

Systematic Analysis of the Functions and Prognostic Values of Rna Binding Protein in Head and Neck Squamous Cell Carcinoma

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Abstract

Background: Dysregulation of RNA-binding proteins (RBPs) plays an important role in controlling processes in cancer development. However, the function of RBPs has not been thoroughly and systematically documented in head and neck cancer. We aim to explore the role of RPB in the pathogenesis of HNSC.

Methods: We obtained HNSC gene expression data and corresponding clinical information from The Cancer Genome Atlas (TCGA) and the GEO databases, and identified aberrantly expressed RBPs between tumors and normal tissues. Meanwhile, we performed a series of bioinformatics to explore the function and prognostic value of these RBPs.

Results: A total of 249 abnormally expressed RBPs were identified, including 101 down-regulated RBPs and 148 up-regulated RBPs. Using least absolute shrinkage and selection operator (LASSO) and univariate Cox regression analysis, the fifteen RBPs were identified as hub genes. With the fifteen RBPs, the prognostic prediction model was constructed. Further analysis showed that the high-risk score of the patients expressed a better survival outcome. The prediction model was validated in another HNSC dataset, and similar findings were observed.

Conclusions: Our results provide novel insights into the pathogenesis of HNSC. The fifteen RBP gene signature exhibited the predictive value of moderate HNSC prognosis, and have potential application value in clinical decision-making and individualized treatment.

Background

Head and Neck squamous cell carcinoma (HNSC) is one of the common types of malignancy, which is characterized by advanced diagnosis, a higher rate of lymph node metastasis, and high recurrence [1-3]. According to the previously published global cancer statistics report, there are approximately 400,000 oral and pharyngeal diseases, 160,000 laryngeal cancers, and 300,000 deaths worldwide each year [4-6]. The prognosis of patients with head and neck cancer stage III/IV is not ideal. The 5-year recurrence-free survival (RFS) and overall survival (OS) are 40% and 60%, respectively [7]. Although the diagnosis and treatment of HNSC have achieved great progress in the past few decades, there is still a lack of effective therapies for patients. Therefore, there is an urgent need for new treatments to enrich the treatment of head and neck cancer and improve the survival rate of head and neck cancer patients, which could further understand the molecular biology basis of HNC.

RNA binding protein (RBP) is essentially a pleiotropic protein and regulates gene expression at the post-transcription level through interaction with target RNA [8, 9]. Through high-throughput screening, a total of 1542 RBPs were identified in human cells, accounting for 7.5% of all protein-coding genes [8]. Studies have shown that many diseases are related to abnormal expression of RBP, including abnormal metabolism, germ cell development, muscular dystrophy, the human fragile X syndrome, nervous system diseases, and cancer [10-14]. However, only a small fraction of RBPs have been studied intensively and found to have key roles in cancers to date. Therefore, we collected HNSC data from the Cancer Genome Atlas (TCGA) database and conducted a systematic analysis to examine the potential molecular function and clinical significance of RBPs in HNSC.

Material And Methods

Data source and data Preprocessing

Firstly, the transcriptome sequencing data with the HTSeq-FPKM format of 546 samples were downloaded from the TCGA database by the GDC tool (<https://portal.gdc.cancer.gov/>) as train group, including 502 HNSC patients and 44

adjacent tumor samples. Besides, the clinical information of each patient, including age, gender, TNM stage, tumor grade, and survival time, is obtained from the GDC portal. Finally, a total of 501 samples were considered eligible in this analysis. Secondly, another HNSC dataset was also downloaded from the GEO dataset (GSE84437[15]) and serves as an external verification group.

Differentially expressed analysis

The differentially expressed RPBs between adjacent tumor samples and tumor samples were screened using the “limma” package in R [16]. The differentially expressed RPBs of data sets with $|\log_2 \text{fold change}| \geq 0.5$ and a P-value less than 0.05 was considered as the selection criteria for subsequent analysis. The heatmap plot was also exhibited.

Functional enrichment analysis

To understand the underlying biological processes and pathways of aberrantly expressed RPBs, the biological functions are systematically analyzed through gene ontology (GO) enrichment, which includes three terms: molecular function, biological process, and cellular composition. The Kyoto Encyclopedia of Genes and Genomes (KEGG) is used to detect potential biological pathways for differentially expressed RPBs. Functional enrichment analysis was conducted by “ClusterProfiler” of R package with the cut-off value of P less than 0.05 [17].

Construction of protein-protein interaction network and identification of sub-modules

The protein-protein interactions (PPIs) among differentially expressed RPBs were detected using the String website, and the PPIs network was constructed by Cytoscape 3.6.2 software. By using the MCODE (Molecular Complex Detection) plugin in Cytoscape, we can identify sub-modules from the PPI network with a score > 9 and a node count > 10 . Functional enrichment analysis for each module was conducted by the “clusterProfiler” of R package with the cut-off value of P less than 0.05.

The screen of prognostic RPBs

Firstly, the univariate COX regression analysis was conducted to screen the prognostic RPBs with the threshold of a P value less than 0.05. Then, the least absolute shrinkage and selection operator (LASSO) was further used to identify the prognostic significance of candidate RPBs [18].

Construction of prognostic prediction model

With the candidate RPBs identified by LASSO, we built a prognostic prediction model. The regression coefficients from the LASSO algorithms were used to create a classification index for each sample, and used the following formula to weight the expression value of the selected miRNA: The Prognostic Index = (Exp mRNA1* Coef1) + (Exp mRNA2* Coef2) + (Exp mRNA3* Coef3) + ... + (Exp mRNA_n* Coef_n).

Based on the median of the prognostic index, the HNSC samples were divided into two groups: high-risk group and low-risk group. We used the Kaplan-Meier analysis and log-rank test to evaluate the difference in overall survival (OS) between the two groups. To evaluate the performance of this prognostic model, receive operating characteristic (ROC) curves were used to compare the sensitivity and specificity of the survival prediction. The ROC curves were plotted using the “survivalROC” package in R. Univariate and multivariate COX prognostic analysis was used to determine whether the established prognostic prediction model was an independent prognostic factor for HNSC.

Validation Hub RPBs in HPA database

The protein expression of the 15 RPBs between HNSC and normal tissues was determined at a translational level using immunohistochemistry (IHC) from the Human Protein Atlas database (HPA, <https://www.proteinatlas.org/>)[19].

Validation of the prognostic prediction model

The established prognostic prediction model was validated in another GEO (GSE65858), and the KM survival curve, ROC curve, and risk curve was plotted. And we also evaluate whether the established prognostic prediction model was an independent prognostic factor for HNSC.

Results

Identification of Differentially Expressed RBPs

The flowchart was shown in Figure 1. A total of 502 LUSC samples and 44 adjacent tumor samples were considered eligible in the study. Differentially expressed RBPs were screened by the “limma” package of R software. In total, 249 differentially expressed RBPs were identified, including 104 downregulated RBPs and 145 up-regulated RBPs (Supplementary Table 1). The heatmap and volcano plot was exhibited in Figure 2A,B.

GO and KEGG Analysis

To uncover the potential function and molecular mechanism of differentially expressed RBPs, they were divided into two groups (upregulated and downregulated group) according to their expression levels. Functional enrichment analysis was carried out via the hypergeometric test using the ClusterProfiler R package. Upregulated differentially expressed RBPs were notably enriched in biological processes, including metabolic process, ncRNA processing, ribonucleoprotein granule, and spliceosomal complex. The downregulated differentially expressed RBPs were enriched in the mRNA processing, RNA splicing, ribosome, and translation repressor activity (Table 1, Figure 3A,B). The results of KEGG analysis for upregulated RBPs revealed that a total of nine KEGG terms was enriched, including Spliceosome, RNA transport and Ribosome biogenesis in eukaryotes, whereas the downregulated RBPs were significantly enriched in RNA transport, mRNA surveillance pathway and RNA degradation (Table 1, Figure 3C,D).

Construction of PPI Network and identification of Key Modules

The PPI network was established using these differentially expressed RBPs in the STRING database, then a protein-protein co-expression network was visualized using Cytoscape. The PPI network for RBPs was exhibited in Figure 4A, which included 223 nodes and 1,094 edges. The two hub networks were identified via the plug-in MODE in Cytoscape. Module I included 22 genes, while Module II included 33 genes (Figure 4B,C,D). The GO and KEGG analysis in Module 1 was mainly enriched in RNA splicing, ncRNA processing, ribosome biogenesis, RNA polymerase, and NOD-like receptor signaling pathway, while the GO and KEGG analysis in Module 2 were mainly enriched in mitochondrial translation, snoRNA metabolic process, nuclear RNA surveillance, Ribosome, and RNA degradation.

Selection of Prognostic-Related RBPs

Among 249 differentially expressed RBPs, 24 RBPs were regarded as prognostic-related RBPs using the univariate Cox regression analysis (Figure 5). Then, we used the LASSO method to build a prediction model, selected non-zero regression coefficients to identify the optimal RBPs gene set, and used the selected RBPs to build RBPs-based predictive model to predict prognosis. In LASSO logistic regression method, fifteen RBPs as optimal features were ultimately recognized, including DNMT1, MRPL33, EZH2, PCF11, RBM24, TRMT112, DZIP1, EIF5A2, MKRN3, DARS2, PSMA6, AZGP1, LENG9, IGF2BP2, and CIRBP. The RBPs-based a predictive model was calculated as the following

formula: The risk score of each HNSC patient was computed according to the following formula: $DNMT1 * (-0.21475) + MRPL33 * 0.177058 + EZH2 * (-0.12256) + PCF11 * (-0.23465) + RBM24 * 0.08662 + TRMT112 * 0.091039 + DZIP1 * 0.025207 + EIF5A2 * 0.147603 + MKRN3 * 0.495024 + DARS2 * 0.259781 + PSMA6 * 0.161435 + AZGP1 * (-0.0518) + LENG9 * (-0.14885) + IGF2BP2 * (-0.027825) + CIRBP * 0.00951$.

To assess the predictive ability of this model, the patients were divided into high- and low-risk groups for survival analysis according to the median risk score. Patients in the high-risk group exhibited better survival rates than those in the low-risk group (Figure 6A). The time-dependent ROC analysis was used to evaluate the prognostic performance of the 15-RBP gene signature. The AUC of the ROC curve for OS was 0.67 (0.62-72) at 1 years, 0.68 (0.63-0.73) at 3 years and 0.67 (0.60-0.74) at 5 years (Figure 6B). The result indicated that the model had moderate diagnostic strength. The expression heat map and survival status of patients with the 15-RBP gene biomarkers in the low- and high-risk subgroups are shown in Figure 6C-E.

Univariate and multivariate Cox regression analysis suggested that the prediction model could become an independent predictor, including age, gender, histological grade, stage, TNM stage, and risk score (Figure 7, Table 3).

Validation of candidate RPBs in the HPA dataset

To further determine the expression of these hub genes in HNSC, we used immunohistochemistry results from the Human Protein Atlas (HPA) database to show that AZGP1, DNMT1, DZIP1, IGF2BP2, LENG9, MRPL33, PSMA6, RBM24, and TRMT112 were significantly increased in head and neck cancer compared with normal tissue (Figure 8).

Validation of established prognostic RPB model

The established prognostic RPB model based on the 15 RPBs was validated in another GEO (GSE84437) dataset using the same risk assessment formula, and the result indicated that the model exhibited better predictive ability. Patients in the high-risk group suffered a lower survival probability (Figure 9A). The area under the curve of the ROC curve for 1, 3, 5 years reached 0.58, 0.55, and 0.58, respectively (Figure 9B). The expression heat map and survival status of patients with the 15-RBP gene biomarkers in the low- and high-risk subgroups are shown in Figure 9C-E. Univariate and multivariate Cox regression analysis suggested that the IRGPM could become an independent predictor, including age, gender, histological grade, stage, TNM stage, and risk score (Figure 10A,B).

Discussion

RNA binding protein (RBP) is a general term for the protein that binds to RNA, which regulates the metabolic process of RNA. RBP accompanies the life of RNA, and its main role is to mediate RNA maturation, transport, positioning, and translation. One RBP may have multiple target mRNAs, and its defective expression may cause various diseases. The intricate network of interactions between RBP and its cancer-associated RNA targets will provide a better understanding of tumor biology and may reveal new cancer treatment targets [8, 20]. Recently, several studies have documented that RBPs are dysregulated in various human cancers [21-23]. However, little is known about the expression pattern and mechanism of action of RBPs in HNSC. At the work, we comprehensively analyzed the key RBPs specifically associated with the overall survival of HNSC using bioinformatics analysis. The differentially expressed RBPs between adjacent tumor samples and tumor samples were identified. Meanwhile, a PPI network was constructed based on the RPBs, and the two hub networks were identified via the plug-in MODE in Cytoscape software. Then, we used the univariate Cox regression analysis to screen prognosis-related RBPs and constructed a risk model to predict HNSC prognosis based on a fifteen-RBPs signature.

The biological processes or pathways that upregulated RBPs were significantly enriched in metabolic process, ncRNA processing, ribonucleoprotein granule, and spliceosomal complex. The downregulated differentially expressed RBPs were enriched in the mRNA processing, RNA splicing, ribosome, and translation repressor activity. These results were lined with previous studies. RNA binding proteins are involved in RNA metabolism[24] and play an important role in regulating RNA stability, alternative splicing, modification, localization, and translation[25]. Changing the expression of RBPs in cells may affect various physiological processes in cells, such as alternative splicing and apoptosis [26, 27]. The RNA binding domain (RBD) is a region of RBP, which has been experimentally or structurally confirmed to directly bind RNA[8]. Meanwhile, a large number of studies have recently reported the role of abnormal RNA metabolism and RNA processing, alternative splicing in various diseases, including cancer development[28, 29]. These results further confirm that RBP can affect the growth and metastasis of tumor cells by regulating various biological processes (such as RNA metabolism and RNA processing, RNA transport, RNA degradation).

Subsequently, the univariate Cox regression analysis and Lasso regression analysis were used to identifying the prognostic RPBs, and fifteen RPBs were defined as hub genes. Among these RPBs, DNMT1, MRPL33, EZH2, PCF11, RBM24, TRMT112, DZIP1, EIF5A2, MKRN3, DARS2, PSMA6, AZGP1, LENG9, IGF2BP2, and CIRBP were considered as candidate genes. The DNMT1 was involved in the development of various tumors, including pancreatic ductal adenocarcinoma[30], breast cancer[31], and cervical cancer[32]. MRPL33 was closely related to the incidence of head and neck cancer[33], gastric cancer[34], and breast cancer[35]. EZH2, PCF11, RBM24, DZIP1, EIF5A2, MKRN3, PSMA6, AZGP1, IGF2BP2, and CIRBP was also reported to involve in the development of tumor[36-39].

Conclusions

our research provides relatively novel insights into the role of RBPs in the occurrence and development of HNSC. Besides, our prognosis model shows good predictive performance in terms of survival, which may help to develop new prognostic indicators for HNSC. Also, RBP-related gene markers show a key biological background, which indicates that these RBPs can be used for clinical adjuvant therapy. Furthermore, survival analysis was also performed based on the univariate and multivariate Cox proportional hazards regression models.

Declarations

Ethics approval and consent to participate

Not applicable

Consent for publication

Not applicable

Availability of data and material

Data availability could be obtained from TCGA website.

Competing interests

The authors declare that they have no competing interests.

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Author contributions:

J.K.S;F.L; B.L.H;X.H.Y wrote the main manuscript text;

J.K.S; K.Z;X.L.C prepared Figures 1-9;

J.K.S; X.F.D contributed to data analysis;

All authors reviewed the manuscript.

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Tables

Table 1: Functional enrichment analysis for differentially expressed RPBs.

Differentially expressed RPBs	Terms	P-value
Down-regulated RPBs		
Biological processes	RNA splicing	P<0.001
	regulation of RNA splicing	P<0.001
	regulation of mRNA processing	P<0.001
	regulation of mRNA metabolic process	P<0.001
	regulation of mRNA splicing, via spliceosome	P<0.001
Cellular component	RNA cap binding complex	P<0.001
	ribosome	P<0.001
	cytoplasmic stress granule	P<0.001
	cytoplasmic ribonucleoprotein granule	P<0.001
	ribonucleoprotein granule	P<0.001
Moleccular function	mRNA 3'-UTR binding	P<0.001
	translation factor activity, RNA binding	P<0.001
	single-stranded RNA binding	P<0.001
	poly(A) binding	P<0.001
	poly(U) RNA binding	P<0.001
KEGG pathway	RNA transport	P<0.001
	mRNA surveillance pathway	P<0.001
	RNA degradation	P<0.001
Up-regulated RPBs		
Biological processes	RNA catabolic process	P<0.001
	defense response to virus	P<0.001
	RNA phosphodiester bond hydrolysis	P<0.001
	response to virus	P<0.001
	regulation of translation	P<0.001
Cellular component	cytoplasmic ribonucleoprotein granule	P<0.001
	ribonucleoprotein granule	P<0.001
	P-body	P<0.001
	telomerase holoenzyme complex	P<0.001
	spliceosomal complex	P<0.001

Molecular function	catalytic activity, acting on RNA	P<0.001
	RNA helicase activity	P<0.001
	helicase activity	P<0.001
	double-stranded RNA binding	P<0.001
	ribonuclease activity	P<0.001
KEGG pathway	Spliceosome	P<0.001
	RNA degradation	P<0.001
	RNA transport	P<0.001
	mRNA surveillance pathway	P<0.001
	Measles	P<0.001

Table 2: Univariate Cox regression analysis to identify prognosis-relatedRBPs.

RPBs	HR	HR.95L	HR.95H	P-value
DNMT1	0.66792	0.506998	0.87992	0.004111
MRPL33	1.568354	1.147441	2.143669	0.004764
EZH2	0.657414	0.505662	0.854708	0.001734
PCF11	0.508046	0.332979	0.775157	0.001681
PPARGC1B	0.576362	0.379043	0.8764	0.009967
RBM24	1.191707	1.011945	1.403401	0.035527
TRMT112	1.442278	1.0616	1.959463	0.019166
NUTF2	1.453638	1.039026	2.033697	0.029005
QARS	0.669615	0.461679	0.971202	0.034516
DZIP1	1.418783	1.023762	1.966223	0.035639
EIF5A2	1.526215	1.169747	1.991313	0.001838
MKRN3	2.306887	1.462073	3.639851	0.000328
DARS2	1.402555	1.018074	1.932236	0.038495
MBNL3	0.676222	0.493429	0.926732	0.014966
PSMA6	1.550197	1.122482	2.140889	0.007781
CPEB3	0.586725	0.344384	0.999602	0.049829
RBM47	0.78889	0.626623	0.993176	0.043566
ADARB2	0.534365	0.307906	0.927381	0.025881
MKRN2	0.651642	0.425701	0.997501	0.048669
AZGP1	0.88812	0.79568	0.9913	0.034363
LENG9	0.721531	0.533861	0.975172	0.033709
SIDT1	0.624264	0.405671	0.960645	0.03215
IGF2BP2	1.208821	1.047008	1.395641	0.009696
CIRBP	0.734818	0.552169	0.977884	0.034571

Table 3: The prognostic effect of different clinical parameters.

Index	Univariate analysis				Multivariate analysis			
	HR	95%Upper CI	95%Lower CI	P-value	HR	95%Upper CI	95%Lower CI	P-value
age	1.019682	1.004522	1.035071	0.010764	1.021692	1.005341	1.038308	0.00913
gender	0.744786	0.530085	1.046448	0.089453	0.867827	0.607693	1.239316	0.435525
grade	1.218909	0.948102	1.567067	0.122532	1.041376	0.791244	1.370583	0.772366
stage	1.678967	1.330204	2.119171	1.29E-05	1.180157	0.79929	1.742511	0.404758
T	1.378889	1.164345	1.632965	0.000197	1.148962	0.895807	1.473659	0.274177
M	1.088512	0.924793	1.281214	0.307814	1.049855	0.887407	1.24204	0.570545
N	1.551352	1.306974	1.841423	5.14E-07	1.39208	1.116608	1.735513	0.003279
riskScore	1.763024	1.470252	2.114095	9.36E-10	1.718394	1.377301	2.14396	1.62E-06

Figures

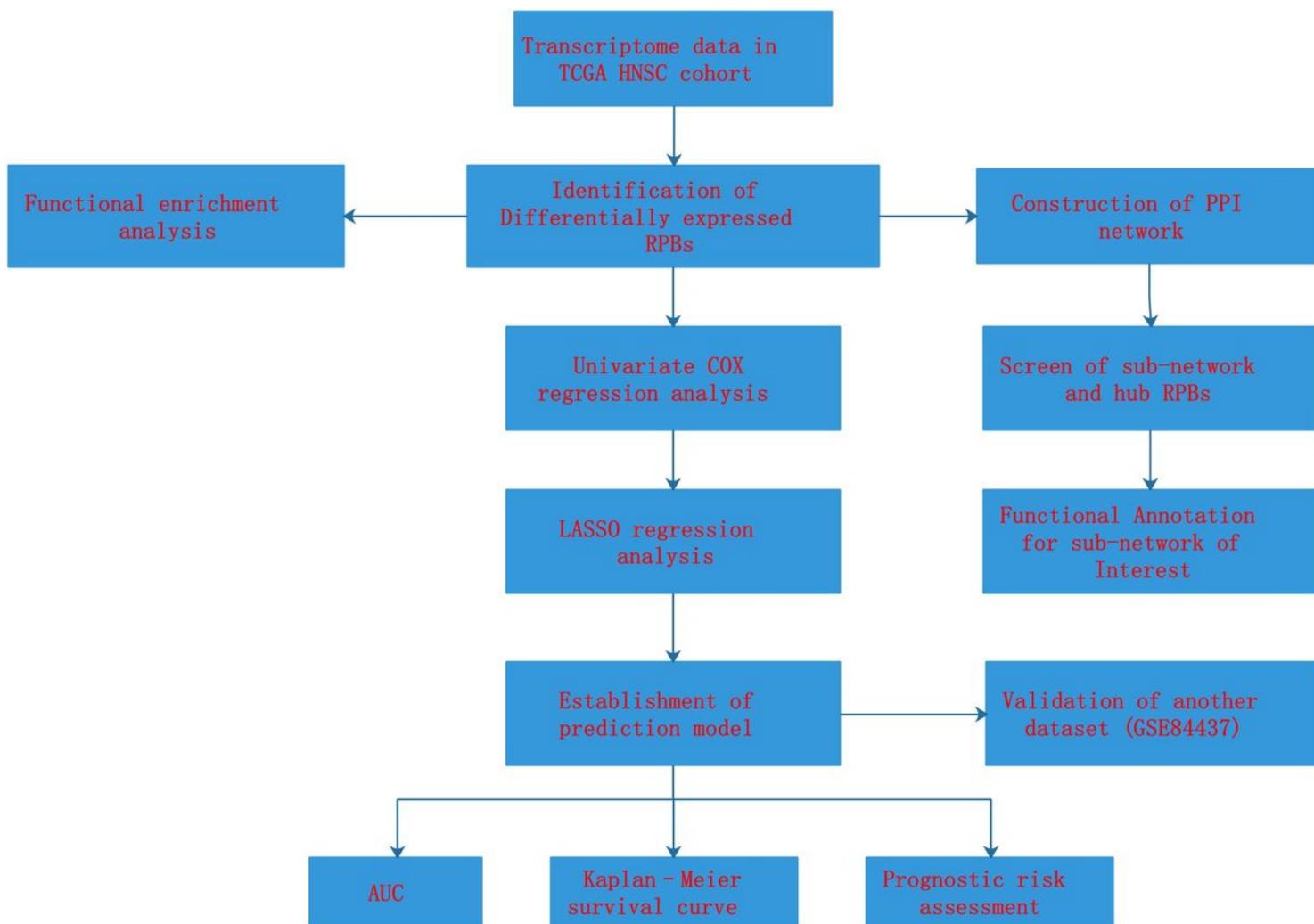


Figure 1

Flowchart delineates the study design and analysis process.

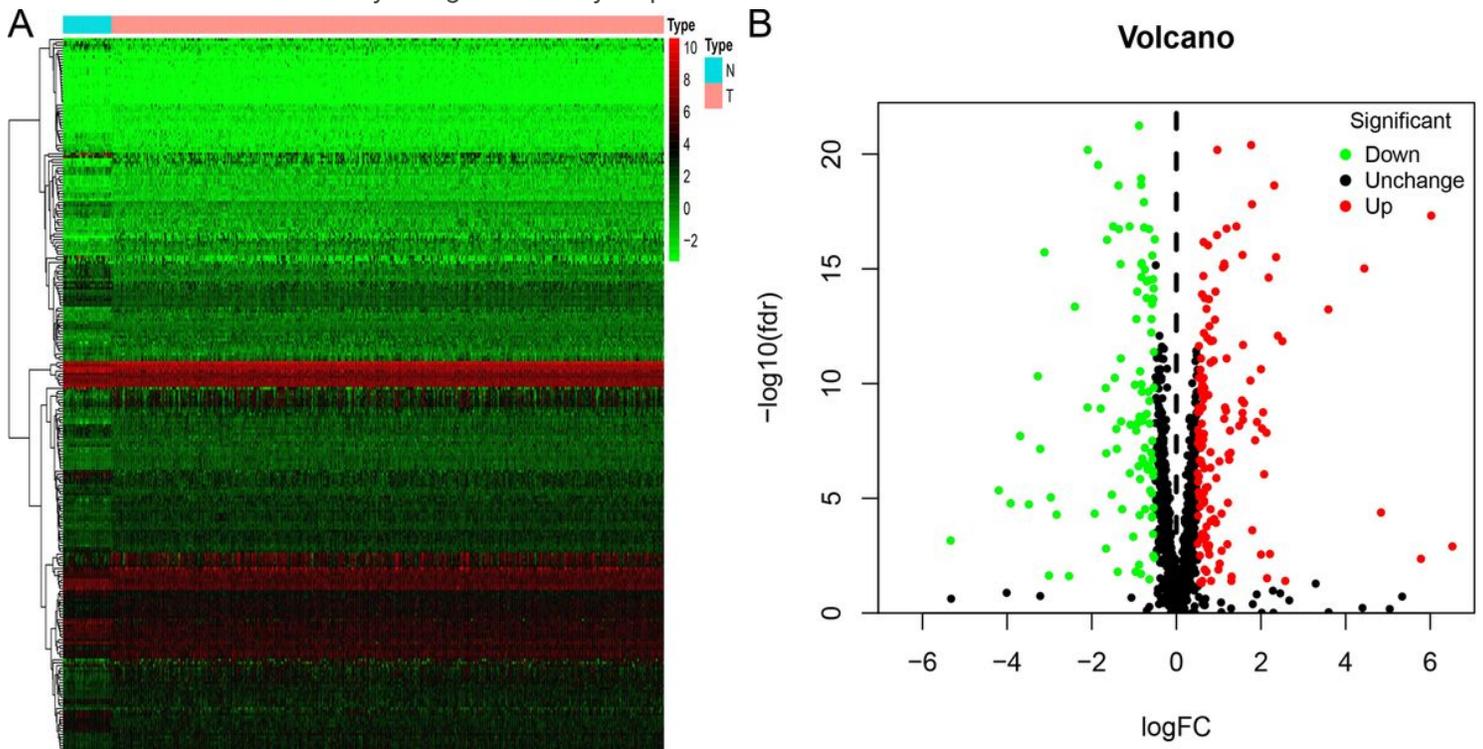


Figure 2

Identification of differentially expressed RPBs between adjacent tumor samples and tumor samples. The heatmap (A) and volcano (B) plot showed the differentially expressed RPBs, a total of 249 differentially expressed RPBs were identified, including 104 downregulated RPBs and 145 up-regulated RPBs.

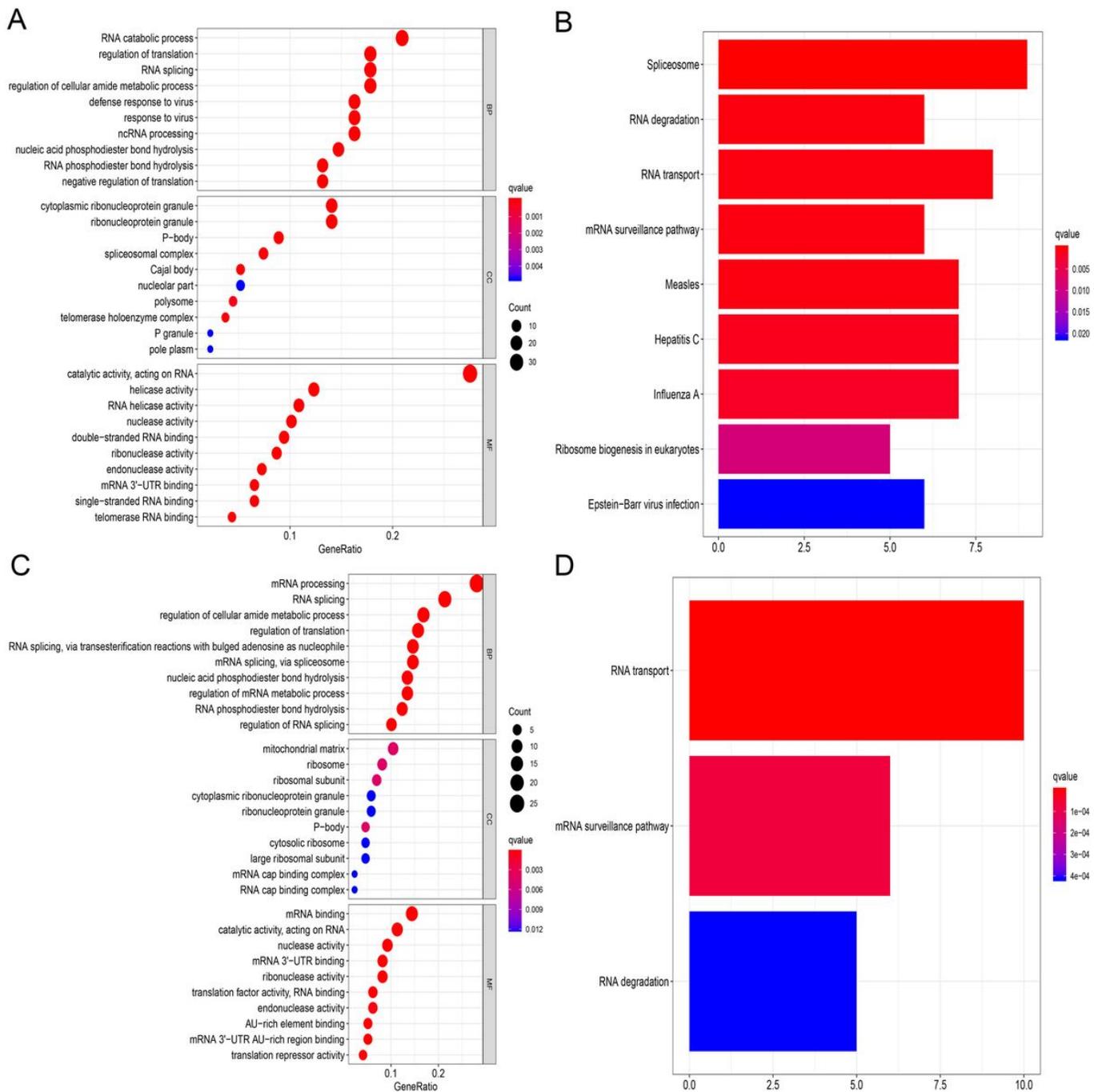


Figure 3

Functional enrichment analysis exhibited functions of the differentially expressed RPBs. A,C is for GO and KEGG analysis among upregulated RPBs; B,D is for GO and KEGG analysis among downregulated RPBs.

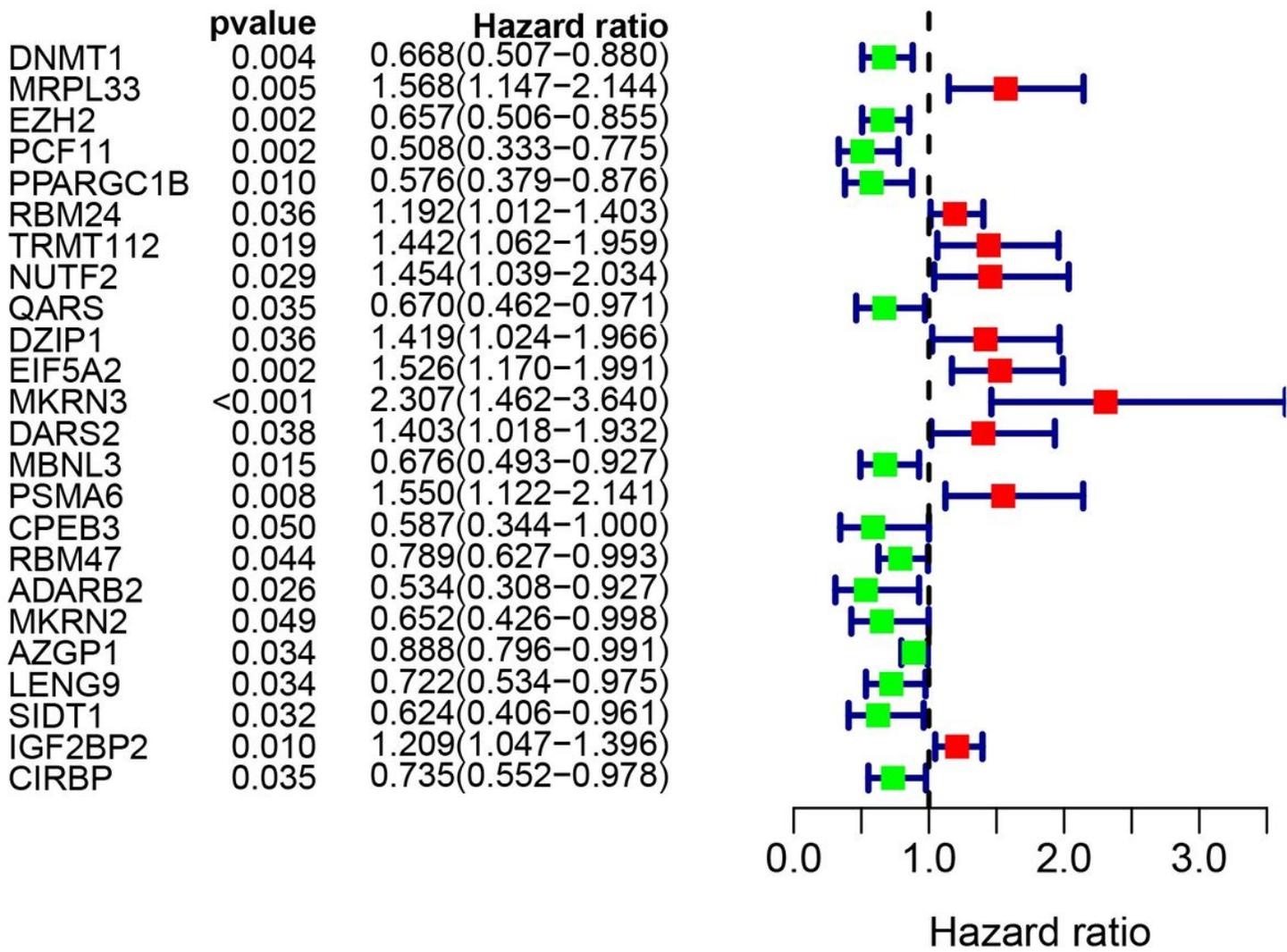


Figure 5

The forest plot of prognosis-related miRNAs between adjacent tumor samples and tumorsamples.

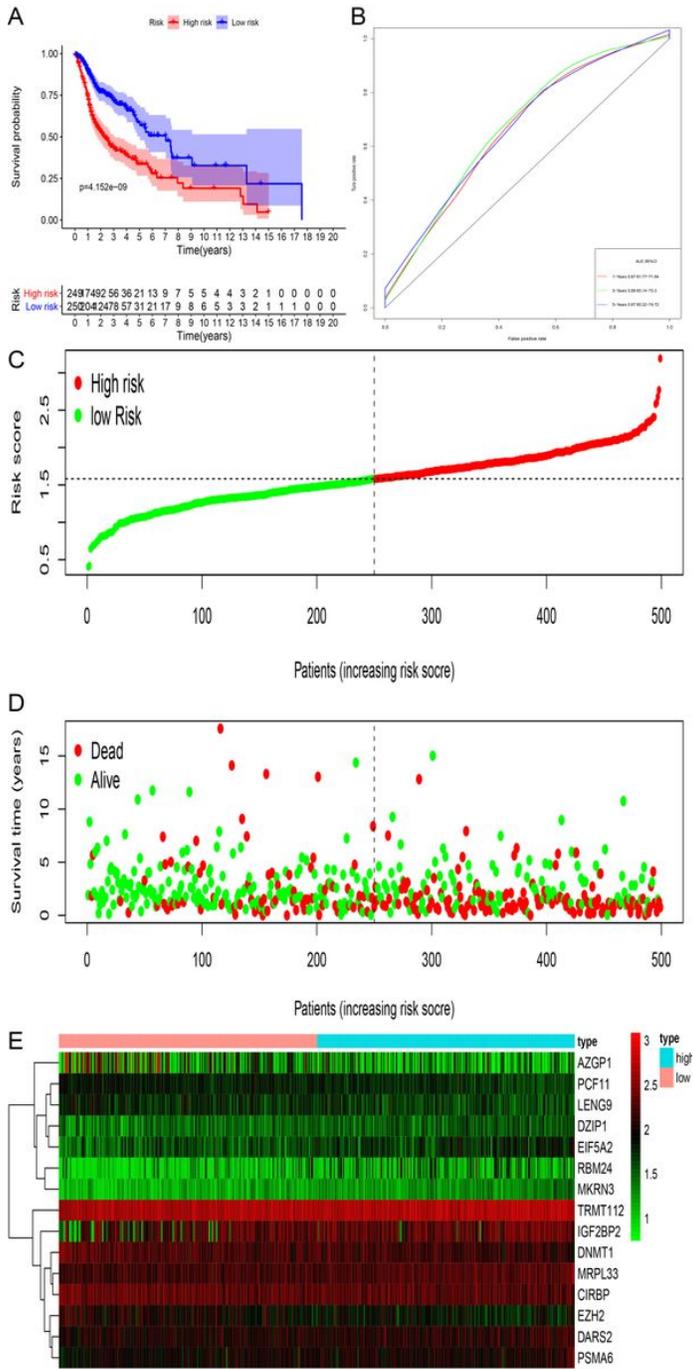


Figure 6

Risk score analysis of fifteen-gene prognostic model in TCGA HNSC cohort. (A) Survival analysis according to risk score; (B) ROC analysis; (C) Risk score of patients; (D) Heat map; (E) Survival status.

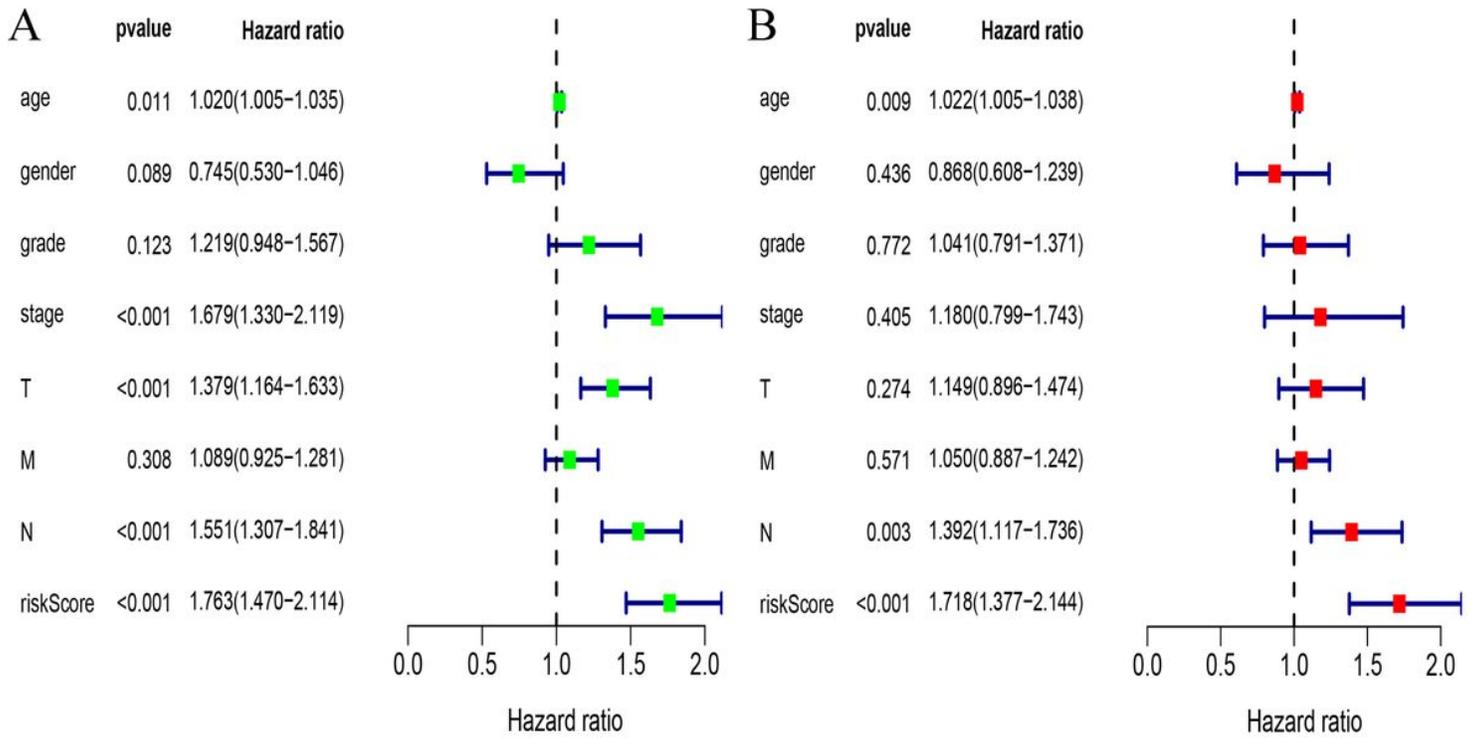


Figure 7

The prediction model could act as an independent predictor in univariate (A) and multivariate regression analysis (B) in TCGA HNSC cohort.

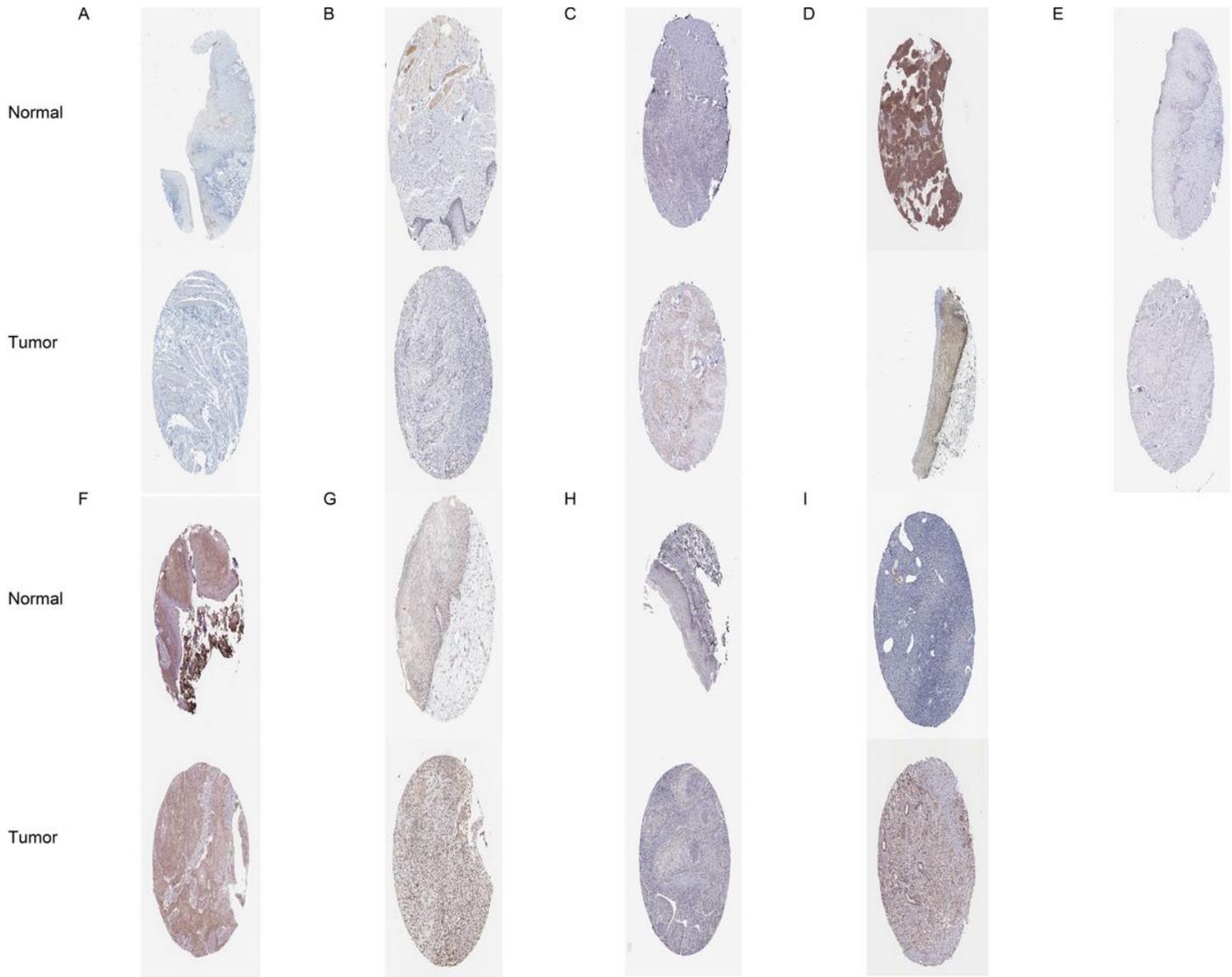


Figure 8

Validation of protein expression of hub genes in normal tissue and HNSC using the HPA database. (A) AZGP1, (B) DNMT1, (C) DZIP1, (D) IGF2BP2, (E) LENG9, (F) MRPL33, (G) PSMA6, (H) RBM24 and (I) TRMT112.

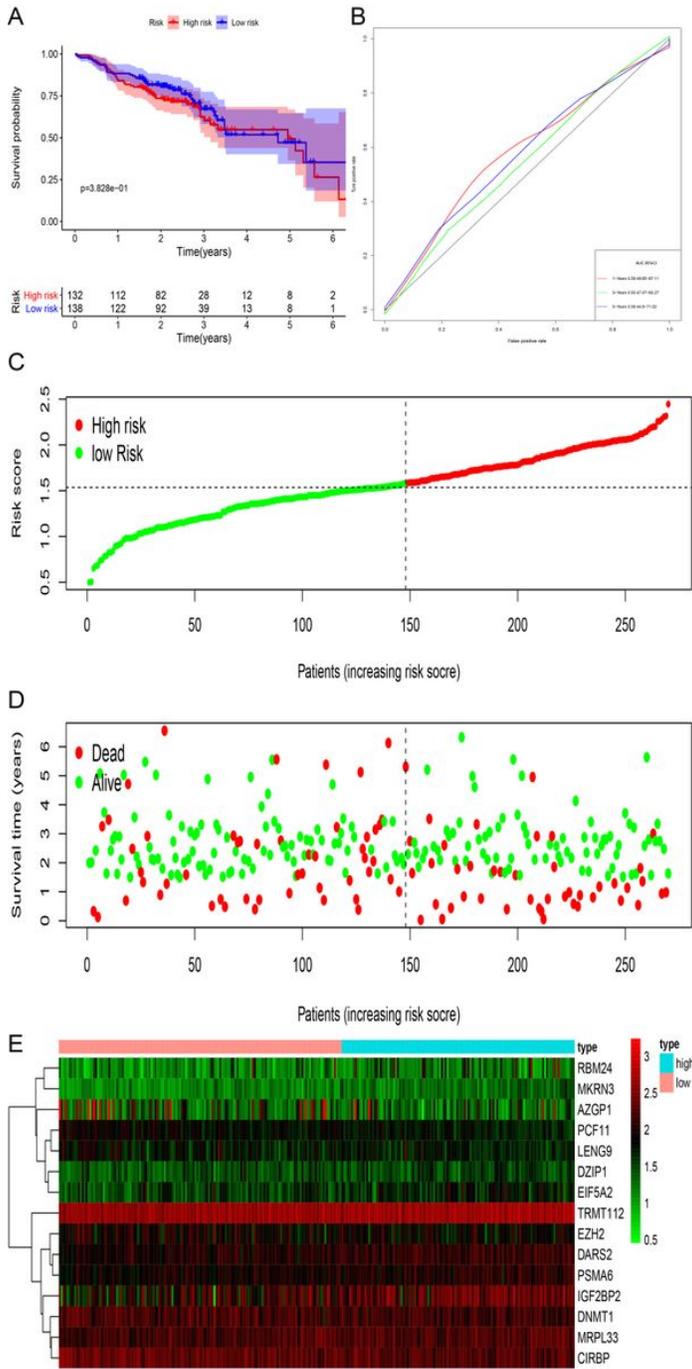


Figure 9

Risk score analysis of fifteen-gene prognostic model in GSE84437. (A) Survival analysis according to risk score; (B) ROC analysis; (C) Risk score of patients; (D) Heat map; (E) Survival status.

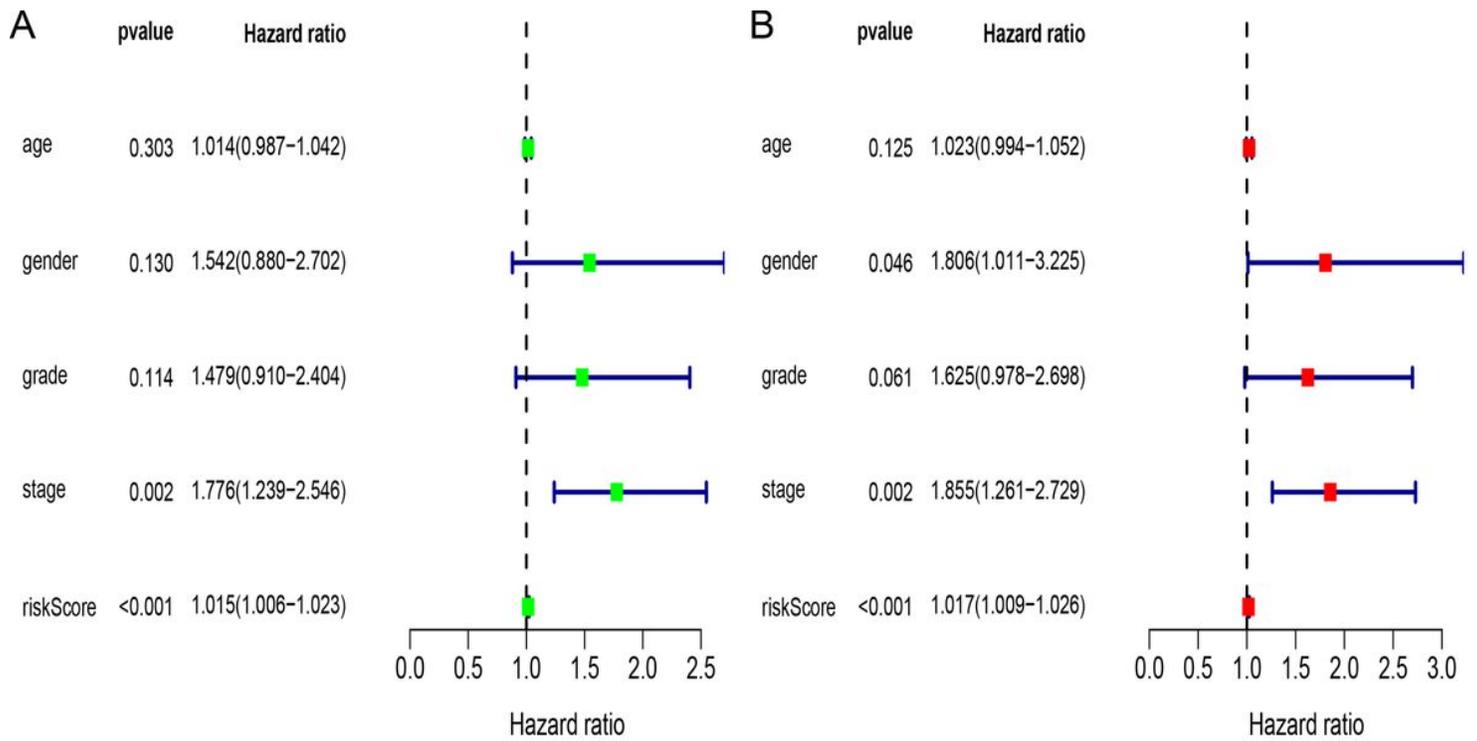


Figure 10

The prediction model could act as an independent predictor in univariate (A) and multivariate regression analysis (B) in GSE84437 dataset.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [SupplementaryTable.docx](#)