

The Effect of Some Soil Amendments (Manure and Biochar) on the Bioaccumulation Capacity of Hexavalent Chromium by Two Species of (*Salicornia Persica*) and (*Salicornia Perspolitana*) From Contaminated Soil

Fahime Ashrafi

Ferdowsi University of Mashhad

Ava heidari (✉ ava.heidari@yahoo.com)

Ferdowsi University of Mashhad

Mohammad Farzam

Ferdowsi University of Mashhad

Alireza Karimi

Ferdowsi University of Mashhad

Malihe Amini

University of Jiroft

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1 **The effect of some soil Amendments (manure and biochar) on the**
2 **Bioaccumulation capacity of Hexavalent Chromium by two species of**
3 **(*Salicornia persica*) and (*Salicornia perspolitana*) from Contaminated soil**
4

5 Fahime Ashrafi¹, Ava Heidari^{1*}, Mohammad Farzam², Alireza Karimi³, Malihe Amini^{4†}

6 ¹Department of Environmental Science, Faculty of Natural Resources and Environment, Ferdowsi University of Mashhad, Mashhad, Iran

7 ²Department of Range and Watershed Management, Faculty of Natural Resources and Environment, Ferdowsi University of Mashhad, Mashhad,
8 Iran

9 ³Department of Soil Sciences, Faculty of Agriculture, Ferdowsi University of Mashhad, Iran

10 ⁴Department of Environmental Science and Engineering, Faculty of Natural Resources, University of Jiroft, P.O. Box: 8767161167, Jiroft,
11 kerman, Iran

12
13 **Abstract:** Heavy metals are among the most dangerous contaminants in the environment.
14 Application of organic compounds and plant species with the ability to accumulate and stabilize
15 heavy metal in their organs is the best option for remediation of these elements in the soil.
16 Therefore, this study aimed to investigate the effects of manure and biochar on the accumulation
17 of heavy metals by *Salicornia* species. Two species of *Salicornia* ,including *S. persica* and *S.*
18 *perspolitana*, were cultivated outdoor in experimental pots. The effects of experimental
19 treatments ,including hexavalent chromium concentrations, manure ,and biochar on the two
20 studied species, were investigated. The results indicated a significant effect (P <0.05) of biochar
21 on the accumulation of heavy metals by two species ,*S. persica*and *S. perspolitana*, so that
22 chromium concentrations in the roots and shoots were 258 and 5.41 mg/kg, respectively. Also,
23 chromium accumulations under manure treatments in the roots and shoots were 334.34 and 9.79
24 mg/kg, respectively. Plant dry weight and height for both species in manure treatment were
25 higher than control and biochar treatments. *S. persica* showed higher growth than *S. perspolitana*
26 species. The content of photosynthetic pigments in both *S.persica* and *S. perspolitana* species

* Corresponding author. heidari@um.ac.ir, ava.heidari@yahoo.com, Phone number: +9838805471

† Co- Corresponding author: m.amini@ujiroft.ac.ir

27 under biochar treatment was higher than control and manure treatments. In general, one can
28 conclude that the accumulation of chromium in *S. perspolitana* was higher than in *S. persica* ,and
29 the application of biochar and manure amendments could stabilize chromium in soil and reduce
30 chromium accumulation in both *S. persica* and *S. perspolitana* species.

31 **Keywords:** Soil contamination, *Salicornia*, phytoremediation, chromium.

32 **1. Introduction**

33 Increased agricultural activities , as well as industrialization in recent decades, have significantly
34 resulted in the accumulation of various contaminants in the environment, especially in soil and
35 water. These contaminants enter into the environment through mining operations, discharge of
36 industrial effluents, pesticide-based agriculture, fertilizers, etc. , and are challenging and
37 problematic due to harmful effects on soil biological systems (Edao 2017). Removal of heavy
38 metals from soil could be a severe issue as heavy metals are special contaminants, which in
39 many cases, could remain in the soil for hundreds (even thousands) of years. Several methods
40 have been documented for removing heavy metals from contaminated soils. Despite being
41 efficient, conventional technologies are costly, time-consuming , and environmentally
42 destructive. Phytoremediation is defined as applying of various plants to reduce the
43 concentration of heavy metals in the soil. Generally, bioremediation has been considered as a
44 cost-effective and environmentally friendly method (Yao et al. 2012). Species store and
45 accumulate high concentrations of heavy metals in their tissues without being poisoned by that
46 metals. During the phytoremediation process, metals are effectively removed from the soil. The
47 efficiency of the phytoremediation process is dependent on the plant functioning as well as the
48 effective transfer of metals from the plant roots to the shoots. Plants that accumulate
49 considerable amounts of metal usually have small shoots, so the critical point for increasing the

50 efficiency of the phytoremediation process is how to change plant biomass to enhance metal
51 uptake (Evangelou et al. 2007). Biochar is a porous carbon product obtained from the pyrolysis
52 of plant-derived organic matter (tree bark, rice husk, pinewood, etc.) or non-plant-derived
53 biomass (cattle manure, poultry manure, etc.) with high cation exchangeable capacity and
54 alkaline properties that improve soil structure. The addition of biochar to soil leads to increased
55 soil biological activities , thereby improved crop yield(Bashir et al. 2018). In addition to biochar,
56 applying of some other materials such as animal manure in soils could also increase the
57 phytoremediation efficiency. Organic fertilizers, especially animal manures, contain large
58 amounts of organic matter compared to chemical ones , which can provide nutrient sources such
59 as nitrogen, phosphorus , and potassium overtime for the plant and improve soil physical,
60 chemical , and biological properties. Increased organic matter in soils leads to the formation of
61 aggregates and improves soil moisture-holding capacity, hydraulic conductivity, bulk density,
62 degree of compaction, fertility , and soil resistance to water and wind erosion (Zebarth et al.
63 1999). A study by Al-Wabe et al.(Al-Wabel et al. 2015) showed that the application of biochar
64 improved the growth of maize (*Zea mays* L.) and increased the soil water holding capacity of the
65 soil. Biochar also leads to alkaline properties in the soil , and this effect varies depending on
66 biochar characteristics, production temperature , and its raw materials.

67 Few plant species, including *Sutera fodina*, *Dicoma niccolifera*, and *Leptospermum scoparium*,
68 have been reported to have the ability to accumulate high concentrations of chromium in their
69 tissues. Also, mustard (*Brassica juncea*) and sunflower (*Helianthus annus* L.) have been reported
70 to accumulate high concentrations of chromium in their tissues(Shahandeh &Hossner 2000).
71 Coupe et al.(Coupe et al. 2013) compared the ability of three plant species , namely *Eucalyptus*
72 *camaldeulensis*, *Brassica juncea* , and *Medicago sativum* , to uptake copper, zinc , and

73 chromium from soil and stated that *Eucalyptus camaldealensis* had the highest capability to
74 uptake these contaminants from the soil. *Salicornia*, a halophyte and salt-tolerant plant, serves as
75 a suitable plant for phytoremediation of heavy metals due to its characteristic roots and the
76 ability to stabilize metals (Van Oosten et al. 2015).

77 Several studies have been carried out on the phytoremediation ability of *Salicornia* plant species
78 for removal of heavy metals other than chromium, but for the first time, we aimed to investigate
79 the ability of *Salicornia persica* and *Salicornia perspolitana* in phytoremediation of chromium-
80 contaminated soils. To do this, we studied the effect of manure and biochar on the capacity of the
81 two mentioned species to accumulate chromium in their tissues from contaminated soil. In this
82 regard, experimental treatments including hexavalent chromium concentrations, manure , and
83 biochar.

84 **2. Materials and methods:**

85 *2.1 Experiments*

86 All steps of this research were performed in 2019-2020 in the laboratory and growth chamber of
87 Faculty of Natural Resources and Environment, the Ferdowsi University of Mashhad, Iran,
88 located at 25 km apart from northeast of Mashhad city (59° 31' 16" E and 36° 29' 92" N). The
89 soil samples were taken from a 0-40 cm and passed through a 2-mm sieve after air-drying.

90 *2.2 Determination of soil physical and chemical properties*

91 Soil texture was characterized by the hydrometric method using a soil texture triangle. Soil pH
92 and EC (electrical conductivity) were measured using pH-meter (20+, Crison, Spain) and EC-
93 meter (4510, Jenway, England) devices, respectively. Soil organic carbon was determined by the
94 dry combustion method (Park et al. 2017). Total nitrogen was measured using a Kjeldal (V50)
95 device. Soil available phosphorus was measured according to the Olsen method (1954) using a

96 spectrophotometer (model DR 5000) at the wavelength of 660 nm. Available potassium was
97 determined using the ammonium acetate method and photoelectric flame photometer (Jenway
98 PFP7).

99 *2.3 Determination of physical and chemical properties of manure and biochar*

100 The completely rotten manure used in this study was prepared from the greenhouse of Ferdowsi
101 University of Mashhad, Iran. The applied biochar was a mixture of the wood obtained from
102 eucalyptus, poplar , and ironwood prepared at 500° C at the laboratory of Tarbiat Modares
103 University, Iran. Before performing the experimental treatments, manure and biochar samples
104 were analyzed using conventional laboratory methods previously applied to measure the soil
105 samples. Calcium and sodium contents were determined by inductively coupled plasma
106 spectroscopy (ICP-OES) (model ICP-OES, SPECTRO ARCOS-76004555) and the percentages
107 (%) of carbon, nitrogen, hydrogen, oxygen , and sulfur in the biochar using elemental analysis
108 (FLASH EA 1112 SERIES, Thermo Finnigan).

109 *2.4 Preparation of soil samples*

110 The soil in the natural habitat was characterized by clay-sandy loam texture and an EC of 1800
111 $\mu\text{s}/\text{cm}$, and we prepared the soil samples containing sand, NaCl , and hexavalent chromium
112 (concentrations of 0-150 mg/kg) in order to simulate the soil of the natural site in terms of
113 texture and salinity. After implementing treatments, the soil samples were completely
114 homogenized and mixed , and stored in plastic bags. The treated soil samples were then
115 moistened to the field capacity and kept at this constant moisture level for 15 days in order to
116 simulate the soil contamination with heavy elements somewhat similar to the natural conditions
117 in the contaminated fields. Then, biochar and manure were added (10 g of amendment per 1 kg

118 of soil) to each sample of the contaminated soils with three replicates. Soil samples were mixed
119 well for each pot to make soil conditions homogeneous in all parts.

120

121 2.5 Germination of *Salicornia* seeds

122 Metal chromium was added to the seeds of *S. persica* , and *S. perspolitana* species as a factorial
123 design in 3 replicates and seven concentration levels (0, 5, 10, 15, 50, 100 , and 150 mg/l). Inside
124 each Petri dish, there were 10 ml of hexavalent chromium solution at different concentrations as
125 well as 25 seeds. Then, all the petri dishes were incubated in a growth chamber at a temperature
126 of 25-30 °C with a photoperiod of 16 h of light and eight h of darkness. The seeds with a radicle
127 of 2 mm were considered as a germinated seed , and all the germinated seeds were counted on a
128 daily basis. Because no germination was observed from the 14th to the 16th day, the counting
129 was stopped on the 16th day. Also, the length of root and stem , as well as the length of seedling
130 (the sum of root length and stem length) was measured on the 16th day by an mm-sized ruler.
131 Germination percentage, germination rate, seedling length, seed vigor index , and allometric
132 index were calculated based on the equations 2-1 to 2-5, respectively(Saberi et al. 2010).

$$133 \quad GP = \frac{G}{N} \times 100 \quad (2-1)$$

$$134 \quad GS = \sum \frac{ni}{Di} \quad (2-2)$$

$$135 \quad TL = SL + RL \quad (2-3)$$

$$136 \quad SVI = \frac{GR \times \text{Mean}(SI+RL)}{100} \quad (2-4)$$

$$137 \quad \text{Allometry} = \frac{\text{root length}}{\text{shoot length}} \quad (2-5)$$

138 Where, GP: germination percentage, G: the final number of germinated seeds N: number of
139 seeds sown (25 seeds in this study), GS: germination rate, ni: number of germinated seeds on

140 counting days and Di: number of days in the experiment, TL: total seedling length, SVI: seed
141 vigor index, SL: stem length, RL: root length

142 2.6 *Cultivation of seeds*

143 When the soil contamination period ended up , and after adding manure and biochar to the
144 contaminated soil, 12 seeds of *Salicornia* were planted at a depth of 0.5 cm for each pot. After
145 one month, due to lack of seed germination observed in the contaminated soil, the seedlings of *S.*
146 *persica* and *S. perspolitana* were planted in a mixture of coco-peat (Coir) and perlite (1: 1) in the
147 growth chamber at a temperature of 25-30 °C and a photoperiod characterized by 16 h of light
148 and eight h of darkness. In the early stages of the seedlings' growth, Hoagland nutrient solution
149 was used every two weeks. After five months, seedlings of the same size were selected and
150 transferred to the contaminated soil in the pots. The pots were incubated in contaminated soil
151 from June to November. The seedlings were died at 50, 100 , and 150 mg/kg, but survived at 0,
152 5, 10 , and 15 mg/kg concentrations. The seedlings of *S. persica* and *S. perspolitana* were
153 harvested after six months of growth in the contaminated soil. After harvesting, the roots and
154 shoots were washed first with tap water and then with distilled water.

155 2.7 *Measurement of plant growth and morphological traits*

156 Root and shoot weight were calculated in g per kg pot. The content of photosynthetic pigments ,
157 including chlorophyll *a* and *b* , and carotenoids in *S. persica* and *S. perspolitana* were read by
158 UV-Visible spectrophotometer (HACH, DR 5000, America). The total phenolic content of the
159 samples was calculated as gallic acid equivalent (mg of gallic acid per g of dry matter). The
160 concentration of hexavalent chromium was determined by a UV-Visible spectrophotometer
161 (HACH, DR 5000, America).

162

163 2.8 Analyzes

164 Scanning electronic microscopy (SEM) (Model VP 1450, LEO - Germany) was applied for
165 determining the morphology of the materials used in this study. Experimental treatments
166 included plant species (2 levels), the addition of metal hexavalent chromium in 7 levels (zero, 5,
167 10 and 15, 50, 100 and 150 mg/kg) , and soil amendments in 3 levels (manure, biochar , and
168 control). The total number of experimental units (number of pots) was 126. A factorial
169 experiment in the form of a completely randomized design with three replicates was used to
170 evaluate the results of this study. Statistical analysis of data including ANOVA was performed in
171 Minitab v.16 software , and Tukey test was applied for comparison of means (both the main
172 effects and interactions) at the significance level of 0.05.

173 3. Results

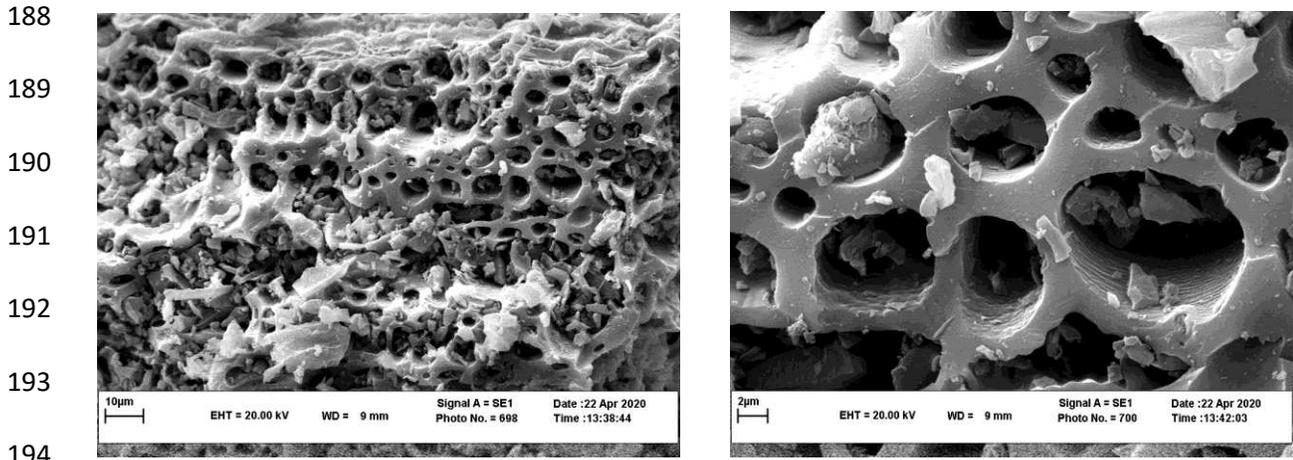
174 3.1 Physical and chemical properties of soil, cattle manure , and biochar

175 The soil used in this study was characterized by heavy texture, low salinity (EC of 1.8) , and
176 approximately alkaline pH (8.14). The soil texture was clayey-sandy loam with the organic
177 carbon content of 2.56%. The percentages of organic carbon, total nitrogen, phosphorus,
178 potassium in the manure were 56%, 2.95%, 0.62% , and 0.94%, respectively. The values for
179 carbon, hydrogen, nitrogen, oxygen and sulfur in biochar were 73.41%, 2.71%, 0.37%, 23.51%
180 and 0%, respectively. The results of heavy metal analysis tests showed that the concentration of
181 hexavalent chromium in the soil was 2.91 mg/kg, while there was no hexavalent chromium in
182 manure and biochar.

183 3.2 Scanning electron microscopy (SEM)

184 Figure 1 shows the Scanning Electron Microscopy images (SEM) of the biochar surface. As
185 shown, biochar is characterized by a large surface area and a relatively regular network of

186 honeycomb-shaped pores on its surface, which leads to the absorption of heavy metals. Also, this
187 honeycomb network represents a carbon skeleton in the biochar structure(Ghani et al. 2013).



195 Figure 1. Scanning electron microscopy (SEM) of biochar surface

196

197 *3.3 Effect of different concentrations of hexavalent chromium on germination traits of S.*
198 *persica and S. perspolitana*

199 The results of the effect of different concentrations of chromium on the germination percentage
200 (%) and rate, and root length, shoot length , and seedling length in the two studied species shown
201 in Table 1 indicated a higher resistance of *S. persica* compared to *S. perspolitana* against metal
202 hexavalent chromium stress so that all germination traits declined with increasing the
203 concentration of chromium from the control (0 mg/l) to 150 mg/l, and eventually led to plant
204 dying.

205

206

207

208

209 Table 1. Results of comparison of the means related to simple effect of different concentrations
 210 of chromium on germination traits of *S. persica* and *S. perspolitana*

Treatment	Germination percentage	Germination rate	Stem length	Root length	Seedling length
species					
<i>S. persica</i>	58.89 ^a	1.05 ^a	10.11 ^a	1.088 ^a	21.11 ^a
<i>S. perspolitana</i>	50.44 ^b	0.900 ^b	8.16 ^b	1.080 ^a	9.24 ^b
Different concentrations of chromium					
Cr₀	67.68 ^a	1.22 ^a	11.38 ^a	1.91 ^a	13.3 ^a
Cr₅	64.67 ^{b a}	1.15 ^{ab}	11.29 ^a	1.50 ^b	12.83 ^a
Cr₁₅	60.67 ^b	1.08 ^b	9.60 ^b	0.91 ^c	10.51 ^b
Cr₅₀	52.67 ^c	0.94 ^c	8.89 ^{bc}	0.85 ^c	9.75 ^{bc}
Cr₁₀₀	45.33 ^d	0.80 ^d	8.03 ^c	0.73 ^c	8.76 ^c
Cr₁₅₀	36.00 ^e	0.64 ^e	5.62 ^d	0.57 ^c	6.2 ^d

211 Different letters indicate statistically significant differences at P > 0.05.

212

213 *3.4 Effect of different concentrations of hexavalent chromium, manure , and biochar on the*
 214 *plant height of two species S. persica and S. perspolitana*

215 As seen in table 2, the effect of different concentrations of chromium on the plant height of the
 216 two studied species was statistically significant (P <0.05). The highest and lowest heights were
 217 related to Cr₀ (no chromium) and Cr₁₅ (chromium concentration of 15 mg/g) with corresponding
 218 heights of 45.33 and 27 cm, respectively. The height reported for *S. persica* was 6.42% higher
 219 than *S. perspolitana*, which can be attributed lower accumulation of chromium in *S. persica*
 220 tissues. As shown in the table, the effect of different amendments on the height of the two
 221 studied plant species was also significant. Manure and biochar treatments indicated an increased
 222 height by 22.42% and 12.51%, respectively, compared to the control treatment.

223

224

225 Table 3. Results of mean comparison of interaction effects between different concentrations of
 226 chromium, manure and biochar on the height of *S. persica* and *S. perspolitana*

treatments	Plant height (cm)	Shoot dry weight (g per pot)	Root dry weight (g per pot)
amendments			
control	31.79 ^c	0.53 ^c	0.06 ^c
biochar	31.79 ^c	0.68 ^b	0.07 ^b
manure	35.26 ^b	0.79 ^a	0.098 ^a
species			
<i>S. persica</i>	92.38 ^a	0.97 ^a	0.11 ^a
<i>S. perspolitana</i>	36.60 ^a	36.0 ^b	0.039 ^b
Different concentrations of Cr			
Cr₀	34.39 ^b	1.12 ^a	0.17 ^a
Cr₅	45.33 ^a	0.81 ^b	0.067 ^b
Cr₁₀	37.28 ^b	0.45 ^c	0.046 ^c
Cr₁₅	32.36 ^c	0.29 ^d	0.02 ^d

227 Different letters indicate statistically significant differences at P > 0.05.

228 3.5 The effect of different concentrations of hexavalent chromium, manure , and biochar on the
 229 root and shoot dry weight of *S. persica* and *S. perspolitana*

230 Table 3 shows the results of the mean comparison of the interaction effects of different
 231 concentrations of chromium, manure , and biochar on the root and shoot dry weight of *S. persica*
 232 and *S. perspolitana*. According to these results, different concentrations of chromium negatively
 233 influenced on the dry shoot weight of the two studied species. Concentrations of chromium as 5,
 234 10 , and 15 mg/kg resulted in decreased shoot dry weight of *S. persica* species by 52.59%,
 235 66.88% , and 88.31%, while the decreases reported for *S. perspolitana* species were found to be
 236 17.07%, 36.58% , and 48.78%, respectively, compared with controls.

237 According to the results, the organic compounds (manure and biochar) applied in this study
 238 showed a positive effect on the dry shoot weight of the two studied species. When applying
 239 biochar treatment at concentrations of 5, 10 , and 15 mg/kg chromium, shoot dry weight
 240 increased by 84.9%, 9.80% and 111.1%, in *S. persica* species, and by 82.8%, 7.69% and 19.04%

241 in *S. perspolitana* species compared to the control (without amendment), respectively. The
242 increases in shoot dry weight for the manure treatment at concentrations of and 5, 10 and 15
243 mg/kg chromium were 115.06%, 49.01% , and 100% for *S. persica* species and 50%, 26.9% ,
244 and 76.1% for *S. perspolitana* compared to the controls, respectively.

245 The results concerning the mean comparison of the interaction effects of different concentrations
246 of chromium, manure , and biochar on the root dry weight of *S. persica* and *S. perspolitana* in
247 Table 3 indicated the negative effect of different concentrations of chromium on the root dry
248 weight of the two studied species , so that root dry weight decreased with increasing the
249 concentration of this element in the soil. The decreases reported at concentrations of 5, 10 , and
250 15 mg/kg chromium were 69.23%, 89.61% , and 94.61% for *S. persica*, and 30%, 63.6% , and
251 63.4% for *S. perspolitana* species compared to the controls, respectively. Again, the organic
252 compounds used in this study , including manure and biochar , positively influenced on the root
253 dry weight of the two species. By applying of biochar into the soil at chromium concentrations of
254 5, 10 , and 15 mg/kg, the root dry weight increased by 8.75%, 137.0% , and 28.5% in *S. persica*
255 and 8.5%, 26.37% , and 1.09% in *S. perspolitana* compared to the controls (without
256 amendment), respectively. The increases for the manure treatment at chromium concentrations of
257 5, 10 , and 15 mg/kg were 50%, 200% , and 321.4% for *S. persica* and 8.5%, 251.6% , and
258 156.8% in *S. perspolitana*, compared to the controls, respectively. For biochar treatment at
259 chromium concentrations of 5, 10 , and 15 mg/kg, the root dry weight decreased by 8.75%,
260 137.0% , and 28.5% in *S. persica* and by 8.5%, 26.37% , and 1.09% in *S. perspolitana* compared
261 to the controls (without amendment), respectively.

262 Table 3. Mean comparison of the interaction effects of different concentrations of chromium,
263 manure and biochar on shoot and root dry weight in *S. persica* and *S. perspolitana*

different concentrations of chromium	Species	amendments	Shoot dry weight (g per pot)	Root dry weight (g per pot)
Cr ₀	<i>S. persica</i>	control	1.54 ^c	0.26 ^b
		biochar	1.82 ^{ab}	0.27 ^b
		manure	1.93 ^a	0.3 ^a
	<i>S.perspolitana</i>	control	0.41 ^{fghi}	0.05 ^{efgh}
		biochar	0.49 ^{efgh}	0.06 ^{defg}
		manure	0.52 ^{defg}	0.062 ^{defg}
Cr ₅	<i>S. persica</i>	control	0.73 ^{de}	0.08 ^{de}
		biochar	1.35 ^c	0.087 ^d
		manure	1.57 ^{bc}	0.12 ^c
	<i>S.perspolitana</i>	control	0.34 ^{fghi}	0.035 ^{fghi}
		biochar	0.37 ^{fghi}	0.038 ^{fghi}
		manure	0.51 ^{defg}	0.038 ^{fghi}
Cr ₁₀	<i>S. persica</i>	control	0.51 ^{fghi}	0.027 ^{ghi}
		biochar	0.56 ^{def}	0.064 ^{def}
		manure	0.76 ^d	0.081 ^{de}
	<i>S.perspolitana</i>	control	0.26 ^{ghi}	0.0182 ^{hi}
		biochar	0.28 ^{ghi}	0.023 ^{hi}
		manure	0.33 ^{fghi}	0.064 ^{def}
Cr ₁₅	<i>S. persica</i>	control	0.18 ⁱ	0.0183 ^{hi}
		biochar	0.38 ^{fghi}	0.0185 ^{hi}
		manure	0.36 ^{fghi}	0.047 ^{efghi}
	<i>S.perspolitana</i>	control	0.21 ⁱ	0.014 ⁱ
		biochar	0.25 ^{hi}	0.018 ^{hi}
		manure	0.37 ^{fghi}	0.059 ^{defg}

264

265 3.6 Effect of different concentrations of chromium, manure , and biochar on chlorophyll a and
266 b of *S. persica* and *S. perspolitana*

267 The results of the mean comparison of the interaction effects of different concentrations of
268 chromium, manure , and biochar on the content of photosynthetic pigments in *S. persica* and *S.*
269 *perspolitana* are shown in Tables 4. According to these results, different concentrations of
270 chromium negatively influenced on the content of chlorophyll a and b in the two studied species
271 so that the content of chlorophyll a and b decreased with increasing chromium in the soil. At
272 chromium concentration of 5, 10 and 15 mg/kg in *S. persica*, the reported decreases were

273 31.37%, 28.23% and 27.45% for chlorophyll *a* and 39.54%, 38.04% and 35.03% for chlorophyll
274 *b*, respectively. In *S. perspolitana*, the contents of chlorophyll decreased by 28.01%, 28.8% , and
275 39.22% , and of chlorophyll *b* by 15.6%, 18.7% and 27.6% compared to controls, respectively.
276 Applying of hexavalent chromium caused a significant reduction in chlorophyll contents. There
277 was a significant difference between different levels of chromium concentrations as 5, 10 , and
278 15 mg/kg in terms of decreases in chlorophyll *a* and *b* (Table 4). The organic compounds
279 (manure and biochar) used in this study also indicated a positive effect on the content of
280 photosynthetic pigments in the two studied species. By applying the biochar treatment on *S.*
281 *persica* species at chromium concentrations of 5, 10 , and 15 mg/kg, chlorophyll content
282 increased by 49.71%, 19.6% , and 11.35% and the values for chlorophyll *b* by 61.4%, 25.4% ,
283 and 0.3%. For *S. perspolitana*, applying of biochar resulted in increases in chlorophyll *a* by
284 29.9%, 35.75 , and 52.48% and in chlorophyll *b* by 26.85%, 36.52% , and 40.66% compared to
285 the controls (without amendment), respectively. Also, in the manure treatment at chromium
286 concentrations of 5, 10 , and 15 mg/kg, the content of chlorophyll *a* decreased by 3.4%, 5.4%
287 and 6.4% , and of chlorophyll *b* by 10.74%, 10.19%, and 15.3% in *S. persica* species,
288 respectively. For *S. perspolitana* species, the decreases in chlorophyll *a* were as 29.3%, 24.24% ,
289 and 47.5% , and in chlorophyll *b* as 24.21%, 9.6% , and 29.9%, respectively, compared to the
290 control (without amendment). In general, the highest content of chlorophyll *a* (2.73 mg/g fresh
291 weight) was found in *S. persica* species at chromium concentration of 0 mg/kg in biochar
292 treatment , and the lowest content (1.41 mg/g fresh weight) was found for *S. perspolitana* species
293 at chromium concentration of 15 mg/kg in the control treatment (without amendment). Also, the
294 highest content of chlorophyll *b* (0.669 mg/g fresh weight) was found in *S. persica* species at
295 chromium concentration of 0 mg/kg in Biochar treatment , and the lowest content (0.36 mg/g

296 fresh weight) has belonged to *S. perspolitana* species at chromium concentration of 15 mg/kg in
 297 the control treatment (without amendment)

298 Table 4. Mean comparison of the interaction effects of different concentrations of chromium,
 299 manure and biochar on the content of photosynthetic pigments in *S. persica* and *S. perspolitana*

different concentrations of chromium	Species	amendments	Chlorophyll a (mg. fresh weight)	Chlorophyll b (mg. fresh weight)	Carotenoid (mg. fresh weight)
Cr ₀	<i>S. persica</i>	control	2.55 ^{bc}	0.665 ^a	160.5 ^c
		biochar	2.73 ^a	0.669 ^a	160.5 ^c
		manure	2.63 ^{ab}	0.661 ^a	160.1 ^c
	<i>S.perspolitana</i>	control	2.32 ^{ef}	0.4989 ^h	147.4 ^k
		biochar	2.47 ^{cd}	0.5100 ^{fg}	163.3 ^b
		manure	2.42 ^{de}	0.5043 ^{gh}	156.1 ^d
Cr ₅	<i>S. persica</i>	control	1.75 ^{mn}	0.402 ^o	115.3 ^s
		biochar	2.62 ^b	0.649 ^b	167.2 ^a
		manure	1.81 ^m	0.4471 ^{jk}	124.9 ^o
	<i>S.perspolitana</i>	control	1.67 ⁿ	0.4208 ^m	131.3 ^o
		biochar	2.17 ^{gh}	0.5338 ^d	149.2 ^e
		manure	2.16 ^{ghi}	0.5227 ^e	148.4 ^f
Cr ₁₀	<i>S. persica</i>	control	1.83 ^{lm}	0.412 ^{mn}	119.7 ^q
		biochar	2.19 ^g	0.517 ^{ef}	137.9 ⁱ
		manure	1.93 ^{kl}	0.454 ^j	128.6 ^m
	<i>S.perspolitana</i>	control	1.65 ⁿ	0.4052 ^{no}	122.1 ^p
		biochar	2.24 ^{fg}	0.5532 ^c	163.6 ^b
		manure	2.05 ^{ij}	0.4442 ^k	134.3 ^j
Cr ₁₅	<i>S. persica</i>	control	1.85 ^{lm}	0.432 ^l	125.2 ^o
		biochar	2.06 ^{hij}	0.445 ^k	129 ^{lm}
		manure	1.97 ^{jk}	0.4983 ^h	126.3 ⁿ
	<i>S.perspolitana</i>	control	1.41 ^o	0.361 ^p	118.4 ^r
		biochar	2.15 ^{ghi}	0.5078 ^g	144.3 ^h
		manure	2.08 ^{hig}	0.4691 ⁱ	129 ^l

300 different letters indicate statistically significant differences at P > 0.05.

301 3.7 Effect of different concentrations of hexavalent chromium, manure , and biochar on
 302 carotenoid content in *S. persica* and *S. perspolitana*

303 Table 4 shows the results of the mean comparison of the interaction effects of different
 304 concentrations of chromium, manure , and biochar on the carotenoid content of *S .persica* and *S.*

305 *perspolitana*. According to these results, the different concentrations of chromium negatively
306 influenced on the carotenoid content in the two species so that the carotenoid content decreased
307 with increasing the concentration of this heavy metal in the soil. At three chromium
308 concentrations of 5, 10 , and 15 mg/kg, the carotenoid content decreased by 28.16%, 25.42% ,
309 and 21.99% in *S. persica*, and by 10.92%, 17.16% , and 19.67% in *S. perspolitana* compared to
310 the controls, respectively. Thus, the organic compounds used in this study indicated a positive
311 effect on the carotenoid content of the two species. In biochar treatment at chromium
312 concentrations of 5, 10 , and 15 mg/kg, the carotenoid content in *S. persica* increased by
313 45.01%, 15.20% , and 3.03% and in *S. perspolitana* by 13.6%, 33.9% , and 21.8% compared to
314 the controls, respectively. Also, in manure treatment at chromium concentrations of 5, 10 , and
315 15 mg/kg, the increases in carotenoid content were 8.32%, 7.43% , and 0.87% in *S. persica*, and
316 13.02%, 9.99% , and 8.95% in *S. perspolitana* compared to the controls, respectively.

317 3.8 *The effect of different concentrations of chromium, manure , and biochar on the total* 318 *phenolic content in S. persica and S. perspolitana*

319 Table 5 shows the results of the mean comparison of the interaction effects of different
320 concentrations of chromium, manure , and biochar on the total phenolic content in *S. persica* and
321 *S. perspolitana*. According to these results, different concentrations of chromium negatively
322 affected on the total phenolic content of the two studied species , and so that the total phenolic
323 content in the plant decreased with increasing the concentration of chromium in the soil. At three
324 chromium concentrations of 5, 10 , and 15 mg/kg , the total phenolic content in *S. persica*
325 species decreased by 39.30%, 39.86% , and 49.73% , and in *S. perspolitana* by 34.75%, 42.44% ,
326 and 60.35% compared to the controls, respectively. Again, the organic compounds used in this
327 study showed a positive effect on the total phenolic content of the two species. In biochar

328 treatment at chromium concentrations of 5, 10 , and 15 mg/kg, the total phenolic content
 329 increased by 62.95%, 67.30% , and 111.16% in *S. persica* species, and by 24.26%, 11.31% , and
 330 36.40% in *S. perspolitana* compared to the controls, respectively. Also, in manure treatment at
 331 concentrations of 5, 10 , and 15 mg/kg, the increases in total phenolic content were 39.14%,
 332 32.87% , and 21.22% for *S. persica*, and 3.54%, 5.26% , and 22.35% for *S. perspolitana*
 333 compared to the controls, respectively.

334 Table 5. Mean comparison of the interaction effects of different concentrations of chromium,
 335 manure and biochar on the total phenolic content in *S. persica* and *S. perspolitana*.

different concentrations of chromium	Species	amendments	total phenolic content (mg gallic acid per g fresh weight)
Cr₀	<i>S. persica</i>	control	37.58 ^{bc}
		biochar	44.15 ^a
		manure	38.57 ^b
	<i>S.perspolitana</i>	control	30.7 ^{de}
		biochar	34.39 ^{cd}
		manure	34.13 ^{cd}
Cr₅	<i>S. persica</i>	control	22.81 ^{fg}
		biochar	37.17 ^{bc}
		manure	31.74 ^{de}
	<i>S.perspolitana</i>	control	20.03 ^{ghij}
		biochar	24.89 ^f
		manure	20.74 ^{ghi}
Cr₁₀	<i>S. persica</i>	control	22.6 ^{fgh}
		biochar	37.81 ^{bc}
		manure	30.03 ^e
	<i>S.perspolitana</i>	control	17.67 ^{ijk}
		biochar	19.67 ^{ghij}
		manure	18.6 ^{hijk}
Cr₁₅	<i>S. persica</i>	control	81.89 ^{ghijk}
		biochar	39.89 ^b
		manure	22.9 ^{fg}
	<i>S.perspolitana</i>	control	12.17 ^l
		biochar	61.60 ^{jk}
		manure	14.89 ^{kl}

336 different letters indicate statistically significant differences at P > 0.05.

337 3.9 The effect of different concentrations of chromium, manure , and biochar on the values of
338 accumulated chromium in roots and shoots of *S. persica* and *S. perspolitana*

339 Table 6 shows the mean comparison of the interaction effects of different concentrations of
340 chromium, manure , and biochar on the accumulation of total chromium, Cr (VI) , and Cr (III) in
341 the roots of *S.persica* and *S.perspolitana* after six months of growing in the contaminated soil.
342 Generally, the accumulation of chromium in the roots of both species was significantly affected
343 by the chromium content in the soil so that the accumulation of chromium in the roots of both
344 species increased by increasing the chromium concentration from 0 to 15 mg/kg in the soil. In *S.*
345 *persica*, at chromium concentration of 0 mg/kg, accumulation of total, hexavalent , and trivalent
346 chromium in the roots of plants grown in contaminated soil (without amendment) were 17.77,
347 5.38 , and 16.86 mg/kg. At chromium concentration of 15 mg/kg, the corresponding values for
348 accumulation of total, hexavalent , and trivalent chromium were 357.61, 6.41 , and 351.19 mg/kg
349 for this species. In *S. perspolitana*, at chromium concentration of 0 mg/kg, accumulation of total,
350 hexavalent , and trivalent chromium in the roots of plants grown in contaminated soil (without
351 amendment) were 21.58, 7.73 , and 13.85 mg/kg. At chromium concentration of 15 mg/kg, the
352 values for accumulation of total, hexavalent , and trivalent chromium were 624.91, 46.08 , and
353 578.83 mg/kg for this species. In total, the accumulation of total, hexavalent and trivalent
354 chromium in the roots of *S. perspolitana* was higher than *S. persica*. The organic compounds
355 used in this study showed a negative effect on the accumulation of total, hexavalent and trivalent
356 chromium in the roots of the two studied species. In biochar treatment, the accumulation of total,
357 hexavalent and trivalent chromium in *S.persica* species decreased by 33.89%, 50.39% and
358 33.60% at chromium concentration of 5 mg/kg, 76.47%, 73.09% and 76.5% at chromium
359 concentration of 10 mg/kg and 25.01%, 81.7% and 23.39% at chromium concentration of 15

360 mg/kg. In *S. perspolitana* species under biochar treatment, the accumulation of total, hexavalent
 361 and trivalent chromium decreased by 53.60%, 19.96% and 54.4% at chromium concentration of
 362 5 mg/kg, 23.72%, 14.54% and 24.0% at chromium concentration of 10 mg/kg and 11.39%,
 363 32.53% and 9.50% at chromium concentration of 15 mg/kg. In the manure treatment, the
 364 accumulation of total, hexavalent and trivalent chromium in *S.persica* species decreased by
 365 23.82%, 36.24% and 23.61% at chromium concentration of 5 mg/kg, 22.73%, 51.85% and
 366 22.13% at chromium concentration of 10 mg/kg and 1.75%, 52.32% and 0.22% at chromium
 367 concentration of 15 mg/kg. In *S. perspolitana* species under biochar treatment, the accumulation
 368 of total, hexavalent and trivalent chromium decreased by 26.36%, 8.11% and 22.96% at
 369 chromium concentration of 5 mg/kg, 21.55%, 2.02% and 22.22% at chromium concentration of
 370 10 mg/kg and 9.48%, 16.96% and 8.81% at chromium concentration of 15 mg/kg.

371 Table 6. Mean comparison of the interaction effects of different concentrations of chromium,
 372 manure and biochar on chromium accumulation in the roots of *S. persica* and *S. perspolitana*

different concentrations of chromium	Species	amendments	Hexavalent chromium (mg.kg)	Total chromium (mg.kg)	Trivalent chromium (mg.kg)
Cr ₀	<i>S. persica</i>	control	1.05 ^p	20.39 ⁿ	19.34 ^o
		biochar	0.8 ^p	16.27 ⁿ	15.47 ^o
		manure	0.85 ^p	16.63 ⁿ	15.78 ^o
	<i>S.perspolitana</i>	control	9.39 ^j	24.54 ⁿ	15.14 ^o
		biochar	6 ^l	19.4 ⁿ	13.4 ^o
		manure	7.8 ^k	20.8 ⁿ	13 ^o
Cr ₅	<i>S. persica</i>	control	7.56 ^k	440.99 ^{fg}	433.42 ^{fg}
		biochar	3.75 ⁿ	291.53 ^k	287.78 ^l
		manure	4.82 ^m	335.91 ^j	331.08 ^k
	<i>S.perspolitana</i>	control	15.03 ^f	589.67 ^c	574.63 ^b
		biochar	12.03 ^h	273.59 ^l	261.53 ^m
		manure	13.81 ^g	434.18 ^g	420.36 ^h
Cr ₁₀	<i>S. persica</i>	control	10.78 ⁱ	532.27 ^d	521.48 ^e
		biochar	9.2 ^{no}	125.21 ^m	122.31 ⁿ
		manure	5.19 ^{lm}	411.27 ^h	406.07 ⁱ
	<i>S.perspolitana</i>	control	19.8 ^d	588.99 ^c	569.19 ^{bc}
		biochar	16.92 ^e	449.27 ^f	432.34 ^g

		manure	19.4 ^d	462.06 ^e	442.66 ^f
Cr ₁₅	<i>S. persica</i>	control	11.6 ^{hi}	392.81 ⁱ	381.21 ^j
		biochar	2.12 ^o	294.53 ^k	292.01 ^l
		manure	5.53 ^{lm}	385.9 ⁱ	380.36 ^j
	<i>S.perspolitana</i>	control	55.18 ^a	671.66 ^a	616.48 ^a
		biochar	37.23 ^c	595.13 ^c	557.89 ^d
		manure	45.82 ^b	607.94 ^b	562.11 ^{cd}

373 different letters indicate statistically significant differences at $P > 0.05$.

374 Table 7 indicates the mean comparison of the interaction effects of different concentrations of
375 chromium, manure , and biochar on the accumulation of total, hexavalent , and trivalent
376 chromium in the shoots of *S. persica* and *S. perspolitana* after six months of growing in the
377 contaminated soil. Like the results obtained for the roots, the accumulation of chromium in the
378 shoots of both species was significantly affected by the chromium content in the soil. In other
379 words, chromium accumulation in the shoot showed an increasing trend with increasing
380 chromium concentration from 0 to 15 mg/kg in the soil for both species. In *S. persica*, at
381 chromium concentration of 0 mg/kg, accumulation of total, hexavalent , and trivalent chromium
382 in the shoots of plants grown in contaminated soil (without amendment) were 2.07, 0.56 , and
383 1.51 mg/kg. At chromium concentration of 15 mg/kg, the corresponding values for accumulation
384 of total, hexavalent , and trivalent chromium were 33.54, 1.12 , and 32.42 mg/kg for this species.
385 In *S. perspolitana*, at chromium concentration of 0 mg/kg, accumulation of total, hexavalent ,
386 and trivalent chromium in the shoots of plants grown in contaminated soil (without amendment)
387 were 1.03, 0.33 , and 0.7 mg/kg. At chromium concentration of 15 mg/kg, the values for
388 accumulation of total, hexavalent , and trivalent chromium were 28.41, 1.25 , and 27.16 mg/kg
389 for this species.

390 Unlike roots, accumulation of total, hexavalent , and trivalent chromium in shoots of *S. persica*
391 was higher than *S. perspolitana*. As shown in table 7, the organic compounds used in this study

392 hurt the accumulation of total, hexavalent , and trivalent chromium in the shoots of the two
 393 species.

394 In biochar treatment, the accumulation of total, hexavalent and trivalent chromium in *S.persica*
 395 species decreased by 91.83%, 39.66% and 93.61% at chromium concentration of 5 mg/kg,
 396 93.22%, 72.53% and 93.78% at chromium concentration of 10 mg/kg and 93.60%, 24.42% and
 397 94.66% at chromium concentration of 15 mg/kg. In *S. perspolitana* species under biochar
 398 treatment, the accumulation of total, hexavalent and trivalent chromium decreased by 73.06%,
 399 29.68% and 75.07% at chromium concentration of 5 mg/kg, 60.05%, 26.08% and 61.29% at
 400 chromium concentration of 10 mg/kg and 70.96%, 61.11% and 71.36% at chromium
 401 concentration of 15 mg/kg. In the manure treatment, the accumulation of total, hexavalent and
 402 trivalent chromium in *S.persica* species decreased by 51.53%, 14.04% and 52.77% at chromium
 403 concentration of 5 mg/kg, 88.50%, 55.44% and 89.40% at chromium concentration of 10 mg/kg
 404 and 90.62%, 19.08% and 91.71% at chromium concentration of 15 mg/kg. In *S. perspolitana*
 405 species under biochar treatment, the accumulation of total, hexavalent and trivalent chromium
 406 decreased by 48.89%, 21.87% and 50.14% at chromium concentration of 5 mg/kg, 33.23%,
 407 21.73% and 33.65% at chromium concentration of 10 mg/kg and 60.90%, 50% and 61.36% at
 408 chromium concentration of 15 mg/kg.

409 Table 7. Mean comparison of the interaction effects of different concentrations of chromium,
 410 manure and biochar on chromium accumulation in shoots of *S. persica* and *S. p*

different concentrations of chromium	Species	amendments	Hexavalent chromium (mg.kg)	Total chromium (mg.kg)	Trivalent chromium (mg.kg)
Cr ₀	<i>S. persica</i>	control	0.65 ^{fg} hi	2.06 ^{lm}	1.41 ^{kl}
		biochar	0.5 ^{hijk}	2.14 ^{lm}	1.64 ^{kl}
		manure	0.54 ^{ghij}	2.03 ^{lm}	1.49 ^{kl}
	<i>S.perspolitana</i>	control	0.37 ^{jk}	1.05 ^m	0.68 ^l
		biochar	0.28 ^k	0.99 ^m	0.7 ^l

		manure	0.33 ^{jk}	1.04 ^m	0.71 ^l
Cr₅	<i>S. persica</i>	control	1.21 ^{bc}	37.24 ^d	36.02 ^d
		biochar	0.73 ^{fgh}	3.04 ^{kl}	2.3 ^{jk}
		manure	1.04 ^{cd}	18.05 ^f	17.01 ^f
	<i>S.perspolitana</i>	control	0.64 ^{fghi}	14.48 ^g	13.84 ^g
		biochar	0.45 ^{ijk}	3.9 ^{jk}	3.45 ^{ij}
manure		0.5 ^{hijk}	7.4 ^h	6.9 ^h	
Cr₁₀	<i>S. persica</i>	control	1.93 ^a	72.73 ^b	70.8 ^b
		biochar	0.53 ^{ghij}	4.93 ^{ij}	4.4 ⁱ
		manure	0.86 ^{def}	8.36 ^h	7.5 ^h
	<i>S.perspolitana</i>	control	0.69 ^{fgh}	20.13 ^e	19.43 ^e
		biochar	0.51 ^{hijk}	8.04 ^h	7.52 ^h
manure		0.54 ^{ghij}	13.44 ^g	12.89 ^g	
Cr₁₅	<i>S. persica</i>	control	1.31 ^b	86.93 ^a	85.62 ^a
		biochar	0.99 ^{cde}	5.56 ⁱ	4.57 ⁱ
		manure	1.06 ^{cd}	8.15 ^h	7.09 ^h
	<i>S.perspolitana</i>	control	1.98 ^a	50.7 ^c	48.71 ^c
		biochar	0.77 ^{efg}	14.72 ^g	13.95 ^g
		manure	0.99 ^{cde}	19.82 ^e	18.82 ^e

411 different letters indicate statistically significant differences at $P > 0.05$.

412 *3.10 Determination of TF¹, BCF², BAC³ indices to evaluate the phytoremediation ability of S.*
413 *persica and S. perspolitana species*

414 In order to evaluate the phytoremediation potential of plants after determining the concentration
415 of heavy metals in plant and soil samples, it should be necessary to calculated indices including
416 TF (Translocation Factor; a ratio of metal concentrations in shoots to roots), BCF (Bio
417 Concentration Factor; ratio of metal concentrations in roots to soil), BAC (Biological
418 Accumulation Coefficient; the a ratio of metal concentration in shoots to soil) because the
419 phytoremediation potential of a species is calculated using these indices. The values of TF
420 greater than 1 indicate that the plant is suitable for extracting the contaminants. Also, plants with
421 values of TF and BAC greater than one are suitable for phytoremediation of the contaminants.
422 Plants with TF value lower than 1 and BCF values higher than one are suitable for
423 phytostabilization(Yoon et al. 2006).

424 The results concerning the effect of the studied treatments on the accumulation of metals in
 425 Table 8 showed that the value of TF for heavy metals was lower than 1 in all treatments.
 426 Therefore, both *S. persica* and *S. perspolitana* would serve as suitable species for
 427 phytostabilization.
 428 Table 8. Mean values of TF, BCF and BAC related to total chromium in *S. persica* and *S.*
 429 *perspolitana*.

different chromium concentrations of	Species	amendments	TF	BAC	BCF
Cr₀	<i>S. persica</i>	control	0.010	0.028	0.280
		biochar	0.13	0.029	0.223
		manure	0.12	0.027	0.228
	<i>S.perspolitana</i>	control	0.043	0.014	0.33
		biochar	0.051	0.013	0.26
		manure	0.050	0.014	0.28
Cr₅	<i>S. persica</i>	control	0.084	0.47	5.67
		biochar	0.010	0.039	3.75
		manure	0.053	0.915	4.32
	<i>S.perspolitana</i>	control	0.02	0.18	7.59
		biochar	0.0142	0.050	3.52
		manure	0.017	0.095	5.59
Cr₁₀	<i>S. persica</i>	control	0.13	0.87	6.43
		biochar	0.039	0.059	1.51
		manure	0.020	0.102	4.91
	<i>S.perspolitana</i>	control	0.034	0.24	7.12
		biochar	0.017	0.097	5.43
		manure	0.029	0.16	5.58
Cr₁₅	<i>S. persica</i>	control	0.22	0.99	4.480
		biochar	0.01	0.063	3.35
		manure	0.021	0.092	4.40
	<i>S.perspolitana</i>	control	0.075	0.57	7.66
		biochar	0.024	0.167	6.78
		manure	0.032	0.22	6.93

430 Cr₀: without chromium, Cr₅: chromium concentration of 5 mg/kg, Cr₁₀: chromium concentration
 431 of 10 mg/kg, Cr₁₅: chromium concentration of 15 mg/kg

432 4. Discussion

433 4.1 The effect of hexavalent chromium on germination traits of *S. persica* and *S. perspolitana*

434 Heavy elements are limiting factors in the germination and growth stages of the seedling.
435 Response to environmental stresses is a complex and undeniable phenomenon in higher
436 plants(Díaz et al. 2001). In this regard, both *S. persica* and *S. perspolitana* species showed a
437 similar response of reduced germination under increased chromium concentration. Seed
438 germination is dependent on protein activity, which decreased under chromium treatment. Both
439 *Salicornia* species indicated decreased growth with increasing hexavalent chromium
440 concentration , and it was also found that chromium negatively influenced on root and stem
441 length. Decreased growth in a plant may be due to reduced growth in the roots and consequently
442 deficient transfer of nutrients to the upper parts of the plant. Also, the transfer of chromium to
443 the plant shoot could directly affect its cellular metabolism, which leads to a decreased seedling
444 length(Shanker et al. 2005).

445 The study of the effect of heavy metals on plant characteristics seems to be necessary to identify
446 suitable plants for phytoremediation of contaminated soils on the one hand, and to identify
447 resistant plant genotypes on the other hand. Bhardwaj et al.(Bhardwaj et al. 2009). demonstrated
448 that the germination (%) of bean plant was not affected at low cadmium concentrations than the
449 control, but prevented germination was observed at higher cadmium concentrations of 3 g/kg
450 soil. Peralta et al. (Peralta et al. 2001). reported a decrease in seed germination, root , and shoot
451 length with increasing concentration of cadmium in soil under alfalfa plant (*Medicago satival*).
452 Seed germination was the same as the control at a cadmium concentration of 5 mg/l, although the
453 stem length reduced by about 17% and the root length increased by about 22% compared to the
454 control treatment, but decreases in seed germination and seedling length was significant at a

455 concentration of 10 mg/l. Also, a cadmium concentration of 40 mg/l led to the death of most
456 alfalfa seedlings. Authors have reported delayed seedling emergence as a sign of cadmium
457 toxicity. Therefore, the results of the mentioned studies are consistent with the findings of this
458 investigation so that a decrease in the traits related to seed germination rate was observed with
459 increasing the concentration of heavy metals in both *S. persica* and *S. perspolitana*.

460 Heavy metals cause serious damage to the cell by inducing a lack of active oxygen production.
461 As vital stages in the plant life cycle, germination and establishment of seedlings can be affected
462 by high levels of heavy metals. Chromium toxicity at concentrations of 5, 25, 100 , and 150
463 mg/kg had a significant effect on seedling growth. Chromium treatment at 100 mg/kg negatively
464 influenced on germination indices of okra (*Hibiscus esculentus*)(Amin et al. 2013).

465 Also, in a study, it was found that the *Vigna radiata* did not tolerate chromium concentrations
466 higher than 50 mg/l (Murtaza et al. 2018). In the present study, the results regarding the seedling
467 growth showed a similar trend, so that stem length also decreased with decreasing root length in
468 both *S. persica* and *S. perspolitana* species, which may be due to reduced nutrients. Therefore,
469 the present study confirmed that seed germination and seedling growth are affected by different
470 concentrations of chromium. Results with different concentrations of chromium in comparison
471 with the control treatment demonstrated a decrease in germination parameters for both *S. persica*
472 and *S. perspolitana* species confirming the findings obtained by other researchers.

473 4.2 *Effect of experimental treatments on plant physiological parameters (plant height, root* 474 *and shoot length)*

475 This study showed that different concentrations of chromium in the soil negatively influenced on
476 growth indices in both species so that the plant height decreased with increasing the
477 concentration of chromium in the soil, which is consistent with the results of other researches. In

478 the case of *Eruca sativa* species, the presence of hexavalent chromium at a concentration of 500
479 mg/kg in the soil(Kamran et al. 2017). Caused a decrease in plant height. By studying a number
480 of 32 plants, Lukina et al.(Lukina et al. 2016) reported that the toxicity of hexavalent chromium
481 (1000 mg/kg) indicated an adverse effect on 94% of the studied species. Decreased growth and
482 shoot height in plants are due to reduced root growth and development under the presence of
483 chromium, which can result in the decreased transfer of water and nutrients to the aerial parts of
484 the plant. Also, increased chromium transport to the shoot can directly affect sensitive plant
485 tissues (leaf), photosynthesis, and cellular metabolism of the shoot, thereby reducing plant
486 height. The effect of soil chromium on plant growth and yield appears in various forms ,
487 including decreased water and nutrient uptake, impaired cell division, imbalance in critical
488 nutrient uptake and transport, plant inefficiency in selective uptake of inorganic nutrients,
489 increased production of free radicals , and oxidative damage to sensitive plant tissues such as
490 mitochondria, pigment content, RNA, DNA, lipids, etc.(Shahid et al. 2017).

491 All of these factors, individually or in combination, can influence on plant growth and
492 development as well as plant functioning at cellular and molecular levels.

493 The negative effect of chromium on root dry weight as a result of damage to root cells has also
494 been reported. Sundaramoorthy et al.(Sundaramoorthy et al. 2010).

495 found that chromium concentrations higher than 20 and 40 mg/l led to an inhibitory effect on
496 plant growth. Hexavalent chromium at a concentration of 200 mg/l also resulted in reduced root
497 growth in Asian rice (*Oriza sativa* L). In another study, the roots of *Zea maize* under hexavalent
498 chromium treatment were characterized by shorter roots, lower dry weight , and brown color
499 (Bhalerao &Sharma 2015). In the present study, different concentrations of chromium caused a

500 decrease in the dry weight of the roots in both studied species, confirming the results of other
501 studies.

502 Our results also showed that the applied amendments had a positive effect on the growth indices
503 for both studied species. addition, addition of organic matter to the culture medium serves as one
504 of the factors improving soil conditions for plant growth. Organic matter can increase soil
505 aeration and improve soil physical structure and cation exchange capacity , which varies
506 depending on the type of organic matter (Moameri et al. 2017).

507 Atiyeh and Lee (2002) in an experiment investigated the effect of organic fertilizers on tomato
508 and cucumber seedlings They indicated that application of these fertilizers had a positive effect
509 on dry shoot weight, which can be attributed to the positive effect of organic matter on the
510 physical properties and nutrient status of the soil.

511 Due to its nutrients and porous structure, specific surface , and strong functional groups, biochar
512 is anion that can increase crop yield by improving soil nutrients and physical and chemical
513 propertiesv(Xiang et al. 2017). The application of manure and biochar provided suitable
514 conditions in terms of nutritional balance for plants , so that plant growth and yield increased
515 significantly under these treatments, which is consistent with the results of other studies. Ahmad
516 et al. (Ahmad et al. 2014). Investigated the effect of biochar application on the biomass of maize
517 (*Zea mays* L.) and reported a significant increase in plant biomass compared to the control
518 sample. Also, Lehmann and Joseph (Lehmann et al. 2009). reported argued adding biochar to the
519 soil would preserve nutrients and thus increase plant growth, which is consistent with the results
520 of the present study so that we observed that amending compounds led to increased growth of the
521 two species of *S. persica* and *S. perspolitana*.

522 4.3 Accumulation of chromium in the roots and shoots of *S. persica* and *S. perspolitana*

523 This study showed that the accumulation of chromium in the roots and shoots of both species
524 increased with increasing the chromium concentration in the soil. Most of the chromium
525 accumulation occurred in the roots , which is consistent with the findings of Shahid et al.(Shahid
526 et al. 2017). who stated that in *Pisum sativum*, the chromium concentration in different parts of
527 the plant was as following: root > stem > leaf > seed. Liu et al.(Liu et al. 2009). observed the
528 highest concentration of chromium in the root cell wall and intercellular spaces of the rhizome.
529 Chromium fixation in plant roots is probably due to the formation of insoluble chromium
530 compounds in the plant. Some authors have reported that chromium storage in root cells may be
531 increased due to its storage in vacuoles of root cells, which may be a plant reaction to heavy
532 metal toxicity to limit the toxic potential of the heavy metals(Shahid et al. 2017). In the present
533 study, the effect of different chromium concentrations on the chromium accumulation in the two
534 studied species at different concentrations of 0, 5, 10, 15 was significantly different , so that the
535 accumulation of chromium in the shoots for both species increased with increasing chromium
536 concentration in the soil. Also, chromium accumulations in the shoots were significantly
537 different among the experimental treatments. During plant growth and development, higher
538 chromium accumulation was observed for the roots. Higher accumulation of chromium in the
539 roots compared to the shoots is a characteristic phenomenon observed in crops. Brunetti et
540 al.(Brunetti et al. 2011). stated that higher accumulation of chromium, copper, lead , and zinc in
541 the roots than the shoots represents the mechanism of plant tolerance and adaptation to high
542 concentrations of metals in the soil. Singh et al.(Singh et al. 2004) attributed the higher
543 accumulation of heavy metals in the roots than that of the shoot to the complexation of these
544 metals with sulfhydryl groups, which prevented the transfer of these metals to the shoot. This

545 prevents the transfer of heavy metals to the food chain. Although the transfer of chromium from
546 the plant roots to the shoots is minimal, the transfer of chromium within the plant tissue depends
547 on its chemical form. The decreased chromium transport to plant shoots may be due to the
548 conversion of hexavalent to trivalent chromium within plants due to the tendency of trivalent
549 chromium to attach to cell walls. As shown in table 7, trivalent chromium has a higher tendency
550 to be absorbed into the plant than hexavalent form, which may indicate the ability of *Salicornia*
551 to convert hexavalent chromium to trivalent form in its tissues. In terms of growth, the plant
552 needs to absorb trivalent chromium.

553 Also, the results obtained from this study showed that the amendments negatively influenced on
554 the accumulation of chromium in the roots and shoots in both species. The highest decrease in
555 chromium uptake was observed in biochar treatment. Biochar can be applied as an effective
556 amendment for stabilizing heavy metals in contaminated soils. Namgay et al.(Namgay et al.
557 2010). showed that adding biochar as an amendment to soils contaminated with heavy metals
558 could reduce the availability and absorption of these metals in plants. Zheng et al.(Zheng et al.
559 2012) also demonstrated that cadmium concentration in rice roots (*Oryza sativa* L.) decreased
560 after the application of biochar.

561 The addition of manure to the soil also resulted in decreased uptake of chromium compared to
562 soil without amendment. By increasing the cation exchange capacity of the soil, manure's
563 application can be effective in reducing the availability of heavy metals in soil and plants.
564 Containing a large amount of humus, manure's addition to the soil can alter the availability of
565 heavy metals by changing soil physical and chemical properties(Pinto et al. 2004). Antoniadis et
566 al.(Antoniadis et al. 2017) proved that the addition of manure would lead to reduced
567 bioavailability of chromium in the chicory (*Cichorium spinosum*) and serves as the best way for

568 minimizing the toxic effect of chromium on the plant. Therefore, adding manure may be the
569 most appropriate option for the stabilization of hexavalent chromium to inhibit the absorption of
570 this heavy metal by the plant. As a result, manure's addition leads to decreased bioavailability of
571 hexavalent chromium and slows down the phytoremediation process.

572 4.4 The effect of chromium toxicity on total phenolic content and photosynthetic pigments

573 The results of the present study showed that total phenolic content and photosynthetic pigments
574 significantly decreased as a result of heavy metal toxicity. This decrease was greater in *S.*
575 *perspolitana* than in *S. persica* and probably due to the higher accumulation of the metal in *S.*
576 *perspolitana*. Also, the organic compounds (biochar and manure) caused an increase in the
577 number of photosynthetic pigments with the higher contribution for biochar than manure, which
578 can be explained by biochar's effect on increasing growth and reducing the accumulation of
579 heavy metals in the plant. The non-enzymatic defense system of plants produces phenolic
580 compounds, and these antioxidant compounds react with free radicals and convert them to stable
581 radicals by giving up electrons to free radicals (Falleh et al. 2012).

582 In the present study, different chromium concentrations strongly affected the phenolic content of
583 the plant, so that the accumulation of chromium at the concentration of 15 mg/kg completely
584 suppressed the antioxidant properties of the plant, and the weakened defense system resulted in
585 decreased plant growth. Levizou et al. (Levizou et al. 2019) showed that the total phenolic
586 content in the marjoram plant (*Origanum vulgare*) decreased with the increasing chromium in
587 the soil. Islam et al. (Islam et al. 2016).

588 Attributed the decreased phenolic content to the malfunctioning of critical enzymes in the
589 biosynthesis of these compounds, which was consistent with the results of the present study
590 regarding the effect of chromium stress on reducing phenolic content.

591 In addition to inhibiting chlorophyll biosynthesis due to the presence of heavy metals, Dhir et
592 al.(Dhir et al. 2008) reported a decrease in iron and magnesium in the chlorophyll structure as a
593 possible reason for the decrease in chlorophyll efficiency. In the present study, the contents of
594 chlorophyll a and b significantly decreased under the effect of metal chromium compared to
595 control plants, which is consistent with the findings of other studies.

596 Carotenoids are lipophilic secondary metabolites and the second most abundant pigment in
597 nature. While chlorophylls are classified as optical pigments, carotenoids in stressful situations
598 mainly contribute to protect the chlorophyll photosynthetic system. To quench the photodynamic
599 reactions that can lead to chlorophyll loss, carotenoids replace peroxides to prevent the
600 chloroplast membrane from collapsing (Duarte et al. 2012). The results of our study showed that
601 chlorophyll *a*, *b* , and carotenoids contents in the shoots of both *S. persica* and *S. perspolitana*
602 decreased with increasing the chromium concentration in the soil, confirming the results reported
603 by many authors regarding the effect of chromium on plant pigments such as maize (*Zea mays* cv
604 704)(Rahmaty &Khara 2011) and wheat (*Triticum aestivum* L) (Subrahmanyam 2008).

605 4.5 *The effect of Salicornia on the phytoremediation process*

606 The results of phytoremediation in the present study showed that *S. persica* and *S. perspolitana*
607 were able to absorb heavy metals from the soil with higher uptake of metals in the roots than
608 shoots for both species. Considering that the TF index was lower than 1 for all treatments, one
609 can conclude that *S. persica* and *S. perspolitana* were not suitable for phytoextraction of metal
610 chromium. Also, the BCF factor for metal chromium was higher than 1, indicating that both *S.*
611 *persica* and *S. perspolitana* stabilize the metal in the soil during the stabilization process. Both
612 species mostly absorb trivalent form of chromium , indicating the high efficiency for converting
613 hexavalent to trivalent form.

614 Yoon et al. (Yoon et al. 2006) found that plants with high accumulation and low transfer factors
615 can stabilize heavy metals. Plant species that have been used to stabilize heavy metals so far
616 include *Festuca rubra* (lead and zinc) and *Brassica juncea* (cadmium) (Ghosh & Singh 2005) ,
617 which is consistent with our findings in this research.

618 **5. Conclusions**

619 This study aimed to investigate the accumulation capacity of chromium metal in shoots and roots
620 of *S. persica* and *S. perspolitana*. In this study, manure and biochar amendments were used to
621 immobilize and stabilize metal chromium in contaminated soils. Chromium concentration in
622 shoots and roots in *S. persica* and *S. perspolitana* were significantly affected by the
623 concentration of this metal in the soil. In all observations, chromium accumulation in shoots and
624 roots of the two studied species increased with increasing soil metal concentration. Both *S.*
625 *persica* and *S. perspolitana* can be used for phytoremediation of soils contaminated with low
626 concentrations of chromium as these two species could not tolerate the accumulation of
627 chromium at high concentrations. The results also indicated higher chromium accumulations in
628 the roots than the shoots for both *S. persica* and *S. perspolitana* species. Therefore, these two
629 species can change the rhizosphere environment of the root and influence on concentration and
630 bioavailability of metal chromium. The results of the present study showed a characteristic
631 feature of converting hexavalent to trivalent chromium for both studied species. Accumulation of
632 total chromium, trivalent chromium , and hexavalent chromium in the shoots was higher in *S.*
633 *persica* than *S. perspolitana*. The photosynthetic pigments and total phenolic content in both
634 studied species also decreased with increasing chromium concentration.

635 Our findings also showed that the use of organic compounds could improve the growth and yield
636 of *S. persica* and *S. perspolitana* and , at the same time , decrease accumulation of chromium in

637 roots and shoots. Application of biochar and manure in *S. persica* and *S. perspolitana* species in
638 the reclamation of soils contaminated with heavy metals caused increases of 49.05% and 28.30%
639 in dry shoot weight, 53.12% and 14.06% in root dry weight, 23.93% , and 13.29% in chlorophyll
640 *a*, 22.72% and 13.63% in chlorophyll *b* and 16.93% and 6.92% in carotenoids content compared
641 to the controls (without amendment). Biochar and manure treatments also reduced the total
642 phenolic content by 11.83% and 13.12% in *S. persica* and *S. perspolitana*, respectively. Biochar
643 and manure treatments caused decreases by 36.69% and 17.98% in accumulation of metal
644 chromium in the roots, and 84.83% and 72.55% in the shoots, respectively, compared to controls
645 (without amendment). In general, the application of biochar treatment had a greater effect on
646 reducing the accumulation of heavy metals than manure treatment because biochar has a higher
647 ability to absorb metals due to its porous structure. As a result, it reduces the bioavailability of
648 metals in both species. Therefore, using organic amendments in soils contaminated with heavy
649 metals can stabilize these metals in the soil and prevent entering them into the food chain.

650

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662 Fahime Ashrafi: Investigation, Project administration, Resources, Validation Writing – original
663 draft, Software, Formal analysis, funding acquisition.
664 Ava Heidari: Supervision, Visualization, Writing – review & editing, Conceptualization, Data
665 curation, Methodology, Project administration, Funding acquisition.
666 Mohammad Farzam: Supervision, Data curation, Validation.
667 Alireza Karimi: Supervision, Data curation, Validation.
668 Malihe Amini: Writing – review & editing, Formal analysis, Validation

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Figures

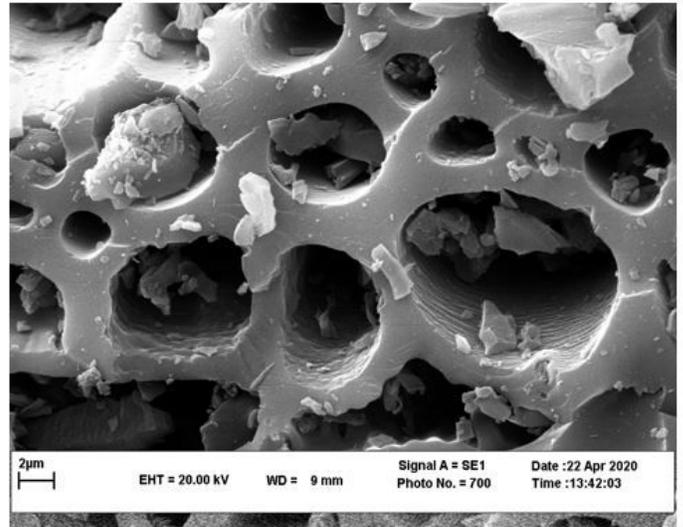
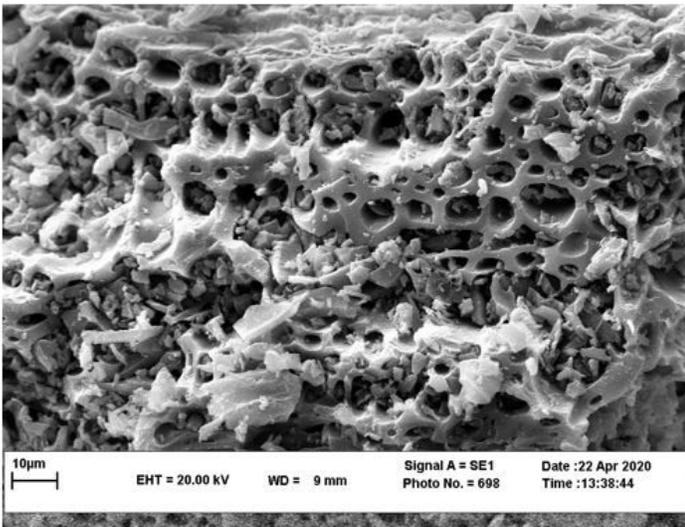


Figure 1

Scanning electron microscopy (SEM) of biochar surface