

Seed Set Patterns in East African Cooking Bananas are Asymmetric in Bunches and Fruits

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Seed set patterns in East African Cooking Bananas are asymmetric in bunches and fruits

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15 **Abstract**

16 **Background:** Low female fertility in bananas is the biggest hurdle for banana breeding. The aim
17 of this study was to determine seed set patterns in East African cooking bananas EACBs to
18 inform future decisions on a more targeted approach of increasing seed set and subsequently
19 banana breeding efficiency. Matooke (AAA) and Mchare (AA) bananas are genetically distinct

20 but belong to the same genetic complex, they referred to as EACBs. Seed set patterns in
21 ‘Enzirabahima’ (AAA), ‘Mshale’ (AA) and ‘Nshonowa’ (AA) all with residual fertility were
22 examined after hand pollination with a highly male fertile wild banana ‘Calcutta 4’ (AA).

23 **Results:** Seed set in ‘Enzirabahima’ is predominant in distal hands. Mchare cultivars have a
24 slightly more even distribution of seeds in their hands compared to ‘Enzirabahima.’ There is a
25 gradual increase in seed set from proximal to distal hands with a slight drop in the last hand. This
26 pattern is more definite in ‘Enzirabahima’ and ‘Mshale’ while ‘Nshonowa’ has a somewhat
27 inconsistent pattern. There is also a drop in seed set per 100 fruits per hand from small to larger
28 bunches. However, larger bunches have a higher pollination success compared to smaller
29 bunches. They therefor set more seed on 100 fruits per hand and per bunch basis if bunches
30 without seed are accounted for. Pollination success rate increases from smaller to larger bunches
31 of EACBs. Seed set is biased toward the distal third part of fruits of examined EACBs as well
32 tetraploid Matooke hybrid ‘401K-1’ (AAAA) and improved diploid ‘Zebrina’ GF (AA) that were
33 used for comparison. In comparison, in the highly female fertile ‘Calcutta 4,’ seed set is along
34 the entire length of the fruit.

35 **Conclusion:** Seed set bias in the distal hands and distal end of fruits suggests a systematic
36 mechanism rather than a random occurrence. It is expected that this information will provide a
37 foundation for increased crossbreeding efficiency in bananas.

38 **Keywords: female fertility, banana breeding, Matooke and Mchare pollination, bunch size,**
39 **pollination success**

40

41 **Background**

42 Bananas (*Musa* ssp.) including plantains are the world's most popular fruit crop grown in about
43 137 tropical and subtropical countries on an estimated 5 million hectares of land (FAOSTAT,
44 2018) Though grown in the tropics and subtropics, their origin is traced back to Southeast Asian
45 and Western Pacific regions where their wild relatives exist in natural forests (Robinson and
46 Saúco, 2010). Bananas are also the only fruit crop that is a staple food for many farming
47 communities in the tropics. Overall banana production (excluding plantains) was estimated at
48 125 million tons in 2017 (FAOSTAT, 2018) with only 14% exported (FAO, 2018) and the rest
49 was locally consumed. Latin America and the Caribbean accounted for 86% of all exported
50 bananas, Africa accounted for 4% and the rest came from Asia (FAO, 2018). The East African
51 region produced close to 10 million tonnes of bananas in 2017 (FAOSTAT, 2018) with
52 negligible export. The dominant banana type in the Great Lakes region of East Africa is the East
53 African Highland Bananas (EAHBs) commonly known as Matooke (AAA) (Karamura et al.,
54 2012). The Mchare subgroup (AA) which is genetically distinct from Matooke is common in
55 Tanzania (Němečková et al., 2018); it is classified as a dessert banana by FAO. But Matooke and
56 Mchare can be collectively referred to as East African Cooking Bananas (EACBs).

57 From the 1970s, banana production in East Africa experienced a drop attributed largely to a
58 complex of diseases (fungal, bacterial, and viral), nematodes, and insect pests (Viljoen et al.,
59 2017). Low production and productivity is also attributed to poor agronomic practices, abiotic
60 stresses including poor edaphic factors and an imminent threat of climate change (Pillay and
61 Tripathi, 2007; Ortiz and Swennen, 2014). Most banana breeding programs therefore focus on
62 these production constraints when defining their breeding goals. The major stumbling block to

63 crossbreeding in bananas is low seed set (Pillay and Tripathi, 2007). Ssebuliba *et al.* (2005)
64 reported an average of 303 ovules per fruit in EAHBs, yet the seed set range was only 0-25 per
65 bunch. This implies that an average sized EAHB bunch with 100 fruits could potentially yield
66 over 30,000 seeds. A total of 78 EAHBs were screen in Uganda of which 37 were considered to
67 be seed fertile (Ssebuliba *et al.*, 2005). On the other hand, Mchares have not been well
68 characterized, especially for female fertility. The reasons for low seed set are a complex array of
69 factors that have hindered efficient banana breeding. For the triploid bananas including the
70 EAHBs, female sterility as a result of meiotic failures has been a major obstacle (Pillay and
71 Tripathi, 2007). Some cultivars including those of the EAHBs are rendered “infertile” (Ssebuliba
72 *et al.*, 2006a) thus; improvement of such cultivars by conventional means is difficult.

73 Nevertheless, female sterility has also been observed among diploid bananas (Ray, 2002),
74 suggesting factors beyond meiotic failures contribute to sterility. For example, successful
75 pollination can be achieved when flowers of ‘Gros Michel’ bananas are pollinated between 7:00
76 am and 10:30 am (Shepherd, 1960b). To unveil the mystery of overcoming sterility in *Musa*,
77 responsible factors have to be dealt with comprehensively to have a better perspective. In most
78 analyses in *Musa*, seed set is considered on a bunch basis. However, bunch size for the same
79 genotype varies depending on environmental and soil fertility factors (Fortescue and Turner,
80 2011). Intriguingly, analyses by Shepherd (1954) strongly suggested a relationship between
81 bunch size and seed set with larger bunches setting more seed on a seed set per 100 fruits basis.
82 Besides, significant differences in seed set for hand position have been observed in EAHBs and
83 this was linked to stigma receptivity (Ssebuliba *et al.*, 2006b). Distal hands were found to have
84 more receptive stigmas than proximal hands thus the higher fertility in distal hands. In plantain,
85 seed set also depends on hand position (Swennen and Vuylsteke, 1989) and there is a general

86 tendency of seed set in the middle hands for both plantain and EAHBs. In addition, seed set in
87 ‘Gros Michel’ was found to be predominant in the distal end of the fruit rather than the proximal
88 end (Shepherd, 1954).

89 These observations have not been made on individual bunch size basis especially in EAHBs and
90 there is a need to look into the behavior of seed set position in hands of different bunch sizes.
91 Correlation coefficients between seed set and weather attributes are mostly less than 0.5
92 (Ssebuliba et al., 2009). This suggests that there are more factors that are yet to be identified and
93 considered. Batte et al. (2019) also found no month effects for pollination success of EAHBs
94 after 21 years of crossbreeding. This raises questions about the underlying causes of observations
95 made and how understanding these factors can ultimately lead to overcoming the fertility crisis
96 in *Musa*. The aim of this study was to investigate seed set patterns in hands of the bunch, number
97 of fruits per hand in different bunch sizes, seed set position in the fruits as well as pollination
98 success in relation to bunch size of EACBs. Seed set patterns were also compared for different
99 pollination techniques used. The data generated will inform future decisions on a more targeted
100 approach of increasing seed set in edible bananas and facilitating banana breeding.

101 **Materials and methods**

102 **Banana genotypes used and study site**

103 *Musa* (AAA group, Matooke subgroup) ‘Enzirabahima’ and *Musa* (AA group, Mchare
104 subgroup) ‘Mshale’ and ‘Nshonowa’ with residual female fertility were used as female parents.
105 *Musa* (AAA group, Matooke subgroup) ‘Nakitembe’ and *Musa* (AA group, Mchare subgroup)
106 ‘Mlelembo’ were also pollinated along with the first three cultivars. Only one bunch of
107 ‘Nakitembe’ set one seed while ‘Mlelembo’ set two seeds in the same bunch, over 200 bunches

108 of each of these two cultivars were made. Because of the extreme low fertility, ‘Nakitembe’ and
109 ‘Mlelembo’ were not considered for seed set patterns in EACBs. Matooke and Mchare banana
110 groups are referred to as EACBs with reference to their utilization. A highly pollen fertile *Musa*
111 *acuminata* ssp. *burmannicoides* (‘Calcutta 4’) wild banana was used as a male parent.

112 Each female parent was planted in a pollination block with 9 mats within columns and 22 mats
113 within rows to yield a total of 198 mats. A spacing of 3 m between rows and 2 m between mats
114 in columns was used which gave a density of 1,666 mats/ha. The male parent ‘Calcutta 4’ was
115 planted in columns separating female parents. Improved diploid ‘Zebrina GF’ (AA) and
116 tetraploid Matooke hybrid ‘401K-1’ (AAAA) from other pollination blocks were used for
117 comparison with EACBs seed set patterns in fruits. The study was conducted at the National
118 Agricultural Research Laboratories (NARL), Kawanda. NARL is located at latitude 0° 25’ N,
119 longitude 32° 32’ E at an elevation of 1,177 m above sea level.

120 **Pollination techniques and procedure**

121 Different pollination techniques were used; these include, hand pollination as described by
122 Vuylsteke *et al.* (1997) (technique 1 or control). Technique 1 was modified by applying pollen
123 germination media (PGM) on stigmas after dusting pollen (technique 2). The third technique
124 involved early pollination by forcing bracts open about a day before opening with PGM
125 application on stigmas after dusting pollen. The third was explored between June 2016 and
126 February 2018. The fourth and last technique was evening pollinations after normal bract
127 opening with PGM application on stigmas. This was also explored between June 2016 and June
128 2017. These pollination techniques were performed on different bunches as they emerged. They
129 were geared towards increasing seed set but the focus here is seed set patterns.



Fig 1. Procedure of early pollination (technique 3) on *Musa* (AA group subgroup Mchare) 'Nshonowa:' (A) Flower bract forced open and tepals being removed; (B) Tepals removed to expose stigmas for pollination and pollen applied; (C) PGM solution applied with hand sprayer (D) Flower bract returned in position (E) Inflorescence re-bagged and labeled. This process from A to E is repeated for pollination of next hand when it is ready. (F) Pollinated bunch left to mature in the open

130 Pollen germination media was applied with the aim of enhancing stigma receptivity for
131 maximum pollen germination as demonstrated in our earlier work (Waniale et al., 2020). Using
132 tap water, PGM was prepared using 30 g/L glucose as a substitute for sucrose along with other
133 compounds as described by Nyine and Pillay (2007) and boric acid at 0.1 g/L. The PGM used for
134 pollinations in 2016 was 30g/liter glucose and complete PGM in the rest of the pollinations. The
135 PGM was applied to pollen dusted on stigmas in a fine mist using a hand spray pump before re-
136 bagging (Figure 1). The three cultivars had varying days to full maturity, ‘Enzirabahima’
137 matured in 98 days, ‘Mshale’ in 131 days and ‘Nshonowa’ in 135 days. On the other hand,
138 ‘Nakitembe’ and ‘Mlelembo’ matured in 96 and 130 days respectively.

139 **Data handling**

140 At full maturity when at least one finger started ripening, bunches were harvested, kept in a
141 ripening room and seeds were hand extracted, washed, air-dried and counted. Total number of
142 seeds per hand was recorded for each cultivar and each bunch. The number of fruits per hand
143 was also recorded. For each cultivar, bunch sizes were categorized based on number of hands per
144 bunch and percentage seed set was calculated for different hands and different bunch sizes as
145 follows;

$$146 \frac{\text{Total number of seeds per hand position for a given bunch size category}}{\text{Total number of seeds for a given bunch size category}} \times 100\%$$

147 Pollination success rate was calculated as follows;

$$148 \frac{\text{Total number of bunches with seed for a given bunch size category}}{\text{Total number of pollinated bunches for a given bunch size category}} \times 100\%$$

149 Analysis of variance (ANOVA) on number of fruits per hand for different bunch sizes of the
150 three cultivars was done using GenStat, 14th Edition (Payne et al., 2011). Fruits per hand were
151 taken as treatment with total bunches as replicates. The number of hands in a bunch category was
152 therefore the total number of treatments and different categories were analyzed separately. The
153 ANOVA model used was as follows;

154 Fruits per hand = Mean + Treatment + Error

155 Seed set per bunch was adjusted to seed set per 100 fruits of each bunch for equal footing
156 comparison of bunches with different total fruits and hands. The adjustment was made as
157 follows;

$$158 \frac{\text{Total seed in a bunch}}{\text{Total number of fruits of that bunch}} \times 100$$

159 To compare seed set of different hand positions on equal footing, average seed set per hand was
160 adjusted to seed set per 100 fruits of that hand. Only bunches with seed were considered in the
161 calculation. The adjustment was made for the three cultivars and all bunch sizes as follows;

$$162 \frac{\text{Average seed of hand position}}{\text{Average number of fruits of that hand position}} \times 100$$

163 Where average seed of hand position was calculated as;

$$164 \frac{\text{Total seed of hand position}}{\text{Number of bunch with seed}}$$

165 Seed set per 100 fruits per hand was plotted against hand position for different bunch size
166 categories of the same cultivar on the same plot.

167 Chi-square goodness of fit (X^2_{cal}) for seed set in hand positions was calculated as follows;

$$168 \sum \frac{(\text{Observed seed per hand} - \text{Expected seed per hand})^2}{\text{Expected seed per hand}}$$

169 Where;

170 Observed seed per hand is the number of seed in given hand position for all fertile bunches.

171 Expected seed per hand of a particular bunch size category and cultivar was calculated as;

$$172 \frac{\text{Average fruits per hand for that hand position}}{\text{Average fruits per bunch}} \times \text{Total seed}$$

173 Degrees of freedom for the Chi-square test are the number of hands in bunch size category minus
174 one. Chi probability (X^2_{prob}) was taken as the right tailed Chi probability.

175 The position of seed set in the fruit pulp was observed by carefully splitting open pollinated ripe
176 fruits of ‘Mshale,’ ‘Nshonowa’ and ‘Enzirabahima’ in comparison to the tetraploid Matooke
177 hybrid ‘401K-1’ (AAAA). Seed set positions of pollinated partially parthenocarpic improved
178 diploid ‘Zebrina GF’ (AA) and in the non-parthenocarpic open pollinated wild banana ‘Calcutta
179 4’ (AA) were also observed for comparison with EACBs. Selected pictures of seed set position
180 in fruits are presented.

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185 **Results**

186 **Percentage seed set per hand and fruits per hand**

187 Irrespective of the pollination technique used, seed set pattern was similar within bunch size
 188 categories among the three female fertile EACBs. Data of different pollination techniques and
 189 same bunch size categories were therefore merged for the respective cultivars (Supplementary
 190 Tables 1, 2, and 3). For ‘Enzirabahima,’ maximal seed set was observed in the second last hand
 191 of the 4, 5, and 7 hand bunches (Table 1).

192 Table 1. Percentage seed set per hand and by number of hands per bunch in EACB

Cultivar	Bunches with seed	Bunch size (hands)	Total seed	Seed set per hand (%)										
				1	2	3	4	5	6	7	8	9	10	
‘Enzirabahima’ (AAA)	6	4	27	3.7	22.2	44.4	29.6							
	23	5	89	0.0	4.5	28.1	34.8	32.6						
	43	6	243	1.6	10.7	15.2	30.5	22.6	19.3					
	34	7	210	0.0	5.2	15.2	14.8	25.7	27.1	11.9				
	7	8	28	0.0	10.7	10.7	32.1	7.1	21.4	10.7	7.1			
‘Mshale’ (AA)	2	4	42	11.9	38.1	31.0	19.0							
	31	5	610	5.7	17.2	24.1	24.6	28.4						
	47	6	961	5.5	8.3	19.7	22.8	24.2	19.5					
	29	7	1,193	10.9	18.3	17.5	17.5	15.8	10.7	9.3				
	4	8	67	0.0	7.5	13.4	9.0	28.4	26.9	10.4	4.5			
1	10	66	6.1	43.9	19.7	12.1	12.1	6.1	0.0	0.0	0.0	0.0	0.0	
‘Nshonowa’ (AA)	1	4	2	0.0	0.0	0.0	100.0							
	10	5	39	0.0	15.4	25.6	20.5	38.5						
	11	6	90	4.4	22.2	22.2	26.7	17.8	6.7					
	17	7	234	15.8	16.2	11.1	12.8	16.2	10.3	17.5				
	14	8	237	3.4	3.4	8.4	10.1	15.6	16.9	35.4	6.8			
	3	9	36	0.0	27.8	0.0	0.0	2.8	25.0	8.3	2.8	33.3		
	3	10	29	3.4	10.3	0.0	6.9	17.2	17.2	24.1	0.0	3.4	17.2	

193 The 6 and 8-hand bunches had maximal seed set in the third last and fourth hand respectively.
194 There was generally less seed set in the first two hands of ‘Enzirabahima’ with no seed in the
195 first hands of 5, 7, and 8 hand bunches. There were no embryos in seeds from the ‘Nakitembe’
196 and ‘Mlelembo’ bunches the set one and two seeds per bunch respectively. There was seed set
197 increase in ‘Enzirabahima’ after pollination with PGM. But the increase happened in the same
198 fertile hand positions as of bunches pollinated without PGM (Supplementary Table 1).

199 In ‘Mshale,’ and ‘Nshonowa,’ on a percentage seed set basis, there was generally a more even
200 seed set among middle hands with the proximal and distal hands having fewer seeds. Mchare
201 cultivars had 7 hand bunches with relatively even seed set across all hands. Unlike
202 ‘Enzirabahima,’ 5 hand bunches of Mchare were observed to set maximal number of seeds in last
203 hand with female fruits. Similar results were obtained by Swennen and Vuylsteke (1989) for
204 seed set in plantain (*Musa* AAB group), especially in cultivar ‘Obino I’Ewai.’ Smaller bunches
205 had the highest seed set in the middle hands whereas larger bunches had a slight shift of maximal
206 seed set to the distal hands (Table 1). Though fewer Mchare bunches were pollinated in
207 comparison with ‘Enzirabahima,’ they had relatively higher pollination success (Supplementary
208 Tables 1, 2, and 3). Generally, there was an increase in average seed set rates per 100 fruits as
209 well as success rate from small to larger bunches in the three EACB cultivars (Supplementary
210 Tables 1, 2, and 3).

211 The Mchare cultivars had a higher seed set range of 0 to 261 per bunch compared to the range in
212 ‘Enzirabahima’ of 0 to 33 (Supplementary Table 1, 2, and 3). ‘Mshale’ had a higher seed set
213 range of 0 to 261 per bunch than that of ‘Nshonowa’ with 0 to 85 seeds. The highest seed set per
214 bunch in ‘Mshale’ was from a 7 hand bunch, from an 8 hand bunch in ‘Nshonowa’ while for

215 'Enzirabahima,' a 6 hand bunch had the highest. Strangely, there were hyper increases of seed
216 set in a few bunches which could not be replicated in same size bunches of the same cultivars
217 pollinated about the same time. This prompted an investigation of the features around the mats
218 where such bunches grew. Interestingly, such mats were on pockets of poor soil with some
219 pebbles.

220 There were significant differences ($P < 0.001$) for number of fruits per hand among
221 'Enzirabahima,' 'Mshale,' and 'Nshonowa' bunches of different sizes (Table 2). Number of
222 fruits in the last hand with female fruits of all different sized bunches evaluated were
223 significantly lower than in other hands. The exception was 4-hand category of 'Nshonowa'
224 which was non-significant even if there was a difference of 5 fruits between the proximal and
225 distal hands. The number of fruits per hand, however, seemed not to account for highest seed set
226 in the distal hands as there was a gradual decrease of fruits per hand from proximal to distal
227 hands. With these differences in fruits per hand position, a better approach of comparing seed set
228 between hands was a prerequisite.

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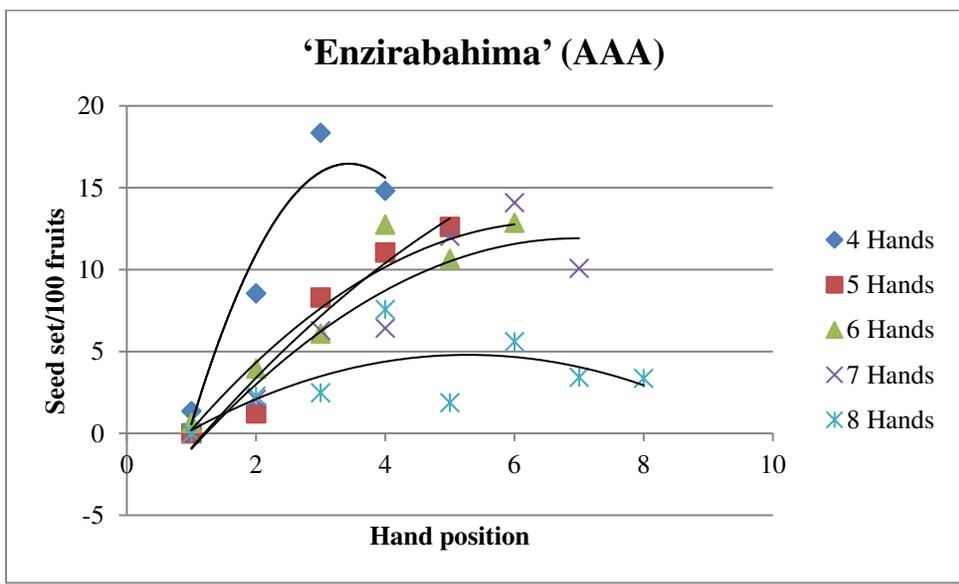
235 Table 2. ANOVA for fruits per hand for different bunch sizes of ‘Enzirabahima,’ ‘Mshale,’ and
 236 ‘Nshonowa’

Cultivar	Bunch size (hands)	Number of bunches	Fruits per hand								F-pro	
			1	2	3	4	5	6	7	8		
‘Enzirabahima’ (AAA)	4	39	12.3a	11.7a	10.9a	9.0b						<0.001
	5	107	14.3a	14.2a	13.1b	12.2c	10.0d					<0.001
	6	103	14.6ab	15.3a	14.1bc	13.5c	12.0d	8.5e				<0.001
	7	52	14.7ab	15.6a	15.0ab	14.2bc	13.2c	11.9d	7.3e			<0.001
	8	4	18.8a	18.8a	17.3ab	17.0ab	15.3b	15.3b	12.5c	8.5d		<0.001
‘Mshale’ (AA)	4	5	15.0a	14.2ab	13.0b	11.2c						<0.001
	5	63	16.1a	15.9a	14.9b	13.4c	11.6d					<0.001
	6	46	15.4a	15.2a	14.2ab	13.4bc	12.4c	11.1d				<0.001
	7	15	14.2a	14.4a	13.7ab	12.9ab	12.3ab	11.3bc	9.7c			0.004
	8	3	17.7a	17.7a	16.3ab	14.3bc	14.7abc	12.7cd	11.7cd	10.7d		0.002
‘Nshonowa’ (AA)	4	9	14.0a	12.7a	11.1a	9.3a						0.143
	5	52	16.8a	16.2a	14.7b	13.4c	11.7d					<0.001
	6	35	17.1a	16.7ab	15.4bc	14.3cd	13.4d	11.1e				<0.001
	7	20	14.7a	14.7a	13.4ab	12.8bc	12.0bc	11.6cd	9.8d			<0.001
	8	12	15.9a	15.5a	14.8ab	14.3abc	13.6abc	12.5bc	11.5cd	9.3d		<0.001

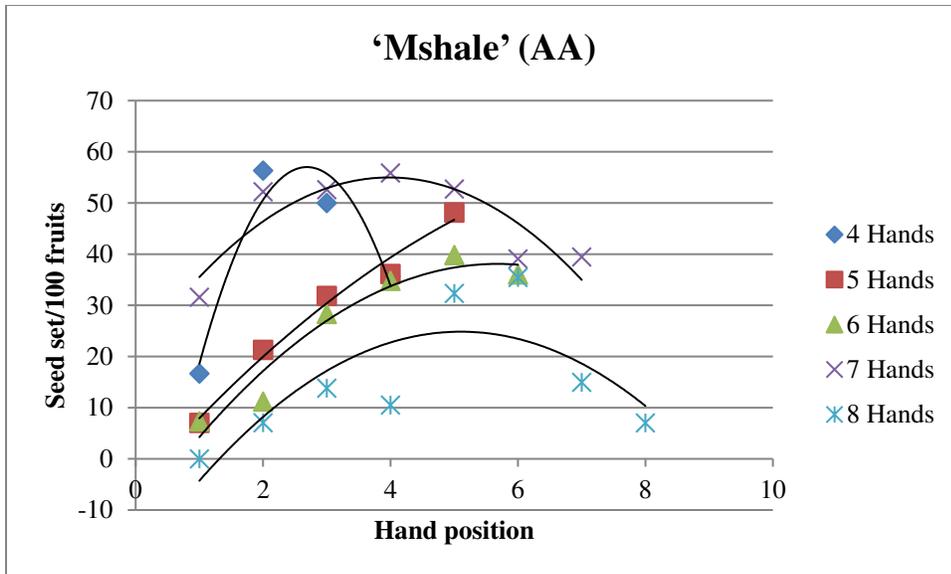
237 **Average seed set per 100 fruits per hand**

238 In ‘Enzirabahima,’ seed set per 100 fruits per hand followed a hyperbolic pattern for all bunch
 239 sizes from proximal to distal hands (Figure 2). For all bunch sizes, proximal hands had less seed
 240 set which gradually increased in hands that followed and slightly dropped in last hand with
 241 female fruits. Small sized bunches had higher seed set per 100 fruits per hands which gradually
 242 reduced with increased bunch size. ‘Mshale’ had a similar pattern as ‘Enzirabahima’ with the

243 exception of 7 hand bunch category which had the most seed in proximal hand (Figure 3). The
 244 ‘Mshale’ curve for 7 hand bunch category had highest averages partly because of a single bunch
 245 that had 261 seeds. For both ‘Mshale’ and ‘Enzirabahima,’ the 4 hand bunches experienced the
 246 biggest drop in seed set in the last hand. ‘Nshonowa’ did not have a consistent pattern as 5, 7,
 247 and 9 hand bunch categories had positive “a” coefficients of the quadratic equation (Figure 4 and
 248 Table 3). With the exception of 5 hand bunch category of ‘Nshonowa,’ bunches with 6 or less
 249 hands had high R² values (Table 3). These observations were as a result of few bunches used to
 250 calculate means especially for the 9 and 10 hand bunch categories which had three bunches with
 251 seed each. The 9 and 10 hands categories were therefore excluded from the plot (Figure 3). A
 252 chi-square test for goodness of fit revealed that observed number of seed was significantly
 253 different from expected number except in ‘Mshale’ of 4 hands per bunch category
 254 (Supplementary Table 4).

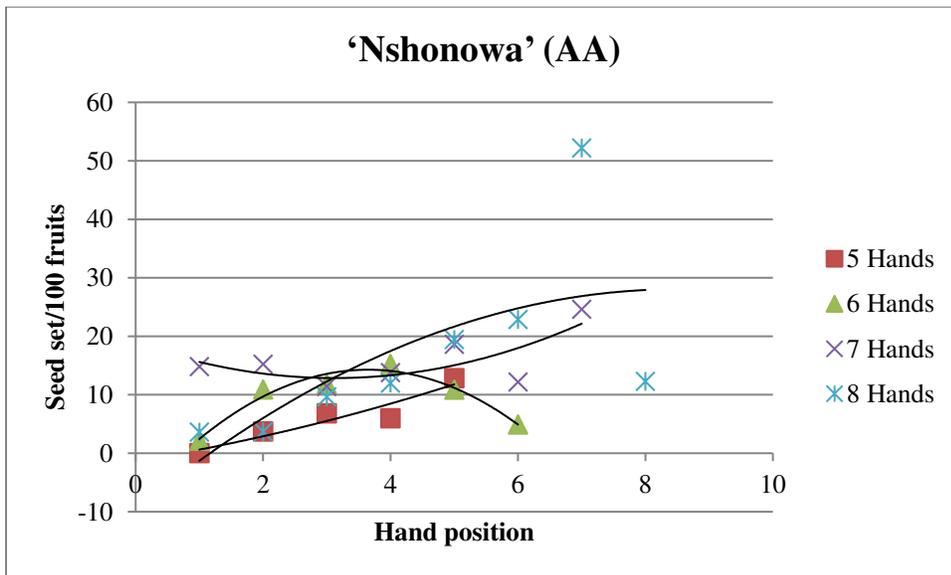


255
 256 Fig 2. Seed set per 100 fruit per hand in different sized bunches of ‘Enzirabahima’



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258 Fig 3. Seed set per 100 fruit per hand in different sized bunches of 'Mshale'



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260 Fig 4. Seed set per 100 fruit per hand in different sized bunches of 'Nshonowa'

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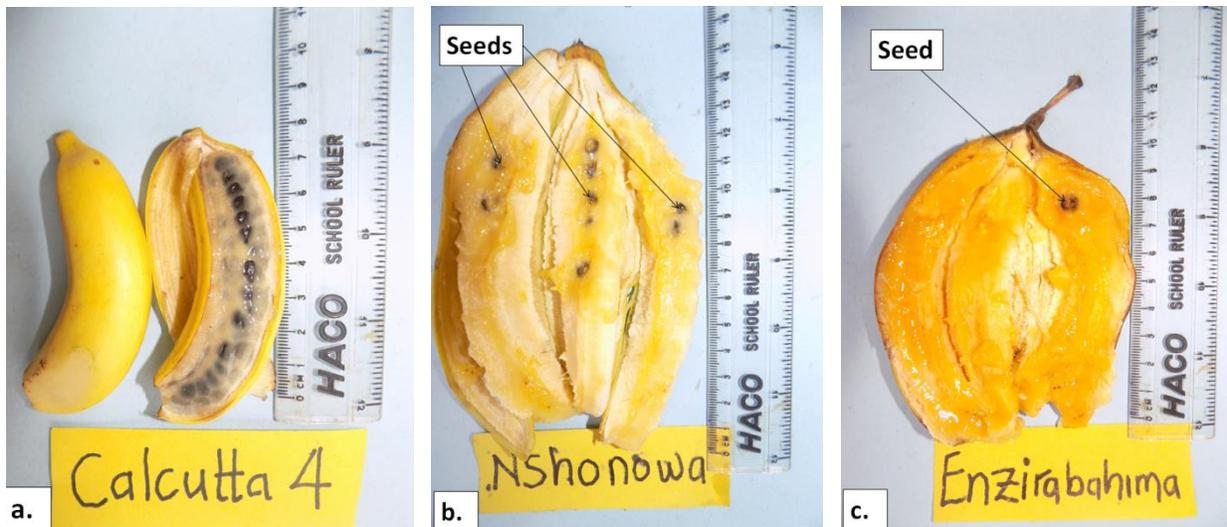
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263 Table 3. Second order polynomial and R² values fitted for seed set per 100 fruits per hand

Cultivar	Bunch size (Hands)	Bunch number	a	b	C	R ²
'Enzirabahima' (AAA)	4	6	-2.68	18.43	-15.19	0.924
	5	23	-0.26	5.07	-5.70	0.943
	6	43	-0.40	5.33	-4.72	0.915
	7	34	-0.36	5.02	-5.61	0.880
	8	7	-0.25	2.65	-2.21	0.447
'Mshale' (AA)	4	2	-13.49	72.53	-40.47	0.922
	5	31	-0.77	14.34	-5.64	0.982
	6	47	-1.55	17.56	-11.77	0.939
	7	29	-2.19	17.46	20.25	0.773
	8	4	-1.73	17.66	-20.21	0.629
'Nshonowa' (AA)	5	10	0.17	1.78	-1.33	0.886
	6	11	-1.69	12.34	-8.22	0.948
	7	17	0.62	-3.84	18.84	0.529
	8	14	-0.52	8.86	-9.63	0.447
	9	3	0.94	-7.06	15.84	0.384
	10	3	-0.26	3.55	-2.31	0.193

264 **Seed set position in fruits**

265 For fruits which set seed among female fertile EACBs predominantly set seed in the distal one
 266 third of the fruit, especially when there were no more than 3 seeds (Figure 5c). The same pattern
 267 was observed in '401K-1' (AAAA) and 'Zebrina GF' (AA) that are used in breeding. In the
 268 event of multiple seed set in a fruit (more than 3), seeds were distributed in the distal half of the
 269 fruit (Figure 5b). This was observed mainly in Mchare. On the contrary, open pollinated wild
 270 banana 'Calcutta 4' set seed over the entire length of the fruit (Figure 5a).



271 Fig 5. Seed set position in fruits as revealed after ripening among selected banana genotypes. a)
 272 Small intact and peeled fruits of non-parthenocarpic open pollinated wild banana ‘Calcutta 4’
 273 with numerous evenly distributed seed. b) Multiple seed in the distal end of the fruit in the
 274 cultivar ‘Nshonowa’ pollinated with ‘Calcutta 4’. c) Single seed set in the distal end of
 275 ‘Enzirabahima’ pollinated with ‘Calcutta 4’.

276 Discussion

277 Getting an in depth understanding of seed set patterns in EACBs is certainly one of the measures
 278 that will help in designing experiments for increasing seed set in *Musa* spp. The end goal is to
 279 have a fast and an efficient banana breeding pipeline with better hybrids for many farmers in the
 280 tropics and subtropical regions globally. Here, there is a deliberate effort to understand the cause
 281 of sterility by looking at seed set patterns in relation of position of the hands, bunch size and
 282 position in the fruit in EACBs. In *Musa* spp. sterility has been understood to arise from
 283 developmental errors in the sporophyte, the gametophyte as well as pistil-pollen interactions. But
 284 more emphasis has been placed on understanding the gametophyte which is said to account most
 285 towards sterility (Fortescue and Turner, 2011). Meiotic errors as a result of chromosome

286 mismatch were reported by Simmonds (1962) to cause embryo sac failure in sterile *Musa* spp.
287 Fortescue and Turner (2005), however, found that about 10% of embryo sacs in ovules of
288 triploid bananas were correctly positioned compared to 75% in *Musa acuminata* ssp. They also
289 discovered that the presence of a B-genome increased the presence of an embryo sac to 96-
290 100%. But these observations do not explain the fact that seed set in EACBs is biased toward
291 distal hands and distal end of the fruit. There is also no documentation of a link between the
292 presence and correct positioning of an embryo sac to seasonal influence of seed set in *Musa* spp.
293 with residual fertility.

294 The observed bias of seed set in distal hands especially in Matooke has been attributed to the
295 high stigma receptivity in distal hands (Ssebuliba et al., 2006b). But if this were entirely true,
296 then the use of PGM on stigmas would have increased seed set in the proximal hands as well
297 since all hands are pollinated. And since ‘Enzirabahima’ was reported to have fairly high stigma
298 receptivity in all hands, seed set should have been distributed in all hands. Instead, seed set
299 increase was observed in the same hand positions as of bunches pollinated without PGM. Use of
300 PGM should have also resulted in seed set in ‘Nakitembe’ and ‘Mlelembo’ if stigma receptivity
301 was a prime contributor of sterility in *Musa*. This implies that there are other factors which come
302 into play after pollen has germinated on the stigmas in different hand positions. The necrosis
303 formed in the prolongation zone of fruits after anthesis especially in triploids (Soares et al.,
304 2014) could partly account for absence of seed set in proximal hands. This necrosis acts as a
305 barrier to pollen tube growth. Shepherd (1960b) observed that pollen tube growth through the
306 stigma in ‘Gros Michel’ looked normal, but was slowed or arrested and the tips developed a
307 swelling resulting in pollen tubes not reaching the ovules. It may be worthwhile to investigate
308 rates of necrosis formation in fruits of different hand positions to find a link to seed set if any.

309 In an attempt to avoid the necrosis formed soon after anthesis in the fruit prolongation zone,
310 early pollinations were made (technique 3) but the technique did not increase seed set. Shepherd
311 (1960a) speculated that physiologically immature stigmas may delay or prevent penetration by
312 pollen tubes and this could have been the case with early pollination. Results in the present study
313 suggest that application of PGM on stigmas enhanced pollen germination (Waniale et al., 2020).
314 But this was most effective on stigmas presumed to be physiologically mature after natural
315 flower opening as suggested by Shepherd (1960a). This observation may also suggest that arrest
316 of pollen tube growth may be a biochemical rather than a physical process. It is supported by the
317 fact that a necrosis is not formed in the prolongation fruit zone of diploids (Soares, et al. 2014).
318 Pollination technique 3 was therefore discontinued in December 2016 since seed set was lower
319 compared with the customary pollination technique described by Vuylsteke et al. (1997). The
320 other plausible cause of low seed set with technique 3 could have been that the PGM did not
321 have all components for *in vitro* pollen germination as 30g/L glucose was used. Evening
322 pollinations were also discontinued in 2016 since they did not increase seed set. This resulted
323 from reduced pollen viability with time of the day coupled with low humidity at the time of
324 pollination between 5 and 6:30 pm. The rationale of evening pollination was to have flowers
325 pollinated soon after opening as *Musa* flowers start opening from evening through the night.
326 However, not all female flowers had fully opened at the time of evening pollination and fresh
327 pollen could not be obtained. Fairly reliable comparisons were made between the control and
328 technique 2 since a considerable number of bunches were fairly distributed in different months of
329 year during the study period (Supplementary Tables 5, 6, and 7).

330 Ssebuliba et al. (2006b) found that bracts of distal female hands opened to a bigger angle than
331 that of proximal hands. This phenomenon was linked to a response of stigma receptivity which

332 increased from proximal to distal hands. But it might be that ovules in proximal hands and the
333 proximal fruit end have higher abortion rates if pollen tubes reach them. Abortive ovules have
334 been reported in other crops like hazelnut whereby some unigenes are upregulated and others
335 down regulated in abortive ovules compared to developing ovules (Cheng et al., 2015). The
336 gradual increase of seed set from proximal to distal hands could suggest a mechanism of ovule
337 abortion rate in the same order. The most likely cause of this observation could be auxins in large
338 amounts from the root tips that reach proximal hands first. Auxins induce formation of edible
339 pulp in both seeded and non-seed bananas as well as partially parthenocarpic types (Simmonds,
340 1953; Simmonds, 1960). And in seeded bananas, synthetic auxins 4-CPA and 2,4,5-T have been
341 reported to hinder seed development (Simmonds, 1960). Auxins therefore play a critical role in
342 parthenocarpy as well as sterility.

343 The effect of weather on seed set in *Musa* spp. could be as a result of a drop in auxin levels with
344 heat and/or moisture stress. This comes with high temperature, high solar radiation and low
345 rainfall. These weather conditions correlate with increase in seed set in Matooke (Ssebuliba et
346 al., 2009). This is likely to slow down evapo-transpiration pull as the plant tries to conserve
347 moisture. Consequently, movement of materials including auxins does not reach the distal hands
348 and fruit tips in adequate amounts. A reduced fruit circumference of 'Gros Michel' correlates
349 with increased seed set (Shepherd, 1954) which can reflect reduced parthenocarpy. This theory
350 also tends to fit the observed increase of seed set per 100 fruits with increase in bunch size; small
351 bunches are saturated easily. A reduction in auxin levels during moisture stress could be caused
352 by salicylic acid which is involved in response to both abiotic and biotic stress (Rivas-San
353 Vicente and Plasencia 2011; An and Mou 2011). Salicylic acid and auxin signalling are mutually
354 antagonist (An and Mou 2011) and production of salicylic acid in response to moisture stress or

355 heat stress could be responsible for reduced auxin levels thus reduced parthenocarpy. This may
356 be linked to the spontaneous seed set increase observed from bunches on mats with poor soils
357 which drain easily thus high salicylic acid production.

358 There were inconsistencies in patterns of seed set in 'Nshonowa' as fewer bunches were available
359 compared with the other two cultivars. Using few bunches with seed to calculate averages for
360 'Nshonowa' implied that a single bunch with high seed set affected the overall shape of the
361 curve. It was also noticed that bunches with high seed set had uneven seed distribution among
362 hands. Sudden seed increase was observed in hand position 7 of 8 hand bunch category. It
363 strongly suggests that there are specific weather conditions for maximum seed set as hands were
364 pollinated on different days with unique weather conditions. High morning temperatures are
365 likely to overcome the issue of pollen tube growth arrest. In citrus (Kawano et al., 2016) and in
366 apples (Yoder, et al. 2009), high temperatures were reported to overcome self-incompatibility
367 and this it could be a similar issue in banana. This could have applied to all the three EACBs but
368 use of many bunches evened out this effect in 'Enzirabahima' and 'Mshale.'

369 Results in the current study revealed that the hands with the highest number of fruits do not
370 necessary produce the highest number of seeds. This clearly suggests that there are other factors
371 that are more important for seed set. Ideally, it is expected that different sized bunches have same
372 pollination success and same seed set per 100 fruits. However, these observations suggest that
373 bunch size has an influence on fertility based on differences of seed set per 100 fruits for
374 different bunch sizes. Shepherd (1954) also observed an increase of seed set on 100 fruit basis
375 from small to larger bunches of 'Gros Michel' as observed in EACBs. Bigger bunches are also
376 more fertile than smaller bunches in terms of total seed and pollination success. The theory of

377 auxins influencing fertility in different bunch sizes seems to fit this observation. Smaller bunches
378 would be easily saturated by auxins compared with larger bunches. This leads to the low seed set
379 (per 100 fruit basis) and low pollination success in smaller compared to larger bunches.
380 However, there is higher seed set per 100 fruit per hand in smaller bunches if only bunches with
381 seed are considered. But large bunches generally set more seed per hand as a result of higher
382 pollination success compared to small bunches.

383 All these observations call for a slightly different approach to better understand sterility and use
384 it profitably as it is a prerequisite in the final hybrids. It may be the right time for banana
385 researchers to start looking in the direction of hormonal manipulations for increased seed set. But
386 since segregation data suggests that parthenocarpy and sterility are independent (Simmonds,
387 1962), this would rule out auxins as the sole cause of sterility in *Musa* spp. A recent genome
388 wide association study in *Musa* found parthenocarpy genes to be potentially linked to
389 seedlessness. But the prime candidate gene was the gene orthologous to Histidine Kinase CKI1
390 (Sardos et al., 2016). Cytokinins have been reported to determine the fate of seed development
391 (Jameson and Song, 2016) and they could be responsible for sterility in *Musa* even if there is
392 successful fertilization. Non-parthenocarpic progeny that has sterile plants could therefore point
393 to ovule abortion. This stems from a mutation in the gene orthologous to Histidine Kinase CKI1
394 that was linked to seedlessness by Sardos et al. (2016).

395 It could be possible that the same gene orthologous to Histidine Kinase CKI1 is responsible for
396 production of poor seed and low embryo rescue rates in some parental combinations. Some
397 edible banana genotypes have been rendered 'infertile' and this could be as a result of hyper
398 production auxin individually or in combination with the gene orthologous to Histidine Kinase

399 CKII which has been linked to seedlessness. It could be that relative contribution these two
400 factors along with other factors can result in high sterility as observed in ‘Nakitembe’ and
401 ‘Mlelembo.’ Other factors such as fruit length are involved in banana fertility as demonstrated in
402 the current study. In ‘Calcutta 4,’ seeds set covers the entire length of the fruit implying pollen
403 tube growth covered an estimated distance of 10 cm. In ‘Nshonowa,’ pollen tube growth covered
404 about 10 cm and about 7 cm of the fruit pulp had no seed. And in ‘Enzirabahima,’ pollen tubes
405 cover a distance of about 5 cm with about 8 cm of pulp without seed. This suggests that pollen
406 tube growth within the fruit is not the reason for a biased seed set towards the fruit tip as the
407 distance covered in ‘Enzirabahima’ is shorter. With the same approach of auxin involvement in
408 *Musa* fertility, it likely that auxins can covers a limited distance during moisture stress thus
409 longer fingers set more seed. This explains the higher seed set observed in Mchare compared
410 with Matooke in the current study. The drop in seed set from the last hand could also be as a
411 result of reduced fruit length in the last hand. Fruits in the proximal hands are longer with bigger
412 circumference which reduces towards the distal end of the bunch. This was reported in plantain
413 and cooking banana types, distal hands have fruits with the least fruit length (Annor et al., 2016).

414 **Conclusions**

415 The results of this study demonstrated that small sized bunches of EACBs have low seed set and
416 pollination success rates. Seed set in EACBs is mainly predominant in the distal hands starting in
417 the mid-section of the bunch. Seed set in fruits is also skewed to the distal end with strong
418 evidence that fruit length and bunch maturity period being involved in fertility. The study also
419 showed that seed set patterns in hand positions are not influenced by stigma receptivity as earlier
420 thought. There is a mechanism that prevents seed set in proximal hands that needs further

421 experimentation to be understood. Auxins are likely to be one of the most important causes of
422 sterility in *Musa* but they are not entirely responsible. A holistic approach will therefore be
423 required to increase seed set and overcome sterility. Efforts have to be devoted to improving *in*
424 *vivo* pollen germination and having agronomic practices that increase bunch size. In future,
425 scientists may have to consider hormonal manipulations especially those that will reduce
426 parthenocarpy. Overcoming sterility will ultimately broaden the parental base to include
427 genotypes whose breeding potential is unknown. A selection criterion of parents has always
428 included male and female fertility trait yet such parents are often non-parthenocarpic. Being able
429 to use parents with parthenocarpy and inherent sterility in crosses will increase the number of
430 parthenocarpic hybrids per cross thus a more efficient breeding pipeline.

431 **Supplementary information**

432

433 **Abbreviations**

434 EACBs: East African cooking bananas; EAHBs: East African Highland bananas; PGM: Pollen
435 germination media; NARL: National Agricultural Research Laboratories; ANOVA: Analysis of
436 variance; 4-CPA: 4-chlorophenoxy acetic acid; 2,4,5-T: (2,4,5-trichlorophenoxy) acetic acid

437 **Declarations**

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444 **Authors' contributions**

445 Conception of research idea and designing experiments: AW, RS, AKT. Performed experiments,
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452 **Availability of data and information**

453 All data generated or analyzed during this study are included in this published article [and its
454 supplementary information files].

455 **Ethics approval and consent to participate**

456 Not applicable.

457 **Consent for publication**

458 Not applicable.

459 **Competing interests**

460 The authors declare that they have no competing interests.

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Figures



Figure 1

Procedure of early pollination (technique 3) on Musa (AA group subgroup Mchare) 'Nshonowa:' (A) Flower bract forced open and tepals being removed; (B) Tepals removed to expose stigmas for pollination and pollen applied; (C) PGM solution applied with hand sprayer (D) Flower bract returned in position (E) Inflorescence re-bagged and labeled. This process from A to E is repeated for pollination of next hand when it is ready. (F) Pollinated bunch left to mature in the open

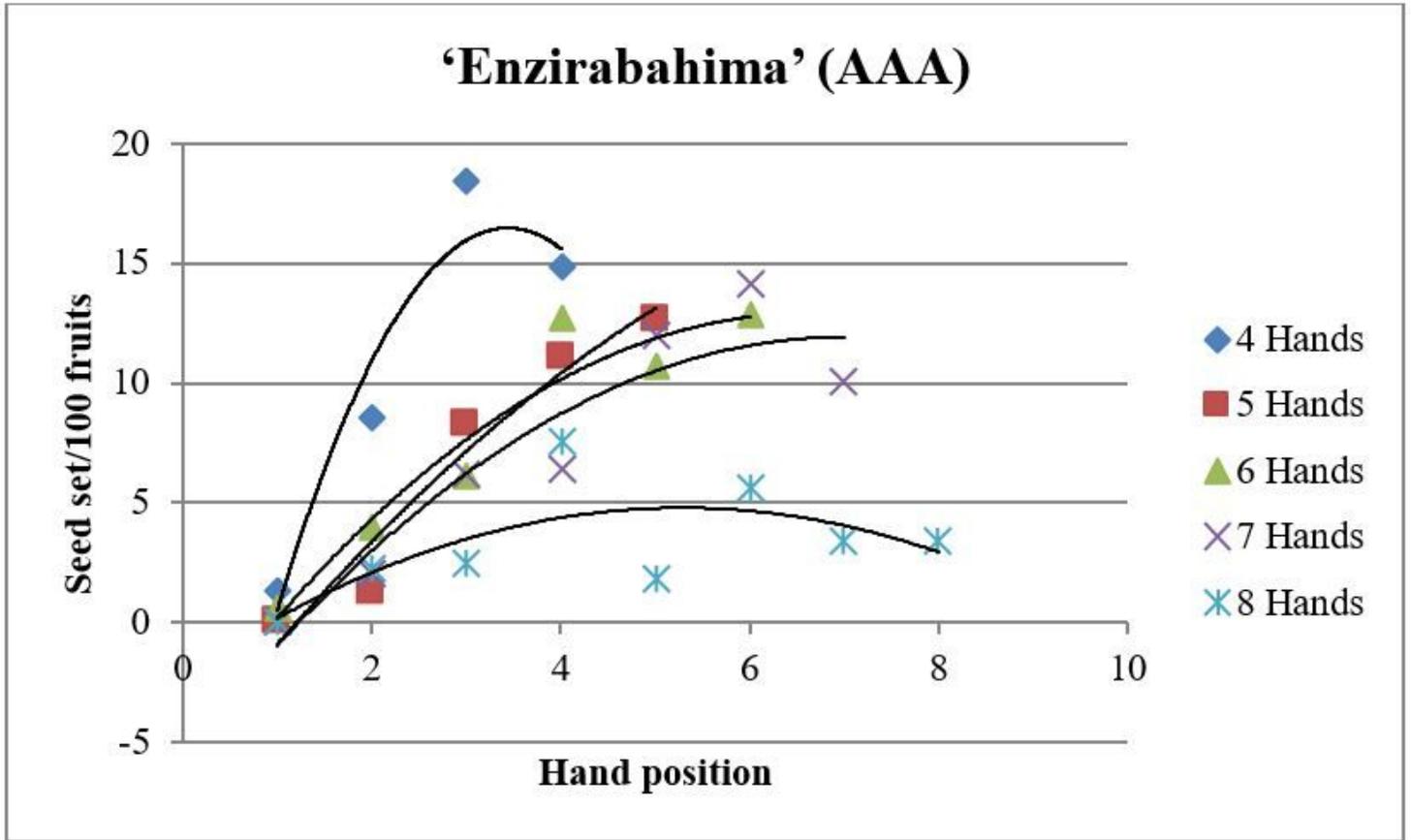


Figure 2

Seed set per 100 fruit per hand in different sized bunches of 'Enzirabahima'

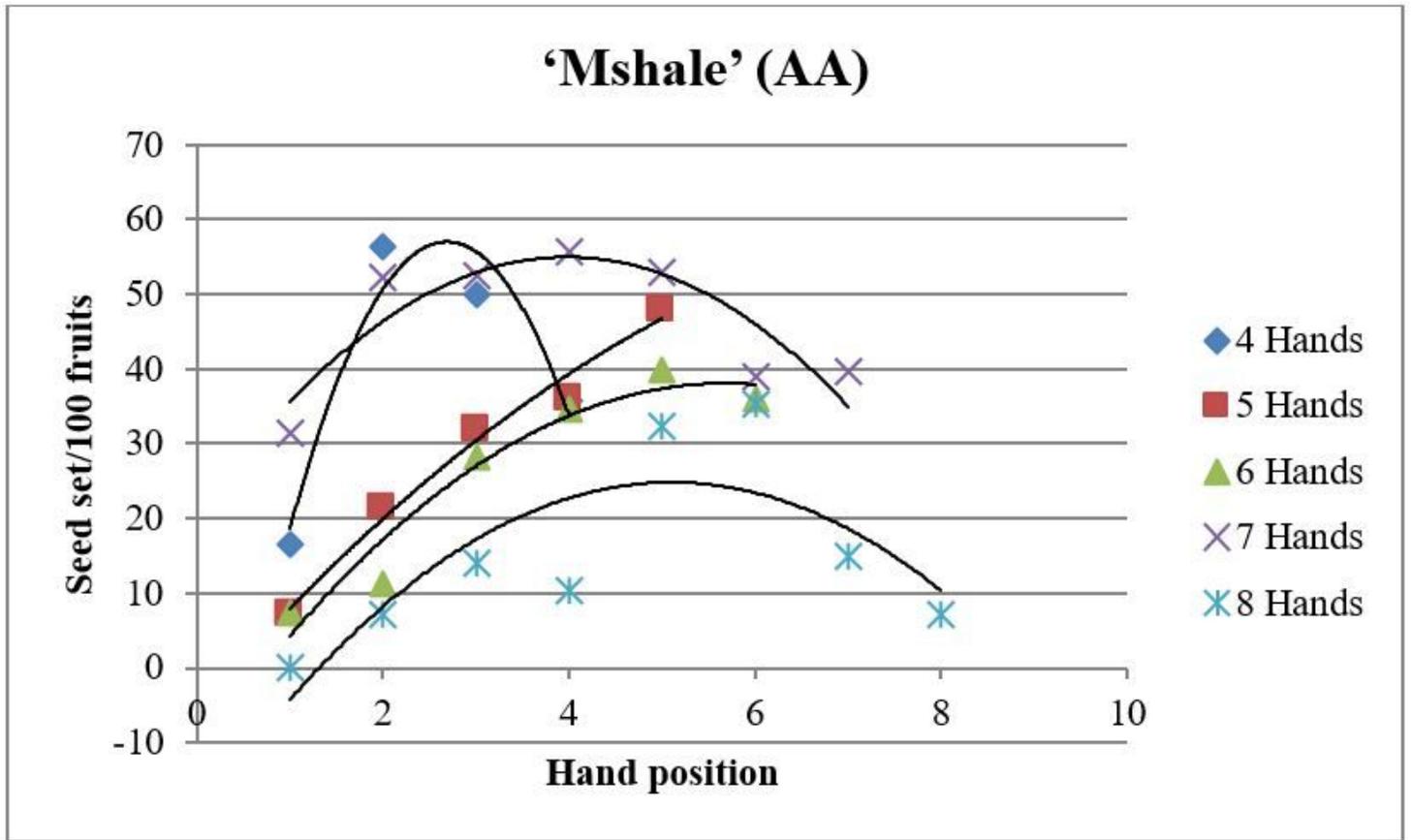


Figure 3

Seed set per 100 fruit per hand in different sized bunches of 'Mshale'

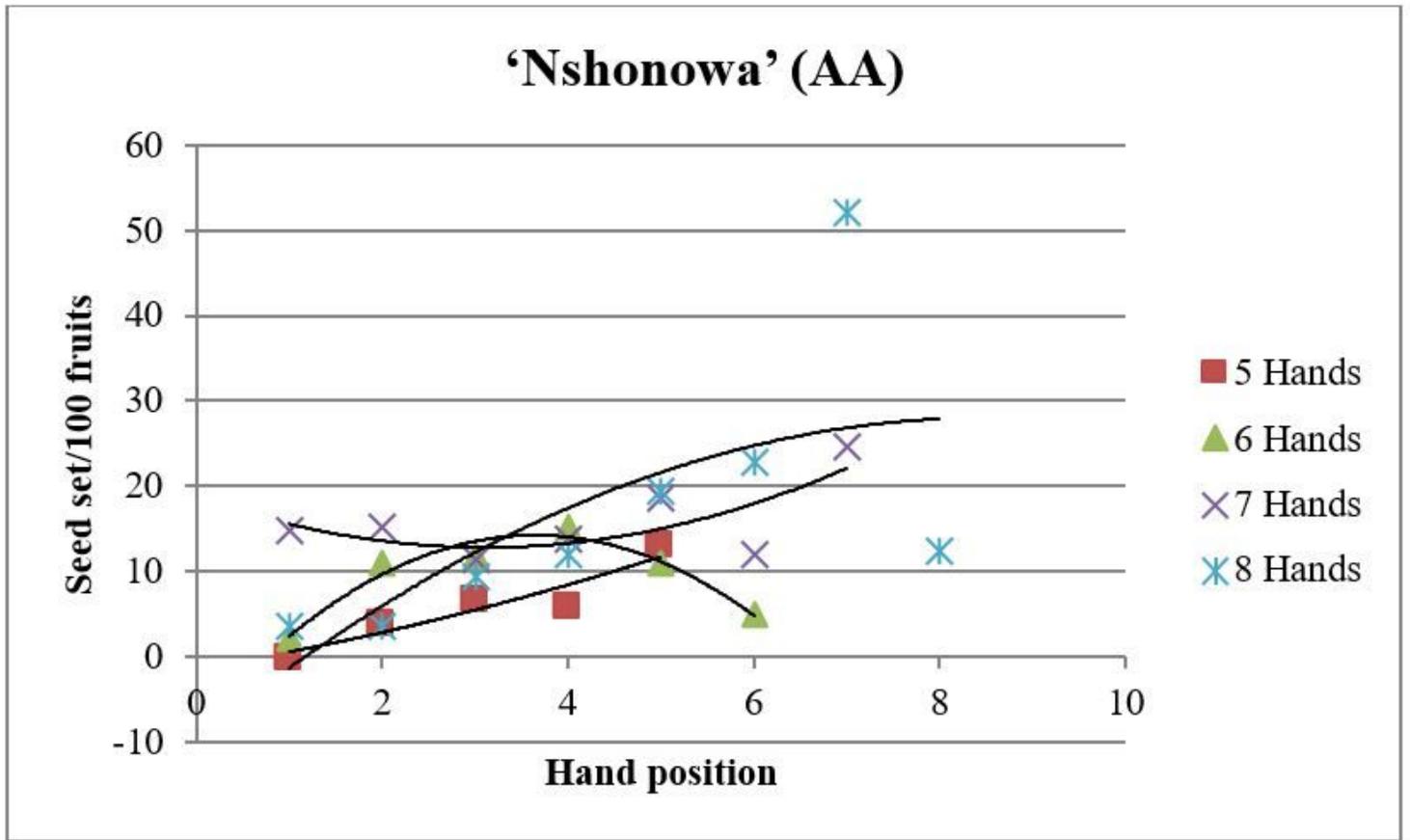


Figure 4

Seed set per 100 fruit per hand in different sized bunches of 'Nshonowa'

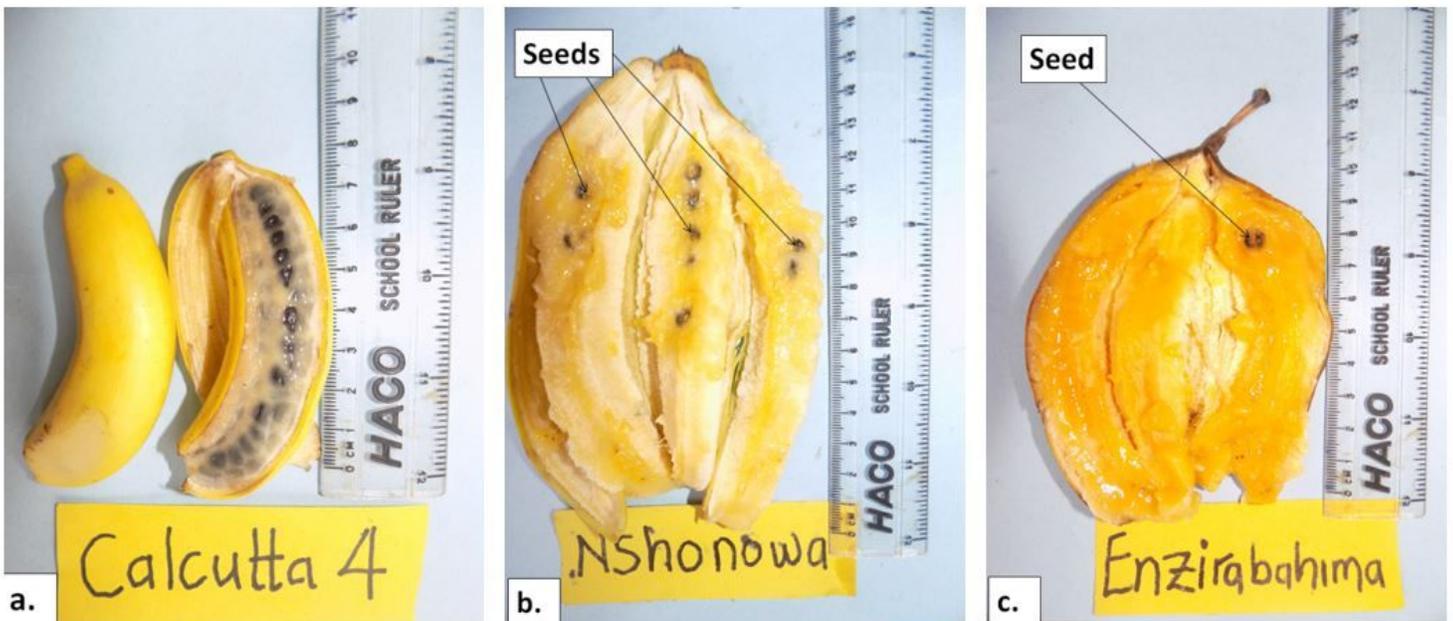


Figure 5

Seed set position in fruits as revealed after ripening among selected banana genotypes. a) Small intact and peeled fruits of non-parthenocarpic open pollinated wild banana 'Calcutta 4' with numerous evenly distributed seed. b) Multiple seed in the distal end of the fruit in the cultivar 'Nshonowa' pollinated with 'Calcutta 4'. c) Single seed set in the distal end of 'Enzirabahima' pollinated with 'Calcutta 4'.

Supplementary Files

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