity in axial spondyloarthritis (axSpA): a multicentre randomised trial of 100 patients. *Br J Sports Med.* 2020;54(5):292–7.
13. Gibala MJ, Little JP, Macdonald MJ, Hawley JA. Physiological adaptations to low-volume, high-intensity interval training in health and disease. *J Physiol.* 2012;590(5):1077–84.
14. Gallo-Villegas J, Aristizabal JC, Estrada M, Valbuena LH, Narvaez-Sanchez R, Osorio J, et al. Efficacy of high-intensity, low-volume interval training compared to continuous aerobic training on insulin resistance, skeletal muscle structure and function in adults with metabolic syndrome: study protocol for a randomized controlled clinical trial (Intraining-MET). *Trials.* 2018;19(1):144.
15. Leiva-Valderrama JM, Montes-de-Oca-Garcia A, Opazo-Diaz E, Ponce-Gonzalez JG, Molina-Torres G, Velazquez-Diaz D et al. Effects of High-Intensity Interval Training on Inflammatory Biomarkers in Patients with Type 2 Diabetes. A Systematic Review. *Int J Environ Res Public Health.* 2021;18(23).

16. Reljic D, Dieterich W, Herrmann HJ, Neurath MF, Zopf Y. HIIT the Inflammation: Comparative Effects of Low-Volume Interval Training and Resistance Exercises on Inflammatory Indices in Obese Metabolic Syndrome Patients Undergoing Caloric Restriction. *Nutrients*. 2022;14(10).
17. Benatti FB, Pedersen BK. Exercise as an anti-inflammatory therapy for rheumatic diseases-myokine regulation. *Nat Rev Rheumatol*. 2015;11(2):86–97.
18. Bay ML, Pedersen BK. Muscle-Organ Crosstalk: Focus on Immunometabolism. *Front Physiol*. 2020;11:567881.
19. Steensberg A, Fischer CP, Keller C, Moller K, Pedersen BK. IL-6 enhances plasma IL-1ra, IL-10, and cortisol in humans. *Am J Physiol Endocrinol Metab*. 2003;285(2):E433–7.
20. Libby P, Hansson GK. From Focal Lipid Storage to Systemic Inflammation: JACC Review Topic of the Week. *J Am Coll Cardiol*. 2019;74(12):1594–607.
21. Kowianski P, Lietzau G, Czuba E, Waskow M, Steliga A, Morys J. BDNF: A Key Factor with Multipotent Impact on Brain Signaling and Synaptic Plasticity. *Cell Mol Neurobiol*. 2018;38(3):579–93.
22. Olfert IM, Baum O, Hellsten Y, Egginton S. Advances and challenges in skeletal muscle angiogenesis. *Am J Physiol Heart Circ Physiol*. 2016;310(3):H326–36.
23. Baraliakos X, Ostergaard M, Lambert RG, Eshed I, Machado PM, Pedersen SJ et al. MRI lesions of the spine in patients with axial spondyloarthritis: an update of lesion definitions and validation by the ASAS MRI working group. *Ann Rheum Dis*. 2022.
24. Dorneles GP, Haddad DO, Fagundes VO, Vargas BK, Kloecker A, Romao PR, et al. High intensity interval exercise decreases IL-8 and enhances the immunomodulatory cytokine interleukin-10 in lean and overweight-obese individuals. *Cytokine*. 2016;77:1–9.
25. Rohnejad B, Monazzami A. Effects of high-intensity intermittent training on some inflammatory and muscle damage indices in overweight middle-aged men. *Apunts Sports Medicine*. 2023;58:100404.
26. Rudwaleit M, van der Heijde D, Landewe R, Listing J, Akkoc N, Brandt J, et al. The development of Assessment of SpondyloArthritis international Society classification criteria for axial spondyloarthritis (part II): validation and final selection. *Ann Rheum Dis*. 2009;68(6):777–83.
27. Astrand PO, Cuddy TE, Saltin B, Stenberg J. Cardiac Output during Submaximal and Maximal Work. *J Appl Physiol*. 1964;19:268–74.
28. Borg G. Borg's perceived exertion and pain scales. *Human kinetics*; 1998.
29. Taylor JL, Holland DJ, Spathis JG, Beetham KS, Wisloff U, Keating SE, et al. Guidelines for the delivery and monitoring of high intensity interval training in clinical populations. *Prog Cardiovasc Dis*. 2019;62(2):140–6.
30. Garrett S, Jenkinson T, Kennedy LG, Whitelock H, Gaisford P, Calin A. A new approach to defining disease status in ankylosing spondylitis: the Bath Ankylosing Spondylitis Disease Activity Index. *J Rheumatol*. 1994;21(12):2286–91.
31. EuroQol G. EuroQol—a new facility for the measurement of health-related quality of life. *Health Policy*. 1990;16(3):199–208.

32. Gillen JB, Martin BJ, MacInnis MJ, Skelly LE, Tarnopolsky MA, Gibala MJ. Twelve Weeks of Sprint Interval Training Improves Indices of Cardiometabolic Health Similar to Traditional Endurance Training despite a Five-Fold Lower Exercise Volume and Time Commitment. *PLoS ONE*. 2016;11(4):e0154075.
33. Ramos JS, Dalleck LC, Tjonna AE, Beetham KS, Coombes JS. The impact of high-intensity interval training versus moderate-intensity continuous training on vascular function: a systematic review and meta-analysis. *Sports Med*. 2015;45(5):679–92.
34. Edwards JJ, Griffiths M, Deenmamode AHP, O'Driscoll JM. High-Intensity Interval Training and Cardiometabolic Health in the General Population: A Systematic Review and Meta-Analysis of Randomised Controlled Trials. *Sports Med*. 2023.
35. Mateo-Gallego R, Madinaveitia-Nisarre L, Gine-Gonzalez J, Maria Bea A, Guerra-Torrecilla L, Baila-Rueda L, et al. The effects of high-intensity interval training on glucose metabolism, cardiorespiratory fitness and weight control in subjects with diabetes: Systematic review a meta-analysis. *Diabetes Res Clin Pract*. 2022;190:109979.
36. Martland R, Mondelli V, Gaughran F, Stubbs B. Can high-intensity interval training improve physical and mental health outcomes? A meta-review of 33 systematic reviews across the lifespan. *J Sports Sci*. 2020;38(4):430–69.
37. Croft L, Bartlett JD, MacLaren DP, Reilly T, Evans L, Matthey DL, et al. High-intensity interval training attenuates the exercise-induced increase in plasma IL-6 in response to acute exercise. *Appl Physiol Nutr Metab*. 2009;34(6):1098–107.
38. Proschinger S, Schenk A, Wessels I, Donath L, Rappelt L, Metcalfe AJ et al. Intensity- and time-matched acute interval and continuous endurance exercise similarly induce an anti-inflammatory environment in recreationally active runners: focus on PD-1 expression in T(regs) and the IL-6/IL-10 axis. *Eur J Appl Physiol*. 2023.
39. Zwetsloot KA, John CS, Lawrence MM, Battista RA, Shanely RA. High-intensity interval training induces a modest systemic inflammatory response in active, young men. *J Inflamm Res*. 2014;7:9–17.
40. Aluganti Narasimhulu C, Singla DK. The Role of Bone Morphogenetic Protein 7 (BMP-7) in Inflammation in Heart Diseases. *Cells*. 2020;9(2).
41. Descamps E, Molto A, Borderie D, Lories R, Richard CM, Pons M, et al. Changes in bone formation regulator biomarkers in early axial spondyloarthritis. *Rheumatology (Oxford)*. 2021;60(3):1185–94.
42. Singla DK, Singla R, Wang J. BMP-7 Treatment Increases M2 Macrophage Differentiation and Reduces Inflammation and Plaque Formation in Apo E<sup>-/-</sup> Mice. *PLoS ONE*. 2016;11(1):e0147897.
43. Kwon JH, Moon KM, Min KW. Exercise-Induced Myokines can Explain the Importance of Physical Activity in the Elderly: An Overview. *Healthc (Basel)*. 2020;8(4).
44. Mahmoud A, Fayez D, Gabal MM, Hamza SM, Badr T. Insight on Bone Morphogenetic Protein 7 in Ankylosing Spondylitis and its association with disease activity and radiographic damage. *Electron Physician*. 2016;8(7):2670–8.

45. Park MC, Park YB, Lee SK. Relationship of bone morphogenetic proteins to disease activity and radiographic damage in patients with ankylosing spondylitis. *Scand J Rheumatol*. 2008;37(3):200–4.
46. Chen HA, Chen CH, Lin YJ, Chen PC, Chen WS, Lu CL, et al. Association of bone morphogenetic proteins with spinal fusion in ankylosing spondylitis. *J Rheumatol*. 2010;37(10):2126–32.
47. Kliszciewicz B, Markert CD, Bechke E, Williamson C, Clemons KN, Snarr RL, et al. Acute Effect of Popular High-Intensity Functional Training Exercise on Physiologic Markers of Growth. *J Strength Cond Res*. 2021;35(6):1677–84.
48. Manferdelli G, Freitag N, Doma K, Hackney AC, Predel HG, Bloch W, et al. Acute Hormonal Responses to High-Intensity Interval Training in Hyperoxia. *J Hum Kinet*. 2020;73:125–34.
49. Appel H, Janssen L, Listing J, Heydrich R, Rudwaleit M, Sieper J. Serum levels of biomarkers of bone and cartilage destruction and new bone formation in different cohorts of patients with axial spondyloarthritis with and without tumor necrosis factor-alpha blocker treatment. *Arthritis Res Ther*. 2008;10(5):R125.
50. Pedersen SJ, Hetland ML, Sorensen IJ, Ostergaard M, Nielsen HJ, Johansen JS. Circulating levels of interleukin-6, vascular endothelial growth factor, YKL-40, matrix metalloproteinase-3, and total aggrecan in spondyloarthritis patients during 3 years of treatment with TNFalpha inhibitors. *Clin Rheumatol*. 2010;29(11):1301–9.
51. Khalafi M, Symonds ME. The impact of high-intensity interval training on inflammatory markers in metabolic disorders: A meta-analysis. *Scand J Med Sci Sports*. 2020;30(11):2020–36.

## Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [AdditionalFile1HIITaxSpAAnderssonetal231105.docx](#)