

Variations in haematological and biochemical parameters in healthy ponies

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Abstract

Background

Breed specific reference ranges for selected blood parameters are recommended for proper interpretation of blood tests, but there are only few reports dealing with ponies. The purpose of this study was to investigate if blood parameters differ among ponies' classes and to check if general normal values for equine species are applicable to ponies.

Results

All, except total protein concentration, biochemical parameter significantly ($p < 0.05$) differed among ponies' classes. The most pronounced difference was noted in blood lactate concentrations, higher ($p < 0.001$) in the smallest ponies (class A). In all groups of ponies muscle enzymes (aspartate aminotransferase and creatine kinase) and urea were high when compared to normal values for equine species, but triglycerides and creatinine were low. Blood lactate concentration was high in comparison with normal values for horses only in class A ponies'.

Conclusions

In healthy ponies, blood lactate concentration significantly differs between height classes. Normal values for equine species should not be directly applied to interpret the lactate, triglycerides, aspartate aminotransferase and creatine kinase values in ponies.

Background

Equine breed traits manifest as unique appearance and disposition. Physiological diversity is more complex and involves blood composition, reflected by the variations in haematological and biochemical parameters. Thus, breed differences should be taken into consideration in establishing reference intervals of blood parameters and interpretation of blood tests. Moreover, the reference intervals reported in the literature may also vary due to the population differences (e.g., geographic location, physical activity, age, sex, biological rhythms, etc.) [1–7].

Horses are usually grouped as “hot-blooded” and “cold-blooded”. The first group generally involves the horses of Arabian ancestry, including Arabians, Quarter Horses, Standardbreds, and Thoroughbreds, whereas “cold-blooded” are basically draft type horses, including Clydesdales, Percherons, and Shires. Several differences between these two groups regarding haematological parameters have been reported, including lower haematocrit in cold-blooded and higher erythrogram values in hot-blooded horses [7]. Currently, normal values for equine species in general, as well as for the most popular breeds, are easily available. However, the data for endemic breeds and ponies are still incomplete.

In the context of haematology and blood biochemistry, ponies have been suggested to be more like “hot-blooded” than “cold-blooded” horses [8], however, other sources include ponies into “cold-blooded” [7]. Reference intervals presented in current textbooks are based primarily on Thoroughbreds and Arabians and ponies are represented by only a single breed: American Miniature Horses, for which only a few values are available [1, 7]. Consequently, clinicians must interpret the common reference intervals with caution, as they may not be directly applicable to the pony practice. Several differences are likely important in assessing general health, fitness, stress, welfare and performance or even critical for proper diagnosis in sick animals. Concentrations of L-lactate (LAC) in blood and peritoneal fluid are the most common examples, used as diagnostic and prognostic indicators of colic severity. The commonly accepted reference interval of blood lactate concentration in healthy horses is approx. 0.7 mmol/L [9, 10]. In sick horses it varies, corresponding with the diagnosis, prognosis and required treatment [9–16]. However, in ponies, data are less clear and the clinical impressions suggest that pony and miniature breeds with gastrointestinal diseases could have higher blood lactate concentration than large breeds [17]. Thus, based on the reference intervals for hot-blooded horses, ponies might have been falsely suspected of having a poorer prognosis or even being subjected to euthanasia. Blood lactate concentrations in healthy ponies have been determined only once in large population [18] but breed differences among ponies still remain unclear.

The commonly discussed differences between horses and ponies are glucose and triglyceride metabolism and cellular response to catecholamines [17, 19, 20], which may promote metabolic diseases [21, 22]. Ponies are less sensitive to insulin, so the rapid development of hypertriglyceridemia resulting from insulin resistance seems more likely [23, 24].

In the global literature, there are only few reports dealing with haematology in the ponies. Thus, the aim of this study was to investigate the differences among blood parameters in ponies of different origin and height, represented by small (class A – up to 121 cm) ponies, Felin ponies and Polish Koniks and to compare these values with commonly accepted norms for horses. Felin ponies and Polish Koniks are homogenous populations. The Polish Konik (*Equus caballus gmelini* Ant.) is a primitive breed, descended directly from the wild Tarpans. They are small in stature (132–136 cm, so belong to height class C), have modest living requirements and high resistance to disease and environmental conditions [25]. Felin ponies represent a relatively new breed, specially dedicated for riding and competitive events for young riders. They originate from Arabians, Polish Koniks, Welsh ponies and Malopolski horses to whom they have the shortest genetic distance [26]. Malopolski horses were created from indigenous horses living in the east and north-west territories of Poland by crossings them with Thoroughbreds, Arabians and AngloArabians so that from genetic point of view, Felin ponies may be treated as hot-blood horses, but according to the height (125–136 cm) they belong to class B or C.

Results

Hematological and biochemical parameters in class A ponies', Felin ponies and Polish Koniks are presented in Table 1 and Table 2.

Table 1

Hematological parameters (median, IQR, range) in ponies class A, Felin ponies and Polish Koniks compared with normal values for warm blood horses (according to ¹Rossdale & Partners, 2011, ²Weiss & Wardrop, 2010, ³Hinchcliff & Geor, 2013)

Parameter	Ponies class A (n = 73)	Felin ponies (n = 28)	Polish Koniks (n = 41)	Normal values for warm blood horses	Normal values for cold blood horses	Normal values for American Miniature Horses
RBC [T/L]	7.4 ± 1.0 (5.6–11.5)	7.9 ± 1.0 (6.3–10.7)	7.7 ± 0.9 (6.0–10.0)	8.2 (6.2–10.2) ¹	5.5–9.5 ²	4.7–9.4 ²
HGB [mmol/L]	6.9 ± 9 (4.9–9.3)	7.1 ± 0.7 (6.0–8.4)	7.0 ± 0.6 (5.5–8.4)	8.4 (6.9–9.9) ¹	4.96–8.69 ²	5.52–10.12 ²
HCT [l/l]	0.35 ± 0.04 (0.3–0.5)	0.36 ± 0.03 (0.3–0.4)	0.35 ± 0.03 (0.3–0.4)	0.37 (0.31–0.43) ¹	0.24–0.44 ²	0.24–0.44 ²
MCV [fL]	46.9 ± 3.4 (36–55)	46.0 ± 3.8 (37–52)	46.0 ± 3.4 (39–51)	46 (40.0–50.0) ¹		37–58 ²
MCH [fmol]	0.92 ± 0.07 (0.7–1.1)	0.91 ± 0.07 (0.7–1.0)	0.91 ± 0.07 (0.8–1.0)	1.03 (0.94–1.18) ¹		0.81–1.43 ²
MCHC [mmol/L]	1.97 ± 0.06 (1.8–2.1)	1.97 ± 0.07 (1.8–2.1)	1.98 ± 0.06 (1.8–2.1)	2.38 (2.07–2.40) ¹		2.17–2.48 ²
WBC [G/L]	7.1 ± 1.2 (4.5–10.7)	6.8 ± 1.4 (4.0–10.4)	7.5 ± 1.1 (5.5–10.3)	7.5 (6.0–10.0) ¹	6.0–12.0 ² 3.9–12.4 ³	5.0–14.97 ² 10 ± 2.5 ³
GRA [G/L]	2.2 ± 0.7 (1.2–4.2) ^a	1.8 ± 0.5 (1.2–3.8) ^a	3.0 ± 0.7 (1.7–4.8) ^b	4.4 (3.4–5.4) ¹	2.6–7.2 ³	3.7 ± 0.8 ³
LYM [G/L]	4.0 ± 1.0 (2.3–8.4) ^a	4.3 ± 1.1 (1.9–6.7) ^{ab}	3.6 ± 0.9 (2.0–5.6) ^{ac}	2.6 (2.0–3.2) ¹	0.89–3.6 ⁴	5.9 ± 0.9 ³
MON [G/L]	0.9 ± 0.3 (0.3–1.5)	0.8 ± 0.2 (0.4–1.2)	0.9 ± 0.2 (0.6–1.5)	0.3 (0.2–0.4) ¹	0–0.62 ⁴	0.04 ± 0.08 ³

^{a-c} groups marked by different letters differ significantly, $p \leq 0.05$

RBC - number of red blood cells, HGB - haemoglobin concentration, HCT - haematocrit, MCV - mean corpuscular volume, MCH - mean corpuscular haemoglobin, MCHC - mean corpuscular haemoglobin concentration, WBC - number of white blood cells, GRA – number of granulocytes, LYM - number of lymphocytes, MON - number of monocytes, PLT – number of platelets

Parameter	Ponies class A (n = 73)	Felin ponies (n = 28)	Polish Koniks (n = 41)	Normal values for warm blood horses	Normal values for cold blood horses	Normal values for American Miniature Horses
PLT [G/L]	334 ± 207 (95–900) ^a	518 ± 243 (181–900) ^b	433 ± 153 (116–900) ^b	156 (100–250) ¹		
^{a-c} groups marked by different letters differ significantly, $p \leq 0.05$						
RBC - number of red blood cells, HGB - haemoglobin concentration, HCT - haematocrit, MCV - mean corpuscular volume, MCH - mean corpuscular haemoglobin, MCHC - mean corpuscular haemoglobin concentration, WBC - number of white blood cells, GRA – number of granulocytes, LYM - number of lymphocytes, MON - number of monocytes, PLT – number of platelets						

Table 2

Blood biochemical parameters (median, IQR, range) in ponies class A, Felin ponies and Polish Koniks compared with normal values for warm blood horses (according to ¹Rossdale & Partners, 2011, ²Hinchcliff & Geor, 2013, ³Latson 2005, Henderson 2013)

Parameter	Ponies class A (n = 73)	Felin ponies (n = 28)	Polish Koniks (n = 41)	Normal values for warm blood horses	Normal values for cold blood horses	Normal values for American Miniature Horses
ALP[U/l]	185 ± 81 (92–756) ^a	149 ± 55 (72–281) ^{ab}	211 ± 46 (126–358) ^{ac}	204 (147–261) ¹		
AST [U/l]	384 ± 83 (228–648) ^a	321 ± 50 (233–423) ^b	395 ± 65 (221–536) ^a	263 (102–350) ¹	234 ± 163(Clydesdales) ² 172±28(Shire) ²	189 ± 33 ²
GGTP [U/l]	19 ± 6 (8–53) ^a	23 ± 9 (14–48) ^a	66 ± 36 (20–198) ^b	16 (1–40) ¹	24.2 ± 6.0 (Shire) ²	11 ± 4.4 ²
CPK [U/l]	510 ± 195 (171–1233) ^a	356 ± 98 (192–609) ^b	628 ± 220 306–1137) ^c	186 (110–250) ¹	56 ± 22 (Clydesdales) ² 58 ± 12 (Shire) ²	273 ± 136 ²
TP [g/l]	66.8 ± 4.9 (56–75)	63.9 ± 5.6 (50–75)	66.1 ± 3.7 (57–75)	66.1 ± 3.7 (57–75) ¹	72 ± 6(clydesdales and Shire) ²	66 ± 6 ²
ALB [g/l]	35.6 ± 3.2 (26–41) ^a	37.8 ± 2.6 (34–46) ^b	36.1 ± 1.9 (32–41) ^a	35 (29–41) ¹	36 ± 5(Clydesdales) ² 28 ± 3(Shire) ²	
Urea [mmol/L]	7.0 ± 1.4 (3.8–10.2) ^a	7.0 ± 1.1 (5.1–9.0) ^a	8.2 ± 1.5 (3.8–13.0) ^b	5.2 (2.5–10.0) ¹	5.2 ± 1.14(Clydesdales) ²	8.5 ± 1.4 ²
Creatinine [µmol/L]	95.8 ± 16.9 (46.8–130.0) ^a	98.8 ± 16.3 (61.9–137.0) ^a	77.6 ± 17.1 (47.7–116.7) ^b	125 (85–165) ¹	151 ± 26.5(Cludesdales) ²	88.4 ± 17.7 ²

^{a-c} groups marked by different letters differ significantly, $p \leq 0.05$

ALP - alkaline phosphatase, AST - aspartate aminotransferase, GGTP - gamma-glutamyl transpeptidase, CPK – creatine kinase, TP – total protein concentration, ALB – albumin concentration, TBIL - total bilirubin, TG – triglycerides.

Parameter	Ponies class A (n = 73)	Felin ponies (n = 28)	Polish Koniks (n = 41)	Normal values for warm blood horses	Normal values for cold blood horses	Normal values for American Miniature Horses
TBIL [μmol/L]	16.9 ± 5.3 (5.3–32.3) ^a	20.6 ± 6.4 (11.1–37.6) ^b	12.3 ± 4.6 (4.4–28.7) ^c	20 (13–34) ¹		
TG [mmol/L]	0.30 ± 0.15 (0.1–0.8) ^a	0.42 ± 0.20 (0.2–0.9) ^b	0.33 ± 0.12 (0.2–0.6) ^{ab}	0.7 (0.2–1.2) ¹		
Glucose [mmol/L]	5.4 ± 0.9 (3.9–8.4) ^a	5.1 ± 1.1 (3.6–9.1) ^a	3.8 ± 0.9 (2.2–6.3) ^b	4.9 (4.3–5.5) ¹	4.21 ± 1.1 (Clydesdales) ²	5.1 ± 0.7 ²
Lactate [mmol/L]	1.19 ± 0.54 (0.1–2.7) ^a	0.18 ± 0.24 (0.1–0.9) ^b	0.19 ± 0.42 (0.1–2.6) ^b	< 0.7 ³		
a-c groups marked by different letters differ significantly, p ≤ 0.05						
ALP - alkaline phosphatase, AST - aspartate aminotransferase, GGTP - gamma-glutamyl transpeptidase, CPK - creatine kinase, TP - total protein concentration, ALB - albumin concentration, TBIL - total bilirubin, TG - triglycerides.						

Table 3
Population involved in the study

Group	n	Height	Age (median, IQR)	Sex		
				mare	stallion	gelding
Class A ponies (8 Shetland ponies, 3 Welsh ponies, 62 crossbreed ponies)	73	up to 121 cm	10.7–15	38 (52%)	3 (4%)	32 (44%)
Felin ponies	28	125–136 cm	11.5–19	20 (71%)	5 (18%)	3 (11%)
Polish Koniks	41	130–140 cm	7.4–11	38 (93%)	3 (7%)	0

There were no differences in erythrogram parameters among the groups of ponies, but all values except HCT and MCV were slightly lower when compared to normal values for non-thoroughbred (hot-blooded) horses [5]. WBC counts did not differ among ponies and were similar to the normal value for hot-blooded horses, but the proportion between GRA and LYM in ponies was inverted when compared to non-thoroughbred horses (the number of GRA was lower and LYM was higher) [5]. The numbers of granulocytes differed significantly among groups of ponies. In Polish Koniks, the number of granulocytes was significantly higher than in class A ponies'

($p < 0.001$) and Felin ponies ($p < 0.001$) and the number of lymphocytes was lower than in Felin ponies ($p = 0.034$). PLT in all groups of ponies were much higher than those reported for hot-blooded horses [5], and differed significantly between class A ponies' and other groups: Felin ponies ($p = 0.003$) and Polish Koniks ($p < 0.001$).

The activities of blood enzymes (ALP, AST and GGTP) in ponies were generally higher than values accepted for hot-bloods [5] and differed also among tested groups. The activity of ALP was significantly lower in Felin ponies than in Polish Koniks ($p = 0.002$). The activity of AST in Felin ponies was significantly lower than in class A ponies' ($p = 0.003$) and Polish Koniks ($p < 0.001$). The activity of GGTP was significantly higher in Polish Koniks than in class A ponies' ($p < 0.001$) and Felin ponies ($p < 0.001$).

Total protein and albumin concentrations of ponies was similar to hot-blood horses [5]. Albumin concentration differed significantly between pony groups and was significantly higher in Felin ponies than in class A ponies ($p = 0.003$) and Polish Koniks ($p = 0.015$).

Blood urea was higher and creatinine was lower in ponies than in hot-blood horses [5] and significantly differed in Polish Koniks when compared with other groups. Urea was higher in Polish Koniks than in class A ponies' ($p < 0.001$) and Felin ponies ($p = 0.002$) and creatinine was lower ($p < 0.001$ for both groups).

Liver parameters were generally lower in ponies than in hot-blood horses [5]. Only TBIL in Felin ponies was the same as the normal value for hot-bloods and it was significantly higher than in other groups of ponies ($p = 0.022$), moreover, TBIL was significantly higher in class A ponies than in Polish Koniks ($p < 0.001$). TG was low in all groups of ponies and significantly lower in class A ponies than in Felin ponies ($p = 0.01$).

Glucose concentrations in class A ponies' and Felin ponies were in the higher limit of the reference interval for hot-blood horses [5], significantly higher than in Polish Koniks ($p < 0.001$ for both groups).

Lactate was significantly higher in class A ponies' ($p < 0.001$ when compared with other groups) and higher than normal value for hot-bloods [9, 10].

Discussion

To the authors' knowledge, current data regarding reference intervals for haematological and blood biochemical parameters in ponies are limited. Additionally, there are no clear recommendations as to how ponies should be classified from a haematological standpoint. Two options exist: 1. ponies are (advised to be treated) referenced and evaluated as hot-bloods [8] or 2. cold bloods [7], but practitioners generally use the values for hot-bloods.

It is commonly accepted that erythrogram values (RBC, HGB, HCT) are lower in cold-bloods than in hot-bloods. It has been stated that ponies' haematocrit may be as low as 24 l/l, moreover, in American Miniature horses MCV, MCH and MCHC were higher [7]. WBC counts have been reported as slightly lower in cold-blooded than hot-blooded horses and neutrophil to lymphocyte ratios were 1.0 in Arabians and Thoroughbreds, 1.7 in cold-blood horses and 0.67 in American Miniature horses [7].

Erythrogram values determined in the ponies examined in our study did not differ among groups and were much lower than accepted for adult Thoroughbreds and moderately lower than recommended for hot-blooded [5] and Hucul horses [27]. Hucul horses are small (132-145cm high), so meet the general height criteria for ponies, but from genetic point are closer to hot-blooded horses. The differences between haematological parameters in Huculs and Felin ponies were quite surprising, due to ancestry where more similarities would be expected. It was also surprising that no pronounced differences were observed in HCT values between the ponies examined in our study and hot-blood horses. Similar hematological data have been reported for Noma and Kiso horses [4, 28], Shetland ponies [29], ponies in regular show jumping and eventing training [30] and Polish Koniks examined previously [31]. Interestingly, Bosnian ponies similar in size to Felin ponies and Polish Koniks had higher erythrogram values [32].

Leukogram values determined in our study were similar to values for hot-blooded horses but the neutrophil to lymphocyte ratios (N:L) were below 1.0, confirming the previous findings in ponies [7]. However, Shawaf et al. [29] have shown that in Shetland ponies WBC was quite high in summer and lower in winter. Regardless of season the values were higher than in our study. Also, the N:L ratio was above 1.0 in summer and winter [29]. In Noma horses WBC count was similar to the values determined in our study, but N:L ratio was above 1.0 [28]. It has been shown that granulocyte activity and the onset of inflammation differs between horses and Shetland ponies, being more robust in the latter [33]. Wilmink et al. [33] did not define the numbers of granulocytes, N:L ratios and WBCs, so we cannot compare our data with their findings. It might be postulated that the N:L ratio is one of breed related difference among ponies. However, it cannot be excluded that these differences were related to geographical region, as the cited studies were performed in Japan [28] and Saudi Arabia [29], not central Europe.

Erythrogram and leukogram features, determined in our study, suggest breed related haematological findings are of limited diagnostic importance. However, observed biochemical differences are diagnostically important

One of the important findings in our study was a higher concentration of lactate in class A ponies, but not classes B and C represented by Felin ponies and Polish Koniks. This is important from the diagnostic standpoint, as blood and peritoneal LAC are commonly used as diagnostic and prognostic indicators in gastrointestinal and ischemic emergencies in horses. It has been reported that LAC measurement at admission and the changes over time discriminate between survivors and non-survivors and indicate the need for surgical intervention [11–13, 15–17, 34]. Determination of LAC in peritoneal fluid has been reported as a better prognostic indicator in horses with colic than blood measurements [9, 15]. However, abdominocentesis was not necessarily recommend for all colic patients [15]. As a result, the majority of studies discuss blood measurements. All the previously referenced studies involved large, but mixed breed populations with various conditions, so the values indicating guarded prognosis varied among reports. Mean blood lactate concentrations at admission for non-survivors were reported as 4.1 mmol/L with repeated measurements recommended [11]. Another studies suggested lower > 2.06 [34] or higher 9.48 mmol/L [13] values for non-survivors and a cutoff value of < 6.0 mmo/L for survivors [13]. Delesalle et al, 2007 [15] stated an LAC value of 6.3 mmol/L for non-survivals, and recently an optimal decision tree has been proposed which identified horses as non-survivors when the LAC at admission was ≥ 4.3 mmol/L [16]. Only in the last study ponies (27.3%) and Icelandic horses (31.8%) constituted the majority of the observed population, but breed differences in LAC

measurements were not investigated. The focus of Dunkel's et al. study [17] where the mean LAC values for non-survivors were 4.1 and 3.1 mmol/L for ponies and horses, respectively, and did not differ significantly. However, the value for survivors was significantly higher in ponies (2.5 mmol/L) than in horses (1.2 mmol/L) and the authors concluded that ponies may be falsely suspected of having a surgical lesion or a poorer prognosis as they might present higher blood lactate concentrations. LAC concentrations in healthy ponies have been rarely determined. Recently, Dunkel et al. [18] published a study involving a large group of healthy ponies and determined that the mean LAC concentration was 0.7 (0.2-2.7) mmol/L and was lower than in full-sized horses: 0.9 (0.2-1.9) mmol/L. The authors concluded that lactate concentrations depend both on body condition and age [18]. Although the investigated population was large (101 animals), ponies were defined only on the basis of height (≤ 148 cm), and no data regarding breed or height classes were given. Our findings seem to substantially complement their study and provide additional clinically important information. In our study L-lactate concentrations, when compared to commonly accepted values for hot-blooded full-size horses were < 0.7 mmol/L [9, 10] or 0.9 mmol/L [18]. LAC was high only in class A ponies' (1.19 ± 0.54 mmol/L) but much lower in Felin ponies (0.18 ± 0.24 mmol/L) and Polish Koniks (0.19 ± 0.42 mmol/L) representing classes B and C. The reason for differences in LAC in ponies are unknown. However, LAC may be associated with either increased production or decreased clearance [12].

Several studies mentioned differences in hepatic functions between horses and ponies [17, 35]. Our study indicates that AST and GGTP in all groups of ponies were high and in Polish Koniks GGTP markedly exceeded the reference intervals for hot-blooded horses. In clinical conditions, elevation of AST and GGTP indicates hepatocellular injury [36]. The horses in our study were clinically healthy and liver parameters were not high enough to indicate or even suggest hepatic disease, but physiological differences regarding the function of hepatocytes may be suspected. This hypothesis seems in line with the differences described in Noma and Kiso ponies [28]. In both Japanese breeds GGTP concentrations were lower than in the ponies examined in our study, but AST in Noma ponies was significantly higher than in Kiso ponies [28] and very similar to the values measured in our study. However, in Shetland ponies in Saudi Arabia, GGTP and AST activities only in winter were comparably high to the values determined in our study [29].

Difference regarding muscle enzymes also seem possible. CPK activity was surprisingly high in class A and Polish Koniks. Such high values have not been reported in Shetland ponies, Noma and Kiso ponies [28, 29]. In horses, high values of CPK and AST are indicators of muscle damage, and slight elevation are nonspecific [36]. CPK activities up to 10 000 U/l are considered small, nonspecific elevations [37], which may be associated with transport or exercise [36]. However, in performance horses much lower muscle enzyme activity occur after exercise [38, 39]. Ponies in our study were not transported and did not exercise. CPK activity never exceeded 1300 U/L, and AST activity was not high enough to suggest any muscle pathology. Thus, we postulate that high CPK values were related to muscle mass and composition, which were not examined in our study, but such differences in ponies have been reported in the literature [40].

Another important finding in our study was the low concentration of triglycerides in all groups of ponies. Values were lower than referenced for hot-blooded horses and the lowest in the smallest class (class A ponies': 0.30 ± 0.15 mmol/l). Metabolic differences between horses and ponies in glucose and triglyceride metabolism and tendencies to develop hyperlipemia in a negative energy balance are widely reported in ponies [17, 19, 20, 23, 41]. It is generally believed that healthy ponies may have higher TG levels [42] and thus,

low TG concentrations are not of diagnostic relevance. Higher TG values 78.5 ± 11 mg/dl (0.89 ± 0.12 mmol/l) have been reported [43] but in more recent studies with ponies lower values predominate. Ono et al. [28] reported 37.2 ± 34.3 mg/dl (0.42 ± 0.39 mmol/L) for Noma horses and Takasu et al. [4] 16.4 ± 11.1 mg/L (0.19 ± 0.13 mmol/L) for Kiso horses. Even in pony mares in late gestation TG values as low as 11.3 ± 5.4 mg/dl (0.13 ± 0.07 mmol/l) have been measured [44].

This is particularly important for practitioners because metabolic disturbances leading to hyperinsulinemia and EMS include a moderate elevation of TG concentrations [22, 23]. If ponies normally have low TG concentrations and practitioners use reference values for hot-blooded horses (RI 0.2–1.2 mmol/L) [5], ponies actually having hypertriglyceridemia may be misdiagnosed, because ponies with mildly elevated TG, interpreted according to the reference values for hot-blooded horses, may in fact have markedly elevated TG. Thus, we postulate that the normal TG value for ponies should be reconsidered or at least treated with caution.

It is also noteworthy that in all groups of ponies examined by us, the concentrations of urea were higher and creatinine lower than recommended for full sized horses. Similar finding has been reported in American Miniature horses [1]. In Shetland, Noma and Kiso ponies creatinine concentrations were also low, but blood urea nitrogen (BUN) was lower than measured in our study [28, 29]. Low creatinine and high urea concentrations appear to be a breed related feature, the most pronounced in Polish Koniks.

Another finding in Polish Koniks was significantly lower glucose concentration than in other groups of ponies and also lower than normal values reported for hot-blood horses [5]. This was also reported in Shetland ponies, regardless of season, [29], Noma and Kiso ponies [28] and mixed population of ponies [18]. However, it was similar to the value reported previously in Polish Koniks [25]. The differences between full-sized horses and ponies in glucose and insulin metabolism, oxidative capacity and response under sympathetic stimulation are widely reported in the literature [21–23, 35, 45]. Although higher glucose concentrations, reflecting metabolic differences have been clearly shown in ponies with gastrointestinal diseases [45], these differences are frequently not noticeable at rest but only during dynamic response testing [18, 21].

The main limitation of our study was the number of ponies in examined groups, thus too small to establish reference intervals. It should also be mentioned that differences in haematological and biochemical values may be associated with the method of the test or even equipment. The methods used in our study are routine in other laboratories, except hand-held lactate and glucose analysers, that are used in field practice. Accusport analyser used in this study has been evaluated in horses and successfully used in other studies. It has been shown that Accusport is reliable and accurately measured plasma lactate concentration in horses so that they are comparable to values from other analysers [46, 47].

Conclusions

In conclusion, our study proved breed related differences in haematological and blood biochemical parameters in the ponies. In our opinion, normal values for ponies should vary at least with the height classes. Special attention should be paid on the interpretation of blood LAC and TG values, especially in cases where ponies are being evaluated for clinical disease.

Methods

The study involved 142 riding ponies, divided into 3 groups (Table. 3.). The first group (n = 73) consisted of animals classified on the basis of height as class A ponies (up to 121 cm) included 8 Shetland ponies, 3 Welsh ponies, 62 crossbreed ponies. Both genders were represented and were 2 to 31 years of age. The second group was composed of Felin ponies (n = 28) aged 2 to 29 years, both genders. The third group was Polish Konik horses (n = 41) aged 2 to 16 years, both genders.

The animals were stabled in various horse facilities in Poland. All owners gave the permissions for the procedure. All horses were dewormed and vaccinated according to standard protocols in the stable. None of these procedures had taken place less than 3 weeks before blood collection. All samples were obtained during routine veterinary procedures. All animals were examined in the morning, when they were in stalls, handled by their keepers to minimize stress. Immediately after the procedure, the animals came back to their routine activities. Standard clinical examination revealed no clinical signs of disease.

All blood samples were obtained by jugular venipuncture using a BD Vacutainer system into K2-EDTA tubes for haematological tests and serum analyses. EDTA blood samples were kept in + 4 °C and examined within 5 h for haematological parameters: total number of white blood cells (WBC), granulocyte counts (GRA), lymphocyte counts (LYM), monocyte counts (MON), haematocrit (HCT), haemoglobin concentration (HGB), the number of red blood cells (RBC), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC) and the total number of platelets (PLT) in an automated analyser calibrated for equine species (ABC Vet, Horiba ABX).

The tubes with no anticoagulant were centrifuged at 4380 g for 5 minutes, serum was aspirated and analysed. None of the collected blood samples had visual signs of haemolysis. Clinical biochemistry analyses included alkaline phosphatase (ALP), aspartate aminotransferase (AST), gamma-glutamyl transpeptidase (GGTP), creatine kinase (CPK), serum concentrations of albumin (ALB), total bilirubin (TBIL), triglycerides (TG), urea and creatinine were provided with automated clinical biochemistry analyser (Miura One, ISE. S.r.l., Italy). Total protein (TP) concentrations were measured by refractometer technique (Reichert Rhino Vet 360). For all measurements Pointe Scientific (USA) reagents, standards, calibrators and controls were used. Blood lactate and glucose concentrations were determined immediately after blood collection using The Accutrend Plus (Roche Diagnostics) and OptiumXido (Abbott Diabetes Care), respectively.

Statistical analysis was performed in TIBCO Statistica 13.3 (TIBCO Software Inc.). Numerical variables were given as the arithmetic mean and standard deviation (SD) or the median and interquartile range (IQR) unless a variable was normally distributed according to Shapiro-Wilk W test. Range was presented in all cases. Between-group comparisons were performed using one-way analysis of variance (ANOVA) with Tukey HSD post-hoc test for unequal groups or Welch ANOVA with Games-Howell post-hoc test if variances were unequal (according to the Levene's test) or Kruskal-Wallis H test with Dunn's post-hoc test unless variable was normally distributed. All tests were two-sided. A significance level was set at 0.05.

Abbreviations

ALB - albumin,

ALP - alkaline phosphatase,
AST - aspartate aminotransferase,
CPK - creatine kinase,
GGTP - gamma-glutamyl transpeptidase,
GRA - granulocyte counts,
HCT - haematocrit,
HGB - haemoglobin concentration,
IQR - interquartile range,
LAC - L-lactate
LYM - lymphocyte counts,
MCH - mean corpuscular haemoglobin,
MCHC - mean corpuscular haemoglobin concentration,
MCV - mean corpuscular volume,
MON - monocyte counts,
N:L - neutrophil to lymphocyte ratio
PLT - number of platelets,
RBC - number of red blood cells,
SD - standard deviation,
TBIL - total bilirubin,
TG - triglycerides,
TP - total protein,
WBC - number of white blood cells.

Declarations

Ethics approval and consent to participate:

Not applicable: All samples were obtained during standard veterinary diagnostic procedures; thus, no approval of the Local Commission for Ethics in Animal Experiments was required, according to the Polish legal regulations: art 1.2 (5) Ust. z dnia 15 stycznia 2015 r. o ochronie zwierząt wykorzystywanych do celów naukowych lub edukacyjnych, Dz.U.2018.0.1207 (Resolution on the animals protection used for scientific and educational purposes) and the European directive EU/2010/6.

Consent for publication:

Not applicable

Availability of data and materials:

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Competing interests:

The authors declare that they have no competing interests

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Authors' contributions:

OWP – contributed to the conception of the work; data acquisition and analysis, and drafting the manuscript; AC – contributed to the conception and design of the work; data analysis and interpretation, have drafted the manuscript; KMP – contributed to the design of the work; data acquisition, analysis and interpretation; MCz – contributed to data analysis and interpretation and drafting the manuscript; KS, AB, MPL – substantially contributed to data acquisition and analysis; MC – revised the manuscript; LW – contributed to the conception and design of the work; data analysis and interpretation and drafting the manuscript

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