

Genomic features and evolution of the Parapoxvirus during the past two decades

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1 **Genomic features and evolution of the Parapoxvirus during the past two decades**

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19
20 **Keywords**

21 Parapoxvirus, *Poxviridae*, nucleotide composition, selection pressure, evolution

22
23 **Abstract**

24 Parapoxvirus (PPV) has been identified in most mammals and poses a great threat
25 to both the livestock production and public health. However, it is still not fully

26 understood the viral prevalence and evolution of PPV coding sequences. Here, we
27 performed a comparative approach integrating viral genetics, molecular selection
28 pressure and genomic structure to investigate the genomic features and evolution of
29 PPVs. We noticed that although there were significant differences of GC contents
30 between ORFV and other three species of PPVs, all PPVs showed almost identical
31 nucleotide bias, that is GC richness. This reflected a common mechanism which
32 determines GC compositions for virus with similar life cycles. The structural analysis
33 of PPV genomes showed the divergence of different PPV species, which may due to
34 the specific adaptation to their natural hosts. Additionally, we estimated the
35 phylogenetic diversity of segmented genome of PPV. Our results suggested that during
36 the 2010 – 2018 outbreak, the orf virus has been the dominant species under the
37 selective pressure of the optimal gene patterns. Furthermore, we found the mean
38 substitution rates were between 3.56×10^{-5} to 4.21×10^{-4} in different PPV segments, and
39 the PPV VIR gene was evolved at the highest substitution rate. In these protein-coding
40 regions, purifying selection was the major evolutionary pressure, while the GIF and
41 VIR genes suffered the greatest positive selection pressure. These results may provide
42 useful knowledge on the virus genetic evolution from a new perspective which could
43 help create prevention and control strategies.

44

45 **1. Introduction**

46 Parapoxvirus (PPV) is a chordopoxvirus that causes nonsystemic skin lesion
47 diseases in wild mammals worldwide. PPV belongs to *Poxviridae* and contains orf virus
48 (ORFV), pseudocowpox virus (PCPV), bovine papular stomatitis virus (BPSV) and
49 parapoxvirus of red deer in New Zealand (PVNZ) [1-3]. The PPV genome is
50 approximately 134~139 kb in size, which is terminally connected by hairpin loops. In
51 contrast to other poxvirus genera, PPVs genomes have an extremely high GC content
52 of 65% evenly [4-6]. The genetic structure of PPVs exhibit the general pattern form of
53 a central conserved region, with essential genes in regard to position, spacing and
54 orientation [7]. The terminal variable regions encode genes with vital roles in viral
55 infection process such as the regulation of host immune response to infection and the

56 determination of host range [6, 7].

57 PPVs have a global distribution, and the ruminant flocks will be disturbed while
58 virus emerged in many continents [8-10]. This virus is known to infect mammals, such
59 as goats, sheep, cattle, camels, reindeer and other ruminants [11, 12]. Human will also
60 be infected by PPVs, following direct exposing to skin lesions from infected mammals.
61 Regular public health controlling and reporting of human PPVs infections are in place
62 in a lot of regions of the world, containing Europe, South America and South America
63 [11, 13-17]. Until now, there are live PPVs vaccines which may control the disease, but
64 it may not entirely prevent this disease [18].

65 ORFV is the prototype species of PPVs, which will cause high mortality rates when
66 lesions occur in lips and breast of suckling and grazing animals [6, 19]. Ruminants
67 could be infected with ORFV more than once, although with a shorter period of time to
68 recovery and less obvious pathological changes than in the primary infection [20].
69 BPSV could infect cattle of all ages, however, clinical signs are usually observed in
70 calves. Like ORFV, repeated infection of ruminants with BPSV is common, indicating
71 that this virus infection may not confer significant immunity [6]. Except for the most
72 important viruses (ORFV and BPSV), PCPV and PVNZ both maintained in mammals.
73 As the zoonotic agents, all of them are worldwide epidemics and threaten the public
74 health. Recently, attenuated vaccines could limit the severity of viral infection but they
75 could not completely prevent the disease [6, 21-25].

76 Vaccines have been indicated to impose powerful selection pressure on the virus
77 evolution [26, 27]. Viruses may acquire greater virulence through genetic variation and
78 recombination, as a result of the rapid transmission in vaccinated populations. And they
79 usually have a high mutation rate with their low accuracy of reproduction, which may
80 assist the viruses to evade host immune attack and evolve further. The PPV has a large
81 genome, comprising nearly 134~139 kb in size. Consequently, mutation is more likely
82 to occur in PPVs under vaccination and selection pressures [2, 7, 28].

83 This work reported the global population dynamics of PPVs regarding the
84 evolutionary process and genetic diversity. On the basis of PPVs genomes, the sequence
85 comparison and phylogenetic analysis were performed to investigate the genomic

86 features of viral strains. From the dataset of PPVs genome sequences, the evolutionary
87 rate was also considered by Bayesian methods. Additionally, we further explored the
88 different genes that have played an important role in PPVs evolution by estimating the
89 genetic diversity and the selection pressure in this study.

90

91 **2. Materials and Methods**

92 **PPVs sequences**

93 PPVs complete genome sequences and nucleotide sequences of the B2L, CBP, F1L,
94 GIF, VIR, VEGF and VLTF-1 genes were downloaded from GenBank
95 (www.ncbi.nlm.nih.gov/genbank/). To explore the emergent time of the earliest PPV
96 isolate, the sequences with an unknown collection date were excluded. The host,
97 collection date and sampling location of each PPV isolates were determined either from
98 GenBank, or were obtained from the related publications. This led to a dataset of 17
99 complete genome sequences and 776 gene sequences, including 268 B2L genes, 38
100 CBP genes, 83 F1L genes, 133 GIF genes, 79 VIR genes, 122 VEGF genes and 51
101 VLTF-1 genes, respectively (Table S1). The resulting sequences were aligned using
102 MUSCLE 3.7 [29].

103 **Gene structure and nucleotide composition**

104 The gene structure of PPV was visualized by comparing their genomic nucleotide
105 sequences using the R program gggenes project. Nucleotide compositional analysis of
106 the PPVs complete genome sequences were measured using BioEdit v7.2.5 [30],
107 including the frequencies of A, T, G, C, AT and GC. The nucleotide occurrence
108 frequencies of the 3rd codon position (T3, G3, C3, and A3) of synonymous codons were
109 measured by the CodonW v1.3.

110 **Compositional change of PPV isolates over time**

111 During 1974 – 2018, 268 B2L genes of PPV strains were downloaded from
112 GenBank and analyzed by maximum-likelihood method in PAML v4.9, with 1000
113 bootstrap replicates. An online tool – the Interactive Tree Of Life v2 was performed to
114 design the tree [31]. The resulting isolates were composed of 4 species in PPVs
115 population, including ORFV, PCPV, BPSV and PVNZ. Isolates were assigned on the

116 basis of the PPVs species and the initial collection country. The variation number of
117 PPV was calculated using statistical methods and the plot was drawn with the project
118 ggplot2 in R program [32].

119 **Evolution substitution rates**

120 For all PPV genes, the molecular evolution rates were estimated by the MCMC
121 program of BEAST v2.4.8. This program gives a maximum-likelihood estimate of the
122 evolution substitution rate, based on a model that considered a constant evolution rate
123 (molecular clock). We also estimated the confidence intervals for all the parameters.

124 To obtain the best-fitting models for each viral gene, jModelTest v2.1.7 was
125 performed by Akaike Information Criterion (AIC) [33], which was estimated for each
126 model tested according to the approach of Newton & Raftery [34]. Here, we used
127 general time reversible model, a strict molecular clock and a coalescent model with
128 constant available population size, as performed in the BEAST package. Bayesian
129 Markov chain Monte Carlo (MCMC) analysis was implemented with 10 million steps
130 and was sampled every 10000 steps with 10% burn-in. Tracer v1.6 was used to analyze
131 the results of running BEAST. The effective sample size was greater than 200 for each
132 parameter assessment in the MCMC analysis. Statistical uncertainty in the dataset was
133 estimated in the 95% HPD values.

134 **Genetic diversity and phylogenetic analysis**

135 The genetic diversity (π) of PPV was evaluated as average pairwise difference
136 between viral sequences, using the Tamura-Nei nucleotide substitution models
137 performed in MEGA 5 [35]. BioEdit was implemented to estimated the sequence
138 identity for different gene segments [30], and the heatmap was plotted by R program
139 pheatmap project. For each PPV gene, bayesian phylogenetic tree was inferred by
140 Bayesian Evolutionary Analysis Sampling Trees Software (BEAST) [36]. For all
141 analyses we performed a coalescent constant population model and the general-time
142 reversible model of nucleotide substitution [37]. Based on the correlation between
143 collection date and evolutionary distance for each gene, a strict molecular clock was
144 selected, as evaluated by TempEst [38]. Here, MCMC analysis was run for 10 million

145 steps, 10% of which was removed as burn-in and parameters sampled every 10000 steps.
146 The effective sample size for all the traces are greater than 200.

147 **Selection pressures in PPVs**

148 Selection pressure was highlighted as the ratio between the average number of non-
149 synonymous nucleotide substitutions (dN) and synonymous (dS) nucleotide
150 substitutions per site (dN/dS), and was implemented in the CODEML program within
151 the PAML software package. Besides, paired models of dN/dS were estimated among
152 nucleotide sequences, containing M0 versus M3, M2a versus M1a and M8 versus M7.
153 We applied the likelihood ratio test to compare the neutral and positive selection models,
154 and chi-square test to indicate the divergence between the compared models. We also
155 calculated the dN and dS values respectively.

156

157 **3. Results**

158 **G and C nucleotide was higher enriched in PPV genomes.**

159 To analyze the basic characteristic of virus genome, the nucleotide contents of PPV
160 whole genomes were calculated. It was shown that the mean compositions (%) of G
161 (32.16 ± 0.32) and C (32.12 ± 0.22) were significantly higher than A (17.95 ± 0.28) and
162 T (17.77 ± 0.25) (Table S2, Fig. 1(A), t-test, $P < 0.01$). The mean percentages of G3
163 (36.92 ± 1.02) and C3 (37.12 ± 1.33) were also significantly higher than A3 (20 ± 1.02)
164 and U3 (19.94 ± 0.89), highlighting that G- and C- ended codons are more likely
165 occurred in the PPV whole genome sequences (Table S2, Fig. 1(B), t-test, $P < 0.01$).
166 According to the species classification, it was important to mention that similar trend
167 of nucleotide composition was observed among different species, also suggesting
168 higher G/C or G3/C3 contents in PPV genomes. However, there were significant
169 differences in the nucleotide composition parameters trends among these four species
170 (Table S2, Fig. 1C-D, t-test, $P < 0.01$). As shown in Fig. 2, We further compared the
171 nucleotide composition among four different PPV species. Strikingly, there were highly
172 unusual patterns of nucleotide contents, and were significantly different from the
173 nucleotide composition patterns in the various PPV species.

174 **Different genome structures exist in various PPVs.**

175 According to the phylogenetic relationship and genomic structure, PPV genera
176 belongs to *Poxviridae* and includes four species: ORFV, PCPV, BPSV and PVNZ(Fig.
177 3). The genome of PPV contains large central protein-coding regions encoding
178 functional proteins, such as major envelope protein (B2L), chemokine binding protein
179 (CBP), immunodominant envelope protein (F1L), GM-CSF/IL-2 inhibition factor
180 (GIF), vascular endothelial growth factor (VEGF), viral interferon resistance protein
181 (VIR) and late transcription factor (VLTF). Besides these major structural proteins,
182 different PPVs encode some special accessory proteins, including Ankyrin protein
183 (ANK), IL-10-like protein (vIL-10) and poxviral anaphase promoting complex
184 regulator/ring H2 protein (PACR) (Fig. 3). This suggested that the right end of BPSV
185 genomes were similar with that of other PPV genomes, except for the missing of VEGF
186 existing in the left terminal region of BPSV genomes. It was also of note that PCPV
187 genomes contained one extra VEGF genes in the left end and lacked the vIL-10 gene,
188 while vIL-10 replaced VEGF in the left end of GQ329669.1/PCPV genome.
189 Additionally, PPVs contained more than one ANK genes that may involve in host range.

190 **Population dynamics of PPVs over the past 20 years**

191 In order to explore the change of PPV strains circulating in the world during the
192 past 20 years, 268 B2L gene sequences of PPVs were retrieved from GenBank in this
193 study (Table S1). All B2L isolates were clustered into separate clades on the basis of
194 their phylogenetic tree, suggesting that ORFV was the major species, which circulated
195 mainly in China and India (Fig. S1). In recent years, most of these viruses were from a
196 single species classified as ORFV (Fig. 4A). Consistent with the pattern observed in
197 the phylogenetic tree, the results of case counts also indicated that most of ORFVs were
198 isolated from China and India, and their prevalence increased sharply during 2013 –
199 2015 (Fig. 4B). Since 2010, ORFV has been the predominant species in circulation
200 throughout the world.

201 **ORFV of genera PPV became predominant species in recent years**

202 To describe the phylogenetic evolution of the prevalent PPVs, the phylogenetic
203 trees were constructed for the seven major structural segments. Phylogenetic trees were
204 partitioned into groups with a 20th percentile cutoff [39]. The PPV strains were not

205 clustered exactly consistently in each tree of seven segments (Fig. 5). Within each
206 prevalent clade, a large number of ORFVs further formed a main group with few PCPV,
207 BPSV and PVNZ (Fig. 5), demonstrating they may share a common ancestor surviving
208 a bottleneck event. Since 2010, the species diversity of PPVs was reduced with
209 widespread outbreak in many countries (Fig. 4 and Fig. 5), which can be reasonably
210 considered by high sequence similarity of PPVs isolated in the specific time span across
211 broad regions.

212 In order to confirm this hypothesis, we estimated the temporal dynamic of gene
213 sequence similarities of PPVs by collecting all available PPV sequences during 2000 –
214 2018. Pairwise identity comparison suggested a decreased diversity in all these seven
215 segments of PPVs, starting mainly in 2010 – 2018 (Fig. 6). At nearly the same time
216 point, the reduced sequence diversity in these major segments suggested that the virus
217 with great fitness was selected, acquired dominance, and led to a widespread outbreak
218 throughout the world. This is the first discovery, as we know, of such genetic bottleneck
219 in the PPV genomes (Fig. 6), despite their prevalence in the world for the last 20 years.

220 **Evolutionary rates and selection analysis of different gene sequences in PPVs**

221 To understand the additional forces affecting PPV evolution, we analyzed the
222 evolutionary rates and the average selective pressure of PPV strains whose isolation
223 date were known. According to the different protein-coding gene sequences, a Bayesian
224 coalescent approach was performed to infer the substitution rates of different segments.
225 Based on the best-fit model, Bayesian calculated the mean evolutionary rates of the
226 seven gene segments, ranging from 3.56×10^{-5} to 4.21×10^{-4} substitutions per site per
227 year (Table 1). The VEGF gene evolved at the slowest rate among the structural
228 segments, with the substitution rate of 3.56×10^{-5} and the highest probability density
229 (HPD) between 4.31×10^{-6} to 9.61×10^{-5} . The VIR gene evolved at the highest estimated
230 mean rate of 4.21×10^{-4} (95 % HPD: $1.28 \times 10^{-4} \sim 7.32 \times 10^{-4}$).

231 The selective pressure for each segment was performed with the phylogenetic
232 analysis by maximum likelihood program (PAML) based on the Model 0 (M0) within
233 codon-based phylogenetic models (CODEML). It supposed a constant substitution rate
234 of all sites and provided average values across all sites. The GIF and VIR genes were

235 found to be more variable, while the VLTF-1 gene was more conserved. On the basis
236 of Bayesian analysis, likelihood ratio tests were performed to compare the neutral
237 models and the positive selection models. To estimate the selective pressure on each
238 protein-coding segment, we applied a model (M3) for the differences among non-
239 synonymous and synonymous rates (dN/dS) of amino acid residues, and several neutral
240 models (M0, M1 and M7) with proportions of selected codons (M2 and M8) (Table 2).
241 The log likelihood values showed that positive selection models (M2, M3 and M8) were
242 more fitted in each gene regions than neutral models (M0, M1 and M7). The nested
243 comparisons were also performed between the neutral models and the positive models,
244 such as M0 vs M3, M1 vs M2 and M7 vs M8, and determined the better fitness of
245 positive models, indicating that except for the VLTF-1, positive selection pressure
246 occurred at different gene segments within the genome during the PPVs evolution
247 process (chi-square test, $P < 0.05$) (Table 2).

248

249 **4. Discussion**

250 PPV infections normally occur in animals and even in humans throughout the
251 world [11, 14, 16, 17, 40, 41]. However, comprehensive studies on PPVs are still lack.
252 In the present study, available whole genome sequences and different structural protein-
253 coding sequences were both utilized to investigate the structural and evolutionary
254 genomics of PPVs. In order to identify the common features and differences of PPVs,
255 it is important to analyze genomic patterns of all representative sequenced PPV
256 genomes. To start with, the general nucleotide contents were calculated for the genomes
257 of different PPV species have been shown in Table S2. The results suggested that the
258 PPV coding sequences were GC-rich, especially on the third location of synonymous
259 codons. This finding was consistent with previous researches wherein G and C
260 frequencies were higher than A and T frequencies for alphaherpesviruses,
261 betaherpesviruses, adenoviruses and Rubella virus (RUBV) [42, 43]. It has been known
262 that viruses could take on a wider range of GC compositions than other organisms [44,
263 45]. Even within the same family, viruses have similar replication or life-cycle
264 strategies, but may show different GC frequencies [46]. For instant, there were

265 significant differences of GC contents between ORFV and other three species of PPV,
266 while viruses showed almost identical nucleotide bias (Fig. 1, Fig. 2 and Table S2). This
267 supported the insight that there was common mechanism which determine GC
268 compositions for virus with similar life cycles.

269 These PPVs, although related, could be divided into four subgroups according to
270 the phylogenetic tree (Fig. 3), which contained ORFV, PCPV, BPSV and PVNZ,
271 respectively. To extend the genomic description of these four subgroups of PPVs, we
272 added the characterization of PPV genomes. PPV with large genomes (134~139 kb)
273 encodes nearly complete replisomes [4, 5]. Analysis of the PPV genomes suggested
274 conservation in four subgroups: genes encoding viral structural components (B2L,
275 ANK and FIL), those participated in virion assembly (VEGF), nucleotide metabolism
276 (PACR, CBP, GIF, vIL-10 and VIR), and DNA replication (VLTF). VEGF was known
277 to play an important role in PPV pathogenesis related to vascularization process and
278 epidermal lesion proliferation [6, 47]. Consistent with previous studies, we identified
279 that VEGF was located in the left terminal region of BPSV contrasting the right
280 terminal position of ORFV, PCPV and PVNZ VEGFs [6, 48], suggesting that the
281 divergence of BPSV VEGF may have distinct functions or binding specificities
282 compared with other VEGFs. Notably, PCPV was divergent from other PPVs with
283 deficiency of vIL-10 or left terminal vIL-10. This feature may indicate specific
284 adaptation to their natural hosts [6].

285 In addition to the genomic characteristics of PPVs, the phylogenetic diversity of
286 segmented genome of PPV may be a crucial contributor to understand the temporal and
287 spatial patterns of viral outbreaks [49, 50]. As shown in this study, the proportion of
288 ORFV strains have undergone a sharply increase in the past 20 years. Our results
289 suggested that PPVs evolved by mutation pressure and natural selection over 20 years
290 of prevalence in animals, and the predominant species, ORFV, was selected recently.
291 Since 2010, ORFV has rapidly increased in many countries, especially in China and
292 India, and is now the predominant PPV species in farmed sheep [51-56]. It indicated
293 that under the selective pressure of the optimal gene pattern by going through the
294 selective bottleneck the ORFV eventually became the dominant species during the 2010

295 – 2018 outbreak. Combining with the reduced diversity of the 2010 – 2018 PPVs and
296 the close phylogenetic clustering of their gene segments, these results demonstrated that
297 the observed changes of viruses were common in the recent PPVs.

298 Furthermore, according to the supposition of rate constancy, the differences of viral
299 isolation dates would provide the knowledge about the substitution rate of molecular
300 evolution. The previous study has suggested that the magnitude of evolutionary change
301 may accumulate since the isolation date [57]. A Bayesian coalescent approach
302 suggested the mean substitution rates were between 3.56×10^{-5} to 4.21×10^{-4} in different
303 gene segments, which was comparable with the studies about Newcastle disease virus,
304 hepatitis A and B virus, and infectious bronchitis virus [27, 58, 59]. The highest
305 substitution rate of PPV VIR gene reflected its less important role in viral synthesis and
306 replication process than other gene segments, which may suffer from great natural
307 selection pressure and subject to more substitutions.

308 When we analyzed the selection profiles of PPV protein-encoding genes, the
309 overall rates of dN/dS were mostly less than 1, except for the GIF and VIR,
310 demonstrating that purifying selection was the major force to drive the evolution of
311 PPVs. However, there were clear differences in this selection profiles of various genes.
312 Among these protein coding genes, GIF and VIR were under greater positive selection
313 than other gene segments. Except for VLTF-1, other structural protein coding genes all
314 had higher dN/dS values, highlighting that positive selection pressure has occurred in
315 most segments of PPVs. Viral fitness could exactly constrain the variability of genes,
316 which was important for virus replication [60]. Therefore, it was reasonable that VLTF-
317 1 mainly suffered from purifying selection in the evolutionary process of PPVs.
318 Additionally, as the angiogenic factor, the VEGF was essential to bind receptor tyrosine
319 kinases and influence embryonic development or tumor neovascularization, note that
320 low mutation rate during the viral evolution in host populations. These results indicated
321 that this selection pressure may improve viral fitness in their infected hosts.

322

323 **5. Conclusions**

324 In conclusion, this study indicated that PPVs share several common features within
325 different species. However, we noticed that GC contents were significantly different
326 between ORFV and other PPV species, while they showed almost identical nucleotide
327 bias. Besides, the genetic characteristics were also not similar among various PPV
328 species, a phenomenon that may be due to their specific adaptation to natural hosts. We
329 further investigated the evolutionary features of PPVs, and found that ORFV has been
330 the predominant PPV species and rapidly increased in many countries since 2010. PPV
331 isolates have been evolving at a sharply increased rate under the natural selection
332 pressure. Therefore, combining genetic mutation of PPVs, positive selection may
333 reshape the PPV genomes during the evolutionary process. The genetic analysis of
334 PPVs could extend our knowledge of the mechanisms that promote viral evolution.

335

336 **Author contributions:** DKC and WTM conceived and designed experiments; XTY
337 and MP performed all experiments. XTY, TXW and XC collected and analyzed the data.
338 XTY, MP and XDT drafted the manuscript. All authors read and approved the final
339 manuscript.

340

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342

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350

351 **REFERENCES**

- 352 1. Becher P, König M, Müller G, Siebert U, Thiel HJ: Characterization of sealpox virus, a
353 separate member of the parapoxviruses. *Arch Virol* 2002, 147(6):1133-1140.
- 354 2. Robinson AJ MA: Parapoxvirus of red deer: evidence for its inclusion as a new member in
355 the genus parapoxvirus. *Virology* 1995 Apr 20, 208(2):812-815.
- 356 3. D B: The Encyclopedia of Virology plus. *Trends Biochem Sci* 1997 Jan, 22(1):34-35.
- 357 4. Wittek R KC, Wyler R: High C + G content in parapoxvirus DNA. *J Gen Virol* 1979 Apr,
358 43(1):231-234.
- 359 5. Büttner M RH: Parapoxviruses: from the lesion to the viral genome. *J Vet Med B Infect Dis*
360 *Vet Public Health* 2002 Feb, 49(1):7-16.
- 361 6. Delhon G, Tulman ER, Afonso CL, Lu Z, de la Concha-Bermejillo A, Lehmkuhl HD, Piccone
362 ME, Kutish GF, Rock DL: Genomes of the parapoxviruses ORF virus and bovine papular
363 stomatitis virus. *J Virol* 2004, 78(1):168-177.
- 364 7. Olivero N, Reolon E, Arbiza J, Berois M: Genetic diversity of Orf virus isolated from sheep
365 in Uruguay. *Arch Virol* 2018, 163(5):1285-1291.
- 366 8. Inoshima Y MA, Sentsui H: Detection and diagnosis of parapoxvirus by the polymerase
367 chain reaction. *J Virol Methods* 2000 Feb, 84(2):201-208.
- 368 9. Hosamani M BV, Scagliarini A, Singh RK: Comparative sequence analysis of major
369 envelope protein gene (B2L) of Indian orf viruses isolated from sheep and goats. *Vet*
370 *Microbiol* 2006 Sep 10, 116(4):317-324.
- 371 10. Chan KW LJ, Lee SH, Liao CJ, Tsai MC, Hsu WL, Wong ML, Shih HC: Identification and
372 phylogenetic analysis of orf virus from goats in Taiwan. *Virus Genes* 2007 Dec, 35(3):705-
373 712.
- 374 11. Chakhunashvili G, Carlson BF, Power L, Khmaladze E, Tsaguria D, Gavashelidze M,
375 Zakhshvili K, Imnadze P, Boulton ML: Parapoxvirus Infections in the Country of Georgia:
376 A Case Series. *Am J Trop Med Hyg* 2018, 98(6):1870-1875.
- 377 12. Hosamani M SA, Bhanuprakash V, McInnes CJ, Singh RK: Orf: an update on current
378 research and future perspectives. *Expert Rev Anti Infect Ther* 2009 Sep, 7(7):879-893.
- 379 13. de Sant'Ana FJ RR, Vulcani VA, Cargnelutti JF, Flores EF: Bovine papular stomatitis affecting
380 dairy cows and milkers in midwestern Brazil. *J Vet Diagn Invest* 2012 Mar, 24(2):442-445.
- 381 14. Nougairede A FC, Salez N, Cohen-Bacrie S, Ninove L, Michel F, Aboukais S, Buttner M,
382 Zandotti C, de Lamballerie X, Charrel RN: Sheep-to-human transmission of Orf virus
383 during Eid al-Adha religious practices, France. *Emerg Infect Dis* 2013 Jan, 19(1):102-105.
- 384 15. Kitchen M MH, Zobl A, Windisch A, Romani N, Huemer H: ORF virus infection in a hunter
385 in Western Austria, presumably transmitted by game. *Acta Derm Venereol* 2014 Mar,
386 94(2):212-214.
- 387 16. Osadebe LU MK, McCollum AM, Li Y, Emerson GL, Gallardo-Romero NF, Doty JB, Wilkins
388 K, Zhao H, Drew CP, Metcalfe MG, Goldsmith CS, Muehlenbachs A, Googe PB, Dunn J,
389 Duenckel T, Henderson H, Carroll DS, Zaki SR, Denison MR, Reynolds MG, Damon IK:
390 Novel poxvirus infection in 2 patients from the United States. *Clin Infect Dis* 2015 Jan 15,
391 60(2):195-202.
- 392 17. Midilli K EA, Kuşkuç M, Analay H, Erkiliç S, Benzonana N, Yildirim MS, Mülayim K, Acar H,
393 Ergonul O: Nosocomial outbreak of disseminated orf infection in a burn unit, Gaziantep,
394 Turkey, October to December 2012. *Euro Surveill* 2013 Mar 14, 18(11):20425.

- 395 18. Tedla M, Berhan N, Molla W, Temesgen W, Alemu S: Molecular identification and
396 investigations of contagious ecthyma (Orf virus) in small ruminants, North west Ethiopia.
397 *BMC Vet Res* 2018, 14(1):13.
- 398 19. Mazur C MR: Detection of contagious pustular dermatitis virus of goats in a severe
399 outbreak. *Vet Rec* 1989 Oct 14, 125(16):419-420.
- 400 20. McKeever DJ JD, Hutchison G, Reid HW: Studies of the pathogenesis of orf virus infection
401 in sheep. *Journal of comparative pathology* 1988 Oct, 99(3):317-328.
- 402 21. de Sant'Ana FJ LF, Rabelo RE, Vulcani VA, Moreira CA Jr, Cargnelutti JF, Flores EF:
403 Coinfection by Vaccinia virus and an Orf virus-like parapoxvirus in an outbreak of vesicular
404 disease in dairy cows in midwestern Brazil. *J Vet Diagn Invest* 2013 Mar, 25(2):267-272.
- 405 22. Schmidt C CJ, Brum MC, Traesel CK, Weiblen R, Flores EF: Partial sequence analysis of B2L
406 gene of Brazilian orf viruses from sheep and goats. *Vet Microbiol* 2013 Feb 22, 162(1):245-
407 253.
- 408 23. Abrahão JS BI, Mazur C, Lobato ZI, Ferreira PC, Bonjardim CA, Trindade GS, Kroon EG:
409 Looking back: a genetic retrospective study of Brazilian Orf virus isolates. *Vet Rec* 2012
410 Nov 10, 171(19):476.
- 411 24. Cargnelutti JF FM, Teixeira FR, Weiblen R, Flores EF: An outbreak of pseudocowpox in
412 fattening calves in southern Brazil. *J Vet Diagn Invest* 2012 Mar, 24(2):437-441.
- 413 25. de Oliveira CH AF, Neto JD, Oliveira CM, Lopes CT, Bomjardim Hdos A, Vinhote WM, Silva
414 AG, Abrahão JS, Kroon EG: Multifocal cutaneous ORF virus infection in goats in the
415 Amazon region, Brazil. *Vector Borne Zoonotic Dis* 2012 Apr, 12(4):336-340.
- 416 26. Petrova VN, Russell CA: The evolution of seasonal influenza viruses. *Nat Rev Microbiol*
417 2018, 16(1):47-60.
- 418 27. Zhao Y, Zhang H, Zhao J, Zhong Q, Jin JH, Zhang GZ: Evolution of infectious bronchitis
419 virus in China over the past two decades. *J Gen Virol* 2016, 97(7):1566-1574.
- 420 28. Matsumoto H, Setoyama H, Matsuura Y, Ohtani A, Shimizu K, Okada A, Inoshima Y:
421 Sequential detection of pseudocowpox virus and bovine papular stomatitis virus in a same
422 calf in Japan. *J Vet Med Sci* 2019, 81(3):440-443.
- 423 29. RC E: MUSCLE: multiple sequence alignment with high accuracy and high throughput.
424 *Nucleic Acids Res* 2004 Mar 19, 32(5):1792-1797.
- 425 30. Hall TA: BioEdit: a user-friendly biological sequence alignment editor and analysis
426 program for Windows 95/98/NT. *Nucl Acids Symp Ser* 1999, 41:95-98.
- 427 31. Letunic I BP: Interactive Tree Of Life v2: online annotation and display of phylogenetic
428 trees made easy. *Nucleic Acids Res* 2011 Jul, 39(Web Server
429 issue):W475-478.
- 430 32. Ito K MD: Application of ggplot2 to Pharmacometric Graphics. *CPT Pharmacometrics Syst*
431 *Pharmacol* 2013 Oct 16, 2:e79.
- 432 33. Darriba D, Taboada GL, Doallo R, Posada D: jModelTest 2: more models, new heuristics
433 and parallel computing. *Nat Methods* 2012, 9(8):772.
- 434 34. Newton MAR, A. E: Approximate bayesian inference with the weighted likelihood
435 bootstrap. *J Roy Statist Soc Ser B* 1994, 56:3-48.
- 436 35. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S: MEGA5: molecular
437 evolutionary genetics analysis using maximum likelihood, evolutionary distance, and
438 maximum parsimony methods. *Mol Biol Evol* 2011, 28(10):2731-2739.

- 439 36. Bouckaert R, Heled J, Kuhnert D, Vaughan T, Wu CH, Xie D, Suchard MA, Rambaut A,
440 Drummond AJ: BEAST 2: a software platform for Bayesian evolutionary analysis. *PLoS*
441 *Comput Biol* 2014, 10(4):e1003537.
- 442 37. Shapiro B, Rambaut A, Drummond AJ: Choosing appropriate substitution models for the
443 phylogenetic analysis of protein-coding sequences. *Mol Biol Evol* 2006, 23(1):7-9.
- 444 38. Rambaut A, Lam TT, Max Carvalho L, Pybus OG: Exploring the temporal structure of
445 heterochronous sequences using TempEst (formerly Path-O-Gen). *Virus Evol* 2016,
446 2(1):vew007.
- 447 39. Prosperi MC CM, Fanti I, Saladini F, Pecorari M, Borghi V, Di Giambenedetto S, Bruzzone
448 B, Capetti A, Vivarelli A, Rusconi S, Re MC, Gismondo MR, Sighinolfi L, Gray RR, Salemi M,
449 Zazzi M, De Luca A; ARCA collaborative group: A novel methodology for large-scale
450 phylogeny partition. *Nat Commun* 2011, 2:321.
- 451 40. de Sant'Ana FJ RR, Vulcani VA, Cargnelutti JF, Flores EF: Bovine papular stomatitis affecting
452 dairy cows and milkers in midwestern Brazil. *J Vet Diagn Invest* 2012, 24(2):442-445.
- 453 41. Kitchen M MH, Zobl A, Windisch A, Romani N, Huemer H: ORF virus infection in a hunter
454 in Western Austria, presumably transmitted by game. *Acta Derm Venereol* 2014,
455 94(2):212-214.
- 456 42. Shackelton LA, Parrish CR, Holmes EC: Evolutionary basis of codon usage and nucleotide
457 composition bias in vertebrate DNA viruses. *J Mol Evol* 2006, 62(5):551-563.
- 458 43. Zhou Y CX, Ushijima H, Frey TK: Analysis of base and codon usage by rubella virus. *Arch*
459 *Viro* 2012 May, 157(5):889-899.
- 460 44. Bronson EC AJ: Nucleotide composition as a driving force in the evolution of retroviruses.
461 *J Mol Evol* 1994 May, 38(5):506-532.
- 462 45. Shackelton LA PC, Holmes EC: Evolutionary basis of codon usage and nucleotide
463 composition bias in vertebrate DNA viruses. *J Mol Evol* 2006 May, 62(5):551-563.
- 464 46. Schachtel GA BP, Mocarski ES, Blaisdell BE, Karlin S: Evidence for selective evolution in
465 codon usage in conserved amino acid segments of human alphaherpesvirus proteins. *J*
466 *Mol Evol* 1991 Dec, 33(6):483-494.
- 467 47. Savory LJ SS, Fleming SB, Niven BE, Mercer AA: Viral vascular endothelial growth factor
468 plays a critical role in orf virus infection. *J Virol* 2000 Nov, 74(22):10699-10706.
- 469 48. Rziha HJ BB, Adam KH, Röttgen M, Cottone R, Henkel M, Dehio C, Büttner M: Relatedness
470 and heterogeneity at the near-terminal end of the genome of a parapoxvirus bovis 1
471 strain (B177) compared with parapoxvirus ovis (Orf virus). *J Gen Virol* 2003 May, 84(Pt
472 5):1111-1116.
- 473 49. Perrin A, Larssonneur E, Nicholson AC, Edwards DJ, Gundlach KM, Whitney AM, Gulvik CA,
474 Bell ME, Rendueles O, Cury J *et al*: Evolutionary dynamics and genomic features of the
475 Elizabethkingia anophelis 2015 to 2016 Wisconsin outbreak strain. *Nat Commun* 2017,
476 8:15483.
- 477 50. Pu J, Wang S, Yin Y, Zhang G, Carter RA, Wang J, Xu G, Sun H, Wang M, Wen C *et al*:
478 Evolution of the H9N2 influenza genotype that facilitated the genesis of the novel H7N9
479 virus. *Proc Natl Acad Sci U S A* 2015, 112(2):548-553.
- 480 51. Ahanger SA PR, Nazki S, Dar Z, Dar T, Dar KH, Dar A, Rai N, Dar P: Detection and
481 phylogenetic analysis of Orf virus in Kashmir Himalayas. *Virusdisease* 2018 Sep, 29(3):405-
482 410.

- 483 52. Bala JA BK, Abdullah AA, Mohamed R, Haron AW, Jesse FFA, Noordin MM, Mohd-Azmi
484 ML: The re-emerging of orf virus infection: A call for surveillance, vaccination and effective
485 control measures. *Microb Pathog* 2018 Jul, 120:55-63.
- 486 53. Bora M BD, Barman NN, Borah B, Bora PL, Talukdar A, Tamuly S: Isolation and molecular
487 characterization of Orf virus from natural outbreaks in goats of Assam. *Virusdisease* 2015
488 Jun, 26(1-2):82-88.
- 489 54. Chan K W LJW, Lee S H, Liao C J, Tsai M C, Hsu W L, Wong M L, Shih H C: Identification
490 and phylogenetic analysis of orf virus from goats in Taiwan. *Virus Genes* 2007, 35 (3):705-
491 712.
- 492 55. Flores C GE, Verna A, Peralta A, Madariaga C, Odeón A, Cantón G: Orf virus in human,
493 confirmation in case report from Chile. *Rev Chilena Infectol* 2017 Dec, 34(6):607-609.
- 494 56. M Ş: Association of two clusters of Orf virus isolates in outbreaks of infection in goat in
495 the Central Anatolian region of Turkey. *Virusdisease* 2017 Sep, 28(3):345-348.
- 496 57. Soylemez O, Kondrashov FA: Estimating the rate of irreversibility in protein evolution.
497 *Genome Biol Evol* 2012, 4(12):1213-1222.
- 498 58. Wang H WX, Zheng HH, Cao JY, Zhou WT, Bi SL: Evolution and genetic characterization
499 of hepatitis A virus isolates in China. *Int J Infect Dis* 2015 Apr, 33:156-158.
- 500 59. Alvarado Mora MV RC, Gomes-Gouvêa MS, Gutierrez MF, Botelho L, Carrilho FJ, Pinho JR:
501 Molecular characterization of the Hepatitis B virus genotypes in Colombia: a Bayesian
502 inference on the genotype F. *Infect Genet Evol* 2011 Jan, 11(1):103-108.
- 503 60. Soderholm J, Ahlen G, Kaul A, Frelin L, Alheim M, Barnfield C, Liljestrom P, Weiland O,
504 Milich DR, Bartenschlager R *et al*. Relation between viral fitness and immune escape within
505 the hepatitis C virus protease. *Gut* 2006, 55(2):266-274.

506

507 Fig. 1. Nucleotide content distribution and composition. (A) The mean frequency for A, T, G and C
508 composition in 17 different PPV sequences are shown, suggests higher GC content than AT on the
509 whole. (B) The mean values of the nucleotide content frequency at the 3rd codon position. (C) Based
510 on 4 various species of PPVs, analysis for the overall A, T, G and C composition frequencies. (D)
511 Based on 4 various species of PPVs, analysis for A, T, G and C composition frequencies at the third
512 codon position. Different asterisks represent statistical significance (* $p < 0.05$, ** $p < 0.01$, *** $p <$
513 0.001 , **** $p < 0.0001$).

514

515 Fig. 2. Nucleotide composition of PPVs for different species. Frequency of each observed nucleotide
516 composition, reconstructed in BioEdit (v7.2.5), is shown for different species of PPVs.

517

518 Fig. 3. Phylogenetic analysis and genome comparison among different species of PPVs. ML
519 phylogenetic tree was reconstructed in phyML (v3.0) and tested with 1000 bootstraps. The predicted
520 genes are represented by block arrows showing the direction of transcription. The genes encoding

521 for structural proteins and replication-associated proteins of PPVs are shown in different colors.

522

523 Fig. 4. Population dynamics of PPVs during the past 20 years. Case counts are presented in the plot.
524 The X-axis represents the year intervals, and the Y-axis represents the number of cases. Different
525 colors represent different species (A) and different collection countries of PPVs (B).

526

527 Fig. 5. Phylogenetic analysis of seven structural genes with PPVs during 2000 – 2018. Clades with
528 all of 2000 – 2018 viruses were fully shown. Color of line at right of each leaf node indicates year
529 of isolation (see color bar). Timescale is in years. Vertical lines mark different species of PPVs: gray
530 represents ORFV, purple represents BPSV, black represents PCPV and yellow represents PVNZ.

531

532 Fig. 6. Decreased genetic diversity of the seven segments of PPVs during the widespread outbreaks
533 in the world (2000 – 2018). Pairwise comparison of nucleotide sequences of PPVs was plotted as a
534 heatmap. Viruses isolated from 2000 through 2018 were ordered by isolation date from left to right
535 of the x axis and from top to bottom of the y axis. On the axes, the ticks indicated the isolation years.
536 Black dotted line represented the year 2010. Color indicated identity levels from $\leq 30\%$ (blue) to
537 100% (red).

538

539 Fig. S1. The phylogenetic tree of PPV B2L genes. The tree was generated by the maximum
540 likelihood (ML) method using PAML v4.9. The tree was designed by using the online tool “iTOL”.
541 Here; different colors represent different collection locations. The color of each leaf node represents
542 different virus species.

543

Figures

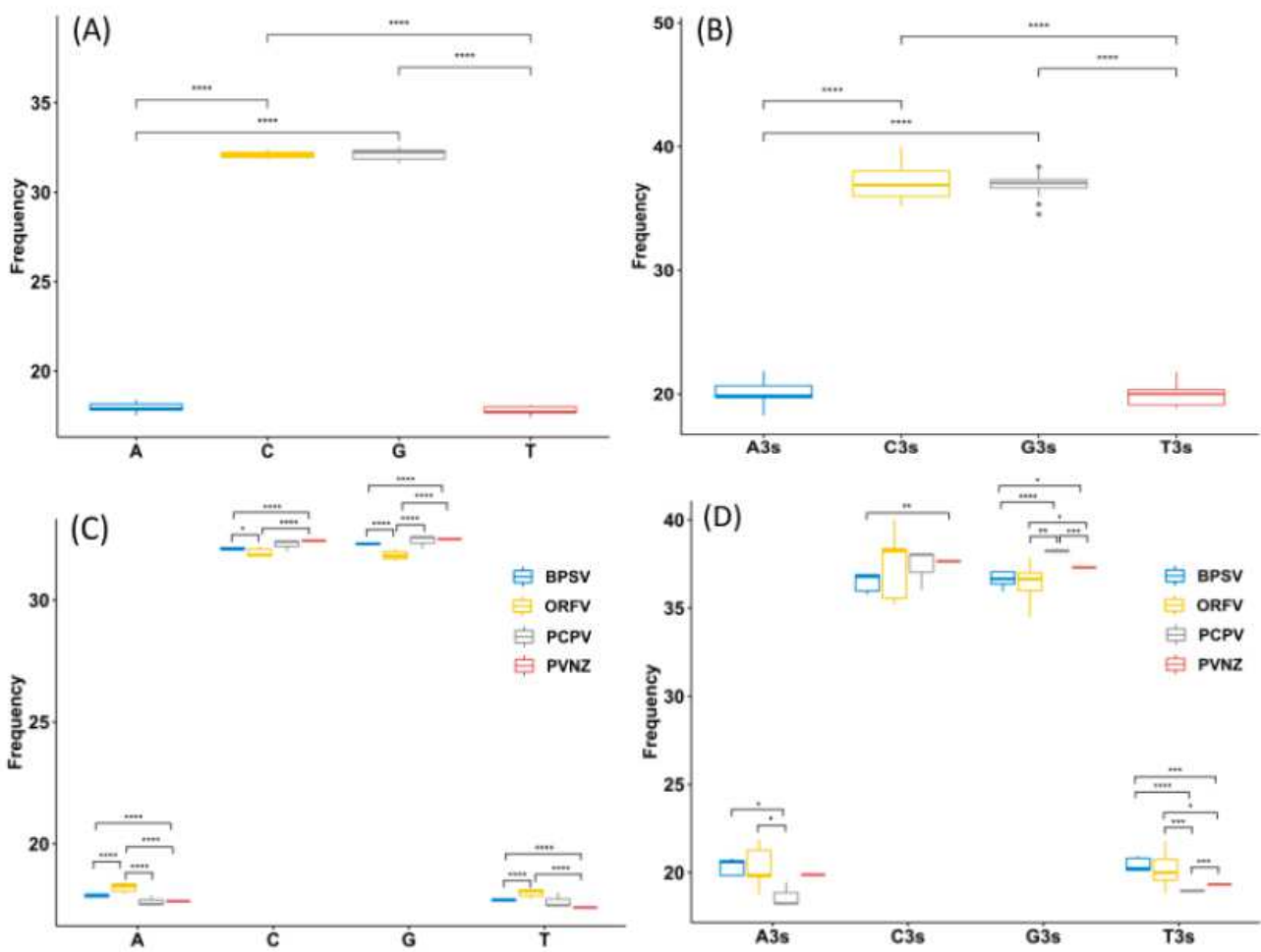


Figure 1

Nucleotide content distribution and composition. (A) The mean frequency for A, T, G and C composition in 17 different PPV sequences are shown, suggests higher GC content than AT on the whole. (B) The mean values of the nucleotide content frequency at the 3rd codon position. (C) Based on 4 various species of PPVs, analysis for the overall A, T, G and C composition frequencies. (D) Based on 4 various species of PPVs, analysis for A, T, G and C composition frequencies at the third codon position. Different asterisks represent statistical significance (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$).

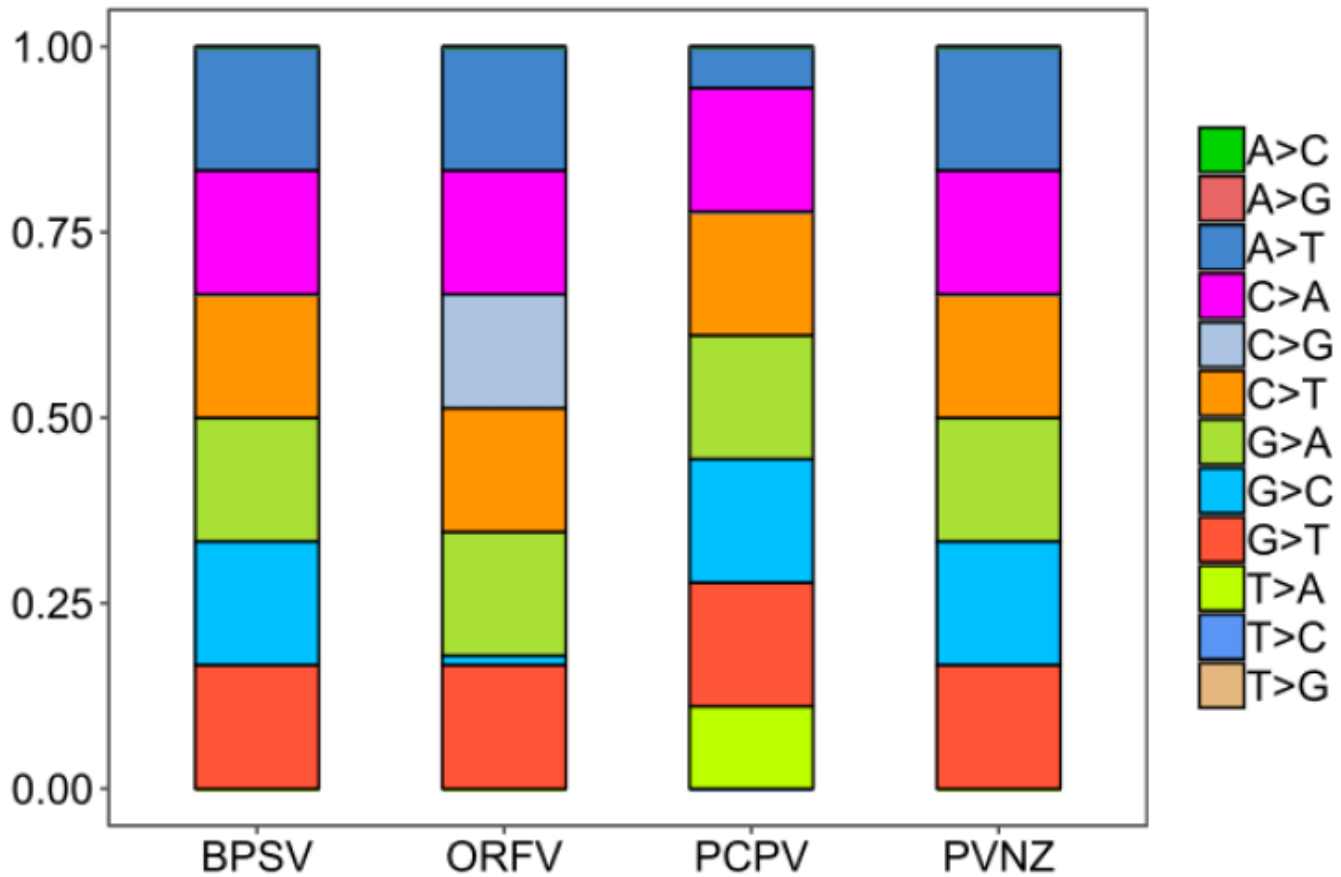


Figure 2

Nucleotide composition of PPVs for different species. Frequency of each observed nucleotide composition, reconstructed in BioEdit (v7.2.5), is shown for different species of PPVs.

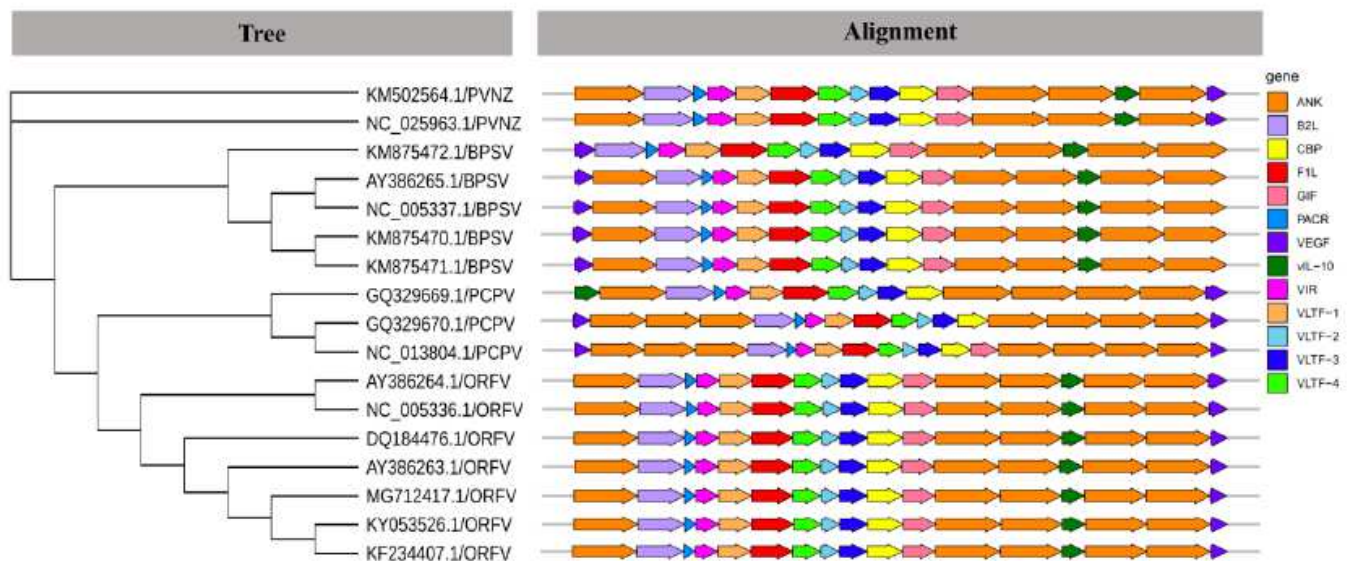


Figure 3

Phylogenetic analysis and genome comparison among different species of PPVs. ML phylogenetic tree was reconstructed in phyML (v3.0) and tested with 1000 bootstraps. The predicted genes are represented by block arrows showing the direction of transcription. The genes encoding for structural proteins and replication-associated proteins of PPVs are shown in different colors.

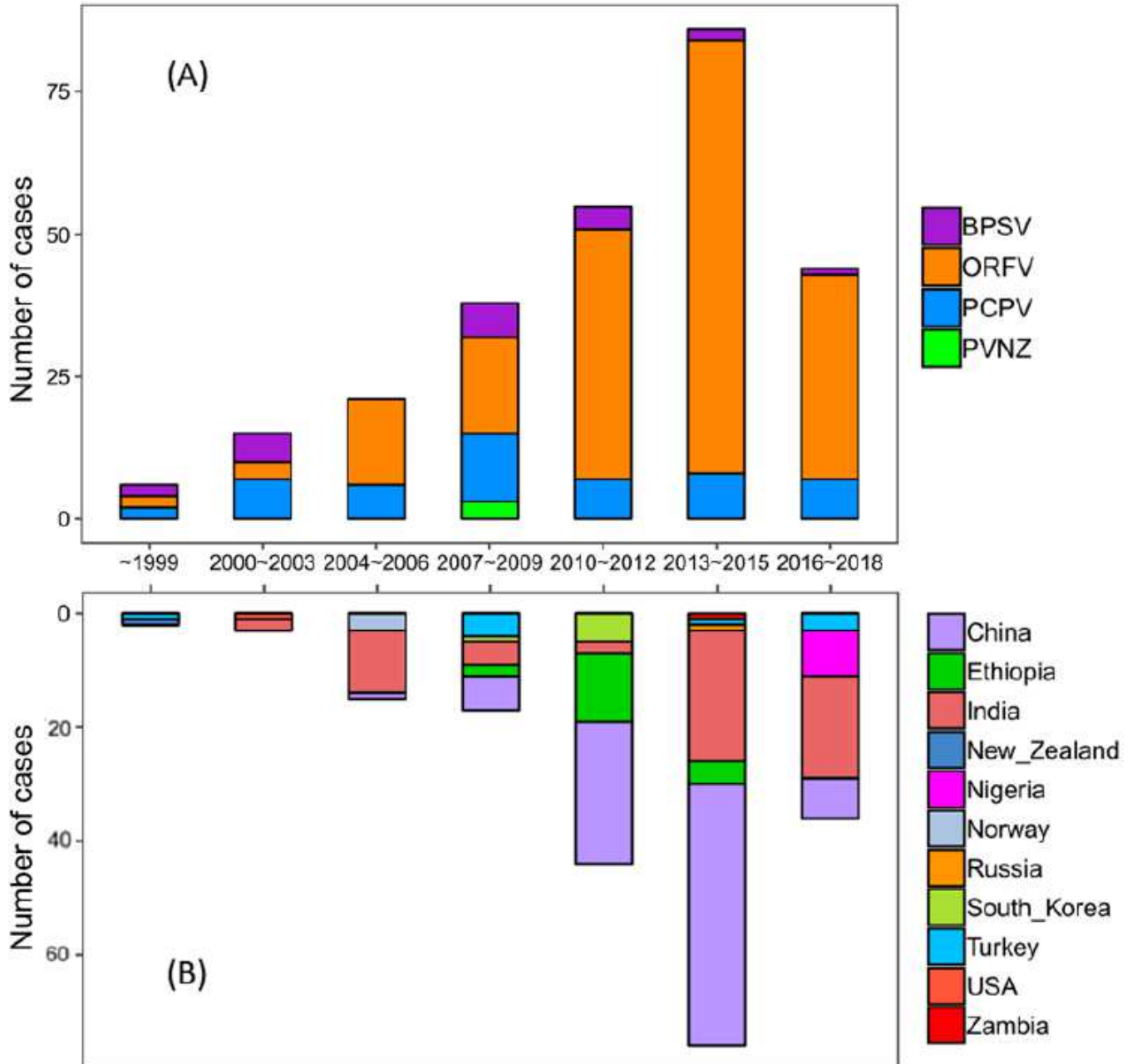


Figure 4

Population dynamics of PPVs during the past 20 years. Case counts are presented in the plot. The X-axis represents the year intervals, and the Y-axis represents the number of cases. Different colors represent different species (A) and different collection countries of PPVs (B).

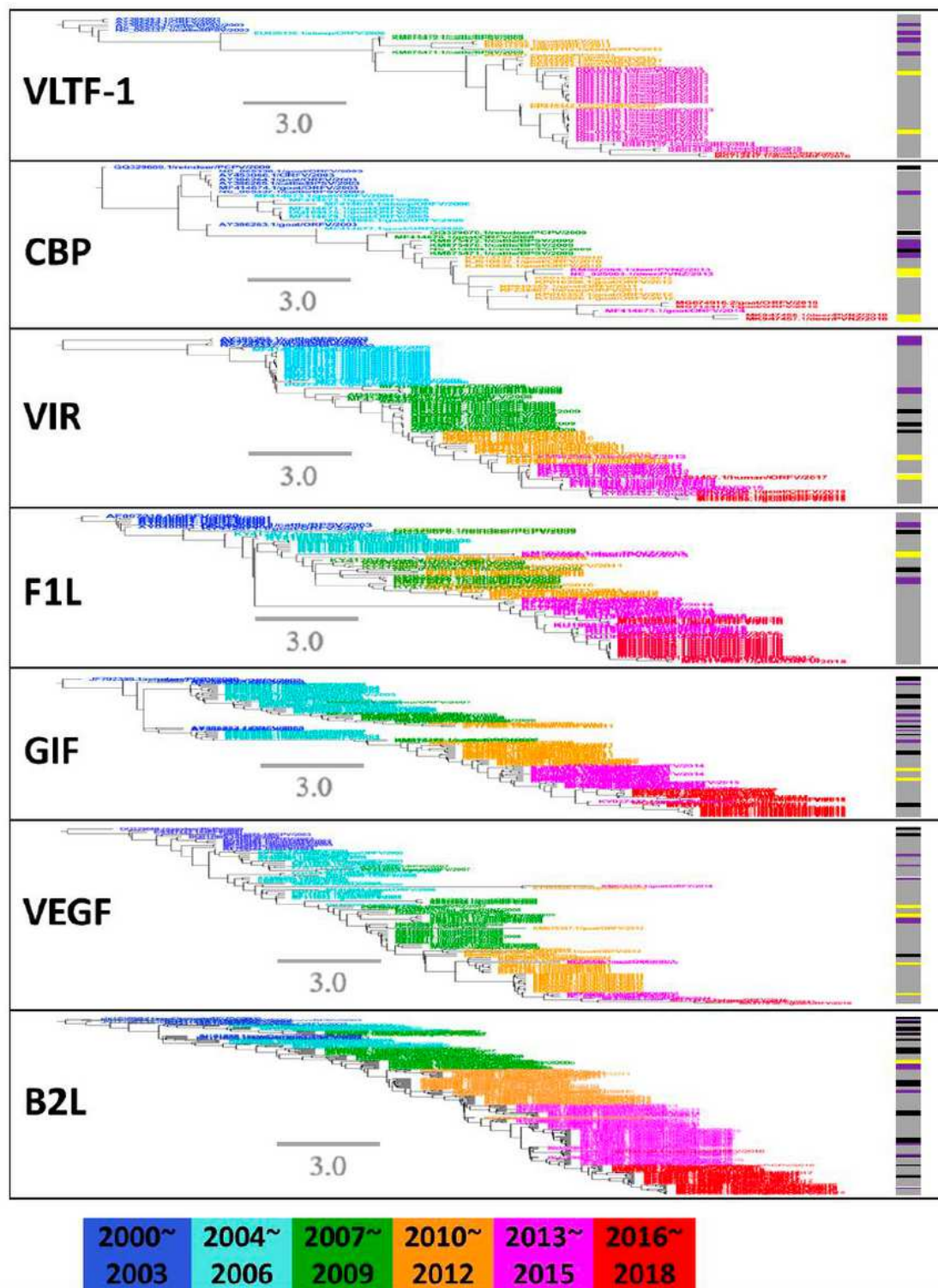


Figure 5

Phylogenetic analysis of seven structural genes with PPVs during 2000 – 2018. Clades with all of 2000 – 2018 viruses were fully shown. Color of line at right of each leaf node indicates year of isolation (see

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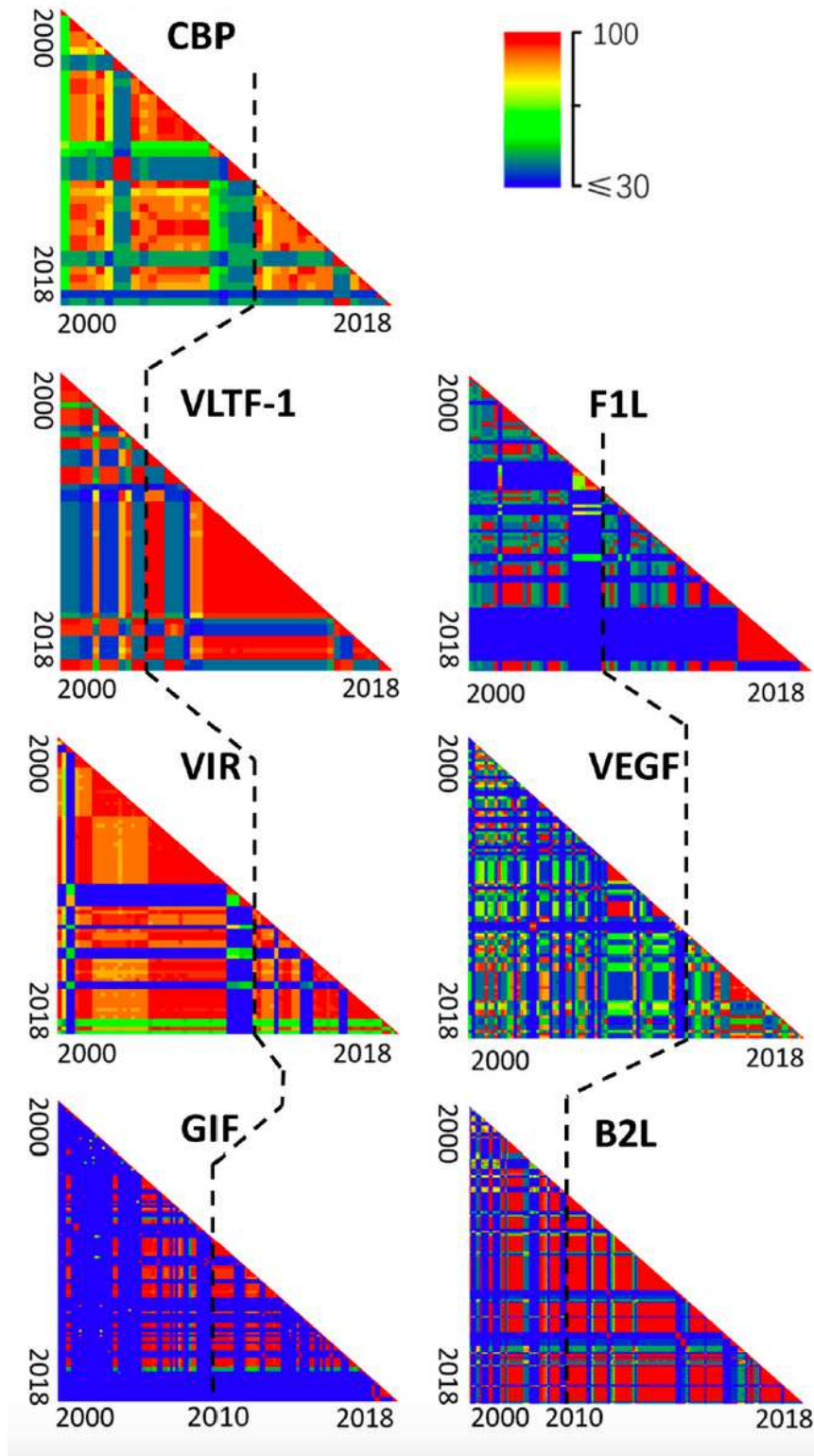


Figure 6

Decreased genetic diversity of the seven segments of PPVs during the widespread outbreaks in the world (2000 – 2018). Pairwise comparison of nucleotide sequences of PPVs was plotted as a heatmap. Viruses isolated from 2000 through 2018 were ordered by isolation date from left to right of the x axis and from

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