

# Multicentre Case-Control Study of Pneumococcal Infections Among Children with Pneumonia in Peninsular Malaysia (MY Pneumo): A Study Protocol

**Nurul Hanis Ramzi**

nurulhanis@imu.edu.my

International Medical University

**Andrew Tan Chun Hoong**

International Medical University

**Nur Alia Johari**

International Medical University

**Anna Marie Nathan**

University of Malaya

**Cindy Shuan Ju Teh**

University of Malaya

**Norhayati Aida Sulaiman**

University of Malaya

**Mohamad Ikram Ilias**

Universiti Sains Malaysia

**Zakuan Zainy Deris**

Universiti Sains Malaysia Health Campus

**Siti Nur Haidar Hazlan**

Universiti Sains Malaysia Health Campus

**Nur Syafiqah Mohamad Nasir**

Universiti Sains Malaysia Health Campus

**Asrar Abu Bakar**

International Islamic University Malaysia

**Muhd Alwi Muhd Helmi**

Kulliyah of Medicine, International Islamic University Malaysia

**Wan Khairunnisa Wan Juhari**

International Islamic University Malaysia

**Norhidayah Kamarudin**

Kulliyah of Medicine, International Islamic University Malaysia

**Chun Wie Chong**

International Medical University

**David W. Cleary**

University of Birmingham

**Stuart C. Clarke**

University of Southampton

**Lokman Hakim Sulaiman**

International Medical University

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**Study protocol**

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**Additional Declarations:** Competing interest reported. DWC was a post-doctoral researcher on GSK funded projects in 2014/15 and has received grant support from Pfizer and the National Institute for Health via the NIHR Southampton Biomedical Research Centre. SCC acts as principal investigator for clinical trials and other studies conducted on behalf of University Hospital Southampton NHS Foundation Trust/University of Southampton that are sponsored by vaccine manufacturers. No personal payments are received from them. SCC has participated in advisory boards for vaccine manufacturers but receives no personal payments for this work. SCC has received financial assistance from vaccine manufacturers to attend conferences. All grants and honoraria are paid into accounts within the respective NHS Trusts or Universities, or to independent charities. All other authors have no conflicts of interest.

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# Abstract

**Background:** *S. pneumoniae* (SPN) is the most common cause of pneumonia. The disease can be effectively prevented through immunisation. Since December 2020, the Malaysian Government has included the 10-valent pneumococcal conjugate vaccine (PCV10) for all infants born on or after 1 January 2020 as part of the National Immunisation Programme (NIP). However, the epidemiology of pneumonia remains poorly understood. To fill the knowledge gap, we established a multicentre surveillance study to understand the burden of pneumococcal pneumonia among young children in Peninsular Malaysia.

**Methods:** MY Pneumo is a multicentre prospective case-control study conducted in three sentinel sites located in three different states of Peninsular Malaysia – Kuala Lumpur, Pahang, and Kelantan. A cohort of at least 500 incident cases and 500 controls is enrolled beginning in October 2021 and matched for age. Cases are hospitalised children <5 years with radiologically confirmed pneumonia, and the controls are children without any features suggestive of pneumonia. Clinical samples, including nasopharyngeal swabs (NPS) and urine, are collected according to the study protocol. Biological fluids such as blood, cerebrospinal fluid (CSF) and pleural fluid are obtained from invasive pneumonia disease (IPD) patients, if available. All children are tested for SPN using polymerase chain reaction (PCR) and pneumococcal urine antigen test (PUAT) using BinaxNow.

**Discussion:** Surveillance data, including carriage rate, serotype variations and the phylogeny data structure of SPN among young children in Malaysia during PCV implementation, will be generated from this study. Trends and patterns of pneumococcal serotypes by different regions are important for targeted public health strategies. Our data will provide baseline information for estimating the impact of PCV10 implementation and will influence policymakers' decisions regarding the upgrade from PCV10 to a higher-valency conjugate vaccine in Malaysia.

**Trial registration:** This project was registered at ClinicalTrials.gov (NCT04923035) on June 11th, 2021. The study protocol was approved by the International Medical University Joint-Committee on Research & Ethics (4.15/JCM-216/2021) and the Institutional Review Board at sentinel sites (USM/JEPeM/21020190, IREC 2021-114, MREC ID No: 2021128-9769) and University of Southampton's Ethics and Research Governance (ERGo II 64844).

## Background

According to the World Health Organization (WHO), pneumonia accounts for a substantial portion of deaths in children under five (1). The burden of pneumonia is high in developing countries with limited access to healthcare resources, overcrowding and vaccines. Preventive measures, such as routine vaccinations, exclusive breastfeeding, good nutrition, and reducing exposure to indoor air pollution, can reduce the risk of pneumonia in children (2–4). Additionally, timely diagnosis and appropriate treatment,

including the use of antibiotics when necessary, are crucial in managing pneumonia and preventing severe outcomes (5,6).

The most common cause of pneumonia among children under five is infection with *Streptococcus pneumoniae* (SPN), also known as pneumococcus. Immunisation with a pneumococcal vaccine is an effective way to prevent pneumonia. In Malaysia, pneumococcal vaccination under the NIP for children has commenced for infants born from January 2020, comprising three doses at 4, 6 and 15 months. The routine use of the 10-valent pneumococcal conjugate vaccine (PCV10) in children is expected to reduce the burden of pneumococcal infections in the country, especially severe infections. The impact of the coronavirus disease 2019 (COVID-19) pandemic may also have implications on the pneumococcal serotype and clinical presentation of infections in the community. There was a 30% to 80% decline in IPD incidence rate observed for all pneumococcal serotype groups in England and the Netherlands populations, respectively (7,8). In Malaysia, children with severe pneumonia due to COVID-19 exhibited a less severe clinical trajectory when compared with cases caused by other respiratory viruses (9). The children manifested early in the disease progression, exhibiting no atypical biomarkers, such as lymphopenia and elevated C-reactive protein (CRP) levels. There were no invasive pneumococcal coinfections despite the low pneumococcal vaccination rates among the study population (9). Therefore, we conducted a prospective case-control study to understand the burden of pneumococcal pneumonia in young children and to provide a baseline assessment to monitor changes in SPN carriage post-PCV implementation in Malaysia.

## Methods/Design

### Study objectives

The study's primary objectives are to provide baseline surveillance of pneumococcal pneumonia in young children in Malaysia by determining the prevalence of SPN nasopharyngeal carriage among children aged 5 years and below with pneumonia and IPD. Secondary study objectives are: 1) to determine SPN serotypes by polymerase chain reaction (PCR) and whole-genome sequencing (WGS), 2) to evaluate the correlation of SPN detection between nasopharyngeal carriage and urine samples, and 3) to establish sensitivity and specificity cutoffs of BinaxNOW for the detection of SPN serotypes in children with pneumonia and IPD.

### Study size power calculation

This is a prospective case-control study in which all children aged 5 years and below with clinically diagnosed pneumonia and healthy controls will be recruited at three study sites. As per assumptions of 95% confidence interval (CI), 80% power and detectable odds ratio (OR) of 1.5, we estimate 164 cases and 164 healthy controls per sentinel hospital site. Thus, we aim to recruit a total of 500 confirmed pneumonia/IPD cases and 500 healthy controls for this study.

### Study sites and design

This prospective, hospital-based, multicentre case-control study is being conducted at three university hospitals in different states within Peninsular Malaysia – Kuala Lumpur, Pahang, and Kelantan (Figure 1). Kuala Lumpur serves as the capital city of Malaysia and is centrally located in the western part of Peninsular Malaysia. Pahang and Kelantan are located on the east coast of Peninsular Malaysia, along the South China Sea, providing them with access to the coastline. Kelantan is situated to the north of Kuantan and shares its eastern border with Thailand. Study enrolment is at 1) University Malaya Medical Centre (UMMC), Kuala Lumpur; 2) Hospital Universiti Sains Malaysia (HUSM), Kota Bharu, Kelantan; and 3) Sultan Ahmad Shah Medical Centre International Islamic University Malaysia (SASMEC IIUM), Kuantan, Pahang. Table 1 summarises the demographics and burden of pneumonia in each state of the participating sentinel site. Cases and controls are age-matched within the range of 6 months. Cases are age-matched children clinically diagnosed with pneumonia and attending the outpatient department or admitted as a hospital inpatient, Invasive Pneumococcal Disease (IPD) (without pneumonia), bacteraemic pneumococcal pneumonia, and chest radiograph (CXR) confirmed pneumonia (by WHO guideline) (10). Controls are healthy age-matched children without any intercurrent respiratory illness. Subject enrollment is over 24 months for each participating site, and a total sample of 400 subjects (200 cases and 200 controls) is targeted for UMMC and HUSM, respectively, and 200 subjects (100 cases and 100 controls) for IIUM. Male and female patients from the three major ethnic groups (Malay, Chinese, and Indian) are being recruited from each sentinel site. The study protocol workflow is as outlined in Figure 2.

Table 1 Demographic characteristics by states of participating university hospitals in MY-Pneumo study.

State	State population type	State population ('000) <sup>a</sup>	State population of children <5 years old, ('000) <sup>a</sup>	No. of deaths of children <5 years old <sup>b</sup>	Under-5 mortality rate per 1000 live births <sup>b</sup>	Deaths of under-5 due to pneumonia, % <sup>b</sup>
<b>Kelantan</b>	Urban/Rural	1829.4	173.9	285	8.2	4%
<b>Pahang</b>	Urban/Rural	1612.5	128.1	173	7.1	5.20%
<b>Federal Territory of Kuala Lumpur</b>	Urban	1945.3	115.9	130	6.2	1.60%

a : 2022 prediction by Department of Statistics, Malaysia. Vital Statistics, Malaysia, 2022

b : Statistics on Causes Of Death Malaysia, 2022, Department of Statistics, Malaysia

## Study participants

The study population is comprised of children under 5 years of age who comply with protocol definitions and inclusion criteria. Eligible participants are identified by study paediatricians and research assistants at each participating site. The following criteria define cases: 1) hospitalised patients aged between 2

weeks and 59 months, 2) clinical features of pneumonia, as described below, 3) radiological confirmation of pneumonia based on CXR findings as per WHO guidelines (Cherian et al., 2005) (10), and 4) an informed consent statement signed by the children's parents or legal guardian. The exclusion criteria for cases are the following: children who 1) do not meet the case definition, and 2) whose parents or legal guardian declined to sign the informed consent statement. Controls are defined by children aged 5 years and below, who are in good health, as determined by a brief medical history and/or clinical judgement of the investigator, and whose parent or legal guardian is willing and able to give informed consent. Exclusion criteria for the controls are: 1) any symptom suggestive of respiratory illness, 2) has nasal surgery, 3) has significant diseases or symptoms, such as febrile illness or a temperature  $\geq 38^{\circ}\text{C}$  on the day of the visit or in the preceding 72 hours that can place the patient at an increased risk of the disease, 4) has a history of antibiotic administration in the month prior to sampling, or 5) minors whose parents or legal guardian decline to sign the informed consent statement. Cases and controls are matched for study site and age ( $\pm 6$  months). General questionnaires are administered to the parents or legal guardians of the participants to obtain demographic and socio-economic data and their medical history. Subjects are identified based on an anonymised identifier. The master list is kept in a password-protected spreadsheet.

## **Definition of pneumonia**

Pneumonia cases are defined as patients with a history of cough and/or difficulty/rapid breathing and/or intercostal recession, with or without fever, and radiological confirmation of pneumonia as per WHO guidelines (10).

## **Biological samples**

Samples are collected in the first 24 hours of patient hospitalisation (Table 2). Nasopharyngeal swabs (NPS) and urine samples are collected from all pneumonia cases and controls following Centers for Disease Control and Prevention (CDC) Pneumococcal Carriage Protocols (11). Each sample is aliquoted as per protocols outlined in this study. Each NPS is inoculated into a medium containing skim milk, tryptone, glucose, and glycerol (STGG) and stored at  $-80^{\circ}\text{C}$  before shipment to the International Medical University Advanced Microbiology Collaborative Research Laboratory (IMU-AMCRL) in Kuala Lumpur, Malaysia, every 4 months by a licensed/registered courier. One NPS and one 15 mL urine sample are collected from each subject at the time of enrolment. The collection procedure is performed by trained clinical staff, nurses, and the research assistant at the sentinel site. The NPS collection procedure involves inserting a nylon-tipped FLOQswab (COPAN Diagnostics Inc., USA) through the nostril into the cavity between the nose and mouth for 5 seconds and rotating it several times. The swab is then inserted into a cryovial with 1 mL of STGG media (2% (w/v) skim milk powder, 3% (w/v) tryptone soy broth powder, 0.5% (w/v) glucose and 10% (v/v) glycerol in water), and immediately vortexed briefly and frozen at  $-80^{\circ}\text{C}$  within 4 hours. Different methods of urine collection are used depending on the age and tolerability of the participant, such as clean catch or void, urine collection bag, and diaper. Briefly, for children using diapers, we used the "urine ball method", which involves placing sterile cotton balls on the subject's diaper. The urine is subsequently collected by placing the urine-soaked cotton balls into a sterile

20 mL syringe, and the fluid is extracted with a plunger. The collected urine is mixed with PIPES buffer and aliquoted into three tubes of 5 ml each. All urine sampling supplies, including tubes, urine bags, buffer and BinaxNOW test kits, are provided to sentinel sites by Merck Sharp & Dohme (MSD)

via Pharmaceutical Product Development (PPD). Urine samples are tested with BinaxNOW at sentinel sites, and the remaining aliquots are shipped to the MSD Central Lab on dry ice. The BinaxNOW® test kit includes a test device (strip or card), a specimen swab, and a buffer solution. The test device is placed on a clean, flat surface before a buffer solution is added from a dropper bottle. A specimen swab is dipped into the urine specimen, removed, and then inserted into the test card. The card is then closed, bringing the specimen into contact with the test strip. Pneumococcal antigen present in the specimen reacts to bind anti-*S. pneumoniae*-conjugated antibody. The resulting antigen-conjugate complexes are captured by immobilised anti-*S. pneumoniae* antibody, forming the Sample Line. Immobilised control antibody captures anti-species conjugate, forming the Control Line. Test results are interpreted by the presence or absence of visually detectable pink-to-purple coloured lines. A positive test result, read in 15 minutes, will include the detection of both a Sample and a Control Line. A negative test result, read in 15 minutes, will produce only a Control Line, indicating that *S. pneumoniae* antigen was not detected in the specimen. Failure of the Control Line to appear, whether the Sample Line is present or not, indicates an invalid assay. Additional blood, cerebrospinal fluid (CSF) and pleural fluid will be taken for IPD patients, if available, at each sentinel hospital site.

Table 2 Laboratory tests on case and control subject samples.

Samples	Collection method	Analyses
Nasopharyngeal swab	Flocked swab in 1 ml STGG cryovial*	Molecular detection using multiplex PCR for SPN serotyping.
		Microbiological culturing and optochin sensitivity
Urine	Sterile cotton balls, Catheter	Pneumococcal urine antigen detection (BinaxNOW)

\*Skim milk powder, Tryptone soy broth powder, Glucose, Glycerol medium.

### Laboratory analysis

Laboratory isolation (including genomic deoxyribonucleic acid (DNA) extraction), identification and confirmation of SPN in each sample are carried out at the IMU-AMCRL. Aliquots of NPS in STGG media include volumes of 400 µl, 350 µl and 250 µl, are stored in -80 °C freezers. The 250 µl NPS-STGG aliquot is the working sample used for blood agar culture, optochin-sensitivity, and DNA extraction for each participant. A 10 µl loopful of sample is taken from the 250 µl aliquot, plated onto Columbia agar with 5% sheep blood (CBA) (Oxoid), and incubated in 5% CO<sup>2</sup> at 37 °C for 24 hours. The plate culture is observed after 24 hours of incubation, and colonies exhibiting SPN morphology are picked for replating onto fresh

CBA plates with 5 mcg optochin disc (HIMEDIA), followed by incubation as in the previous step. SPN isolates are identified as small, greyish, alpha-hemolytic culture growths showing Draughtsman morphology, and are optochin sensitive. Genomic DNA from the remaining 240 µl NPS-STGG aliquot is extracted using the New England Biolabs (NEB) Monarch® Genomic DNA Purification Kit (USA). The extracted genomic DNA is then subjected to multiplex conventional PCR analysis according to the CDC protocol (US scheme protocol) to detect SPN serotypes by amplifying the capsular polysaccharide biosynthesis gene A (*cpsA*) targets. SPN isolates will be transferred to the University of Southampton (UoS), United Kingdom (UK), for WGS analysis. Isolates will be sequenced using a MiSeq (Illumina, UK) to generate 2 × 300 paired-end data. Assembly will be done using SPAdes with assembly improvement and QC as described previously (12).

### **Data sources and quality control**

Clinical site monitoring for research conduct and management is conducted yearly by the Principal Investigator team from the International Medical University (IMU). The main purpose of clinical site monitoring is primarily to ensure protocol adherence, source data verification, investigator training, site performance, data quality assurance, and maintaining data integrity at each participating site. Tasks and responsibilities are based on standard research guidelines, including standard operating procedures. Data sources are monitored and evaluated for case and control definition conformity, errors, and missing data at each sentinel site. Vaccination records, underlying diseases, medical history, radiological findings, and demographic characteristics are recorded prospectively for each patient on a case report form (CRF) (Table 3). Data quality reporting is redacted for each site to ensure the confidentiality and conformity of all study data variables. This process will be applied to data analysis of each enrolled case and control subject. Data accuracy will be assessed by comparing the recorded values with source documents. The principal investigator at each site is contacted for queries regarding this quality assessment and is involved in resolution.

Table 3 Overview of questionnaire used in MY-Pneumo study.

### **Data analysis**

An anonymous database will be built, and clinical data will be linked to laboratory data. Quantitative variables will be described and categorised according to their distribution in the study population. Descriptive analysis will address each covariate for the entire population and will be stratified by site. Patients' characteristics and laboratory data for cases and controls will be compared. The associations between risk factors and carriage will also be examined by estimating the relative risk in the study population. Data analysis will be conducted using software such as R and SPSS. The absence/presence of SPN will be modelled using a mixed effects logistic regression model and random forest. The validity of the model will be tested using cross-validation. Sensitivity analysis will also be carried out to evaluate the impact of selected clinical and demographic parameters on the BinaxNOW test outcome.



Category	Information		
		Subject interview	Hospital records
Demographic	Ethnicity	Main caretaker	Date of Birth
	Residence		Weight at Birth
	Parents' Education Level	Feeding habit	Height at Birth
	Parents/Family member smoking habit		Gestational term Current Weight and Height
Underlying diseases	Immunodeficiency	Liver disease	
	Kidney disease	Respiratory disease	
	Cardiac disease	Malnutrition	
	Blood disease		
Medical history			Prior ear infections
			Prior respiratory illness
			Prior antibiotic treatment
			Prior hospitalisations
Vaccination history (Dates, Number of Doses)	BCG, HepB, DTap, Hib, MMR, IPV, Tetanus	PCV	
Medical intervention			Date of admission
			Date of discharge
			Symptoms
			Level of care
			Antibiotic treatment
			Radiological findings

## Discussion

MY Pneumo study collects data on potential risk factors that may influence trends in respiratory disease, as well as pneumococcal carriage among healthy controls. This study incorporates different study designs which will help us to interpret the changes over time in younger children with pneumonia following PCV10 introduction. The results obtained from this study will be reported in due course, with the hope that the information gained will contribute to a better understanding of pneumococcal disease in the Malaysian context. Regional site analysis, focusing on specific interests, will also be described and published at a later date. Study analysis will generally focus on the prevalence of pneumonia for the entire population and by the sentinel site. Insufficient data regarding pneumonia aetiology in developing countries is evident from the geographical distribution of study sites worldwide (13–15). It has been suggested that further carriage and disease studies are required in middle-income countries, especially in Malaysia, to assess the effectiveness of pneumococcal vaccination to monitor serotype changes in the population (16–18). The Pneumonia Etiology Research for Child Health (PERCH) study was the largest, most comprehensive pneumonia aetiology study conducted in seven countries in Africa and Asia: Gambia, Mali, Kenya, Zambia, South Africa, Bangladesh, and Thailand from 2011 to 2014 (19). The study setting involved severe childhood pneumonia cases (4,232 children between 1 month and 5 years), and each country had different characteristics that may have influenced the causes of pneumonia. Yet, the research study was pivotal in advancing our understanding of the causes and contributors to childhood pneumonia in developing countries. By investigating a diverse range of variables and risk factors, the PERCH study contributed to driving transformative changes in child health outcomes, reducing the burden of pneumonia-related morbidity and mortality. A more comprehensive understanding of the pathogens responsible for childhood pneumonia holds the potential to significantly enhance both preventive measures, including the implementation of effective vaccine policies, and therapeutic interventions. This collective effort towards a better understanding of pneumonia causality has the power to drive improved management strategies, ultimately resulting in a remarkable reduction in the burden of morbidity and mortality associated with childhood pneumonia.

The present study's main strength is the prospective multicentre case-control study design that will permit the description of paediatric pneumococcal pneumonia in study-related locations, especially in urban and rural areas. The sampling and laboratory methods used are consistent with other research studies that have described nasopharyngeal carriage in young children (19,20). Gold-standard molecular methods have been utilised to detect pneumococcus in the nasopharynx (21). The use of the PUAT assay, which detects pneumococcal polysaccharides in the urine of young children, will enable rapid detection of pneumococcal antigens, allowing for timely intervention and reducing the risk of complications.

Whilst our study includes three states in both urban and rural community areas of Peninsular Malaysia, we did not include other states in East Malaysia. Therefore, our findings may not be applicable to represent the diversified population and demographics of Malaysia. The COVID-19 pandemic and co-infection status for pneumonia cases may impact study outcomes as we do not have administrative data available for both. However, we plan to incorporate this information whenever available into the

descriptive interpretation of study results. Moreover, we do not have pneumococcal carriage data for the retrospective period and have therefore chosen clinical endpoints as our primary study objective.

This study will contribute to the incomplete evidence available on the burden of pneumococcal pneumonia in children below the age of 5 years in low-resourced countries, particularly in the Asia-Pacific region. Data on the burden of pneumococcal pneumonia and the impact of PCV can guide decisions related to vaccine prioritisation, resource allocation, and healthcare strategies. As PCV is being introduced in various low-resource settings to combat pneumococcal diseases, understanding its effectiveness and potential benefits is critical for public health strategies. The study is one of the few active population-based pneumonia surveillance programs evaluating the indirect impact of PCV in a resource-limited setting. With the increasing introduction of PCV in low-resourced settings, the methods, experiences, and lessons learned from our study may guide the development of such systems in other countries.

## Abbreviations

CBA: Columbia agar with 5% sheep blood

CDC: Centers for Disease Control and Prevention

CI: confidence interval

COVID-19: coronavirus disease 2019

cpsA: the capsular polysaccharide biosynthesis gene A

CRF: case report form

CRP: C-reactive protein

CSF: cerebrospinal fluid

CXR: Chest radiograph

DNA: Deoxyribonucleic acid

HUSM: Hospital Universiti Sains Malaysia

IMU-AMCRL: International Medical University Advanced Microbiology Collaborative Research Laboratory

IPD: Invasive pneumococcal disease

NIP: National Immunisation Programme

NPS: nasopharyngeal swab

OR: odds ratio

PCR: Polymerase chain reaction

PCV: Pneumococcal conjugate vaccine

PCV10: 10-valent pneumococcal conjugate vaccine

PERCH: Pneumonia Etiology Research for Child Health

PUAT: pneumococcal urine antigen test

SASMEC IIUM: Sultan Ahmad Shah Medical Centre International Islamic University Malaysia

SPN: *Streptococcus pneumoniae*

STGG: skim milk, tryptone, glucose, and glycerol

UMMC: University Malaya Medical Centre

WGS: whole-genome sequencing

WHO: World Health Organization

## Declarations

Ethics approval and consent to participate

The study protocol, informed consent statement, case report form, any amendments and all other study documents have been submitted to and approved by the institutional ethics committee of each site:

1. The International Medical University Joint-Committee on Research & Ethics (4.15/JCM-216/2021),
2. The Human Research Ethics Committee, Universiti Sains Malaysia\_(USM/JEPeM/21020190),
3. The International Islamic University Malaysia\_Research Ethics Committee (IREC 2021-114), and
4. The Medical Research Ethics Committee, University Malaya Medical Centre (MREC ID No: 2021128-9769)
5. University of Southampton's Ethics and Research Governance (ERGo II 64844)

Consent for publication

Not required because no individual identifiers have been included in the manuscript.

Availability of data and materials

Not applicable – manuscript does not contain any data.

Competing interests

DWC was a post-doctoral researcher on GSK funded projects in 2014/15 and has received grant support from Pfizer and the National Institute for Health via the NIHR Southampton Biomedical Research Centre. SCC acts as principal investigator for clinical trials and other studies conducted on behalf of University Hospital Southampton NHS Foundation Trust/University of Southampton that are sponsored by vaccine manufacturers. No personal payments are received from them. SCC has participated in advisory boards for vaccine manufacturers but receives no personal payments for this work. SCC has received financial assistance from vaccine manufacturers to attend conferences. All grants and honoraria are paid into accounts within the respective NHS Trusts or Universities, or to independent charities. All other authors have no conflicts of interest.

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## Authors' contributions

All authors designed and co-authored the protocol. NHR and ATCH were responsible for preparing the manuscript. LHS and SCC conceived the idea. LHS, SCC, CCW, NHR, and NAJ drafted the original proposal. CSJT, SNHH, ZZD, and NK supported microbiological aspects-based protocols. AMN, NAS, MII, ZZD, NSMN, AAB, MAMH, and WKWJ coordinated study implementation at sentinel sites. DWC supported the WGS analysis. All authors read, commented on, and approved the final manuscript version.

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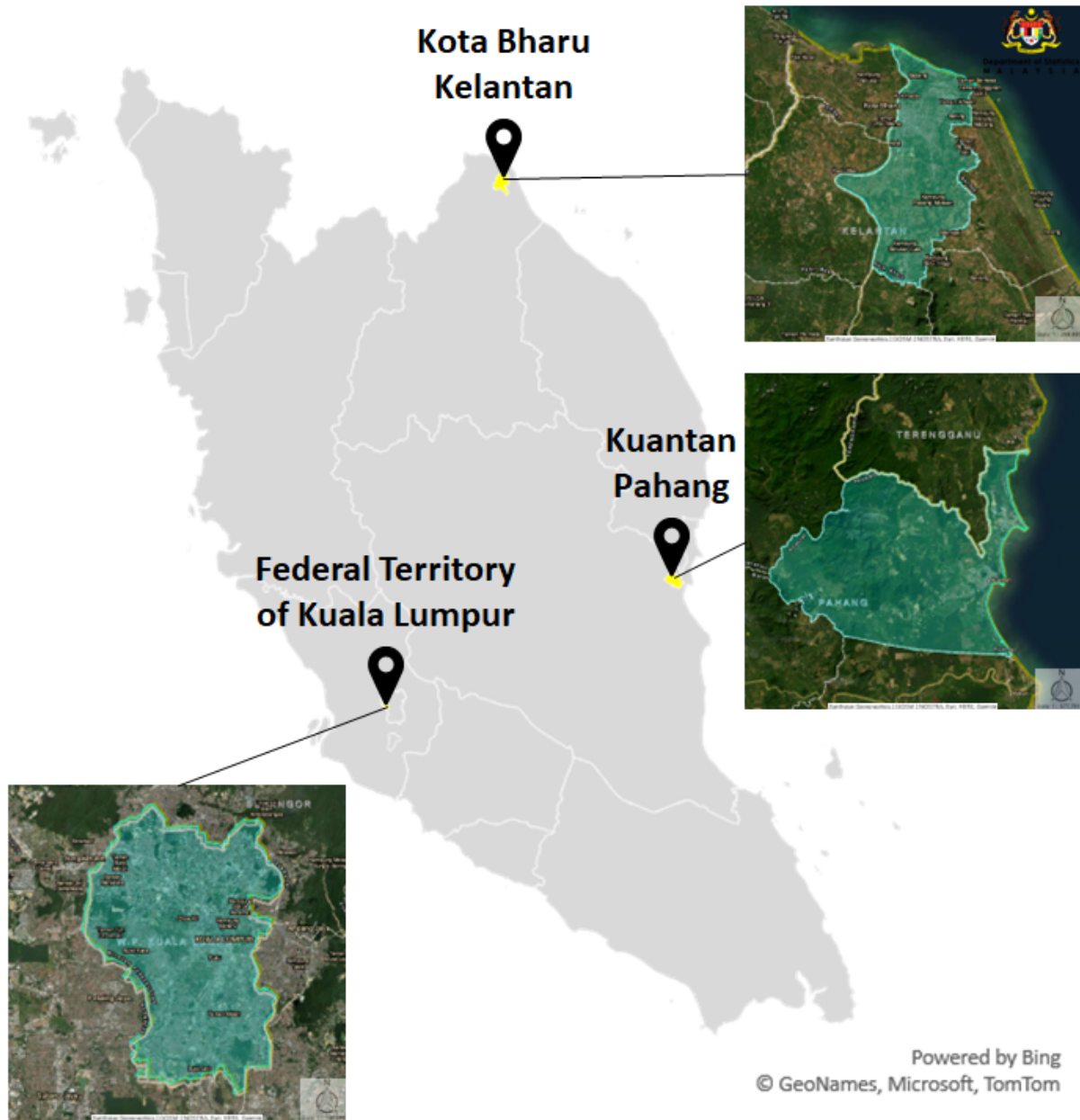
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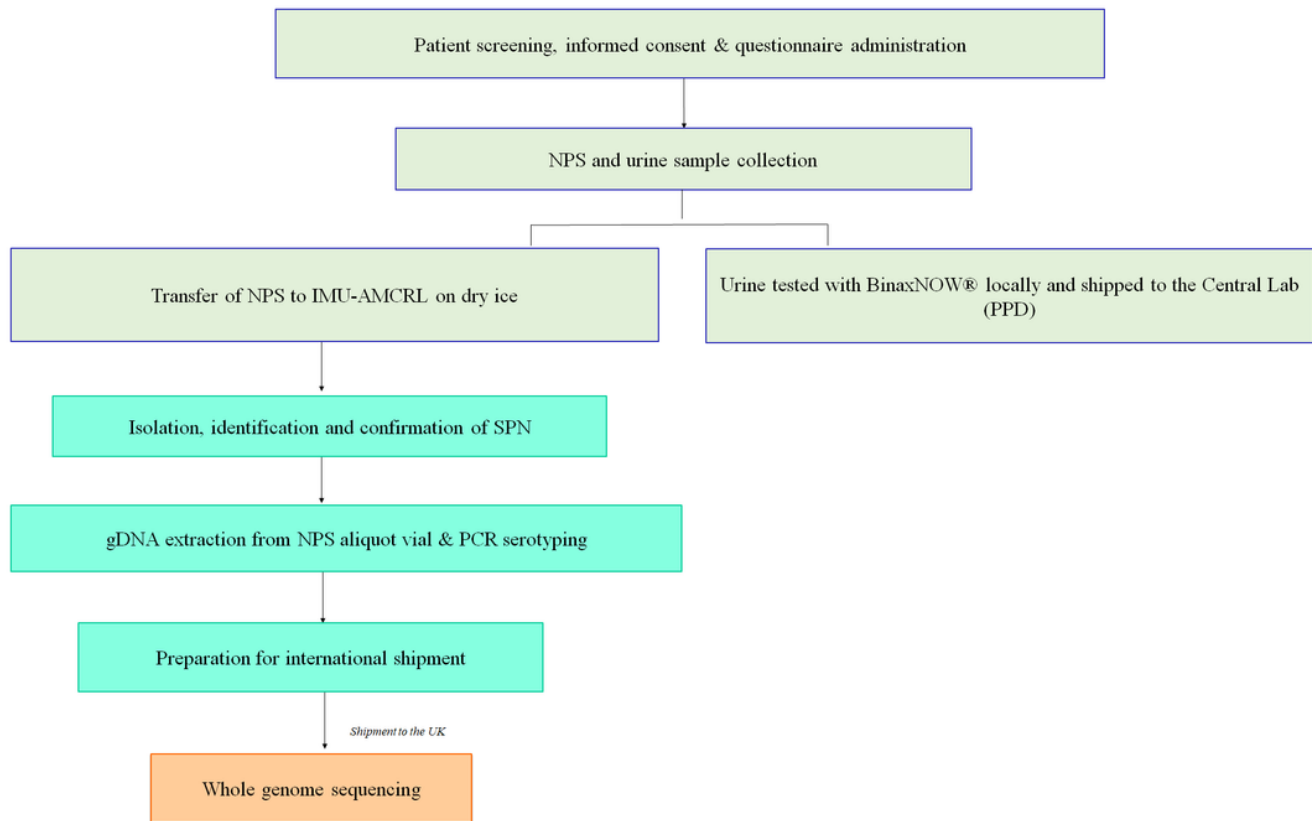
## Figures



**Figure 1**

Geographical locations of participating university hospitals in Peninsular Malaysia: Kuala Lumpur, Pahang, and Kelantan. Source: Adapted from Department of Statistics, Malaysia. (2023, August 16). <https://statsgeo.mycensus.gov.my/geostats/report.php>





**Figure 2**

Summary of the study protocol workflow. The specimen collection, processing, and storage of nasopharyngeal swabs (NPS) were adapted from CDC Pneumococcal Carriage Protocols. Clinical samples were taken according to local policy, including nasopharyngeal swabs (NPS) and urine samples. Different methods were used to collect the urine depending on the age and tolerability of the participant i.e. clean catch/void, urine collection bag, and diaper. Additional blood, cerebrospinal fluid (CSF) and pleural fluid were taken for invasive pneumococcal disease (IPD) patients, if available. NPS was inoculated into a medium containing skim milk, tryptone, glucose, and glycerol (STGG) and stored at  $-80^{\circ}\text{C}$  before shipment to IMU-Advanced Microbiology Collaborative Research Laboratory (AMCRL). General questionnaires/case report form (CRF) were administered to the parents/caregivers of the patients to obtain socio-demographic, vaccination history and medical history of the child. The gene targets for pneumococcal identification, including the main pneumococcal capsular biosynthesis gene A (*cpsA*), were analysed through PCR. *Streptococcus pneumoniae* (SPN) isolates will be transferred to the University of Southampton (UoS), United Kingdom (UK), for whole-genome sequencing (WGS) analysis..