

Remediation of Arsenic-contaminated Soil in Northwestern China by *Pseudomonas Taiwanensis*

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1 **Remediation of arsenic-contaminated soil in northwestern China by *Pseudomonas taiwanensis***

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27 **Abstract**

28 The physiological resistance of *Pseudomonas taiwanensis* to heavy metals was studied and used to
29 repair heavy metal contaminated water and soil. The results showed that the suitable pH conditions for
30 the growth of *P. taiwanensis* were 5-9, and the salt tolerance was 6%. The tolerance concentrations for
31 heavy metals As(V) and Mn(II) were 500 mg L⁻¹ and 120 mg L⁻¹, respectively. The strains were
32 enriched by nutrient broth(NB) medium. After logarithmic phase, the bacteria liquid was mixed with
33 ATCCTM#279 medium in proportion, and a certain amount of Mn(II) was added. The results of
34 removing heavy metals As, Pb and Cd in the composite polluted water phase were 22.09%, 30.75% and
35 35.33%. The molar ratio of manganese and iron will affect the removal efficiency of single pollution
36 heavy arsenic. When the ratio of iron and manganese is 1:5, the highest removal efficiency is 68%.
37 However, when the remediation method is added to the soil, it cannot fix all metals, such as Cu and Zn.
38 This remediation method provides a reference for the practical engineering application of microbial
39 remediation.

40 **Keyword:** Soil pollution; heavy metal; microbial remediation; manganese-oxidizing bacteria;
41 *pseudomonas taiwanensis*; biogenic manganese oxides

42

43 **Declarations**

44 Ethics approval and consent to participate

45 Not applicable

46 Consent for publication

47 Not applicable

48 Availability of data and materials

49 The datasets used or analysed during the current study are available from the corresponding author
50 on reasonable request.

51 Competing interests

52 The authors declare that they have no competing interests.

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55 Authors' contributions

56 All authors made an indispensable contribution to this study. The technical guidance and methods
57 were provided by Shengli Wang. The writing and analysis of the article were completed by Mengbo
58 Liu. The data processing and image production were carried out by Xiang Ning. The experimental
59 scheme and operation were carried out by Meng Yang. The experimental materials and funds were
60 carried out by Zhongren Nan.

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65

66 **1. Introduction**

67 Arsenic is ubiquitous in the natural environment and causes worldwide concern because of its
68 health problems(Zhu Y et al, 2014). Arsenic generally exists in the atmosphere, soil and rock, natural
69 water and organisms, and migrates through natural processes (such as weathering, biological activities
70 and volcanic emissions) and a series of human activities (such as mining and pesticides)(Smedley P&
71 Kinniburgh D, 2002; Yun S et al, 2018). Arsenic in soil environment enters the human body through the
72 food chain under the action of bioconcentration. Long-term consumption of contaminated food will
73 increase the risk of carcinogenic and non-carcinogenic health hazards(Qu C et al, 2012; Zhang J et al,
74 2015; Mao X-L, Li D, Zhu X-H, 2016).

75 Arsenic affects biological safety in the form of trivalent arsenate and pentavalent arsenate as its
76 main oxidation state(Garbinski LD, Rosen BP& Chen J, 2019). Biologically, the biological activity of
77 trivalent arsenic is higher than that of pentavalent arsenic, so it is easy to migrate to plants and affect
78 their normal growth and development(Liu SX et al, 2001). When applied to organisms, arsenic binds to
79 specific tissue proteins and phosphates with similar structures, thereby interfering ATP synthesis(Ali W
80 et al, 2020). Therefore, reducing the biological activity of arsenic in soil has become the focus of many
81 scholars.

82 Arsenic fixation in can be fixed by adding additives such as biochar(Wang Y et al, 2020)、
83 Phytoremediation(Lombi E et al, 2001), or the remediation methods of compost and super accumulator
84 can be used to reduce their active(Cao X, Ma LQ& Shiralipour A, 2003), but these methods have poor
85 applicability, high price, low efficiency, long repair time and may cause secondary pollution, which can
86 not be effectively used in practical engineering. Studies have shown that iron oxide (iron spots) forms
87 on the root surface of plant roots (rice as an example), which can prevent arsenic from further
88 migration to the upper part of the plant(Liu W et al, 2004). Some iron-manganese oxides with poor
89 crystallinity provide major reaction sites for arsenic oxidation and adsorption, which have been used
90 for arsenic immobilization in soils(Wang Y et al, 2020). Similarly, the presence of manganese oxides
91 can also achieve effective results for arsenic fixation(Yun S et al, 2018; Zhang J et al, 2015). Therefore,
92 remediation of arsenic contaminated soil by iron and manganese oxides is a promising method to
93 improve soil environmental quality.

94 Microbes play a key role in soil element cycling. For arsenic, it has been found that many bacteria
95 can migrate and transform arsenic in the environment directly or indirectly(Kumari N, Rana A&
96 Jagadevan S, 2019). Trivalent arsenic can be converted into pentavalent arsenic in a certain pH range.
97 Some scholars have studied the biotransformation, bioaccumulation, and biosorption of arsenic by
98 *D.arsenite*, which can adapt to a large pH range and can be repaired in a high arsenic
99 environment(Zhang Z et al, 2016). However, this method can not be guaranteed to be effective for a
100 long time, and it does not take the situation of compound pollution into account. There is a great hidden
101 danger of desorption. Sundman A et al(Sundman A et al, 2020) showed that the magnetite produced by
102 iron oxide bacteria could remove arsenic and chromium efficiently by surface adsorption. However,
103 compared with primary magnetite, the results of biological modification were not satisfactory, and the
104 adsorption effect on mineral surface was greatly affected by the environment and was not stable
105 enough. Study on *Pseudomonas Taiwan* by Satapute P et al(Satapute P et al, 2019), metal stress
106 promoted the synthesis of cellular proteins, the surface of *pseudomonas taiwanensis* cells interacts with
107 metal cations (hydroxyl, carboxyl, phosphate, amino) to complete the adsorption of heavy metal ions.

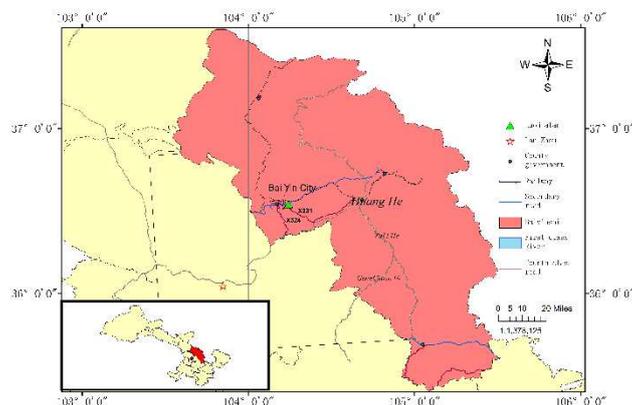
108 The results by Bao L L et al(Bao L-L, 2020) show that *Pseudomonas taiwanensis* can produce
109 ferromanganese minerals under ATCC™#279 culture and can wrap heavy metal ions in lattice, which
110 greatly reduces the mobility of heavy metal ions in soil. This method of indirect in situ remediation by
111 microorganisms is cheap and the removal rate of heavy metals is very high, which is a feasible method.

112 In order to improve the removal rate of heavy metals and reduce the toxicity of heavy metals in
113 the environment, the optimum growth conditions of *Pseudomonas taiwanensis*, the ability to resist
114 heavy metals and the role of iron-manganese ratio in the mineralization process were discussed. Then,
115 under the condition of combined pollution, the effect of iron-manganese ratio on the repair of
116 *Pseudomonas taiwanensis* was discussed.

117 2. Experimental materials and instruments

118 2.1 Materials and Methods

119 The soil in this experiment was collected from the surface soil (0-20 cm) of Luojiatan Village
120 (104.24E, 36.54N) (Fig. 1). Baiyin City is located inland, perennial drought and rain. Since the 1960s,
121 sewage has been used to irrigate farmland soil. Luojiatan Village is located in the upstream of the
122 sewage irrigation area, and the sewage irrigation has not been stopped until 2000. For 40 years, heavy
123 metals continue to accumulate and migrate in farmland soil, causing pollution(Liu W-J, 2012). Remove
124 impurities from soil samples by air drying, grinding and sieving (2 mm and 0.149 mm), standby.



125

126 **Fig. 1** sampling point location

127 Soil physical and chemical properties (pH, EC, organic matter, carbonate) were determined
128 according to the method in soil agrochemical analysis by Bao et al. Electric conductivity (EC) and pH
129 were measured with EC electrode (HI98311) and pH meter (pHS-3E) in deionized water (1:5, m/v),
130 respectively. EC reflects the salinity of the sample(Liu J et al, 2019). Organic matter (SOM) content
131 was determined by potassium dichromate volumetric method -dilute heat method and carbonate (CO₃²⁻)
132 content by neutralization titration method(Bao S-D, 2000).

133 Atomic fluorescence spectrometer (AFS-2880) was used to determine the concentration of total as,
134 and atomic absorption spectrometer (Analytikjena, ZEE nit 700p) was used to determine the
135 concentration of Pb, Cd, Cu, Zn, etc.

136 2.2 Analysis of pollution degree

137 The single factor index method is used to analyze the parity of single metal (metal-like) index one
138 by one, so as to determine the pollution degree of main heavy metals (metal-like) in soil. The

139 calculation formula is as follows:

$$140 \quad P_i = C_i/S_i$$

141 Where: P_i is the single pollution index of metal (metalloid) I; C_i is the measured concentration of soil
142 heavy metal (metalloid) I, mg kg^{-1} ; S_i is the evaluation standard value of soil heavy metal (metalloid) I,
143 mg kg^{-1} .

144 The Nemerow index method is used to reflect the degree of heavy metal (metalloid) compound
145 pollution by taking into account the average value and maximum value of single factor index. The
146 calculation formula is as follows(Guan Y et al, 2014):

$$147 \quad P_z = \sqrt{\frac{P_{i-\max}^2 + P_{i-\text{ave}}^2}{2}}$$

148 Where: P_z is Nemerow's comprehensive pollution index; $P_{i-\max}$ is the maximum value of each single
149 pollution index in soil; $P_{i-\text{ave}}$ is the average value of each single pollution index in soil.

150 2.3 Manganese Oxidizing Bacteria Strains

151 *Pseudomonas taiwanensis* (P4) was provided by Pan research group of Zhejiang University of
152 Technology. The strain was inoculated into Nutritional Broth (NB) medium (peptone 10 g L^{-1} , beef
153 extract 3 g L^{-1} , sodium chloride 5 g L^{-1} , $\text{pH}=7.0$) for expansion culture. After 12 hours of growth, the
154 solid ATCC™#279 medium containing 5ppm Mn(II) (ammonium ferrous sulfate 0.15 g L^{-1} , yeast
155 extract 0.075 g L^{-1} , sodium pyrophosphate 0.05 g L^{-1} , sodium citrate 0.15 g L^{-1} , $\text{pH}=6.8$, agar 1.5%)(He
156 Z et al, 2019) was coated on the plate. The plate was placed in a constant temperature incubator at 30°C
157 for 4-5 days. The growth of the surface strain was observed, and the well-growing plaque was picked,
158 then expanded and stored in the solid NB medium (agar 1.5%).

159 Preserved strains with universal primers 27F: 5'AGAGTTTGATCCTGGCTCAG3'; 1492R:
160 5'TACGGCTACCTTGTTACGACTT3' was used as the primers for PCR amplification of 16S rRNA
161 (primers were synthesized by Beijing Qingke Biotechnology Co. Ltd Xi'an Branch). Genomic DNA
162 was extracted from by plant genomic DNA extraction kit of Beijing Qingke Biotechnology Co. Ltd.
163 Xi'an Branch. PCR reaction system (50 μl): MIX 46 μl , 27F primer ($10\text{ pmol } \mu\text{l}^{-1}$) 1 μl , 1492R primer
164 ($10\text{ pmol } \mu\text{l}^{-1}$) 1 μl , genomic DNA 2 μl , PCR procedure: 98°C , 30 min; 98°C , 10s; 55°C , 10s; 72°C ,
165 20s; Cycle 30 times; 72°C , 2min. The amplified fragment was about 1400 bp-1700 bp, and the
166 sequencing was completed by Beijing Qingke Biotechnology Co. Ltd. Xi'an Branch. The splicing
167 results were compared on NCBI (<https://www.ncbi.nlm.nih.gov/>), and then the phylogenetic tree of
168 *Pseudomonas Taiwan* was established by Neighbor-Joining method in MEGA (6.06), and phylogenetic
169 analysis was carried out. Bootstrapping method was used to evaluate the phylogenetic tree.

170 2.4 Tolerance experiments

171 In order to find the best growth conditions, acid and alkali resistance and salt tolerance were tested.
172 First, prepare the NB medium and adjust the pH to be 3, 5, 7, 9, 11, the other group, using sodium
173 chloride as salt, set the salinity gradient to 2%, 4%, 6%, 8%, 10%(the amount of salt added in this
174 experiment is based on the configured medium, NB the original salinity of the medium is 0.5%),
175 sterilize the two groups of culture medium and inoculate 100 μl bacteria solution under relatively sterile
176 conditions, set two groups parallel and sample at 0, 6, 12, 24, 48, 72, 96, 120, 144, 168h. OD600(600
177 nm ultraviolet spectrum analysis) was determined in time to show the growth of bacteria.

178 Cadmium nitrate and sodium arsenate as stress sources. The arsenic tolerance experiment was set

179 to 0, 100, 200, 300, 400, 450 mg L⁻¹, add arsenic source to NB medium, adjust pH=7, inoculate 100
180 bacteria solution after sterilizing in high pressure sterilizer. In Cd experiment, the Cd gradient was set
181 to 0, 8, 15, 30, 50, 80 mg L⁻¹, after the same treatment, both groups were set parallel, sampling 2 ml, at
182 0, 6, 12, 24, 48, 72, 96, 120 hours, respectively spectrophotometry.

183 2.5 Heavy Metal Removal Experiment in Water Solution

184 In order to study the removal effect of P4 bacteria on Pb, As and Cd in aqueous solution, heavy
185 metal removal experiments were carried out. P4 strain was inoculated in NB medium and cultured to
186 logarithmic growth phase (12-16 h). The following groups were established: 1) group (Pb-P4); 2) group
187 (As-P4); 3) group (Cd-P4); 4) group (composite-P4) and blank control (without adding
188 microorganisms). The 25 ml P4 bacterial solution was transferred to 250 ml conical flask containing
189 Mn(II), target heavy metal with initial concentration of 5 ppm and 225 ml ATCCTM#279 medium. At 0,
190 6, 12, 24, 36, 48, 72, 96, 120, 144, 168h, 1 ml sample was taken, and 9 ml deionized water was added
191 to measure the concentration of heavy metals and pH of culture medium.

192 2.6 Soil testing

193 After the P4 strain was cultured for 14h, the mixed solution was prepared according to the ratio of
194 bacterial solution to ATCCTM#279=1:10. Adding 6ml of the mixture in 10 g soil, shaking 30 min at
195 25°C, 150 rpm, placed in the ventilation, 14 days later, crushed 0.149 mm sieve, weighed 1.0 g for
196 Tessier continuous extraction (Tessier A, Campbell PGC& Bisson M, 1979). Finally, the contents of
197 heavy metals in the extracted solution were tested respectively

198 2.7 Research on the Optimal Ferromanganese Ratio

199 Using the characteristics of microbial mineral passivation of heavy metals, mineral production
200 will undoubtedly affect the passivation effect of heavy metals. In this study, iron and manganese as a
201 mineral element. The addition and proportion are the key factors determining the ore production.
202 According to the tolerance of bacteria to manganese, we keep the content of iron in ATCCTM#279
203 medium unchanged, by changing the amount of manganese added to prepare different ratios of iron and
204 manganese culture medium. The specific method is to prepare Fe/Mn molar ratio of 5:1, 3:1, 1:1, 1:3,
205 1:5, add 1000 mg L⁻¹ Mn(II) to manganese ion concentration of 4.21 mg L⁻¹, 7.02 mg L⁻¹, 20.98 mg L⁻¹,
206 63.13 mg L⁻¹, 105.26 mg L⁻¹, each group to add the configured arsenic solution to arsenic concentration
207 of 50 mg L⁻¹. After inoculation of Pseudomonas Taiwan, at 30 °C, 180rpm shaking culture for 14 days,
208 take 2 ml sample solution, with 0.22 um water phase filter head filter out supernatant, determination of
209 arsenic concentration. The same method of soil remediation, weighing 30.0 g soil, solid-liquid ratio set
210 to 3:1, every 7 days take a sample, a total of 5 times.

211 2.8 Representation analysis

212 The resulting precipitation was cooled and dried for 24h by a cold dryer (Gansu Jimei Instruments
213 & Equipment Technology Co., Ltd., LGJ-12A), and then ground into powder by a mortar. X-ray
214 diffraction (XRD) was performed by X-ray diffractometer (PANalytical, Netherlands, X'PertProMPD)
215 to analyze the change of solid crystal phase before and after the addition of heavy metals. Scanning
216 electron microscopy-X-ray energy spectrum (SEM-EDS) (Hitachi, Japan, S-3400N) was used to
217 analyze the mineral morphology characteristics, element types and contents. Fourier transform infrared
218 spectrometer (FTIR) (NICOLET, NEXUS 670) was used to determine the chemical bonds and

219 functional groups in the molecules.

220 2.9 Statistical analysis

221 Statistical analysis of experimental data using Microsoft Excel 2016 software, drawing using
222 Origin 8.0 and ArcGIS 10.2.

223 3. Results and discussion

224 3.1 Soil condition

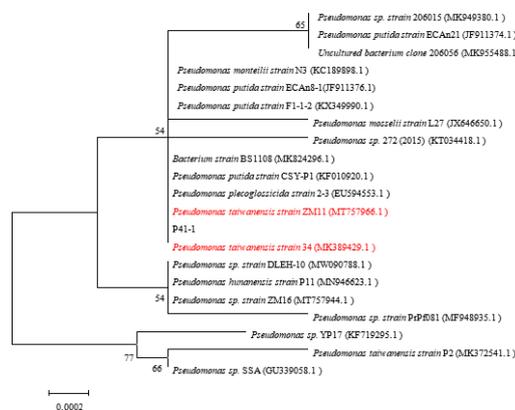
225 The physicochemical properties and heavy metal content of soil were determined, and the results
226 are shown in Table 1.

227 The criteria shown in Table 3 are the Agricultural Land Pollution Risk Control Standards (GB
228 15618-2018) issued by the Ministry of Ecological Environment of China (Ministry of Ecological
229 Environment, 2018). It can be seen from the data that the soil is neutral soil. According to the risk
230 screening value given by this standard, the single factor pollution index and the comprehensive
231 pollution index are calculated, and the results are shown in Table 4.

232 Combining Table 2, we know that the soil pollution was mainly heavy pollution caused by Cd and
233 Pb, and slight pollution caused by As and slight pollution caused by Zn and Cu. Among them, As, Cd,
234 Pb, Zn, Cu, Ni, Cr and Hg were 1.59, 284.68, 6.59, 2.24, 2.46, 0.42, 0.23 and 0.41 times of the standard
235 risk screening values, respectively.

236 3.2 Sequencing of strains

237 According to the splicing results, BLAST and similarity analysis were performed using its 16S
238 rRNA partial sequence (1433 bp). The results showed that the strain was identified as *Pseudomonas*
239 *taiwanensis* (similarity 99%, accession number : MT757966) and named as *Pseudomonas taiwanensis*
240 strain ZM11. Its characteristics were Gram-negative, rod-shaped, Athletic, and non-sporogenous, and it
241 was classified as G-protein bacteria (Wang L et al, 2010) and could grow at 5-42 °C, but could not grow
242 at higher temperature (45-60 °C). The optimal growth temperature was 30-37 °C, the strain could grow
243 at pH=4-9, and the optimum pH was 6-8 (Li L-L et al, 2019). According to the development tree (Fig. 2),
244 P4-1 strain had the closest relationship with Taiwan *Pseudomonas*, which was consistent with the
245 identification results.



246

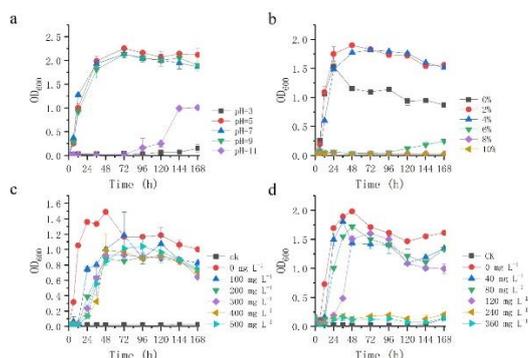
247

Fig. 2 Phylogenetic tree of P4 strain

248 3.3 Tolerance of strains

249 In order to study the environmental adaptability of bacteria and better understand the toxicity

250 mechanism of heavy metals, the concentration of heavy metals related to the environment and the
251 physical and chemical properties were treated, and the OD value was used to describe the growth of
252 bacteria. The results are shown in the Fig. 3.



253

254 **Fig. 3** Growth curve of bacteria under different factors(A: pH; B: salt; C: As(V); D: Mn(II))

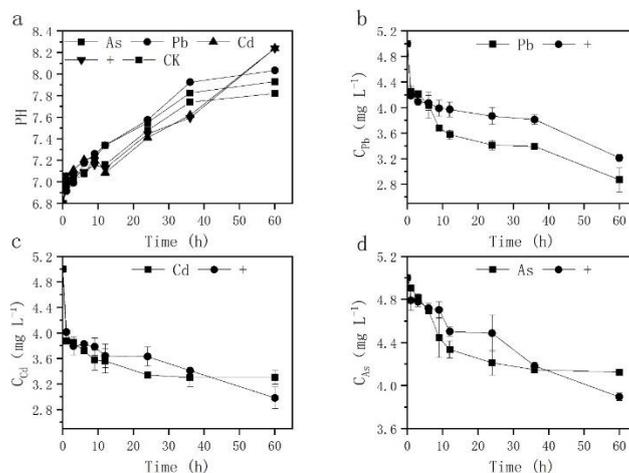
255 The Fig. 3a showed that the strain could not grow normally under the condition of strong acid (pH
256 < 3) and strong alkali (pH > 11). Moreover, under the condition of weak acid, the strain grew
257 vigorously, and the optimum pH was 5. When the initial pH was 11, the growth of the strain increased
258 after 84 hours, and the growth speed was accelerated. In contrast, when the initial pH was 3, the growth
259 of the strain was always inhibited. Soil pH value is the main factor affecting the structure and
260 composition of bacterial community(Wan W et al, 2020). The pH range of optimal growth of bacteria
261 may directly affect its proportion in soil microbial community(Rousk J et al, 2010), thereby affecting
262 its role in soil. In terms of salinity, when the added NaCl content was more than 4%, the growth of the
263 strain was almost completely inhibited, and when 2% NaCl was added, the growth of the strain was
264 more than that without NaCl (Fig. 3b). Similar to pH, salinity will change the abundance of
265 microorganisms in the soil and affect their role in the soil(Zhang W et al, 2019). So that bacteria on pH
266 and salinity tolerance determines the size of its role in the soil. The pH and EC values of the soil
267 selected in this study were 7.38 and 1922, respectively, which were all within the tolerance range of P4
268 indicating that the soil physical and chemical conditions were suitable for the growth of this strain.

269 As the key point of the tolerance of bacteria, the resistance of bacteria to As, Cd and Pb is the key
270 to their survival in contaminated soil. We used trisodium arsenate as the stress source, with 0-500 mg
271 L⁻¹ different gradient added to the bacterial liquid and observed its growth. Results as shown in the Fig.
272 3c, under arsenic stress, the growth of *Pseudomonas Taiwan* was significantly inhibited, and with the
273 increase of arsenic concentration, the growth rate of bacteria in the initial stage decreased. When the
274 concentration of arsenic reached 500 mg L⁻¹, in 60 h, the growth of bacteria and arsenic concentration
275 in 200, 300, 400 mg L⁻¹ at the same time. After 100 h, the highest concentration of bacteria growth
276 began to decline. In contrast, in the blank condition and under the condition of low concentration of
277 arsenic, the decline time of strain growth was estimated after 48 h and 60 h, respectively. It can be seen
278 that arsenic, as a poison, delayed the growth of *P. taiwanensis* to a certain extent(Halan B et al, 2017).
279 In fact, arsenic and cadmium stress on the cell membrane interaction, may be with the membrane
280 hydroxyl, amino, phosphate. etc(Raja CE, Anbazhagan K& Selvam GS, 2006), and will promote cells
281 to produce proteins such as superoxide dismutase (SOD), catalase (CAT), glutathione transferase
282 (GST)(Satapute P et al, 2019), these feedbacks from microbes enable them to adsorb heavy metals in
283 soil (Cavalcanti luna MA et al, 2015). Manganese as a resource of microbial mineralization, excessive
284 manganese will play a toxic role on microorganisms. Therefore, we explored the manganese tolerance
285 of microorganisms. The results (Fig. 3d) showed that when the concentration of manganese was in the

286 range of 0-120 mg L⁻¹, the growth of microorganisms was not strongly affected, and when the
 287 concentration of manganese reached 240 mg L⁻¹, the bacteria almost did not grow. Compared with that,
 288 under the condition of 120 mg L⁻¹ manganese concentration, the bacteria reached the logarithmic phase
 289 after 36 hours, which was relatively delayed by 1 day compared with the blank. The toxic effect is
 290 relatively small, and if the manganese content is more than 120 mg L⁻¹, it can not guarantee the high
 291 survival rate of *P. taiwanensis*, let alone the ore production under this condition.

292 3.4 Removal of heavy metals in aqueous solution

293 The water system is relatively simple to the soil, adding heavy metal sources in water, and adding
 294 it to the mixed bacterial liquid for interaction. In order to study the performance of microorganisms in
 295 the removal of heavy metals in water, and then predict its remediation of soil. According to the data(Fig.
 296 4), the removal rate of complex heavy metals by bacteria seems to be lower than that by single metal
 297 source in liquid phase. When bacteria were added to the culture medium containing heavy metals, the
 298 pH of the culture medium gradually increased with time. Within 60 hours, the pH of the blank culture
 299 medium increased from 6.8 to 7.82. When heavy metals were added to the culture medium, the pH of
 300 the culture medium remained the same. The difference was that when arsenic and lead were added
 301 respectively, the pH of the culture medium seemed to increase faster, after 60 hours, the pH of the
 302 culture medium increased by 1.13 and 1.23 units respectively. When the heavy metals were added to
 303 cadmium, the pH of the culture medium changed slowly before 36 hours. When the time came to 60
 304 hours, the pH of the culture medium reached 8.24. It is worth noting that when the three heavy metals
 305 were added at the same time, the pH of the culture medium increased by 1.44 units after 60 hours.



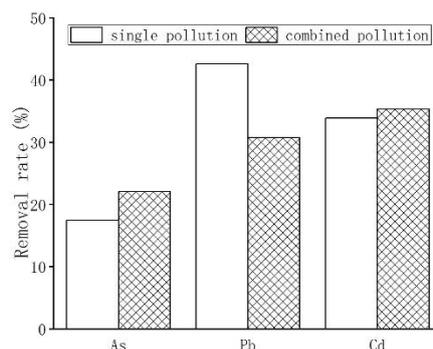
306

307 **Fig. 4** Concentration and pH changes of heavy metals in single or combined polluted aqueous
 308 solution (A: pH; B: Pb; C: Cd; D: As;)

309 The increase of pH value will affect the growth of bacteria and the precipitation of heavy metal
 310 ions. Acidic conditions are not conducive to the oxidation of Mn(II), while Fe(III) is easy to form
 311 Fe(OH)₃ precipitation under alkaline conditions(Akob DM et al, 2014; Emerson D, Fleming EJ&
 312 Mcbeth JM, 2010), which is consistent with the results of Hou et al(Hou D et al, 2020),and Pacini VA
 313 et al(Pacini VA, María ingallinella A& Sanguinetti G, 2005). Whether it is due to the consumption of
 314 CO₂ by FeOB metabolism(Pacini VA, María ingallinella A& Sanguinetti G, 2005)or the environmental
 315 changes caused by MnOB(Akob DM et al, 2014), it is beneficial for bacterial mineral production.

316 The contents of heavy metals in the four media were tested at different time points to obtain the
 317 variation curve shown in the Fig. 4b, c, d. It can be seen from the image that when three kinds of heavy

318 metal sources are added alone, the decline rate of its concentration in the early stage is generally faster
 319 than that when three kinds of heavy metal sources are added at the same time. The Fig. 5 shows us the
 320 removal rates of three heavy metals after 60 hours. For arsenic and cadmium, it seems that the
 321 treatment rate of microorganisms in the compound pollution is higher, reaching 22.09% and 40.34%,
 322 respectively. On the contrary, when lead is added to the culture medium alone, the effect of
 323 microorganisms is relatively large, and in a short period of 60 h, the treatment efficiency can reach
 324 42.61%. When it is added to the mixed liquid with the other two metals, the treatment efficiency
 325 decreases to 35.75%. Among the three heavy metals, cadmium has the highest treatment efficiency
 326 under combined pollution, and lead has the highest treatment efficiency when added alone. In a
 327 complex environment, the proteins produced by cells are related to external interference factors, and
 328 the stress of heavy metals promotes the production of proteins. When proteins bind to a variety of
 329 heavy metals, the affinity of heavy metals to these products and their own properties determine the
 330 order of their binding. On the other hand, the properties of Fe-Mn oxides, such as specific surface area,
 331 formation process, morphological structure and interaction with different heavy metals are also the key
 332 factors to determine the removal efficiency(Nelson YM et al, 2002; Xiao A, Li WC& Ye Z, 2020). The
 333 homogeneous chemical adsorption of Pb by BMO is faster than that of Cd, so that the remediation
 334 efficiency of Pb is higher(Zhou D, Kim D& Ko S, 2015). The presence of Cd and As reduces the
 335 adsorption of Pb by BMO.



336

Fig. 5 Removal rate of heavy metals in aqueous solution after 60 hours

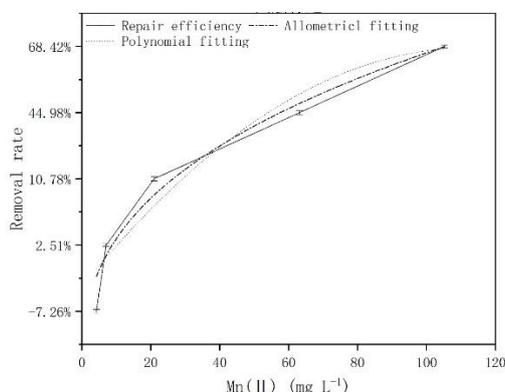
337

338 This may be due to the greater retention tendency of Cd at the exchange site. Under this pH
 339 condition, Cd interacts with the exchange site, competes with Pb, and the adsorption amount is
 340 large(Serrano S et al, 2009). The existence of Fe(II) may play a certain role in the reduction of As(V).
 341 Studies have shown that the effect of BMO on As is relatively slow, and the adsorption of As(V) is
 342 carried out after oxidation(Watanabe J et al, 2013). In the presence of a variety of metals, the
 343 adsorption of divalent metal ions is considered to inhibit the charge repulsion between the oxide
 344 surface and As(V), thereby promoting the adsorption of As(V)(Tani Y et al, 2004; Watanabe J et al,
 345 2012). In any case, under the simultaneous stress of composite metals, biological ferromanganese ore
 346 has a certain adsorption effect on various metals in the formation process.

347 3.5 Effects of different iron-manganese ratios

348 The experiment is based on ATCCTM#279 medium formula, by changing the content of Mn(II), so
 349 as to explore the conditions of different manganese iron ratio, microbial production and, at the same
 350 time, adding the same amount of arsenic stress in the medium, determine the removal of arsenic in the
 351 process of microbial production in different iron manganese ratio. The results showed that the addition
 352 amount of manganese ions was significantly positively correlated with the remediation efficiency of

353 microorganisms for arsenic. With the increase in the addition amount of manganese, the concentration
354 of arsenic in the mixture after 14 days of culture decreased. When the concentration of manganese
355 reached 105 mg L^{-1} , the remediation efficiency reached more than 68%, and the iron-manganese ratio
356 was 1:5. Origin Pro2018 is used to fit the data and the result is shown(Fig. 6).



357

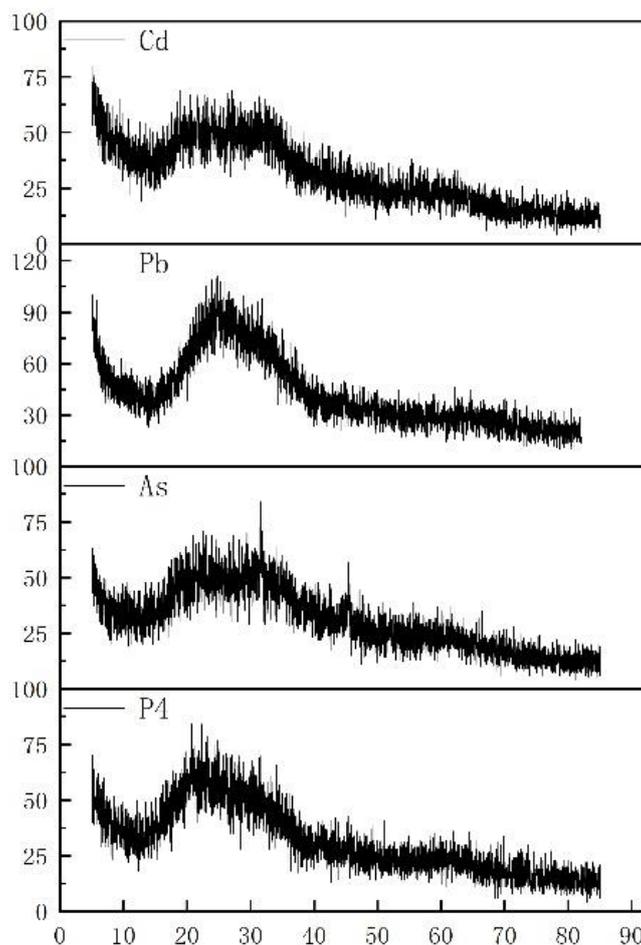
358 **Fig. 6** Removal rate of arsenic under different manganese additions

359 According to the comparison of the obtained fitting curve data, the power index function is more
360 suitable for the curve, so it can be roughly considered that the addition amount of manganese in this
361 experiment is positively correlated with the removal efficiency of arsenic. In any case, the results
362 showed that the removal efficiency of arsenic was the highest when the iron-manganese ratio was 1:5,
363 which was not consistent with the results of Bai et al(Bai Y et al, 2016), because of the different forms
364 of arsenic and the total amount of iron and manganese. During the formation of Fe-Mn oxides, As(V)
365 and Mn(II) are adsorbed at the same time, and the adsorption mode of As(V) on the generated Fe-Mn
366 oxides changes with time(Watanabe J et al, 2012). The increase in Mn content first increases the
367 production of Mn oxides in the solution, so the adsorption amount of heavy metals is increased.

368 3.6 Morphological analysis

369 XRD

370 As shown in Fig. 7, under different heavy metal stress conditions, the diffraction characteristics of
371 bio-manganese oxide combined with heavy metals are basically the same, with low peak and wide peak
372 width, which is consistent with the results obtained in many studies(Hou D et al, 2020; Zhou D, Kim
373 D& Ko S, 2015). It is reported(Zhang Z et al, 2014), compared with the chemical synthesis of
374 manganese oxide materials, biological manganese oxide materials have poor crystallization and cannot
375 be arranged in an orderly manner. This characteristic may be beneficial to the formation of adsorption
376 conditions for heavy metals. Irregular crystals have larger specific surface area, which increases the
377 surface effective binding sites and exchange sites. Under the stress of different heavy metals, the
378 diffraction peak changed slightly, but did not show a stable peak.



379
380 **Fig. 7** XRD images of precipitates under different heavy metal treatments

381 FTIR

382 The minerals produced by *P. taiwanensis* were analyzed by Fourier transform infrared
383 spectroscopy(FTIR), and the results are shown in the Fig. 8. The band in the range of 450-800 cm^{-1} can
384 be attributed to the Mn-O vibration of manganese oxides(Feng Q et al, 1999). The absorption bands of
385 1059-1070 cm^{-1} , 1115-1124 cm^{-1} and 1165-1171 cm^{-1} belong to the inner Mn(III)-OH vibration.
386 Therefore, the peaks at 696.188 cm^{-1} and 615.191 cm^{-1} are the Mn-O lattice vibration in the mineral,
387 and the peaks at 1118.529 cm^{-1} and 1384.662 cm^{-1} are the inner Mn(III) -OH vibration(Chen X et al,
388 2017; Kang L et al, 2007). The peaks at 3399.943 cm^{-1} , 2929.389 cm^{-1} and 2345.054 cm^{-1} correspond
389 to the vibration of hydroxyl and crystal water(Chen X et al, 2017). In summary, the mineral is
390 consistent with the characterization of biological manganese oxides.

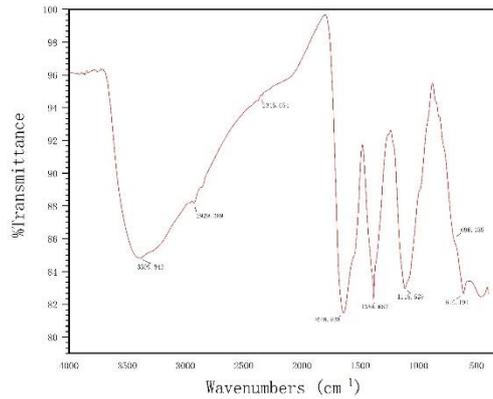


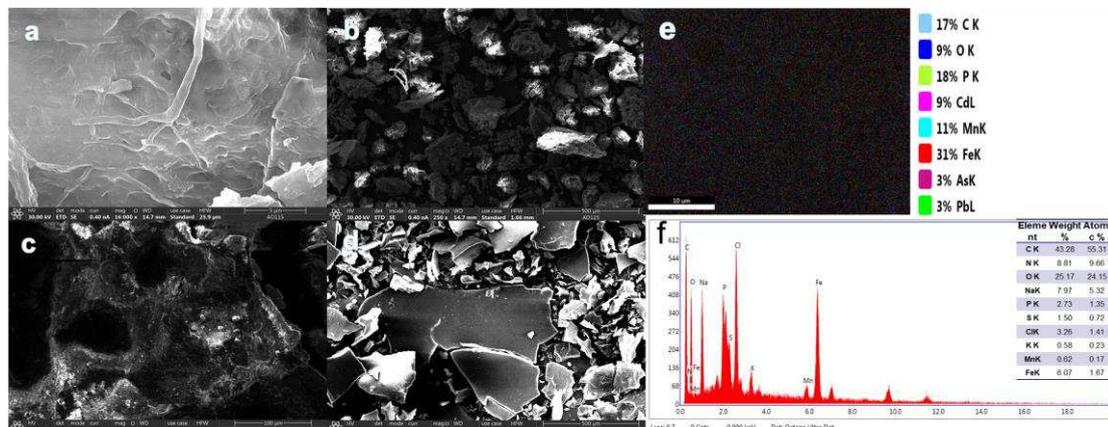
Fig. 8 FTIR spectra of minerals produced by *Pseudomonas taiwanensis*

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393 SEM

394 The scanning electron microscope images of heavy metals adsorbed by bio-manganese oxide
 395 produced during the growth of *Pseudomonas taiwanensis* were shown in Fig. 9. The irregular mineral
 396 surface generated by bacteria is uneven, and relatively flat surface can be formed under the stress of
 397 heavy metals. From the energy spectrum(Fig. 9f), it can be seen that the mineral surface contains a
 398 large number of carbon(C), oxygen(O), nitrogen(N), phosphorus(P), iron(Fe) and manganese(Mn)
 399 elements, which indicates that bio-manganese oxide is closely integrated with the organic groups
 400 generated by cells. Some scholars(Cömert S& Tepe O, 2020) proposed that bio-manganese oxide exists
 401 in the form of complex between manganese oxide and bacterial cells, which can increase the
 402 availability of organic nutrients and protect microorganisms from potential toxic substances, reactive
 403 oxygen species and ultraviolet radiation(Moura HM& Unterlass MM, 2020). From the mapping
 404 image(Fig. 9e), it can be seen that heavy metals adsorbed on its surface, and some studies have shown
 405 that(Zhang Y et al, 2019), the removal of heavy metals may also be due to the intracellular complex
 406 detoxification. In addition, it is worth noting that the surface phosphorus content is very high,
 407 phosphate and arsenate show similar physical and chemical behavior, which will replace arsenic and
 408 increase the mobility of arsenic in soil(Cao X, Ma LQ& Shiralipour A, 2003). We believe that
 409 phosphorus-containing organic matter in the mixture will compete with heavy metal ions for mineral
 410 binding sites, and the use of phosphate by bacteria may reduce this effect(Wang J et al, 2020).



411

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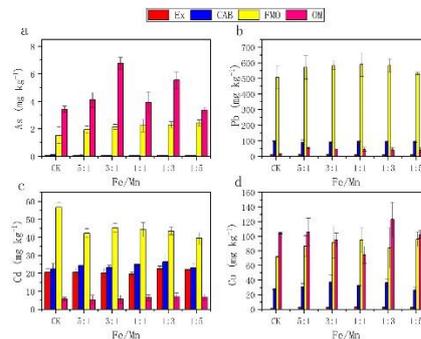
Fig. 9 SEM images and analysis results of minerals produced under different stresses
 a: blank; b: Pb; c: As; d: Cd; e/f: Mapping image and eZAF Smart Quant Results under combined

414

pollution.

415 3.7 Soil remediation

416 The remediation efficiency of *P. taiwanensis* on arsenic in Luojiatan farmland soil is shown in the
417 Fig. 10. The morphology of single heavy metal is analyzed. Before and after remediation, the changes
418 of heavy metal morphology are different under different iron-manganese ratios. The contents of As and
419 Pb Fe-Mn oxides are increased, and the forms of Cd and Cu are mostly decreased. For As, the content
420 of As bioavailable fraction changed little at low concentration, and the organic fraction increased more,
421 which may be due to the role of microorganisms. Pb mainly existed in Fe-Mn oxidation state, while its
422 total content of bioavailable fraction was low, and the Pb in organic fraction was the largest after
423 remediation, which could be doubled. For the most polluted Cd, the addition of microorganisms seems
424 to have failed to make a great change in its pollution status. The metal mainly changes with the
425 increase of carbonate state and the decrease of Fe-Mn oxide state. Similarly, under microbial
426 remediation, the increase of Fe-Mn oxide Cu content cannot avoid the overall activation, which is
427 mainly manifested in the maximum increase of its exchangeable state to twice. In a variety of high
428 concentrations of heavy metals combined pollution, the role of microorganisms is not large in the
429 aqueous phase, the different iron-manganese ratio will affect the change of iron-manganese oxidation
430 state content, for the biologically effective state, when the iron-manganese ratio is 1:5, the removal of
431 heavy metals is the best, respectively, 42% ; Pb, 6% ; Cu, 1% ; However, the maximum removal
432 efficiency of bioavailable Cd was 1.78%, and the Fe/Mn ratio was 5:1.



433

434 **Fig. 10** Morphological changes of heavy metals under different Fe / Mn ratios

435

436 According to the data in the water phase experiment, the microbial remediation effect becomes
437 worse under the combined stress of heavy metals. However, from the results of soil remediation, the
438 combined stress of microorganisms on a variety of heavy metals does not seem to play a certain role,
439 and heavy metals such as copper are activated instead. The migration and transformation of heavy
440 metals in the soil are the results of various mechanisms. The detoxification effect of bacteria on heavy
441 metals can only play a limited role, and the role of the medium should be considered when the
442 bacterial solution is added to the soil. Although the phosphorus-containing medium is the growth
443 demand of microorganisms, it may react on the passivation of heavy metals. Some studies have shown
444 that some phosphates can promote the dissolution of minerals(Yan DY& Lo IM, 2012) and reduce the
445 adsorption density of heavy metals on minerals(Qin W et al, 2012), which is undoubtedly contrary to
our purpose. Therefore, the utilization rate of phosphate by microorganisms is also crucial.

446 4. Conclusion

447 All the results above indicate that *Pseudomonas* ZM11 is tolerant to heavy metals such as arsenic,

448 lead and cadmium, and the bio-manganese oxide produced by *Pseudomonas* ZM11 has a certain
449 removal effect on heavy metals in aqueous phase, which is related to the proportion of iron and
450 manganese added in the culture of microorganisms. The strain can be used as an effective strain for
451 remediation of heavy metal pollution and can be used as a basis for remediation of heavy metal
452 contaminated soil. However, the technology of bioremediation of heavy metal contaminated soil is
453 limited. Considering the co-contamination of multiple heavy metals, its toxicity may increase. The
454 interaction between heavy metals and their interaction with microorganisms may reduce the activity of
455 some heavy metals and may also activate other heavy metals.

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Figures

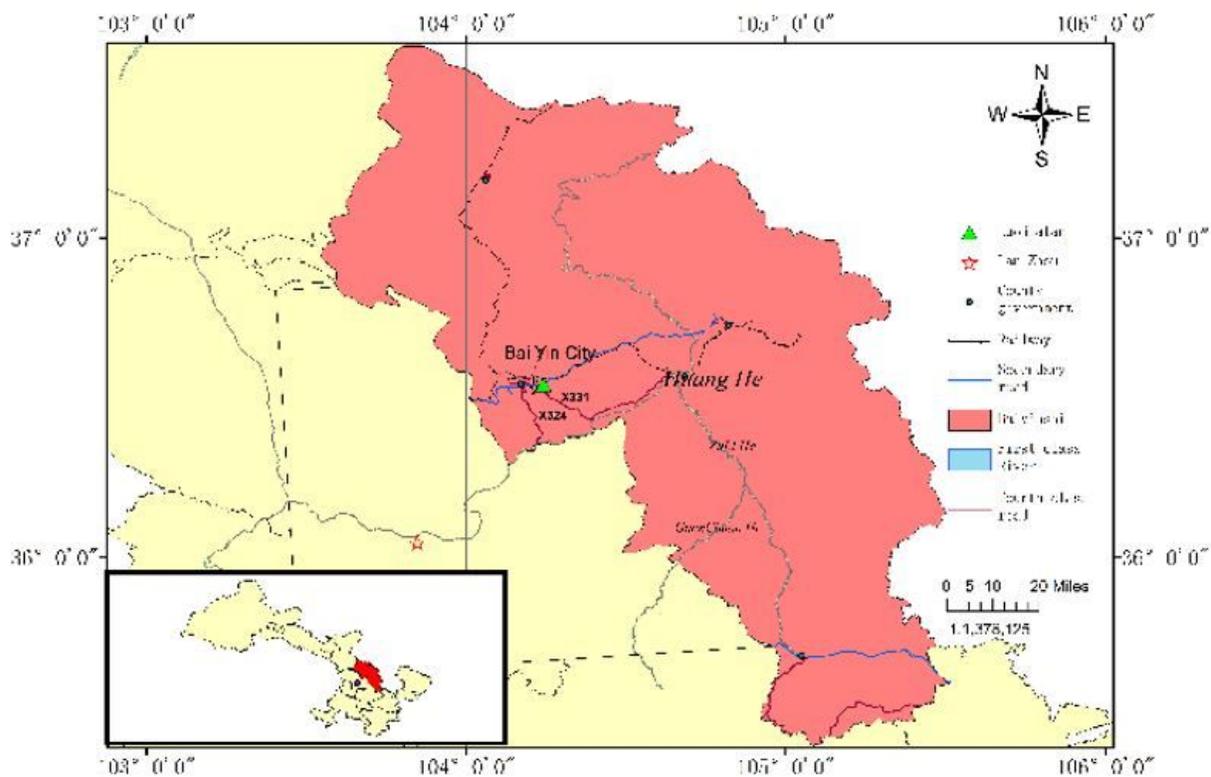


Figure 1

sampling point location Note: The designations employed and the presentation of the material on this map do not imply the expression of any opinion whatsoever on the part of Research Square concerning the legal status of any country, territory, city or area or of its authorities, or concerning the delimitation of its frontiers or boundaries. This map has been provided by the authors.

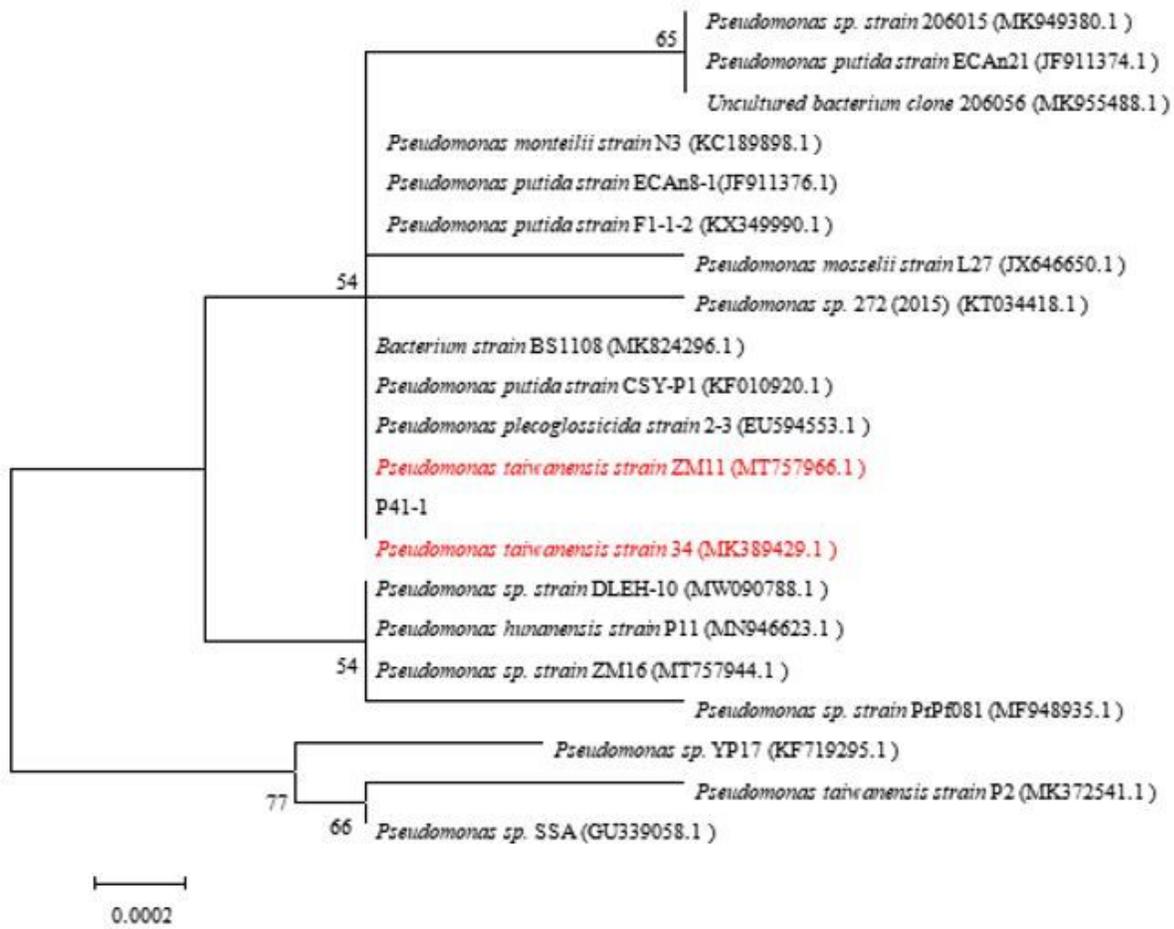


Figure 2

Phylogenetic tree of P4 strain

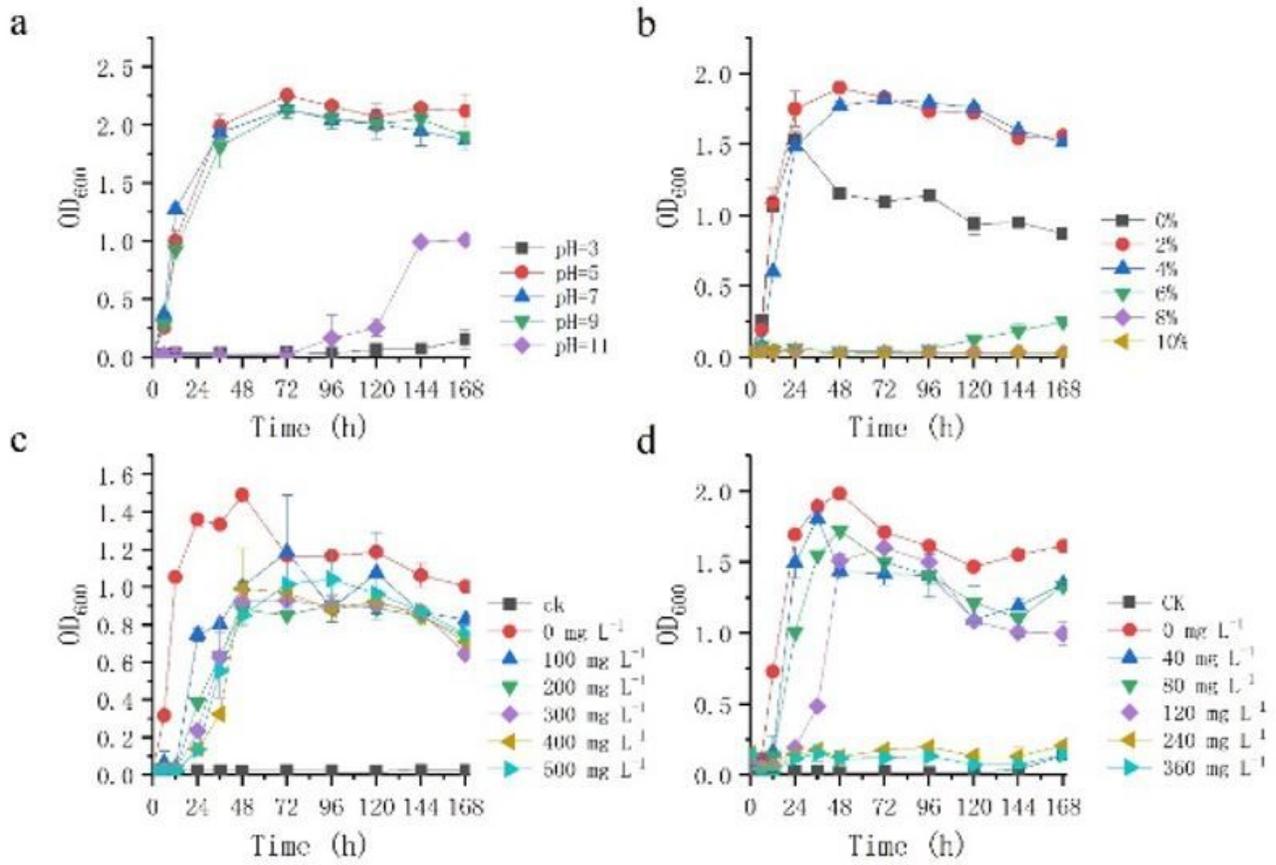


Figure 3

Growth curve of bacteria under different factors(A: pH; B: salt; C: As(V); D: Mn(II))

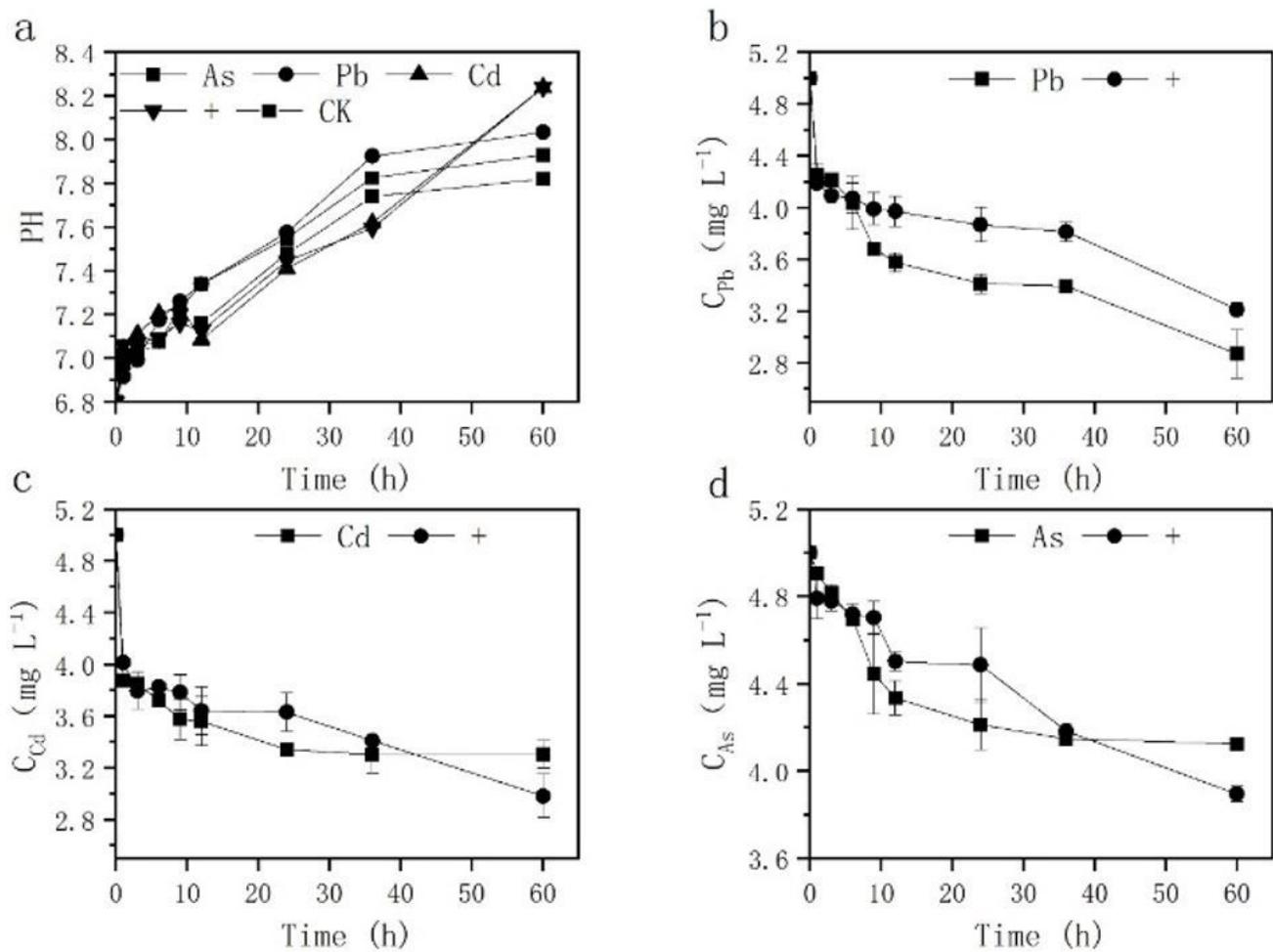


Figure 4

Concentration and pH changes of heavy metals in single or combined polluted aqueous solution (A: pH; B: Pb; C: Cd; D: As;)

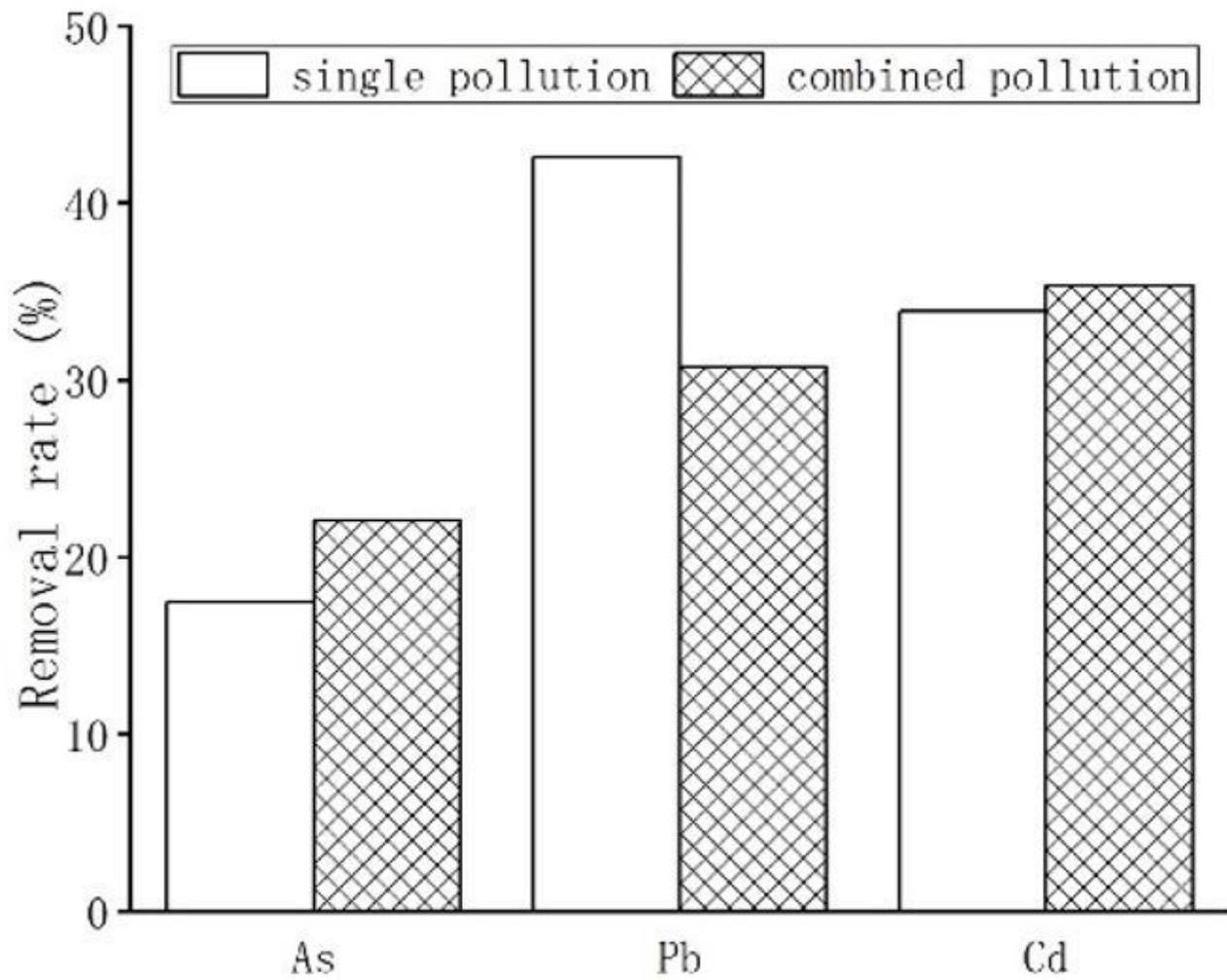


Figure 5

Removal rate of heavy metals in aqueous solution after 60 hours

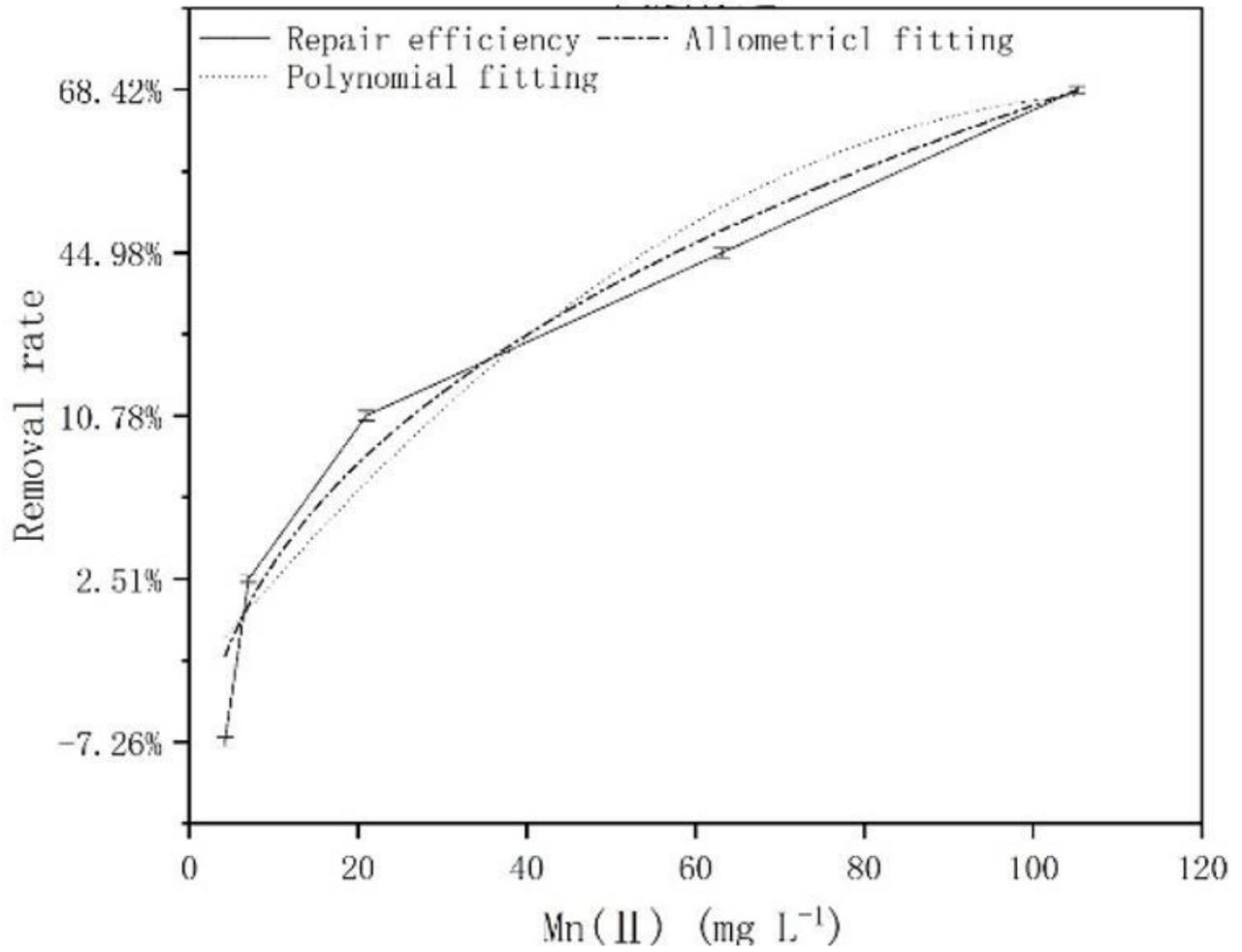


Figure 6

Removal rate of arsenic under different manganese additions

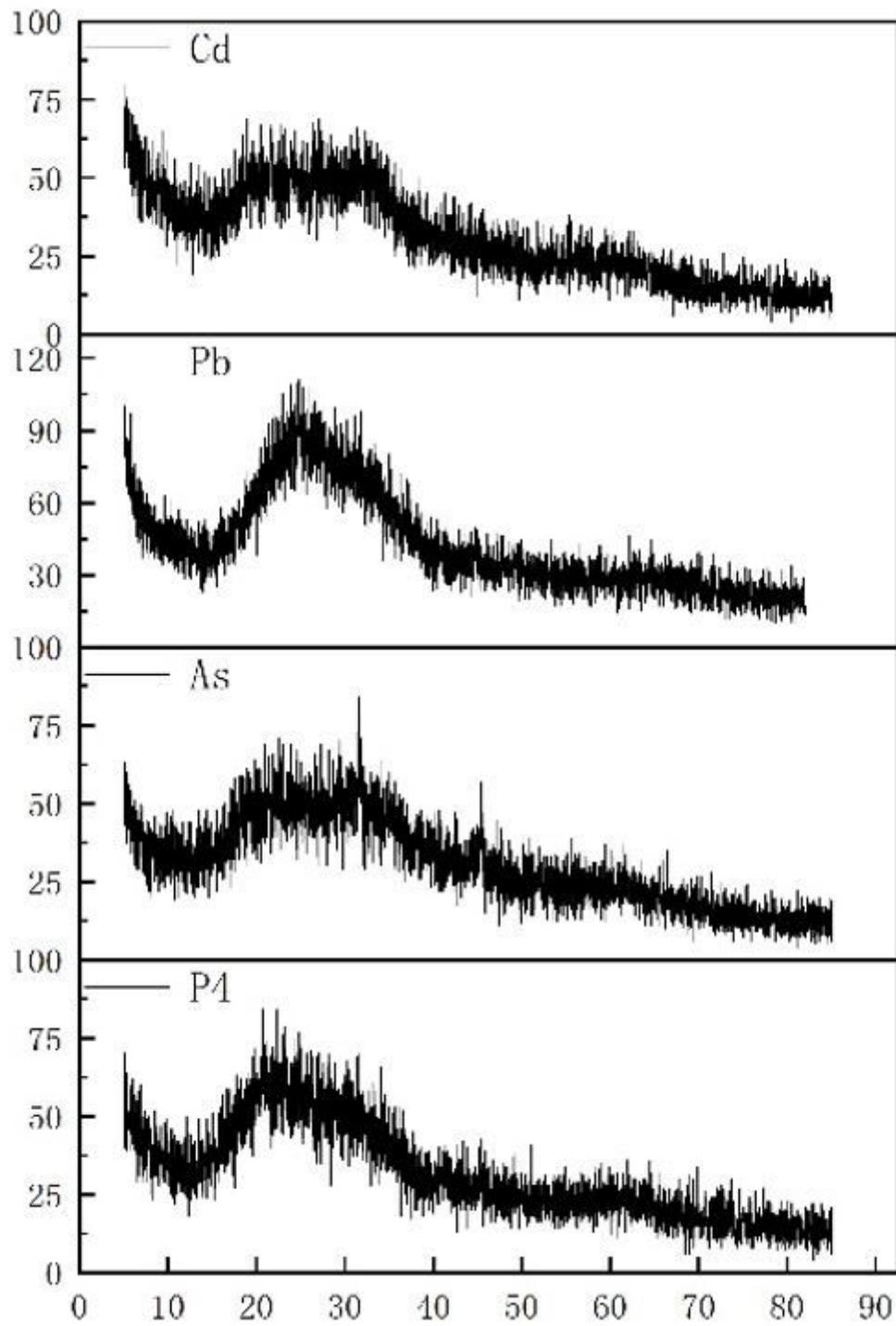


Figure 7

XRD images of precipitates under different heavy metal treatments

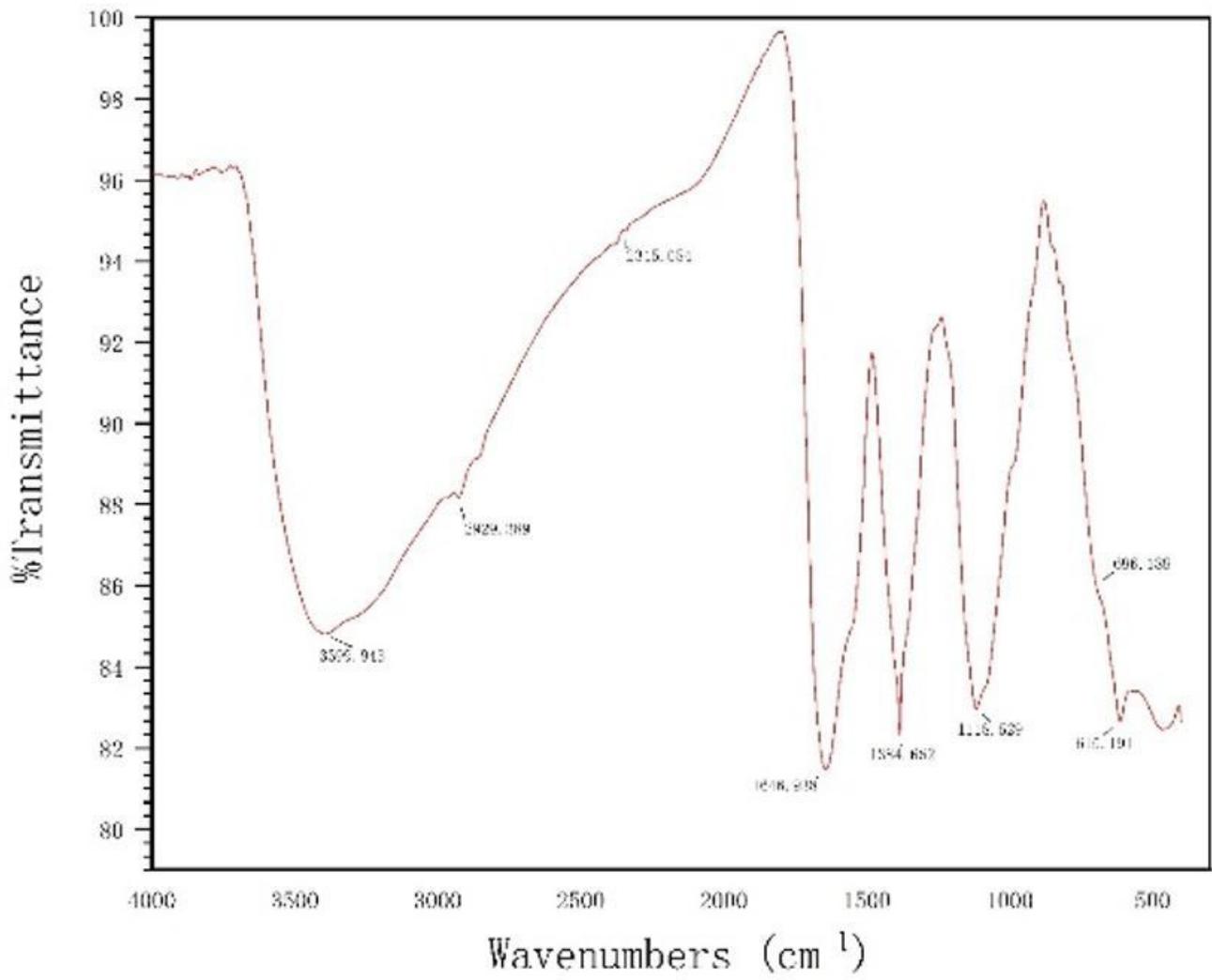


Figure 8

FTIR spectra of minerals produced by *pseudomonas taiwanensis*

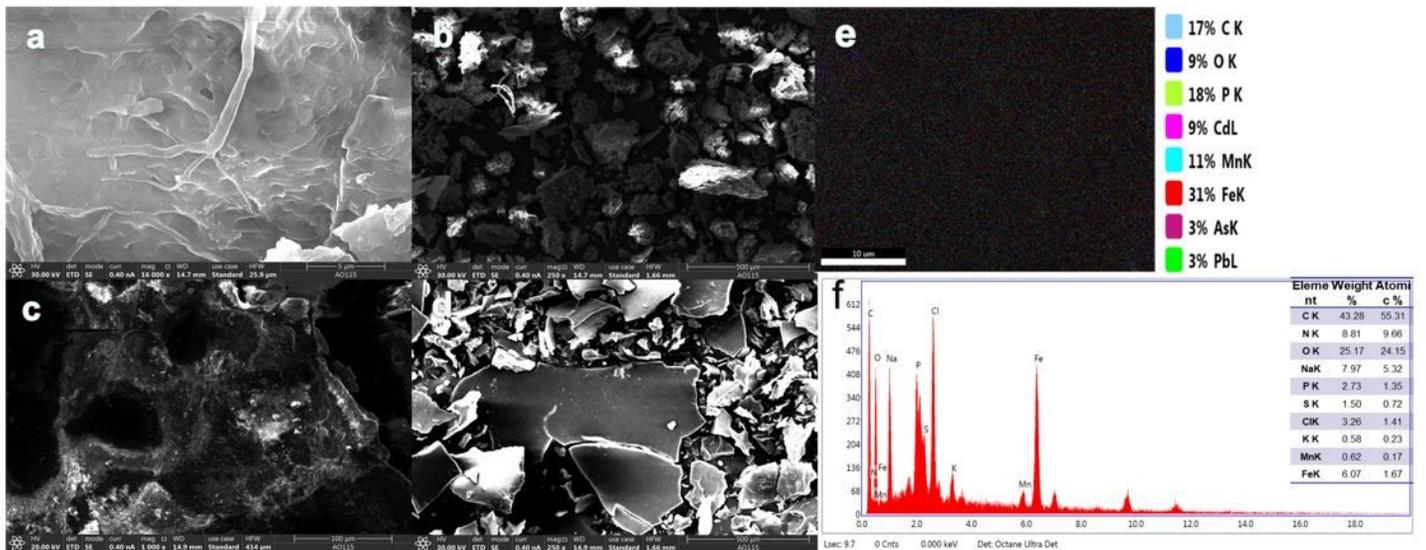


Figure 9

SEM images and analysis results of minerals produced under different stresses a: blank; b: Pb; c: As; d: Cd; e/f: Mapping image and eZAF Smart Quant Results under combined pollution.

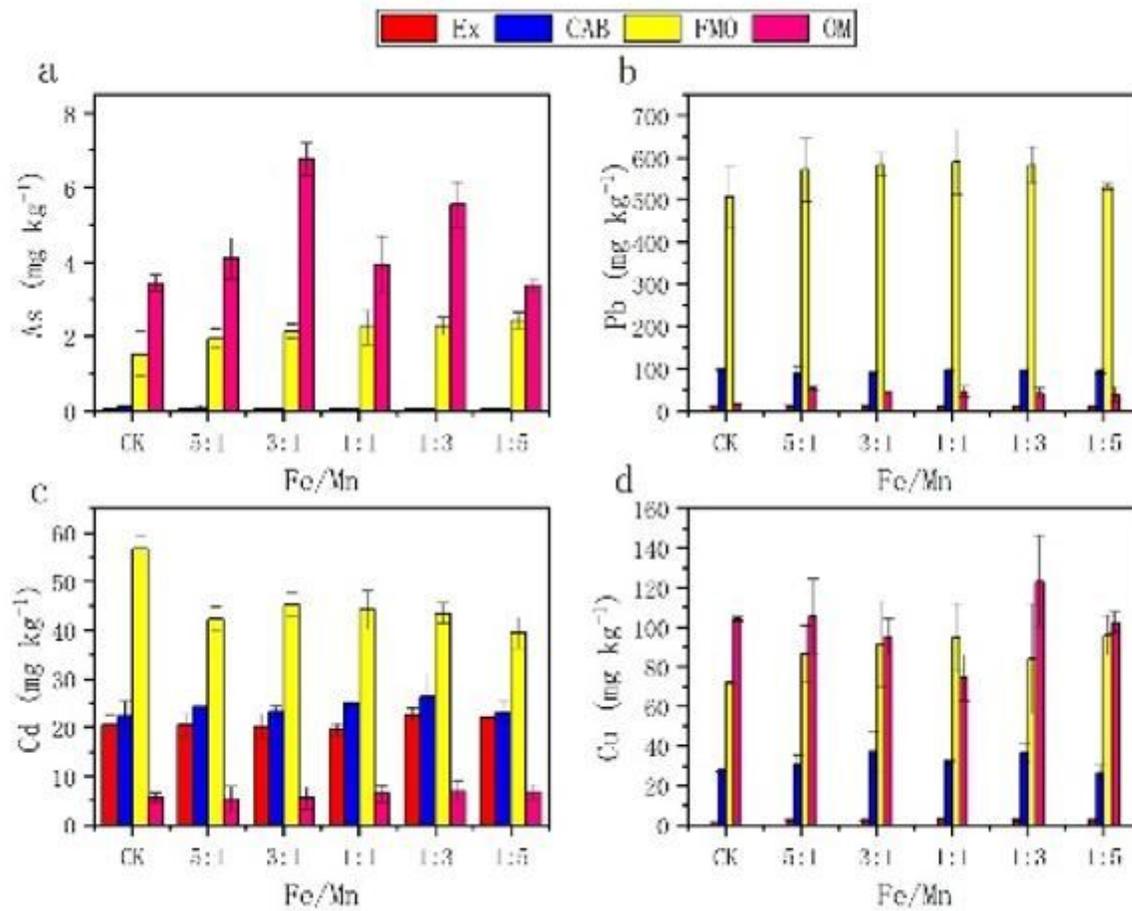


Figure 10

Morphological changes of heavy metals under different Fe / Mn ratios