

# Coagulation-Flocculation: A potential Application for Mosquito Larval Source Management (LSM) with No Insecticide Resistance

#### XinRan Li

East China Normal University https://orcid.org/0000-0002-3494-7636

PeiEn Leng

Shanghai Municipal Center for Disease Control and Prevention

XinYu Lu

East China Normal University

Bing Xiao

East China Normal University

#### Research

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#### **Abstract**

**Background:** Mosquito control is an essential step to eliminate mosquito-borne diseases. Larval mosquitoes have a more limited home range and lower resistance to adverse environment than adults, thus can be ideal targets for vector control in some cases. Coagulation-flocculation technology, which could be used for water treatment in breeding sites of several vector mosquito species, can significantly change both the distribution of organic particles and surface sediment characteristics in water environment. The aim of this study was to explore the effect, principle and possibility of using coagulation-flocculation technology in immature mosquitoes killing.

**Method:** In laboratory, chlorine-free tap water was treated with Poly Aluminum Chloride (PACI, sewage treatment using). The oviposition preference of gravid *Culex pipiens pallens*, the hatching of mosquito eggs and the survival amount of mosquito larvae were observed, and the pupa amounts were recorded each day.

**Results:** Coagulation-flocculation treatment could improve the oviposition preference of *Culex pipiens* pallens to some extent (compared with ordinary chlorine-free tap water), but not significantly (p=0.345). After treatment, mosquitoes laid eggs in chlorine-free tap water were 31.88% more than those laid in untreated water. Coagulation-flocculation affected the larvae's survival by physical means:  $\square$  alum floc layer increases the difficulty of larvae foraging, leads larvae starving to death;  $\square$  the little floc particles adhere to the surface of larvae, which stops larvae from floating upward to breathe. As a result, the alum floc layer had a good killing effect on the mosquito larvae, presented the half lethal time ( $LT_{50}$ ) of 2d, the 90% lethal time ( $LT_{90}$ ) of 8.7±7.3 ~ 14±4.5 d, and the pupation rate of 0 ~ (6.5±0.5)%, respectively.

**Conclusions:** PACI coagulation-flocculation produced lots of alum flocs, which may attract more gravid mosquitoes for laying eggs, and was shown to be highly active against 1<sup>st</sup>~2<sup>nd</sup> instar larvae. The principle of this technology illustrates this method won't develop insecticide resistance. In this study, coagulation-flocculation technology is considered to be a new potential approach to a sustainable, lowimpact and low-cost mosquito control method.

### **Background**

Mosquitoes are vectors for many pathogens, millions of people are estimated to be infected through mosquito biting globally and annually [1-3]. Some of these epidemic mosquito-borne diseases could be fatal for lack of licensed vaccines and effective therapeutics [4]. Therefore, increasing attention has been focused on vector mosquito control [5], particularly for now mosquitoes are expanding their scope of residence and extending their activity time under the condition of global warming [6, 7].

Three of four periods through mosquito's life (egg, larva and pupa) are in the water environment, during which their survival depends on various factors such as competitors [8], light [9], temperature [10], food availability [11, 12], predator [13, 14] and characteristics of water environment (such as pH, ammonia, chloride, dissolved oxygen, etc.) [15–18]. Larval mosquitoes have a more limited home range and lower

resistance to adverse environment than adults, thus can be ideal targets for vector control in some cases [19, 20].

Larval Source Management (LSM), one of the oldest tools in the fight against mosquito-borne diseases such as malaria and dengue, has been the main focus of mosquito control programs for decades in the United States of America (US), Canada, throughout Europe, Brazil and Singapore [21, 22]. Despite of now efforts are made to develop new biological or botanical larvicides [20, 23, 24], rather than routine chemical larvicides, in attempts to overcome the common disadvantages of larviciding, however, these weaknesses still remain largely unsolved, including: (M) toxic to human and other non-target species; (M) polluted to water environment; (M) costly in long-term using; (M) vulnerability to the evolution of target resistance [20]. Therefore, researches in introducing an existing and mature technology of water treatment into mosquito-control field could be attractive, if it has the possibility to avoid the weaknesses above.

Coagulation-flocculation, a proven technology in surface water remediation and sewage treatment, now has been widely used in plenty of water treatment occasions [25–27]. Addition of coagulants could promote agglomeration of suspended particles (including microorganisms), further deposit the latter to the bottom by gravity. Therefore, coagulation-flocculation technology can significantly change both but not limited to the distribution of organic particles and surface sediment characteristics in water [28]. Based on the roles which coagulation-flocculation played in water environment, it's convincing to believe that larval mosquitoes would be influenced deeply by it. In this study, coagulant Poly Aluminum Chloride (PACI) was used to treat chlorine-free tap water, through simulating this treatment process,we measured the influence of coagulation-flocculation on: (X) oviposition preference of gravid mosquitoes, (X) the development and survival of immature mosquitoes.

Culex pipiens pallens from Culex pipiens complex was selected. Mosquitoes in this complex prefer slightly or moderately contaminated water as breeding habitats [29], and these breeding habitats are the application objects of coagulation-flocculation technology. Species in the Culex pipiens complex are world widely distributed [4, 30, 31], meanwhile important vectors to plenty of virus and parasites, including West Nile virus, Wuchereria bancrofti and Dirofilaria immitis [29, 32, 33]. Culex pipiens pallens, commonly found north of 37°N in China, Japan and Korea, is one of the major mosquito species natively responsible for biting human and disease transmission [34].

#### Method

# Biological bioassays

Culex pipiens pallens adults and eggs in this research were obtained from Department of Vector Control, Shanghai Municipal Center for Disease Control & Prevention. Adults were maintained under conditions of 27.5 °C ± 1.5 °C, 13 h light/11 h dark (photoperiod), and continuously provided with a 10% glucose solution. Mosquito larvae were reared in chlorine-free tap water with the same temperature and

photoperiod. Since eggs were hatched, immature mosquitoes were fed with powdered, sterilized rat diet (QianMin SiLiao, China). 0.01 g of sterilized rat food was added within 24 hours after the hatching of mosquito eggs, and 0.03 g of sterilized rat food was added every 48 hours, increasing or decreasing appropriately according to the age and quantity of larvae. During the feeding and experimenting, larvae were removed with a straw after pupation and put into a mosquito cage (L:30 cm, W:30 cm, H:30cm) for cultivation. Female adults were fed on blood for 3 ~ 4 h, and then after 24 h mosquito spawning test was taken within the same cage. Blood source: SPF KM mouse (Shanghai Jiesjie laboratory animal co. LTD).

## Coagulation-flocculation treatment

Poly Aluminum Chloride (PACI) for sewage (Shanghai Ruixin industrial co. LTD, China), contained 28% of  $Al_2O_3$ , was used as coagulant. Coagulant was added as a suspension of 10 g/L.

The procedure of coagulation-flocculation was performed as follows: a predetermined amount of coagulant (60 ml/L, determined by pre-experiments) was dosed into the reactor (1L jar), after the water quality reached steady state, then the suspension was stirred at 300 rpm for 15 seconds, followed by 100 rpm for 2 min. JJ-1 digital speed measuring electric mixer (Changzhou Jintan Youlian Instrument Research Institute) was used to provide stir conditions.

## Data collection, processing, and statistical analysis

The results of mosquito oviposition preference test were expressed by number of valid oviposition times. Mann-Whitney U non-parametric test (IBM SPSS Statistics 20) was used for data analysis, and p < 0.05 was taken to indicate statistical significance. To evaluate how coagulation-flocculation take effect on the development and survival of immature mosquitoes, the mean number of survival mosquito larvae ( $\pm$  SE), the mean number of pupation ( $\pm$  SE), the half lethal time (LT<sub>50</sub>) and the 90% lethal time (LT<sub>90</sub>) were used to describe the results.

#### **Results**

#### Oviposition response of Culex pipiens pallens

Adult mosquitoes chosen for mosquito oviposition preference test were mosquitoes which had been 72 h after eclosion. 24 hours after blood feeding, ovitrap 1 (filled with 200 ml chlorine-free tap water + 20 ml PACI suspension, denoted as 01) and ovitrap 2 (filled with 220 ml chlorine-free tap water, denoted as 02) were placed in the cage. Every 24 hours 01 and 02 were observed, mosquito eggs in 01 and 02 were removed as well.

In the experiment, female mosquitoes laying eggs in two cups (d = 11 cm, h = 5 cm, transparent plastic material) were considered as valid oviposition. After each biting, all of the eggs laid by one single female *Culex pipiens pallens* would gather together as an egg raft and float on the water [35], thus we can recognize the number of valid oviposition times easily. Each observation and removal of all mosquito eggs in O1 & O2 is considered as an independent observation.

In order to exclude the influence of the placement order of ovitrap on the experimental results, the experiment was divided into two parallel control groups which only differed in the placement of O1 and O2, remaining other environmental factors consistent. Apparatus is shown in Fig. 1.

#### **Table 1** Results of two replications in mosquito oviposition preference test

In this test, O1 and O2 had equal oviposition opportunities for female mosquitoes, and finally with a total of 320 valid oviposition times (Table 1). Mosquitoes showed the same oviposition preference in the two replications, so we take the results of R1 and R2 calculated in combination. Of the two replications, O1 had 182 valid oviposition times, which is 31.88% more than that of O2 (138 valid oviposition times), indicating that O1 was more attractive to oviposition, but not significant (n = 15, p = 0.345, p > 0.05).

Among the 15 independent observations with a duration of about 30 days (data not shown), there were 11 times to prove mosquitoes prefer 01 as their breeding habitat, only in 3 times there were more mosquitoes chose 02 rather than 01 to lay eggs, remaining one time equally. This suggests that the attraction of 01 to female mosquitoes' oviposition was persisted and didn't decrease over time.

Numerous studies indicated that the process of selecting breeding sites for female mosquitoes is mediated by multiple factors such as vision [36], smell [37], and pheromone [38]. We speculated in this study mosquitoes' oviposition behavior is mainly influenced by vision. Studies have shown that yellow is more attractive to *Culex pipiens pallens* for laying eggs than white [39], which in this experiment after coagulation-flocculation, yellow alum floc deposited at the bottom of the ovitrap, which is considered to be nutrient-rich for larvae growing, therefore more female *Culex pipiens pallens* mosquitoes preferred O1 as breeding sites [40].

# Effects of coagulation-flocculation on mosquito egg hatching performance and subsequent development of mosquito larvae

This phase was carried out in a jar containing 1L chlorine-free tap water. 0.01 g sterilized rat food was added in chlorine-free tap water in advance, for it would provide basic nutrition for larvae after egg hatching. The surface scums were scoured by deionized water, until most of scums dropped down to the bottom.

Pre-experiment confirmed that the appropriate dosage of PACI suspension was 60 ml/L, which means the mass concentration of PACI to water was about 0.6 mg/L. It was normal dosage in water treatment [41, 42]. The complete coagulation-flocculation operation includes the addition of coagulants and the mixing process, so the experiment was divided into four parallel control groups, group1  $\sim$  4 (Fig. 2). Groups could be divided into the coagulating groups and the non-coagulating groups, according to the addition of coagulant or not. Experiment started after mosquito eggs were added in Group 1  $\sim$  4, and then treated with stirring and/or coagulant adding.

Table 2 shows the comparison of water quality parameters of each group before (t = 0) and after (t = 30 min) treatment. No significant changes occurred in water quality parameters within non-coagulating groups before and after the treatment; group 3 exhibited obvious flocculation 30 minutes after the coagulation, while pH decreased slightly ( $7.36 \pm 0.49$  to  $7.21 \pm 0.43$ ), and turbidity increased significantly ( $1.23 \pm 0.18$  NTU to  $2.99 \pm 0.38$  NTU), for without stirring, alum floc produced by coagulation didn't have enough opportunity for collision and aggregation [43], resulting in the size of some flocs being too small to settled, these floc fragments suspended in supernatant; the pH of group 4 decreased slightly ( $7.37 \pm 0.48$  to  $7.11 \pm 0.36$ ) after treatment, since other factors were basically unchanged. The addition of coagulant increased the salinity (Sal) in the water, and the Electronic Conductivity (EC) and Total Dissolved Solids (TDS) of the coagulating groups (group 3 & group 4) were significantly improved after treatment.

**Table 2** Water quality parameters of each group when t = 0 and t = 30 min

The pupation time of larvae in 4 groups (Fig. 3 (b)) were compared. The first pupa in coagulating groups (group 3) appeared significantly later than that of non-coagulating groups, except for there was no pupa in Group 4. The delay time varied in different batches of experiment, but all exceeded 7 days. Moreover, the pupation time of last larvae in coagulating groups (group 3) was also later than that of the non-coagulating groups, suggests that coagulation-flocculation treatment may delay the development of mosquito larvae. It may be a response of food-lacking, which could delay larvae development [44, 45].

**Table 3** Comparison of pupation rate, lethal time of 50% ( $LT_{50}$ ) and lethal time of 90% ( $LT_{90}$ ) in 4 groups

In table 3, differences between pupation rate,  $LT_{50}$  and  $LT_{90}$  in 4 groups indicated what degree could coagulation-flocculation affect larvae survival in this experiment. The pupation rate of non-coagulating groups was up to 50% in average, while in coagulating groups was below 10%, suggests that by treating with PACl coagulation-flocculation, the number of larvae that become pupae would be definitely reduced. In addition,  $LT_{50}$  of coagulating groups were 2d and  $LT_{90}$  were over 7d, indicating that PACl coagulation-flocculation had a stronger killing effect on newly hatched 1st instar larvae and a limited killing effect on late instars.

# 7d-continuously cultivation to explore the main reason coagulation-flocculation cause 1st ~ 2nd instar larvae death

To explain why PACl coagulation-flocculation could definitely kill 1st  $\sim$  2nd instar larvae, there are two possible reasons: 1) *Culex pipiens pallens* larvae may be sensitive to the water-soluble products produced by PACl coagulation, and died due to toxicity; ( $\mathbb{S}$ ) alum floc layer hide the food from immature mosquitoes, increases the difficulty of larvae feeding, results in larvae starving to death, moreover, these little floc particles could adhere to the surface of larvae, which prevent larvae to float upward to breathe.

In order to find out which from above the main reason is caused the high mortality of 1st ~ 2nd instar mosquito larvae, a 7d-continuously cultivation of *Culex pipiens pallens* was conducted. From the egg

stage, *Culex pipiens pallens* were raised in ordinary chlorine-free tap water (control group, recorded as "group C"), chlorine-free tap water after coagulation-flocculation (coagulation-flocculation group, recorded as "group C-F"), and filtered supernatant of group C-F (recorded as "group SC-F"), respectively.

Results revealed that survival of the larvae in group C and group SC-F was consistent and in good condition, while the number of larvae in group C-F reduced by 75% within 2d after the hatching, and made zero within 5d (Table 4). Therefore, the possibility of *Culex pipiens pallens* larvae died for the water-soluble products produced by PACI coagulation can be excluded, similarly the changes of pH, for larvae favor a neutral pH or slightly alkaline environment [46]. By observing the distribution of the dead larvae in group C-F, it can be found that the location of dead larvae is mainly in the surface and upper part of the floc sediments, and there is no food remains in the digestive tract of larvae (Fig. 4). This phenomenon illustrates that alum floc layer is the main cause of 1st ~ 2nd instar larvae's death.

**Table 4** Results of 7d-continuously cultivation in different groups

#### **Discussion**

Coagulation-flocculation technology is widely used in waste water treatment and water environment remediation, even in drinking water treatment [47, 48], relevant theories are abundant. To our knowledge, this is the first study to raise the possibility that coagulation-flocculation could be applied to the vector mosquito control. In this study, we found that under laboratory conditions, PACI coagulation-flocculation could promote oviposition preference of female *Culex pipiens pallens* to some extent (compared with ordinary chlorine-free tap water), and has a very high lethal effect on the newborn mosquito larvae after hatching. The best chance what coagulation-flocculation could achieve the highest lethal effect, is a lentic environment to keep the alum flocs at the bottom as long as possible, and it is exactly the environment what mosquito larvae need [17, 49–51].

It is worth nothing that coagulation-flocculation could efficiently kill 1st ~ 2nd instar larvae. Firstly, this treatment reduces the amount of larvae's food (organic suspended solids, bacteria and algae) in the water phase, impels larvae to forage from sediment, where is hidden by alum flocs; besides, coagulation-flocculation produces a large number of alum flocs with different size and irregular shapes, not only could it adhere to the surface of larvae and prevent them from floating upward to breathe, but also reduces the opportunity for larvae to get fed.

According to this study, the killing of immature mosquitoes by coagulation-flocculation is mainly achieved by physical means, which makes coagulation-flocculation have obvious advantages over the commonly used chemical insecticide: ( $\mathbb{N}$ ) non-toxic, coagulation-flocculation could be applied in drinking water treatment, that means its environmental impact is very weak; ( $\mathbb{N}$ ) it won't develop insecticide resistance when put into long-term use; ( $\mathbb{N}$ ) it could trap gravid mosquitoes to spawn and kill the larvae simultaneously.

According to the principle of killing mosquito larvae by coagulation-flocculation technology,we believe there is a failure time of coagulation-flocculation to kill larvae, at this time the alum floc layer is compressed to dense enough by gravity, larvae cannot enter it. In a lentic environment, factors which influence the failure time of coagulation-flocculation include coagulation process (type of coagulant, and how it is used), parameters of water environment (pH, salinity, temperature, etc.), and the moving ability of mosquito larvae ("strong" larvae may rescue themselves from alum flocs). We believe that, on the premise that other variables do not change, the adoption of appropriate methods (such as vibrating slightly) to promote the resuspension of alum flocs is helpful to delay the arrival of the failure time of this larvae-killing technology.

The coagulant use in this study was the inorganic coagulant PACI. The dosage of PACI was determined to be 0.06%, according to the best turbidity removal effect in pre-experiment, and it was added in the form of 10 g/L suspension at one time. In the out-door study, not only the dosage of coagulant should be calculated according to the water volume, but also the bottom area should be taken into consider, for alum floc layer need to be thick enough to trap mosquito larvae. The available methods include: adding coagulant at plural points, and increasing (or decreasing) the dosage of coagulant.

Though in laboratory, coagulant PACI could promote oviposition preference of female *Culex pipiens* pallens to some extent, however, considering the limitation of laboratory, and actually female *Culex pipiens pallens* prefer ovitrap in deep color as breeding sites [39], PACI may avoid mosquitoes' oviposition if used out-doors. Still, coagulant modification may help to solve.

Larval Source Management (LSM) is a strategy to prevent the spread of vector-borne diseases by targeting the larval stage of vectors, including habitat modification, habitat manipulation, larviciding and biological control. World Health Organization (WHO) lists LSM as a supplementary vector control method for malaria vector control [19, 52]. At present, commonly used larvicides are chemical insecticides with certain biological toxicity [53, 54], which are difficult to be accepted by some communities. Therefore, non-toxic technology coagulation-flocculation could help LSM being generalized in these communities.

What's more, the increasing insecticide resistance of mosquitoes are threatening the results in eliminating mosquito-borne diseases in the past decades, such as malaria and dengue [55]. Issued in May 2012, the WHO Global plan for insecticide resistance management in malaria vectors (GPIRM) is a plan of action for all stakeholders engaged in the fight against malaria. The document provides comprehensive technical recommendations to manage insecticide resistance in different situations [56]. One of the key actions called by GPIRM and experts to be undertaken is to "develop new, innovative vector-control tools" [56, 57], we believe that coagulation-flocculation technology has the potential to be one of them. It requires more researches about coagulation-flocculation applying in Integrated Vector Management (IVM), Larval Source Management (LSM), and attract-and-kill control strategy. These includes the effects on different mosquito species with different ecological habits, the application of other coagulants, the R&D of new coagulants, the optimization of the dosage and way of use, the boundary and potential risks and so on.

#### **Conclusions**

This study observed how PACI coagulation-flocculation affect: ((I)) *Culex pipiens pallens* gravid female oviposition site selection; ((I)) development of *Culex pipiens pallens* from eggs to pupae. Results exhibited gravid adults prefer to choose chlorine-free tap water under treatment as breeding sites, rather than ordinary chlorine-free tap water. And alum floc could kill newly hatched 1st ~ 2nd instar larvae efficiently. The alum floc produced by coagulation-flocculation mainly affected *Culex pipiens pallens* larvae by physical means, which not only reduced the feeding opportunities of the larvae, but also limited the ability of larvae to float on the water to breathe. Thus it won't develop insecticide resistance. In this study, coagulation-flocculation technology is considered to be a new potential approach to a sustainable, lowimpact and low-cost mosquito control method.

#### **Abbreviations**

**PACI** 

Poly Aluminum Chloride;  $LT_{50}$ :the half lethal time;  $LT_{90}$ :the 90% lethal time; IVM:Integrated vector management; LSM:Larval Source Management; WHO:World Health Organization; GPIRM:Global plan for insecticide resistance management in malaria vectors

#### **Declarations**

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#### Availability of data and materials

All relevant data are included within the article.

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#### **Authors' contributions**

Huang-MS proposed the research ideas and assisted in study designing. Li-XR designed the complete experimental scheme, implemented and was responsible for writing the manuscript. Miss Xiao-B assisted in the breeding and caring *Culex pipiens pallens* adults and larvae. Mr Lu-XY assisted in the photograph and making figures. All authors read and approved the final manuscript.

#### Ethics approval and consent to participate

Not applicable.

#### Consent for publication

Not applicable.

#### Competing interests

The authors declare that they have no competing interests

#### References

- 1. Rezza G. Aedes albopictus and the reemergence of Dengue. BMC Public Health. 2012;12:72.
- 2. Organization WH. The World malaria report 2018. Geneva: World Health Organization; 2019.
- 3. Schaffner F, Medlock JM, Van Bortel W. Public health significance of invasive mosquitoes in Europe. Clin Microbiol Infect. 2013;19:685–92.
- 4. Weaver SC, Reisen WK. Present and future arboviral threats. Antiviral Res. 2010;85:328–45.
- 5. Bhatt S, Gething PW, Brady OJ, Messina JP, Farlow AW, Moyes CL, Drake JM, Brownstein JS, Hoen AG, Sankoh O, et al. The global distribution and burden of dengue. Nature. 2013;496:504–7.
- Robert MA, Christofferson RC, Weber PD, Wearing HJ. Temperature impacts on dengue emergence in the United States: Investigating the role of seasonality and climate change. Epidemics. 2019;28:100344.
- 7. Shin HR, Kneitel JM. Warming interacts with inundation timing to influence the species composition of California vernal pool communities. Hydrobiologia. 2019;843:93–105.
- 8. Reiskind MH, Wilson ML: Interspecific Competition Between Larval < I > Culex restuans</I > Theobald and < I > Culex pipiens</I > L.(Diptera: Culicidae) in Michigan. Journal of Medical Entomology 2008, 45:20-27.
- 9. Muturi EJ, Mwangangi J, Shililu J, Jacob BG, Mbogo C, Githure J, Novak RJ. Environmental factors associated with the distribution of Anopheles arabiensis and Culex quinquefasciatus in a rice agroecosystem in Mwea, Kenya. Journal of Vector Ecology. 2008;33:56–63.
- 10. Grech MG, Sartor PD, Almiron WR, Luduena-Almeida FF. Effect of temperature on life history traits during immature development of Aedes aegypti and Culex quinquefasciatus (Diptera: Culicidae) from Cordoba city, Argentina. Acta Trop. 2015;146:1–6.
- 11. Reiskind MH, Walton ET, Wilson ML. Nutrient-dependent reduced growth and survival of larval Culex restuans (Diptera: Culicidae): laboratory and field experiments in Michigan. J Med Entomol. 2004;41:650–6.
- 12. Yahouedo GA, Djogbenou L, Saizonou J, Assogba BS, Makoutode M, Gilles JR, Maiga H, Mouline K, Soukou BK, Simard F. Effect of three larval diets on larval development and male sexual performance

- of Anopheles gambiae s.s. Acta Trop. 2014;132(Suppl):96-101.
- 13. Shaalan EA-S, Canyon DV, Muller R, Younes MWF, Abdel-Wahab H, Mansour A-H. A mosquito predator survey in Townsville, Australia, and an assessment of Diplonychus sp. and Anisops sp. predatorial capacity against Culex annulirostris mosquito immatures. Journal of Vector Ecology. 2007;32:16–21.
- 14. Segev O, Verster R, Weldon C, Strecker A. Testing the link between perceived and actual risk of predation: mosquito oviposition site selection and egg predation by native and introduced fish. J Appl Ecol. 2017;54:854–61.
- 15. Vanlalruia K, Senthilkumar N, Gurusubramanian G. Diversity and abundance of mosquito species in relation to their larval habitats in Mizoram, North Eastern Himalayan region. Acta Trop. 2014;137:1–18.
- 16. Gardner AM, Anderson TK, Hamer GL, Johnson DE, Varela KE, Walker ED, Ruiz MO. Terrestrial vegetation and aquatic chemistry influence larval mosquito abundance in catch basins, Chicago, USA. Parasit Vectors. 2013;6:9.
- 17. Soleimani-Ahmadi M, Vatandoost H, Zare M. Characterization of larval habitats for anopheline mosquitoes in a malarious area under elimination program in the southeast of Iran. Asian Pac J Trop Biomed. 2014;4:73–80.
- 18. Dias ACA, Rodrigues MMS, Silva AA. Effect of acute and chronic exposure to ammonia on different larval instars of Anopheles darlingi (Diptera: Culicidae). J Vector Ecol. 2019;44:112–8.
- 19. Fillinger U, Lindsay SW. Larval source management for malaria control in Africa: myths and reality. Malar J. 2011;10:353.
- 20. Workman MJ, Gomes B, Weng JL, Ista LK, Jesus CP, David MR, Ramalho-Ortigao M, Genta FA, Matthews SK, Durvasula R, Hurwitz I. Yeast-encapsulated essential oils: a new perspective as an environmentally friendly larvicide. Parasit Vectors. 2020;13:19.
- 21. Floore TG. Mosquito Larval Control Practices: Past and Present. J Am Mosq Control Assoc. 2006;22:527–33.
- 22. Carlson DB. Source Reduction in Florida's Salt Marshes: Management to Reduce Pesticide Use and Enhance the Resource. J Am Mosq Control Assoc. 2006;22:534–7.
- 23. Kim SI, Ahn YJ. Larvicidal activity of lignans and alkaloid identified in Zanthoxylum piperitum bark toward insecticide-susceptible and wild Culex pipiens pallens and Aedes aegypti. Parasit Vectors. 2017;10:221.
- 24. Choi L, Majambere S, Wilson AL. Larviciding to prevent malaria transmission. Cochrane Database Syst Rev. 2019;8:CD012736.
- 25. Dong L, Lin L, Li Q, Huang Z, Tang X, Wu M, Li C, Cao X, Scholz M. Enhanced nitrate-nitrogen removal by modified attapulgite-supported nanoscale zero-valent iron treating simulated groundwater. J Environ Manage. 2018;213:151–8.
- 26. Fosso-Kankeu E, Webster A, Ntwampe IO, Waanders FB. Coagulation/Flocculation Potential of Polyaluminium Chloride and Bentonite Clay Tested in the Removal of Methyl Red and Crystal Violet.

- Arabian Journal for Science Engineering. 2016;42:1389–97.
- 27. Rishel KL, Ebeling JM. Screening and evaluation of alum and polymer combinations as coagulation/flocculation aids to treat effluents from intensive aquaculture systems. J World Aquaculture Soc. 2006;37:191–9.
- 28. Xu Y, Chen T, Liu Z, Zhu S, Cui F, Shi W. The impact of recycling alum-humic-floc (AHF) on the removal of natural organic materials (NOM): Behavior of coagulation and adsorption. Chem Eng J. 2016;284:1049–57.
- 29. Cabanova V, Miterpakova M, Valentova D, Blazejova H, Rudolf I, Stloukal E, Hurnikova Z, Dzidova M. Urbanization impact on mosquito community and the transmission potential of filarial infection in central Europe. Parasit Vectors. 2018;11:261.
- 30. Farajollahi A, Fonseca DM, Kramer LD, Marm Kilpatrick A. "Bird biting" mosquitoes and human disease: a review of the role of Culex pipiens complex mosquitoes in epidemiology. Infect Genet Evol. 2011;11:1577–85.
- 31. Nebbak A, Willcox AC, Koumare S, Berenger JM, Raoult D, Parola P, Fontaine A, Briolant S, Almeras L. Longitudinal monitoring of environmental factors at Culicidae larval habitats in urban areas and their association with various mosquito species using an innovative strategy. Pest Manag Sci. 2019:75:923–34.
- 32. Ciota AT, Chin PA, Kramer LD. The effect of hybridization of Culex pipiens complex mosquitoes on transmission of West Nile virus. Parasit Vectors. 2013;6:305.
- 33. Simonsen PE, Mwakitalu ME. Urban lymphatic filariasis. Parasitol Res. 2013;112:35-44.
- 34. Kim K, Tsuda Y. Sporogony and sporozoite rates of avian malaria parasites in wild Culex pipiens pallens and C. inatomii in Japan. Parasit Vectors. 2015;8:633.
- 35. Shin D, O'Meara GF, Civana A. Size of Openings in Water-Holding Containers: Impact on Oviposition by Culex (Culex) Mosquitoes. *Insects* 2019, 10.
- 36. Dieng H, Satho T, Suradi NFB, Hakim H, Abang F, Aliasan NE, Miake F, Zuharah WF, Kassim NFA, Majid AHA, et al. Presence of a predator image in potential breeding sites and oviposition responses of a dengue vector. Acta Trop. 2017;176:446–54.
- 37. Turnipseed RK, Moran PJ, Allan SA. Behavioral responses of gravid Culex quinquefasciatus, Aedes aegypti, and Anopheles quadrimaculatus mosquitoes to aquatic macrophyte volatiles. J Vector Ecol. 2018;43:252–60.
- 38. Faierstein GB, Lu W, Sena A, Barbosa RMR, Leal WS. Conspecific and allospecific larval extracts entice mosquitoes to lay eggs and may be used in attract-and-kill control strategy. Sci Rep. 2019;9:13747.
- 39. Gunathilaka N, Ranathunge T, Udayanga L, Wijegunawardena A, Abeyewickreme W. Oviposition preferences of dengue vectors; Aedes aegypti and Aedes albopictus in Sri Lanka under laboratory settings. Bull Entomol Res. 2018;108:442–50.
- 40. Buxton M, Cuthbert RN, Dalu T, Nyamukondiwa C, Wasserman RJ. Cattle-induced eutrophication favours disease-vector mosquitoes. Sci Total Environ. 2020;715:136952.

- 41. Shewa WA, Dong T, Mu W, Murray K, Dagnew M. The impact of chemically enhanced primary treatment on the downstream liquid and solid train processes. Water Environ Res. 2020;92:359–68.
- 42. Zhang W, Wei Q, Xiao J, Liu Y, Yan C, Liu J, Sand W, Chow CWK. **The key factors and removal** mechanisms of sulfadimethoxazole and oxytetracycline by coagulation. *Environ Sci Pollut Res Int* 2020.
- 43. Du P, Li X, Yang Y, Fan X, Zhang T, Wang N, Li H, Ji S, Zhou Z. **Effect of rapid-mixing conditions on the evolution of micro-flocs to final aggregates during two-stage alum addition**. *Environ Technol* 2020:1–10.
- 44. Puggioli A, Carrieri M, Dindo ML, Medici A, Lees RS, Gilles JR, Bellini R. Development of Aedes albopictus (Diptera: Culicidae) Larvae Under Different Laboratory Conditions. J Med Entomol. 2017;54:142–9.
- 45. Kaufman MG, Wanja E, Maknojia S, Bayoh MN, Vulule JM, Walker ED. Importance of algal Biomass to growth and development of Anopheles gambiae larvae. J Med Entomol. 2006;43:669–76.
- 46. Abdullah MAR, Merdan Al. Distribution ane ecology og the mosquito fauna in the southwestern Saudi Arabia. J Egypt Soc Parasitol. 1995;25:815–37.
- 47. Zhang Y, Diehl A, Lewandowski A, Gopalakrishnan K, Baker T. Removal efficiency of micro- and nanoplastics (180 nm-125 mum) during drinking water treatment. Sci Total Environ. 2020;720:137383.
- 48. Chua S-C, Chong F-K, Malek MA, Ul Mustafa MR, Ismail N, Sujarwo W, Lim J-W, Ho Y-C: **Optimized Use of Ferric Chloride and Sesbania Seed Gum (SSG) as Sustainable Coagulant Aid for Turbidity Reduction in Drinking Water Treatment**. *Sustainability* 2020, **12**.
- 49. Calhoun LM, Avery M, Jones L, Gunarto K, King R, Roberts J, Burkot TR. Combined sewage overflows (CSO) are major urban breeding sites for Culex quinquefasciatus in Atlanta, Georgia. Am J Trop Med Hyg. 2007;77:478–84.
- 50. Vazquez-Prokopec GM, Vanden Eng JL, Kelly R, Mead DG, Kolhe P, Howgate J, Kitron U, Burkot TR. The risk of West Nile Virus infection is associated with combined sewer overflow streams in urban Atlanta, Georgia, USA. Environ Health Perspect. 2010;118:1382–8.
- 51. Grech MG, Manzo LM, Epele LB, Laurito M, Claverie AN, Luduena-Almeida FF, Miserendino ML, Almiron WR. Mosquito (Diptera: Culicidae) larval ecology in natural habitats in the cold temperate Patagonia region of Argentina. Parasit Vectors. 2019;12:214.
- 52. **Supplementary**. **vector control methods**https://www.who.int/malaria/areas/vector\_control/complementary\_methods/en/.
- 53. McAllister JC, Porcelli M, Medina JM, Delorey MJ, Connelly CR, Godsey MS, Panella NA, Dzuris N, Boegler KA, Kenney JL, et al. Mosquito Control Activities during Local Transmission of Zika Virus, Miami-Dade County, Florida, USA, 2016. Emerg Infect Dis. 2020;26:881–90.
- 54. Santos VSV, Limongi JE, Pereira BB. Association of low concentrations of pyriproxyfen and spinosad as an environment-friendly strategy to rationalize Aedes aegypti control programs. Chemosphere. 2020;247:125795.

- 55. Tabbabi A, Daaboub J, Ben Cheikh R, Laamari A, Feriani M, Boubaker C, Ben Jha I, Ben Cheikh H. The potential role of urbanization in the resistance to organophosphate insecticide in Culex pipiens pipiens from Tunisia. Afr Health Sci. 2019;19:1368–75.
- 56. Organization WH. **Global plan for insecticide resistance management in malaria vectors.** Genevaand2012.
- 57. Benelli G, Beier JC. Current vector control challenges in the fight against malaria. Acta Trop. 2017;174:91–6.

#### **Tables**

Table 1 Results of two replications in mosquito oviposition preference test

	number of valid oviposition times		
	01	02	
R1	71	58	
R2	111	80	
total	182	138	

**Table 2** Water quality parameters of each group when t = 0 and t = 30 min

	Pupation rate (%)	Lethal Time of 50% (d)	Lethal Time of 90% (d)
group1	56.5 ± 23.5	-	-
group2	53 ± 6	-	-
group3	6.5 ± 0.5	2	14 ± 4.5
group4	0	2	8.7 ± 7.3

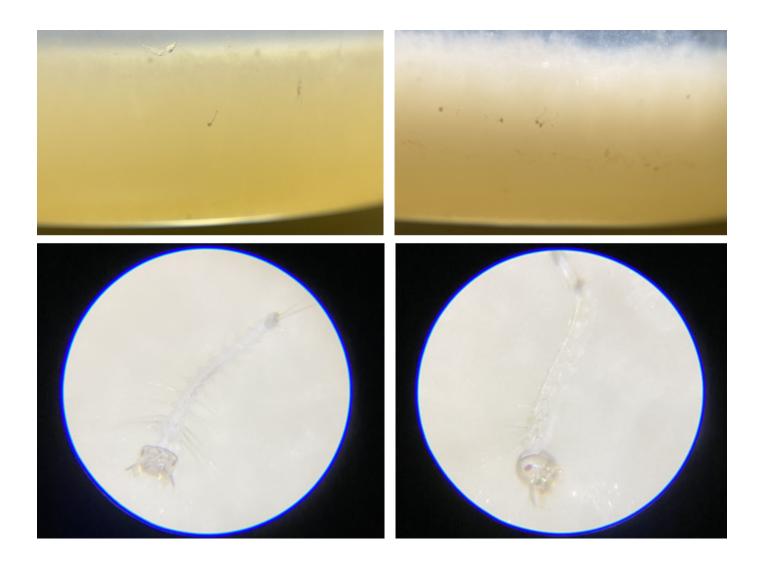
**Table 3** Comparison of pupation rate, lethal time of 50% ( $LT_{50}$ ) and lethal time of 90% ( $LT_{90}$ ) in 4 groups

Time (d)	Number of larvae in each group				
	Control (Group C)	Coagulating-Flocculation (Group C-F)	Supernatant of Group C-F (Group SC-F)		
1	100	100	100		
2	100	25	100		
3	100	23	100		
4	100	15	100		
5	98	0	100		
6	95	0	100		
7	89	0	94		

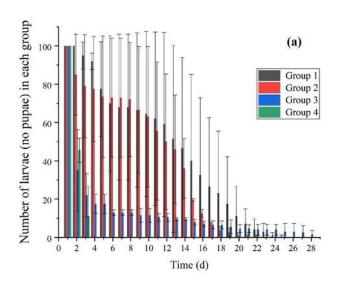
Table 4 Results of 7d-continuously cultivation in different groups

Group number	рН	Turbidity(NTU)	EC(ms/cm)	TDS(ppm)	Sal(g/L)			
Before the treatment (t = 0)								
1	$7.42 \pm 0.46$	1.26 ± 0.22	18.55 ± 0.62	9278 ± 312	10.98 ± 0.40			
2	7.37 ± 0.49	1.09 ± 0.18	19.09 ± 1.00	9545 ± 496	11.32 ± 0.63			
3	7.36 ± 0.49	1.23 ± 0.18	19.03 ± 0.93	9513 ± 466	11.27 ± 0.59			
4	$7.37 \pm 0.48$	1.11 ± 0.15	19.05 ± 0.93	9526 ± 467	11.29 ± 0.59			
30 min after the treatment (t = 30)								
1	7.47 ± 0.49	1.09 ± 0.13	18.90 ± 0.87	9452 ± 437	11.2 ± 0.56			
2	$7.46 \pm 0.48$	1.68 ± 0.92	19.16 ± 1.04	9583 ± 523	11.37 ± 0.67			
3	7.21 ± 0.43	2.99 ± 0.38	23.00 ± 1.07	11503 ± 533	13.87 ± 0.70			
4	7.11 ± 0.36	1.18 ± 0.20	23.68 ± 0.88	11840 ± 439	14.31 ± 0.57			

# **Figures**



**Figure 1**Dead mosquito larvae in alum floc layer of group C-F



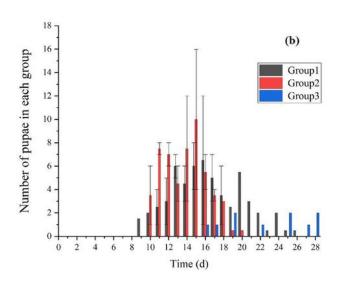


Figure 2

The survival of larvae and the number of pupation over time, group 1~4. (a): number of larvae (no pupae) in each group over time; (b): number of pupae in each group over time.

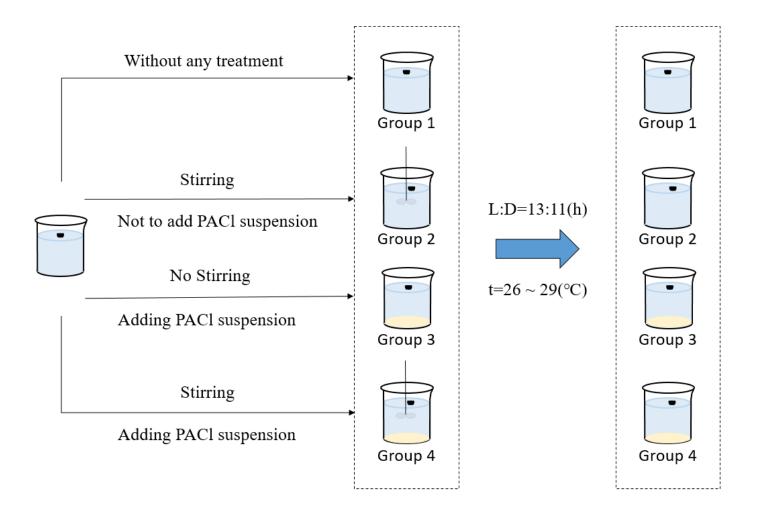


Figure 3

Setting of mosquito egg hatching and larvae developing experimental groups (Stirring was consisted of two stages: 300 rpm rapid stirring for 15s, 100 rpm slow stirring for 2 min)

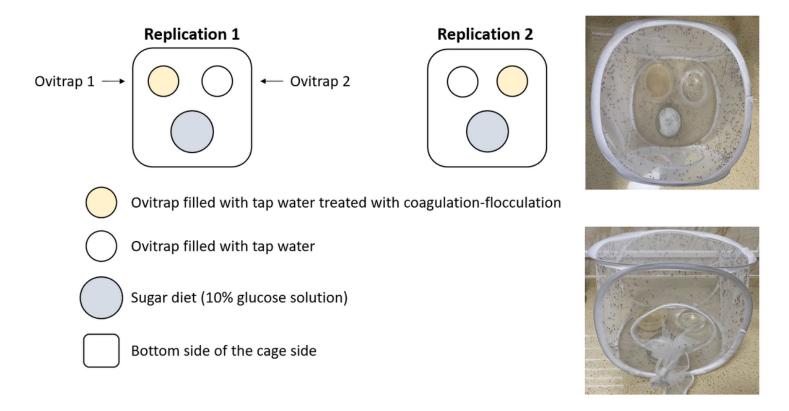


Figure 4

Replication 1 and Replication 2 in mosquito oviposition preference test. (Two replications differed in ovitrap order only).