

Acute phase proteins and total antioxidant capacity in free-roaming cats infected by pathogenic leptospire

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Abstract

Background: Leptospirosis is a neglected but widespread zoonotic disease throughout the world. The vast majority of mammals are hosts of *Leptospira spp.*, including domestic cats, species in which no consensus has been reached on the clinical presentation or diagnosis of the disease. The study of acute-phase proteins (APPs) and biomarkers of oxidative status would contribute to knowledge about the course of the disease in cats. In this report, we evaluated four APPs: Serum amyloid A-SAA, Haptoglobin-Hp, albumin and Paraoxonase 1-PON1 and the antioxidant response through Total Antioxidant Capacity-TAC, in 13 free-roaming cats naturally infected by pathogenic leptospires and 19 leptospires-free cats, through a Principal Component Analysis (PCA).

Results: The PCA retained two principal components (PC1 and PC2), explaining 60.1% of the observed variability of the inflammatory proteins and the antioxidant TAC. PC1 was mainly associated with an inflammatory process, whereas PC2 correlated with the antioxidant status. The most contributing variables in PC1 were albumin (27.46%), SAA (24.71%), Hp (21.56%) and PON1 (20.69%). Variables with significant contribution to the PC2 were the antibody titre against *Leptospira spp.* (48.41%) and TAC (35.04%). Overall, the PCA revealed differences in inflammatory and antioxidant biomarkers in cats naturally infected with leptospires compared to leptospires-free cats.

Conclusions: Increases in Serum SAA, Hp, and decreases in serum albumin concentrations and PON1 activity indicate an acute phase response in infected, DNA positive cats. Moreover, we found an increase in TAC serum concentrations indicating an antioxidant response in this infection, which was proportional to the antibody titre and not to the presence of bacterial DNA.

Background

Leptospirosis caused by a spirochaetal bacterium of the genus *Leptospira* is a common and widely distributed zoonoses affecting livestock, domestic, and wild animals throughout the world [1]. A wide variety of mammalian species may be susceptible to *Leptospira spp.* infection including domestic cats [2] in America [3-5], Asia [6-7], Australia [8] and Europe [9-10].

Whilst, *Leptospira spp.* infection in cats has been suspected for some time [11], however, it was two decades before confirmed by microagglutination test (MAT) and PCR [12-13]. Nowadays, the role of cats as a reservoir host to *Leptospira spp.*, is well established [12,14-17].

Leptospires spread throughout the entire body, reproducing in different organs such as the kidneys, liver, central nervous system, eyes, and reproductive system. In cats, most of the *Leptospira spp.* infections are asymptomatic and unspecific and rarely results in severe organ damage [18]. Few works have described clinical signs related to the infection in cats (e.g., renal injury) [4,12,16,19], most of them being nonspecific [17,20-21]. As a result, there is no consensus on the clinical presentation, development and diagnosis of leptospirosis in domestic cats. Hence ancillary tests inflammatory proteins and biomarkers of oxidative status could, useful to identify the active state of the infection in cats.

Acute-phase proteins (APPs) are increasingly used as tools for the detection of inflammatory diseases in veterinary clinical practice. Use of APP profiles involving at least a major protein (proteins that increase 10-100-fold during the inflammatory response), a moderate protein (increase two-10 fold) and a negative protein (fall in concentration) is highly recommended to differentiate between pathologic states [22]. APPs are species-specific; in cats, Serum Amyloid A (SAA) is a major acute protein, Haptoglobin (Hp) is a positive moderate one acute phase protein, and albumin and paraoxonase-1 (PON1) [22-26] are negatives ones acute phase proteins, with the latest also considered a biomarker of oxidative stress [27]. Several investigations have been carried out concerning the usefulness of APPs in infectious diseases in cats, providing valuable information [28-30].

To date, no work has observed the relationship between leptospirosis and APPs in cats. In dogs, however, C-reactive protein/ Haptoglobin ratio (CRP/Hp) and albumin serum concentrations increase after *Leptospira interrogans* Australis serogroup infection [31].

Biomarkers of redox status have been studied in cats since they may contribute to morbidity in many diseases [30,32-35]. Oxidative stress is defined as reactive oxygen species (ROS) over antioxidant defence mechanisms. It can result from an excess of ROS, a reduction in antioxidants, or both [35]. The measurement of the redox status in companion animals has been addressed with novel approaches in the last few years using the TAC assay [35-36]. TAC represents the sum of the activities of the different antioxidants and the antioxidative effects provided by the interactions between individual antioxidants [37-38], also used to evaluate the antioxidant response. Cats seem to be more susceptible to oxidative stress and damage, probably due to the presence of eight reactive and fragile sulfhydryl groups on its haemoglobin molecule and to the particular splenic structure of the species [39-40].

The objective of this paper was to evaluate a panel of acute Phase Proteins (APPs: Serum Amyloid A –SAA, Haptoglobin – Hp, Paraoxonase1 -PON1 and Albumin) and Total Antioxidant Capacity -TAC in free-roaming cats naturally infected by pathogenic leptospires using a principal component analysis (PCA).

Methods

Selection of cases

Thirty-two serum samples from domestic short-haired cats that had been collected during a previous study on *Leptospira spp.* prevalence in cats in Spain were used in the current study [10]. Cats were part of a free-roaming cat spay program and shelter's animals neutering program. Blood samples were taken under anaesthesia for neutering. Sampling collection was performed under the guidelines of the Ethical Committee Animal Care and Research, Autonomous University of Barcelona, approval number CEEAH, code 2939. Besides, written informed consent was obtained from the shelters to use the animals in the study.

Serum samples were stored at -80°C until analysis. MAT was performed by direct reading following a technique described before [41]. DNA extracted from cats' biological materials were tested with TaqMan real-time PCR described by Ahmed et al., 2020 [42].

Samples were divided into three groups: Group 1 ($n = 8$), cats positive to anti-leptospiral antibodies detected by a MAT consisting of 27 serovars, belonging to 20 serogroups and 8 species of *Leptospira*. Pathogenic *Leptospira spp.* DNA in urine or blood was not detected in these cats. Group 2 ($n = 5$), cats positive to *Leptospira spp.* DNA detected by PCR in blood or urine. These cats had no antibodies against *Leptospira spp.* Group 3 or control group ($n = 19$), sera from leptospires-free cats. The absence of *Leptospira spp.* infection in the animals of this group was verified by serology (negative antibodies against *Leptospira spp.* by MAT) and PCR of blood and urine (negative leptospiral DNA amplification). All animals in the study $n = 32$, were tested against feline immunodeficiency and leukaemia viruses (SNAP FIV/FeLV Combo Test®). According to the medical record, all animals in the present study were free-roaming cats and had unremarkable findings upon physical exam.

APPs analysis

SAA: Serum amyloid A concentrations were determined by a human turbidimetric immunoassay (LZ-SAA; Eiken Chemical Co., Tokyo, Japan), adapted to an automated analyser (Olympus 2700). This method had been previously validated for use in cats [43]. Serum concentrations lower than 5 µg/ml were considered normal for cats; the limit of detection was set at 0.38 µg/ml [44].

Hp: Serum Haptoglobin concentrations were determined by the use of the haemoglobin-binding method with a commercial kit (Tridelta Development Ltd., Brey, Ireland). The method was previously validated for use in cats [45]. Serum concentrations lower than 3 g/L were considered normal; the limit of detection considered was 0.0088 g/L.

Albumin: Serum albumin was determined using a commercially available kit (Albumin OSR 6102; Olympus Life and Material Science Europe GmbH, Irish branch, Ennis, Ireland) following instructions of the manufacturer.

PON1. Serum PON1 activity was determined by measuring the hydrolysis of p-nitrophenyl acetate to p-nitrophenol, following a previously described method [46], validated in cats [45]. The rate of formation of p-nitrophenol was determined at 405 nm after 250 seconds in an automated chemistry analyser (Olympus 2700). The limit of detection was 0.3 IU/ml. Serum concentrations between 3.8 to 7.3 IU/ml were considered normal for cats [27,45].

Antioxidant analysis

TAC: Total antioxidant capacity was determined by the assay Trolox equivalent antioxidant capacity (TEAC1). It is based on the principle that when ABTS (2,2'-azino-bis (3-ethylbenzo-thiazoline-6-sulfonate acid) is incubated with a proper chemical an ABTS radical (ABTS*) is formed, previously described for humans [36] and validated for cats [27]. Serum concentrations higher than 0.35 mmol/L were considered normal; the limit of detection considered was 0.02 mmol/L.

All acute phase proteins and antioxidant analyses were performed on an automated biochemistry analyser (Olympus AU600, Olympus Diagnostic, GmbH).

Statistical analysis

Principal component analysis (PCA) is a dimensionality reduction technique using a linear transformation applied to multidimensional data. The original set of variables (e.g., our set of inflammatory markers, the antioxidant biomarker and antibody titres against *Leptospira spp.*), is reduced to a smaller number of derived variables that may be readily visualised in 2- or 3- principal components containing the highest observed variance. These derived variables can then be compared between categorical variables (e.g., *Leptospira spp.* infection status), through a Student t-test. Median, SD, interquartile range and range (min-max) of our quantitative variables were also calculated. PCA analysis was performed using the libraries "car" 3-0-6 version [47] "FactomineR" 2.2 version [48], "factoextra" 1.0.6 version [49] and "ggplot2" 3.2.1 version [50] of the R Statistical software 3.6.2 version [51].

Results

Baseline characteristics of the cats

The 32 serum samples included in the study comprised 11 females and 21 males between 6 months to 12.5 years old. The final distribution in the groups was 1 female and 7 males (ranging from 6 months to 7 years old) in Group 1 (anti-leptospiral antibodies); 3 females and 2 males (ranging from 6 months to 1 year old) in Group 2 (positive *Leptospira spp.* DNA in blood or urine) and 7 females and 12 males (ranging from 6 months to 12.5 years old) in Group 3 (leptospire-free cats or control group). Only one cat, included in Group 1, was FIV positive; all others were negative for both FIV/FeLV. More details about the distribution of the groups and the involved serovars (Group 1) are shown in Table 1 and Table 2.

Table 1. Distribution of groups in cats naturally infected by leptospire and leptospire-free cats.

		Total population n: 32	Group 1 n: 8	Group 2 n: 5	Group 3 n: 19
Gender	Male	21	7	2	12
	Female	11	1	3	7
Age (y.o.)	Mean	2.6	2.7	0.63	3.0
	Min-Max	0.5-12.5	0.5-7	0.5-1	0.5-12.5
	SD	3	2.43	0.21	3.46

y.o.= years old, (group 1) = anti-leptospiral antibodies (group 2) = positive *Leptospira spp.* DNA in blood or urine, (group 3) = leptospires-free cats or control group.

Table 2. Description of leptospiral serovars and PCR results by group of cats

	Cat	Gender	Group 1 n: 8	Group 2 n: 5	Group 3 n: 19
Infecting leptospiral serovars			Serovar and titre	Seronegative	Seronegative
	1	Female	Ballum 1:20		
	2*	Male	Bratislava 1:20 Cynopteri 1:20		
	3	Male	Cynopteri 1:20		
	4	Male	Cynopteri 1:20		
	5	Male	Cynopteri 1:40		
	6	Male	Ballum 1:20		
	7*	Male	Pomona 1:20 Proechimys 1:20		
8	Male	Sejroe 1:20			
PCR	5 cats	3 Female 2 Male	Negative	1 in blood 4 in urine	Negative

(group 1) = anti-leptospiral antibodies (group 2) = positive *Leptospira spp.* DNA in blood or urine, (group 3) = leptospires-free cats or control group. *Cat positive to more than one serovar.

Serum concentrations of APPs, TAC and antibody titre against *Leptospira spp.* obtained in the different groups of the study are shown in Table 2. No concentrations lower than the limit of detection for any of the APPs and TAC studied were obtained.

Table 3. Serum concentrations of APPs, TAC and anti-leptospiral antibody titre. (mean, minimum and maximum and IQR) in cats naturally infected by leptospires (Groups 1 and 2) and leptospires-free (Group 3).

	SAA (µg/ml)	Hp (g/l)	Albumin (g/l)	PON1 (IU/ml)	TAC (mmol/l)	Antibody titre
Group 1 <i>n</i> = 8	0.62	4.05	2.86	3.91	0.64	22.5
MEAN						
Min-Max	0.10-1.60	2.49-6.29	2.31-3.3	1.87-6.02	0.46-0.80	20-40
IQR	0.82	1.23	5.3	1.33	0.15	0
Group 2 <i>n</i> = 5	0.17	4.47	2.70	3.05	0.53	0
MEAN						
Min-Max	0.1-0.20	1.97-8.11	2.12-3.22	0.14-4.84	0.43-0.70	0
IQR	0.05	2.03	5.2	0.86	0.1	0
Group 3 <i>n</i> = 19	1.43	3.60	3.20	5.35	0.58	0
MEAN						
Min-Max	0.10-6.00	1.64-4.89	2.48-4.49	3.40-7.41	0.44-0.7	0
IQR	1.25	1.68	4.4	1.43	0.12	0

Min = Minimum, Max = Maximum, IQR= Interquartile range, SAA= serum amyloid A, Hp= haptoglobin, PON1= Paraoxonase1, TAC= Total antioxidant capacity, (group 1) = anti-leptospiral antibodies (group 2) = positive *Leptospira spp.* DNA in blood or urine, (group 3) = leptospire-free cats or control group.

Principal component analysis

Our PCA retained two principal components (PC1 and PC2), explaining 60.1% of the observed variability of our set of inflammatory proteins and antioxidant biomarker. PC1 and PC2 accounted for 35.2% and 24.9% of the observed variance and were mainly associated with the inflammatory process and the antioxidant response, respectively (Table 4 and Fig. 1). The most contributing variables in the PC1 were albumin, SAA, HP, and PON1, whereas anti-leptospiral antibodies and serum concentration of TAC did not contribute significantly to the first dimension of our PCA (Table 4). Regarding PC2, anti-leptospiral antibodies and the total antioxidant capacity were the essential variables in terms of contribution to the component (Table 4 and Fig. 2).

The presence of pathogenic *Leptospira spp.* DNA in blood or urine (positive or negative) was a supplementary variable significantly related to PC1 but not to PC2 (Table 4). On the other hand, the presence of antibodies against *Leptospira spp.* by MAT (positive or negative) was a supplementary variable correlated to PC2 but not to PC1 (Table 4).

Table 4. Contribution of variables to PCA and dimensions description

	PC1			PC2		
<u>Active variables</u>	Contribution (%)	Correlation	P-value	Contribution (%)	Correlation	P-value
Albumin	27.46	0.76	4.07e-07	0.28	-0.06	> 0.05
SAA	24.71	-0.72	2.9e-06	11.72	-0.42	1.7e-02
Hp	21.56	-0.67	2.26e-05	0.16	0.05	> 0.05
PON1	20.69	0.66	3.71e-05	4.38	-0.26	> 0.05
Antibody titre	3.07	-0.25	> 0.05	48.41	0.85	7.64e-10
TAC	2.51	0.23	> 0.05	34.05	0.72	2.95e-06
<u>Supplementary variables</u>	Contribution (%)	R ²	P-value	Contribution (%)	R ²	P-value
MAT (+/-)	na	na	> 0.05	n/a	0.62	0.005
PCR (+/-)	na	0.22	0.006	n/a	>0.05	>0.05

Contribution of the inflammatory biomarkers (Albumin, Hp and SAA and PON1), antioxidant biomarker (TAC) and anti-leptospiral antibodies (titre) to the first (PC1) and second (PC2) dimensions of principal component analysis, exploring the response of cats naturally infected by pathogenic leptospires and leptospires-free cats. The outcomes of the MAT test (+/-) and the amplification of pathogenic *Leptospira spp.* DNA through a PCR analysis (+/-) were considered supplementary variables, i.e., not considered for the construction of the factorial axes but used to test statistical differences in the PCA scores. The “na” acronym indicates not applicable.

Discussion

It is known that cats can sporadically suffer from leptospirosis [12,16,17], but while the clinical presentation of the disease is rare, it may be missed [2]. Asymptomatic cats can shed the bacteria through urine [9,15], and unlike infected dogs, antibody titres against *Leptospira spp.* detected in sub-clinically infected animals are low in the species [6-7,20]. Cats infected by *Leptospira spp.* may become an incidental or reservoir host. The incidental host develops an acute disease state with mild to moderate clinical signs, leptospiraemia (1-7 days/sub-acute state), leptospires urinary shedding (10 day-about a month/late acute state) and antibodies titres. Reservoir cats usually exhibit a chronic state of infection and have no clinical signs of disease or leptospiraemia, but they do have long-term and intermittent leptospires urinary shedding and low or absent antibodies titres [21].

APPs analysis has been studied in cats with infectious diseases [27-28, 52-53] and its use is highly recommended to determine the active state of disease. TAC as an antioxidant biomarker can contribute to the morbidity of many diseases and is likely to become, along with APPs, an important component in the active state detection of diseases, including leptospirosis in cats.

A PCA statistical approach is based on the fact that cats naturally infected by *Leptospira spp.* may be grouped into different patterns that may be associated with different serum concentrations of APPs and TAC as antioxidant biomarkers. The information obtained could facilitate the understanding of the pathogenesis of *Leptospira spp.* and infection course in cats.

PC1, accounting for 35.23% of the data variance, was significantly associated with the APPs profile or inflammatory pattern. The supplementary qualitative variable PCR (negative/positive) was significant in the PC1 (R^2 0.22, P -value 0.006), suggesting a direct relationship between the presence of the bacteria in blood or urine and the APPs responses.

Patterns of inflammation described in preliminary works were determined and confirmed by our PCA analysis in PCR positive cats (blood or urine). They include a positive correlation found between SAA and Hp (-0.72 and -0.67 respectively), albumin and PON1 (0.76 and 0.66 respectively) [30,54], and negative correlation between SAA-Hp and albumin-PON1 [54-55]. It is well known that albumin falls gradually with extensive inflammation, with the reduction in concentration being more noticeable in chronic inflammatory disease [56]. Serum PON1 activity was associated with a negative APP and did not play a role as a biomarker of oxidative status in PC1 or PC2, in contrast to previous studies [27,57].

Likewise, based on *Leptospira spp.* pathogenesis, it is possible to assume that PCR positive animals had a sub-acute infection status (*Leptospira spp.* present in blood/ early stage of the infection) or were mostly reservoirs (*Leptospira spp.* present in urine/intermittent urinary shedding). Based on the PC1 results, animals of group 2 reflected an active infection, compared to those infected with anti-leptospiral antibodies (group 1), which did not reflect an active state of infection. The difference in inflammatory patterns between the two groups of infected cats is very likely due to differences in bacterial load and the infecting serovar involved, among others. In diseases such as FIV, the relationship between antigenic load and high serum levels of APPs, and more specifically SAA, has been shown [58]. Likewise, *Leptospira* serovars involved in the infection have also been reported to influence the inflammatory response in other species. In dogs, it has been shown that serogroup Pomona [59] and Icterohaemorrhagiae [2] trigger the strongest inflammatory responses and have the worst prognosis. In cats, this remains poorly understood [21]. In our study, the infecting serovar or serogroup was not detected in the PCR positive cats as they had no anti-leptospiral antibodies.

PC2, accounting for 24.9% of the data variance, was significantly associated with the oxidative status pattern. A positive correlation was found between antibody titre and serum concentration of TAC (0.85 and 0.72, respectively) with PC2; thus, it is very likely that as antibody titres against *Leptospira spp.* increased, endogenous antioxidant synthesis increased as well. Likewise, the supplementary qualitative variable MAT (+/-) was significant in the PC2 (R^2 0.62, P -value < 0.05).

Cats with low anti-leptospiral antibodies titre (ranging from 1:20 to 1:40), probably at a resolution stage of infection, had increases in serum concentrations of TAC, most likely, to counteract the oxidative state associated with the inflammation. In human and companion animals, it has been reported that high serum concentrations of TAC are due to counteracting the increases in oxidants, while decreases are attributed to a persistent state of oxidative stress [27,38, 60].

Seropositive cats seemed to be at risk of developing oxidative stress as antioxidant response measured by the TEC1 assay was above the cut-off point, unlike DNA positive animals. Considering that one of the 8 seropositive cats, also tested positive for FIV infection, it is likely that in this cat the TAC serum values were also increased by the co-infection with the virus.

Antibody titres, in which the antioxidants become exhausted, showing a decrease in serum concentration of TAC and leading to an oxidative status [30], are not yet established. The little information available for cats has used serum concentrations of TAC to assess the antioxidant components globally. Some authors have reported a decrease in serum concentrations of TAC associated with oxidative stress [27, 30, 55, 61].

Despite the positive correlation found in our study between antibody titres against *Leptospira spp.* and serum concentrations of TAC with PC2, it is not possible to establish whether the variation in the serum concentration of TAC obtained in the two groups of animals naturally infected by *Leptospira spp.* (Group 1 and 2) is due to the inflammatory response caused by *Leptospira spp.* infection or whether this is due to the differences in the available assays for measuring TAC [57].

Infectious diseases other than FIP/ViLEF were not ruled out. As mentioned above, diseases such as feline infectious peritonitis (FIP) [27,28], FIV [34] and haemoplasmas [53] have also shown an increase in some of these biomarkers. As well as non-infectious inflammatory diseases [32, 43, 54]. In the present study, Cats health status was based on physical exam findings and biochemistry, so these results need to be interpreted with caution.

One of the limitations of our study is the small sample size (total and by groups), and the lack of follow up of the serum values of the biomarkers measured, due to the nature of the animals in the study.

Further studies should be undertaken to elucidate, the *Leptospira* serovars role

in cats and their involvement in the inflammatory response through serum concentrations of APPs and TAC as oxidative stress markers by using different assays and enzymes implied in the antioxidant response. To the authors' knowledge, this is the first report measuring the serum concentration of APPs and TAC in cats naturally infected by pathogenic leptospires.

Finally, based on PCA analysis results and the PON-1 arrangement on the inflammatory PC1, further experimental studies are needed to estimate its importance as a negative APP in cats.

Conclusions

The use of APPs in *Leptospira spp.* DNA positive cats help to identify the active state of infection. *Leptospira spp.* DNA positive, are not at risk of developing oxidative stress. Cats with anti-leptospiral antibodies did not reflect an active inflammatory pattern but instead indicate an antioxidant response in this infection, which is proportional to the antibody titre.

To conclude, this study provides additional information to the limited data available on the use of acute phase proteins and TAC in cats naturally infected by leptospires, which could be helpful for the understanding and diagnosis of leptospirosis in cats.

Abbreviations

APPs: Acute phase proteins, **CBC:** Complete blood count, **CRP/Hp:** C reactive protein-Haptoglobin ratio, **DNA:** Deoxyribonucleic acid, **FeLV:** Feline leukaemia virus, **FIV:** Feline infectious virus, **GPox:** glutathione peroxidase, **Hp:** Haptoglobin, **IQR:** Interquartile ranges, **LPS:** Lipopolysaccharides, **MAT:** Micro agglutination test, **PC1:** Principal component 1, **PC2:** Principal component 2, **PCA:** Principal components Analysis, **PCR:** Polymerase chain reaction, **pNA:** p-nitrophenyl acetate, **PON1:** Paraoxonase1, **ROS:** Reactive oxygen species, **SAA:** Serum amyloid A, **SOD:** Superoxide dismutase, **TAC:** Total antioxidant capacity, **TEAC1:** Trolox equivalent antioxidant capacity.

Declarations

Ethics approval and consent to participate

Sampling collection was performed under the guidelines of the Ethical Committee Animal Care and Research, Autonomous University of Barcelona, approval number CEEAH, code 2939. Besides, written informed consent was obtained from the shelters to use the animals in the study.

Consent for publication

Not applicable

Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Competing interests

The authors declare that they have no competing interests

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Authors' contributions

All authors read and approved the final manuscript. Study design: AM, RC, JP. Data collection: AM, MG, AA, AT, JC. Statistical analysis: AM, ES, JP. Technical review, interpretation of the data and preparation of this manuscript: AM, RC, ES, JP, AT, JC.

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Figures

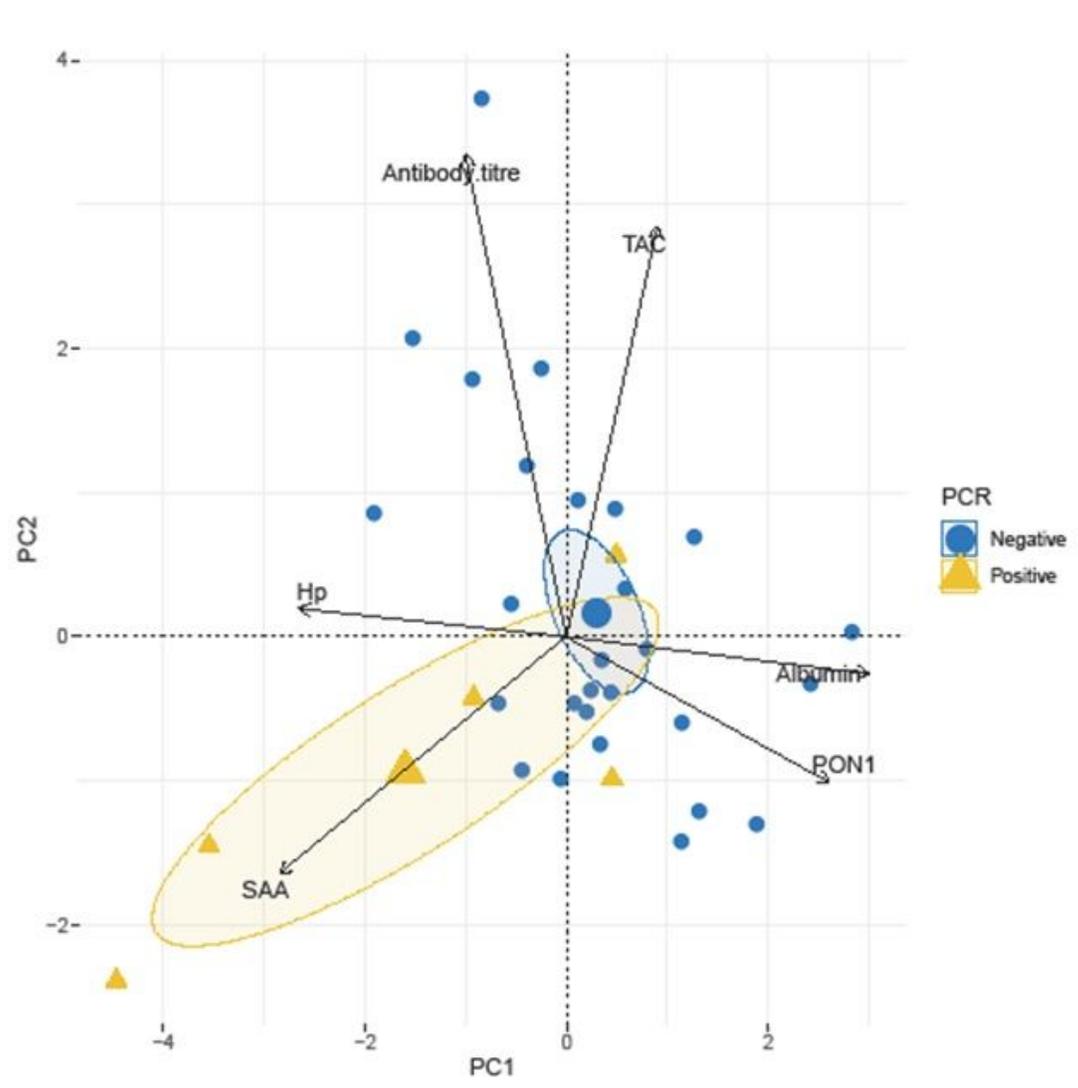


Figure 1

Biplot of PCR as supplementary variable in the PCA. Biplot representing the relationships between the PCR outcome of the pathogenic leptospire (positive/negative), and PC1 and PC2 dimensions of a PCA exploring the relationships among inflammatory biomarkers (Albumin, PON1, HP and SAA), an antioxidant biomarker (TAC), and antibody level against *Leptospira* spp. (antibody titre) in 13 cats naturally infected by pathogenic leptospire and 19 leptospire-free cats from Spain. Neither TAC (total antioxidant capacity) nor the antibody titres against *Leptospira* spp. contributed to the PC1.

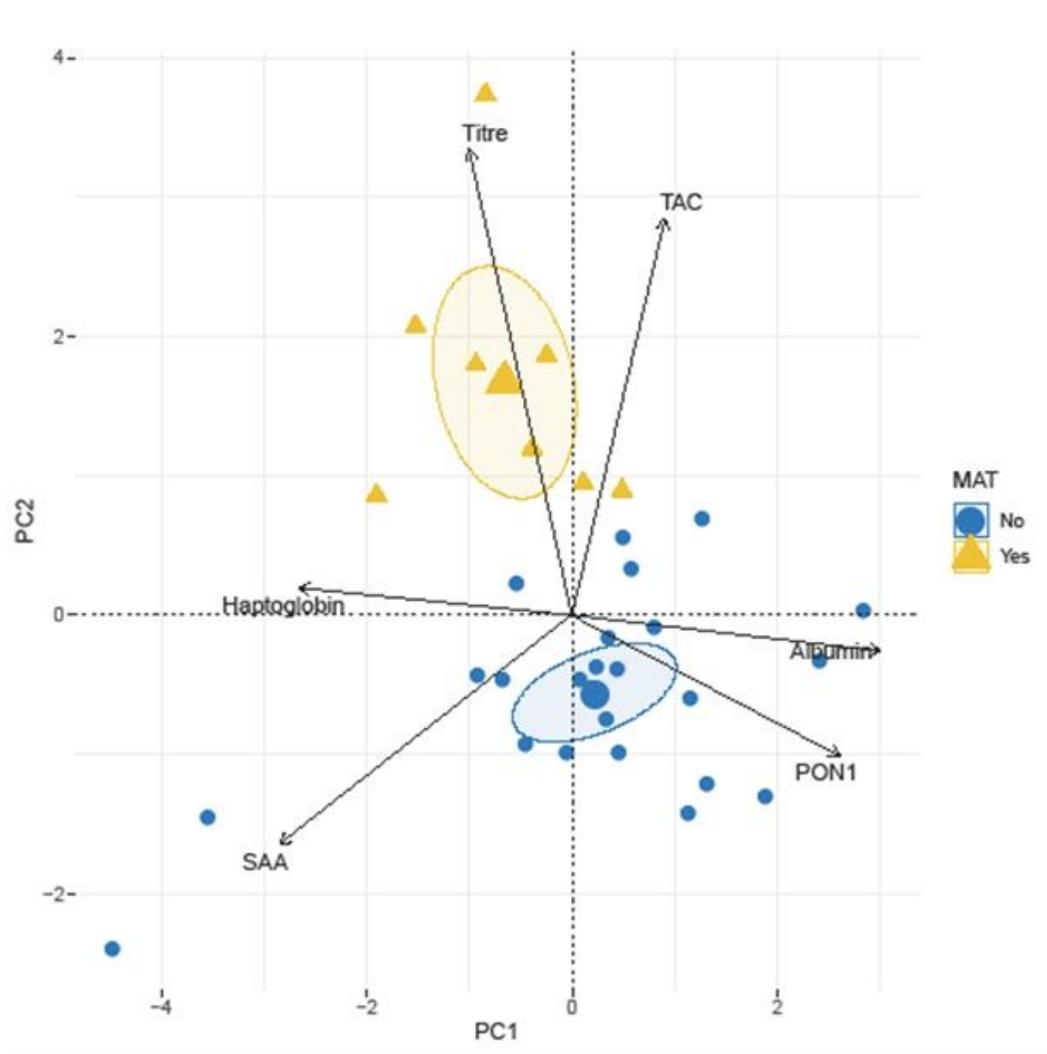


Figure 2

Biplot of MAT as supplementary variable in the PCA. Biplot representing the relationships between MAT outcome (positive/negative), and PC1 and PC2 dimensions of a PCA exploring the relationships among inflammatory biomarkers (Albumin, PON1, HP and SAA), an antioxidant biomarker (TAC), and antibody level against *Leptospira* spp. (antibody titre) in 13 cats naturally infected by pathogenic leptospires and 19 leptospires-free cats from Spain. Inflammatory biomarkers did not contribute to PC2.