Genetic and clinical phenotypic analysis of familial stapes sclerosis caused by a *NOG* mutation

Rong Yu

First Affiliated Hospital of Nanchang University <https://orcid.org/0000-0002-1689-0226>

HONGQUN JIANG

First Affiliated Hospital of Nanchang University

WU GEN LUO (✉ luowugen@126.com)

First Affiliated Hospital of Nanchang University

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Abstract

Background: The noggin protein encoded by the *NOG* gene can interfere with the binding of bone morphogenetic protein to its receptor, thus affecting bone and joint development. The symptoms include abnormal skeletal development and conductive deafness.

Methods: In a retrospective study, clinical data of the proband and her family members, including 8 people and 50 healthy normal controls, were collected. Second-generation sequencing was performed on peripheral blood samples from them.

Results: The sequencing analysis indicated that in the proband, the *NOG* gene had a c.532T>C mutation (cytosine deletion, NM_005450.6:c.532T>C), leading to an amino acid change. The proband's father, grandmother, second sister, and third sister also had this mutation, whereas family members with normal phenotypes did not have the mutation.

Conclusion: Analysis of this family showed that the novel presentation of the c.532T>C mutation in the *NOG* gene resulted in syndrome-type autosomal dominant inheritance reflected in a mild clinical phenotype, which is of great importance for further studies of the clinical phenotype and pathogenesis of stapes sclerosis.

Background

Conductive deafness is a type of hearing loss caused by lesions in the outer and middle ear. The passage of external sound waves into the inner ear is obstructed by pathological factors affecting the sound transmission system in the ear. Congenital causes are common malformation of the outer ear, hypoplasia of the tympanum, malformation of the ossicular chain, familial otosclerosis, etc. Acquired causes are commonly seen in external auditory canal obstruction by cerumen, foreign bodies, inflammation, scars, tumours, etc., or by tympanic membrane trauma, perforation, thickening and adhesion, middle ear effusion or pus, cholesteatoma, fracture of the auditory chain, ear sclerosis, tumour, etc. Stapes ankylosis is rare but can cause early conductive deafness [1-4]. It is difficult to differentiate from the common aetiology of conductive deafness and otosclerosis and is often overlooked, delaying treatment. Stapes ankylosis may be associated with skeletal dysplasia, such as osteogenesis imperfecta type I (OMIM 166200) [5-7]. Skeletal dysplasia is a common congenital malformation and a common phenotype in many inherited disorders.

Abnormal bone and joint development are the most common congenital malformations. Mutations in the *NOG* gene (OMIM 602991) on chromosome 17q21–q22 can lead to multiple autosomal dominant genetic diseases. The noggin protein encoded by the *NOG* gene can interfere with the binding of bone morphogenetic protein to its receptor, thus affecting bone and joint development, which may manifest as skeletal dysplasia and conductive deafness[8-11]. Skeletal dysplasia includes proximal interphalangeal joint fusion syndrome type A (OMIM 185800), multiple osseous junction syndrome type 1 OMIM 186500, tarsal and carpal joint fusion syndrome (OMIM 186570), short finger syndrome type B2 (OMIM 611377),

and stapes stiffness with a wide thumb and toes (OMIM 184460)[8]. To improve the understanding of this rare disease, *NOG* gene analysis was performed in a family with stapes ankylosis in China.

Few studies have investigated the mechanism and correlation of stapes ankylosis in China, and a gold standard for pathological diagnosis is still lacking. Clinical diagnosis is mainly based on family history, genetic history, genetic testing, and phenotypic signs. Currently, surgery is the only specific and effective treatment. Therefore, a thorough understanding of the disease is important. In this study, we performed a clinical examination and genetic mutation analysis of a Han Chinese family with a suspected clinical manifestation of stapes ankylosis.

Methods

Members of the family analysed in this study were from Jiangxi Province, China. The ten-member family, two of whom had died, included three generations. A detailed medical history was taken of the eight living family members, and all suspected patients underwent a detailed physical examination, including a specialist examination of the ear, height measurement, hand X-ray, and hearing tests. Other family members were identified as patients through detailed questioning. Peripheral venous blood was collected from the proband and other family members. The control group included venous blood samples from 50 normal controls in the molecular genetic database of our hospital. All adult subjects signed informed consent forms.

The proband and her three sisters, father, mother, grandmother, and uncle were screened for testing. Peripheral venous blood samples were collected from the subjects with 3–5 mL of EDTA anticoagulant, and DNA was extracted using the Qiagen Genomic DNA Extraction Kit (Qiagen, Inc., Hilden, Germany) following the manufacturer's instructions. The DNA was then quantified, tested for purity using a NanoDrop 1000 (Thermo Fisher Scientific, Waltham, MA, USA) and then stored at -80°C. After purity testing, the extracted DNA was sent to the company's laboratory for relevant genetic testing. The target genes of the samples were captured by liquid-phase capture technology, and high-throughput sequencing was performed on the Illumina HiSeq 2000 platform (Illumina, San Diego, CA, USA) at an average depth of >200X.

The sequencing results were compared to the Human Gene Mutation Database (HG19), Locus Specific Mutation Database ([HTTP://www.hgvs.org/dblist/glsdb.htm](http://www.hgvs.org/dblist/glsdb.htm)), dbSNP (v144), US National Center for Bioinformatics, SPIDEX Database, and other databases. The Netgene2 server was used to analyse and detect the pathogenicity of splicing mutations and other potentially pathogenic sites.

Results

The proband's parents denied inbreeding. The proband reported 4 years of progressive hearing loss with no history of otitis media, ototoxic drug use, or noise exposure. The proband was 168 cm in height and hyperopic, with broad thumbs and toes, difficulty bending thumbs, and limited range of motion in the

neck. Eardrums on both sides were intact. The Rinne and 512 Hz tests using a tuning fork were negative, the Weber test was biased to the side with severe deafness, the Schwabach test showed bone conduction extension, and the Gelle test was negative, suggesting stapes plate fixation. Pure-tone audiometry indicated bilateral conductive deafness. No obvious abnormalities were found in acoustic conductivity tests or on ear computed tomography (Fig 1). The clinical diagnosis was stapes ankylosis.

Six family members had conductive hearing loss, stapes rigidity, hyperopia, and other phenotypic traits, such as broad thumbs and/or toes, difficulty bending thumbs, decreased extremity and neck (range of motion) ROM. The proband had conductive hearing loss, hyperopia, broad thumbs and difficulty bending thumbs, decreased extremity and neck ROM. The proband's first sister had no obvious problems. The proband's second sister had the same symptoms as the proband except difficulty bending thumbs. The proband's third sister had the same symptoms as the proband except hyperopia. The proband's father showed the same symptoms as the proband, and her mother was congenitally deaf and mute. One of the proband's uncles, who had died in an accident, had hyperopia and conductive deafness. Another uncle was normal. The proband's grandmother had the same symptoms as the proband except decreased extremity and neck ROM. The patients in the family were tall except the proband's grandmother, and their walking function was normal. There was no other family history. Autosomal dominant genetic characteristics noted in this family are shown in Fig 2.

The sequencing results were compared to the Human Gene Mutation Database (HG19), Locus Specific Mutation Database ([HTTP://www.hgvs.org/dblist/glsdb.htm](http://www.hgvs.org/dblist/glsdb.htm)), dbSNP (v144), US National Center for Bioinformatics, SPIDEX Database, and other databases. The Netgene2 server was used to analyse and detect the pathogenicity of splicing mutations and other potentially pathogenic sites. The DNA sequencing analysis indicated that the *NOG* heterozygous mutation, namely, c.532t>c (code no. 532 nucleotide variation for cytosine by thymine, NM_005450.6:c.532T>C), was present in the proband's father and grandmother. The proband's two sisters also carried the mutation. To verify that this was the pathogenic mutation that caused stapes sclerosis, we screened 50 individuals with normal hearing from the Chinese population for *NOG* gene mutations. None of them had the mutation. Therefore, the c.532T>C mutation does not belong to the polymorphic site and is considered pathogenic (Fig 3).

We searched the Chinese literature and PubMed database to find relevant literature on *NOG* gene mutations in a Chinese population. The results showed that nearly 40 mutations in the *NOG* gene have been identified, most of which are missense mutations located in the evolutionarily conserved region. Within this region is a functionally critical domain, suggesting a correlation between the region and the *NOG* protein. The c.532T>C mutation identified in this family has not been previously reported. The only clinical symptoms that were similar to those reported in our pedigree were those of Brown et al. 2, who showed stapes ankylosis accompanied by broad thumb *NOG* was absent in patients with fingers, hyperopia and syndactyly. In addition, the clinical characteristics of the affected individuals in this lineage are highly heterogeneous.

Discussion

Congenital stapes ankylosis syndrome is rare, and the only finding consistent with this family's pedigree is described in a study by David J. Brown in an Italian family[2]. However, stapes ankylosis can cause early-onset conductive hearing loss. Congenital stapes ankylosis syndrome is difficult to distinguish from otosclerosis, the most common cause of conductive deafness, and is therefore prone to delayed diagnosis. The skeletal abnormalities it causes are mild, making the syndrome harder to recognize. Mutations in the *NOG* gene can lead to stapes ankylosis, conductive hearing loss, and joint damage. In this study, we report a familial stapes stiffness syndrome in one family with conductive hearing loss. Further genetic testing in this study revealed that the patient and the affected family members all carried the same *NOG* gene mutation. The clinical diagnosis was stapes ankylosis syndrome, and all patients in the family had conductive deafness. All of the patients were hyperopic except one. Some family members are unable to bend thumbs and broad thumbs and/or toes. Some of them have decreased extremity and neck ROM. The overall clinical phenotype of the patients in this family was relatively mild.

The c.532T>C mutation in this family has not been reported before, but its clinical symptoms are similar to other *NOG* mutations[12-14]; however, previous studies did not report the ophthalmological findings in detail. The key features that differentiate SYM1 and SYNS1 from stapes ankylosis with broad thumbs and toes and hyperopia include a characteristic physiognomy, hyperopia, and the absence of cervical vertebral fusion and symphalangism.

The reported results for *NOG* mutation sites and clinical phenotypes show that the diversity of *NOG* is reflected in the diversity of mutation sites, the diversity of clinical manifestations, and the diversity of disease courses and prognoses. Ideally, genotype–phenotype relationship analysis is performed on a large sample of patients, but due to the diversity of gene mutations, patients with identical mutations are limited in number. Therefore, it is difficult to determine the relationship between the mutations and clinical phenotype. Bayat et al. proposed a new mutation in *NOG* in a Danish family in which cysteine 230 was replaced, but none of the affected family members suffered hearing loss[15]. Another study reported that mutations in cysteine 184 caused no indication of hearing impairment. In the family reported here, both cysteine 184 and 230 were affected, resulting in stapes rigidity, hearing loss, and phalangeal dysfunction[16]. These findings confirm that mutations with the same *NOG* coding sequence can lead to different phenotypes and inter-family variation [17-25]. This suggests that disease expression can be independent of the location and type of *NOG* mutation. The spectrum of *NOG* mutations contains a variety of syndromes.

The reported effects of otological surgery for stapes fixation caused by *NOG* protein mutations vary[26-30]. In this study, surgical treatment was adopted in five ears (one patient), and hearing continued to improve after the operations. The follow-up period was 6 months. The patient's hearing improved by approximately 20 dB after surgery. Some reports describe progressive hearing loss due to bone regrowth 2–5 years after surgery. In this study, the number of patients was small, and it was difficult to determine statistical significance or make treatment recommendations. The outcome of stapedectomy may also depend on genetic variation. Our current results were all based on the pure-tone audiogram; the early

audiogram did not assess the speech reception threshold. This threshold is also an important parameter and should be considered in the evaluation of surgical results.

Conclusion

Conductive hearing loss is most often caused by otitis media or other middle ear lesions. *NOG* mutations can lead to early-onset conductive hearing loss. In combination with family history, genetic testing may help in identification. The initial use of clinical presentation and a family history of conductive deafness in the clinic require further evaluation. In the long term, the literature is inconsistent with regard to surgical outcomes, but the results of this study suggest that stapedectomy may be successful and can lead to lasting improvements in hearing. This family showed autosomal dominant inheritance related to the *NOG* gene, which is significant for understanding the pathogenic characteristics and clinical features of stapes ankylosis syndrome.

Abbreviations

NOG: Noggin,

EDTA: Ethylene Diamine Tetraacetic Acid

SYM1: Symphalangism 1

SYNS1: Synostosis syndrome 1

ROM : Range of motion

Declarations

Acknowledgments

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Authors' contributions

R Y: Experimental design, analysis, data interpretations, wrote and edited the manuscript; H J: sample preparation, genotyping and data acquisition, manuscript editing and revision; W L: experimental design, recruitment, clinical diagnosis, surgery, data interpretation, manuscript editing and revision. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

The study adhered to the tenets of the Declaration of Helsinki and had received approval from the Institutional Review Board and Research Ethics Committee of The First Affiliated Hospital of Nan Chang University, Nanchang, Jiangxi (approval number 34352). Written, informed consent was obtained from all participants before their inclusion in this study.

Consent to publish

Written informed consent for publication of identifying images or other personal or clinical details was obtained from all of the participants.

Competing interests

The authors declare that they have no competing interests.

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Not applicable

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Figures

Figure 1

The audiological and physical examination of the probands (A) Cervical vertebra degener ation by X-ray examination; (B-C) Joint space ambiguity by X-ray examination (D) Thickened bulb in both eyes (E) The vitreous body of both eyes is punctate turbid by ultrasonography (F) no obvious abnormality by fundus camera (G) Binaural conductive hearing by PTA; (H) "A" type by acoustic immittance measurement; (I) the temporal bone CT;

Figure 2

Pedigree of family. The arrow denotes the proband. Square symbols denote male patients, and circles denote female patients; blackened symbols denote affected individuals, and unblackened symbols denote unaffected individuals; each quadrant defines a phenotypic element or a set of phenotypic elements, and blackened quadrants indicate the presence of the corresponding phenotypic element(s)

Figure 3

NOG c.532T>C(chr17:54672116) (A) Sequencing results of NOG gene individual total exon detection of the proband; (B) Sequencing results of NOG gene individual total exon detection of father of the proband (C) Sequencing results of NOG gene individual total exon detection of grandma of proband (D) Sequencing results of NOG gene individual total exon detection of father of elder sister of proband (E) Sequencing results of NOG gene individual total exon detection of father of younger sister of proband