

High prevalence of pre-existing HBV polymerase mutations in pregnant women do not limit the telbivudine treatment efficacy

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Abstract

Background: HBV resistant mutants in treatment-naïve patients may lead to antiviral treatment failure. It's not clear if HBV resistant mutants present in the pregnant women before the antiviral treatment and the influence of the pre-existing resistant mutants on the short-term Telbivudine (TBV) therapy during the late pregnancy.

Method: We enrolled 73 pregnant women with high HBV DNA load and TBV treatment during the pregnancy from 2012 to 2015 in this retrospective study. All subjects were followed at least 52 weeks postpartum. The UDPS was used to detect the HBV mutations before and after the TBV treatment.

Results: Before TBV treatment, the complexity of HBV quasispecies of all subjects was 0.40 ± 0.09 . At primary drug resistance mutation sites, 41.1% (30/73) and 53.4% (39/73) subjects had rtM204I/V and rtN236T/A detected, respectively; 9.6% (7/73) patients had more than 20% frequency mutation of rtM204I/V, which was also similar with high frequency of rtN236T/A mutation (41.1% vs. 53.4%, $P = 0.136$; frequencies > 20%: 9.6% vs. 5.5%, $P = 0.347$). After TBV treatment, 71.2% (52/73) subjects still had HBV DNA load $\geq 10^3$ IU/mL at delivery. Among them, 75.0% patients with rtM204I positive had HBV DNA load $\geq 10^3$ IU/mL at delivery, which was comparable with the subjects without rtM204I (75.0% vs. 70.8%, $P = 0.710$). No changes were found in the frequencies and the complexity of HBV quasispecies of rtM204I mutation after the TVB treatment.

Conclusion: The prevalence of pre-existing drug resistant mutations among the pregnant women was high using UPDS. However, the pre-existing HBV mutation had limited influence on the efficacy of short-term TBV treatment, and TBV treatment during late pregnancy seemed not to increase the risk of emerging HBV resistant mutants.

1. Introduction

Current guidelines recommend that pregnant women with high HBV DNA levels should accept antiviral prophylaxis in gestation [1–3]. It was recommended for pregnant women to decrease the HBV DNA load below a relative safe threshold for the prevention of HBV mother-to-infant transmission (MTIT) during the third trimester [4].

The nucleoside/nucleotide analogues (NAs) are able to suppress HBV replication by inhibiting the viral reverse transcriptase (RT), however, HBV RT has no proofreading activity, it increases the HBV mutations and promotes genetic diversity, which may cause drugs resistance [5, 6]. Studies showed that some resistance mutations related to NAs therapy might already be present in treatment-naïve patients [7–9]. It is reported that YMDD mutations was present in a subgroup of NA-naïve patients with a frequency ranging from 3–27% [10–13]. HBV resistant mutants in treatment-naïve patients may lead to drug resistance and treatment failure [14]. It's not clear if HBV resistant mutants present in the pregnant women before the antiviral treatment.

Telbivudine (TBV) classified as category B is one of the NAs that recommended by the Asian-Pacific clinical practice guidelines and widely used to prevent MTIT. On the other hand, TBV and lamivudine (LAM) are considered with low genetic barrier to HBV resistance. LAM was observed drug-resistant viral variants among mothers with high HBV load received LAM treatment from 22 to 88 days during the pregnancy [15], while the study of TBV is still limited. Yingxia Liu *et al.* reported that one of 50 high HBV DNA loads subjects developed rtM204I drug-resistance mutation after receiving TBV treatment, but the time duration that the patient received TBV treatment in the study was not clear [16]. Another prospective study did not find the rtM204 mutations among the participants started TBV 600 mg/day at week 20 to week 32 of gestation and stopped TBV one month postpartum [17]. The clinical impact of short-duration TBV usage should be studied further in the high risk pregnant women.

The objective of this study is to assess the prevalence of HBV pre-existing resistant mutants in pregnant women and explore the influence of the pre-existing resistant mutants on the efficacy of short-term TBV therapy during the pregnancy. We used ultra-deep pyrosequencing (UDPS) to sequence HBV and detect low level (< 1.0%) clinically relevant variants within complex viral populations.

2. Materials And Methods

2.1 Participants

This is a retrospective study, all data were collected from another cohort study[18]. 73 chronic HBV infected pregnant women with high HBV DNA load undergoing routinely consultation from March 1st, 2012 to May 31st, 2015 were recruited from the First Affiliated Hospital of Xi'an Jiaotong University, Shaanxi, China. Inclusion criteria included pregnant women aged from 18 to 40 years, had serum HBsAg positive for more than 6 months and HBV DNA load greater than 10^6 IU/ml, started taking TBV (600 mg/day) from the 24th week of gestation and stopped TBV 12 weeks postpartum. Exclusion criteria were if patients were serologic HIV or hepatitis C or Hepatitis D virus positive, or if patients had anti-HBV treatment before the 24th week of gestation during the pregnancy, took the immunosuppressive agents during the pregnancy, and were diagnosed as any of the following diseases: gestational diabetes, arrhythmia, anemia or proteinuria. All patients were evaluated every 4 weeks from 24th week of gestation, at delivery, postpartum week (PPW) 4, 12, 24, and 52. The study was approved by the ethics committee of the First Affiliated Hospital of Xi'an Jiaotong University. Informed consent was obtained from each participant.

All infants born to the chronic HBV infected mothers received combined immunoprophylaxis, 200 IU of hepatitis B immunoglobulin and 10 µg of recombinant HBV vaccine within 12 h postpartum, at 1 month, and at 6 months.

2.2 Ultra-deep pyrosequencing data

To evaluate the risk of HBV drug resistance generated by the short duration of TBV in pregnancy, polymerase gene analysis was conducted by using UDPS prior to (at the 24th week of gestation) and

after (at the last time point of follow-up) TBV treatment. The HBV RT was amplified (697 bp) with the primers Seq2 (5'-TTGGCCAAAATTCGCAGTC-3') and OS2 (5'-TCTCTGACATACTTTCCAAT-3') [15]. The PCR products were purified using an Omega gel extraction kit (Omega Bio-tek, USA) and quantified by a Nanodrop 1000 (Thermo Scientific, Wilmington, USA). UDPS was performed on the 454 Life Science platform (GS FLX platform, Roche).

The sensitivity of UDPS on the 454 Life Science platform for detecting low-level viral variants at 0.1–1% has been confirmed by the use of standard cloning methods [19–21], the variants with prevalence larger than 1% were classified as high-confidence variants.

The UDPS generated sequence reads were filtered using the following criteria: 1) mismatched base number of 5' primers greater than 1; 2) no undetermined bases; 3) continuous same bases greater than 8; 4) 150 bases in length or less; 5) chimera sequence. The average number of reads generated for each sample was 9766 (range: 1913 to 21909). The filtered sequence reads were aligned to their respective consensus sequences, the Smith-Waterman algorithm and mutations in corresponding sites were used to calculate Sanger sequences.

2.3 HBV quasispecies complexity of AA

The HBV quasispecies complexity of AA level was estimated for each site using Shannon entropy (S_n) [22, 23], which can be calculated with the formula $S_n = -\sum (p_i \ln p_i) / \ln N$, where N is the total number of clones and p_i is the frequency of each clone in the viral quasispecies population [24]. The mean viral complexity in each sample was calculated by the ratio of total amounts of the S_n at each position and the total length AA number. Mutations of rtL80, rtL82, rtV84, rtS85, rtI91, rtI169, rtV173, rtL180, rtA181, rtT184, rtA194, rtA200, rtS202, rtM204, rtV207, rtS213, rtV214, rtQ215, rtL217, rtE218, rtF221, rtL229, rtI233, rtN236, rtP237, rtN/H238, rtY245, rtM250 and rt S/C256 were analyzed in this study.

2.4 Other measurements

Data of age, parity, antiviral treatment history before pregnancy, HBV family history, patients HBVDNA load, HBV serum markers titer including HBsAg and HBeAg, alanine transaminase (ALT) level and creatinine kinase (CK) at 24th, 28th, 32th, 36th week of gestation, delivery and postpartum week (PPW) 4, 12, 24, and 52, corresponding safety data of infants were collected from the medical records of the hospital.

2.5 Statistical Analysis

Continuous variables were presented as means \pm standard deviations and categorical variables were presented as counts (percentages). Paired t-tests were used to test the changes of the complex of HBV quasispecies before and after TBV treatment. The frequency of the mutations at rtM204 were compared using t-tests between patients with and without plasma HBV DNA $< 10^3$ IU/mL at delivery. All tests were two-side tests and p values < 0.05 were considered statistically significant. All analyses were performed with SPSS software 24.0 (SPSS Inc., Chicago, IL, USA).

3. Results

3.1 Baseline maternal characteristics

Total 73 HBsAg (+) and HBV DNA load > 10⁶IU/mL pregnant women were enrolled in the current study. Subjects accepted TBV from 24th week of gestation to PPW 12, and then were followed up at least to PPW 52. The median follow-up time was 76 weeks (range: 52–152 weeks). The baseline demographics and clinical characteristics of the mothers were summarized in Table 1. Six pregnant women (8.2%) accepted antiviral treatment before pregnancy, 3 had interferon treatment and 3 had LAM treatment.

Table 1
Demographics and baseline characteristics

Variable	Value
Age, years*	27.78 ± 3.89
Parity*	1.14 ± 0.35
Previous use of antiviral, number (%)	6 (8.22)
HBV family history, number (%)	32 (43.84)
ALT levels, U/L*	39.15 ± 43.97
ALT > 40 U/L, number (%)	22 (30.14)
ALT > 80 U/L, number (%)	7 (9.59)
ALT > 200 U/L, number (%)	1 (1.37)
HBV DNA load, Log ₁₀ IU/mL*	7.91 ± 0.70
HBsAg titer, Log ₁₀ IU/mL*	4.38 ± 0.47
HBeAg titer, Log ₁₀ s/co*	2.45 ± 1.26
HBeAg(+), number (%)	63 (86.30)

*The values are expressed as means ± standard deviations for continuous variables and number of patients (percentages) for categorical variables. Abbreviations: ALT, alanine transaminase; HBeAg, hepatitis B e antigen; HBsAg, hepatitis B surface antigen; HBV, hepatitis B virus.

3.2 Dynamics of maternal HBV DNA load

As showed in Fig. 1, compared to baseline, TBV treatment declined HBV DNA level (4.36 ± 2.03, range 1.84 to 8.95 Log₁₀IU/mL) in all mothers. There were 52 out of 73 (71.2%) women had serum HBV DNA load more than 10³IU/mL at delivery. Viral breakthrough was not observed during TBV treatment. After TBV withdrawal, HBV DNA levels rebounded in all mothers, and reached to a mean of 7.21 ± 1.34 Log₁₀IU/mL after 3 months of the withdrawal (PPW 24).

3.3 Viral quasispecies complexity and NAs-resistant mutations before TBV treatment

The complexity of viral quasispecies of the samples was calculated as described in Methods section. The complexity of quasispecies (S_n) before TBV treatment in all patients was 0.40 ± 0.09 . The S_n value of treatment-naïve patients was 0.40 ± 0.10 . The S_n value of the three patients who accepted LAM before pregnancy were 0.31, 0.36 and 0.36, respectively. The S_n value of the three patients who accepted interferon treatment before pregnancy were 0.45, 0.34 and 0.41, respectively (Fig. 2).

The 29 known NAs-resistant mutations [25, 26] were analyzed in the 73 pregnant women. At primary drug resistance mutation sites, rtM204I/V associated with resistance to LAM and TBV, also known as classical YMDD mutation, presented in 41.1% (30/73) patients before TBV treatment, 9.6% patients had mutation frequencies greater than 20%; RtA181T/V was in 5.5% (4/73) patients, which involves in the LAM, TBV and ADV shared resistance pathway; and rtN236T/A mutation, which was reported to decrease the sensitivity to TDF, presented in 53.4% (39/73) pregnant women, 5.5% patients had mutation frequencies greater than 20%. The proportions of patients with rtN236T/A mutation has no difference with those of patients with rtM204I/V mutation (41.1% vs. 53.4%, $P = 0.136$; frequencies greater than 20%: 9.6% vs. 5.5%, $P = 0.347$). At the compensatory mutation sites, 15.1% (11/73) participants had L80I/V mutation that is associated with resistance to LAM. In addition, the patients prior to TBV treatment also had other putative antiviral resistance mutations (Table 2). However, rtI169T, rtA194T, rtV173L, rtL180M, rtL82M, rtS85A, rtV207I, rtL217R and rtS/C256G mutations were not present before TBV treatment. Two patients who accepted LAM before pregnancy had pre-existing rtM204I mutation.

Table 2

Potential NAs mutation at 29 positions of HBV reverse transcriptase analyzed in the 73 pregnant women

Mutations type	Relationship with therapy	The proportion of the patients with mutations, n (%) (n = 73)	The frequency of the mutations* (%)	Patients with mutations frequency > 20%, n (%)
Primary resistance mutations				
rtI169T	ETV	0	0	0
rtA181T/V	LAM, TBV, ADV, TDF	4 (5.5)	0.023 ± 0.020	0
rtT184A/C/F/G/I/L/M/S	ETV	52 (71.2)	0.13 ± 0.14	14 (19.2%)
rtA194T	ADV, TDF	0	0	0
rtS202C/G/I	ETV	1 (1.4)	0.01	0
rtM204I/V	LAM, ETV, TBV	30 (41.1)	0.13 ± 0.11	7 (9.6%)
rtN236T/A	ADV, TDF	39 (53.4)	0.10 ± 0.13	4 (5.5%)
rtM250I/L/V		41 (56.2)	0.11 ± 0.08	5 (6.8%)
Compensatory mutations				
rtL80I/V	LAM	11 (15.1)	0.02 ± 0.01	
rtV173L	LAM	0	0	0
rtL180M	LAM, ETV, TBV	0	0	0
Putative NAs mutations				
rtL82M	LAM	0	0	0
rtV84M	ADV	2 (2.7)	0.01 ± 0.001	0
rtS85A	ADV	0	0	0
rtI91L	LAM	40 (54.8)	0.82 ± 0.30	36 (49.3%)
rtA200V	LAM	1 (1.4)	0.02	0
rtV207I	LAM	0	0	0
rtS213T	ADV	3 (4.1)	0.11 ± 0.16	1 (1.4%)

* The prevalence of mutations were expressed as number of patients (percentages) and the mutation frequencies were expressed as mean ± standard deviation. Abbreviations: ADV, adefovir dipivoxil; ETV, entecavir; HBV, hepatitis B virus; LAM, lamivudine; n, number; NA, nucleoside/nucleotide analogues; TBV, telbivudine; TDF, tenofovir disoproxil fumarate.

Mutations type	Relationship with therapy	The proportion of the patients with mutations, n (%) (n = 73)	The frequency of the mutations* (%)	Patients with mutations frequency > 20%, n (%)
rtV214A	ADV	3 (4.1)	0.01 ± 0.004	0
rtQ215P/S	LAM, ADV	12 (16.4)	0.07 ± 0.08	2 (2.7)
rtL217R	ADV	0	0	0
rtE218D	ADV	1 (1.4)	0.52	1 (1.4)
rtF221Y	ADV	30 (41.1)	0.29 ± 0.25	16 (21.9)
rtL229G/V/W	LAM	24 (32.9)	0.05 ± 0.06	1 (1.4)
rtI233V	ADV	30 (41.1)	0.12 ± 0.11	7 (9.6)
rtP237H	ADV	22 (30.1)	0.02 ± 0.02	0
rtN/H238D/S/T/A	ADV	46 (63.0)	0.15 ± 0.12	7 (9.6)
rtY245H	ADV	1 (1.4)	0.13	0
rtS/C256G	LAM, ETV	0	0	0
* The prevalence of mutations were expressed as number of patients (percentages) and the mutation frequencies were expressed as mean ± standard deviation. Abbreviations: ADV, adefovir dipivoxil; ETV, entecavir; HBV, hepatitis B virus; LAM, lamivudine; n, number; NA, nucleoside/nucleotide analogues; TBV, telbivudine; TDF, tenofovir disoproxil fumarate.				

As showed in Table 3, 34.3% (25/73) patients had rtM204I mutation and 27.4% (20/73) had rtM204V mutation. Multi-base mutations combined with rtM204I/V was analyzed, rtM204I + rtN236T and rtM204V + rtN236T appeared to be the most common ones (16.4% and 17.8%, respectively). RtM204I/V + rtL80I/V and rtM204I + rtA181T/V may affect the sensitivity to LAM, TBV and ADV, they also presented but the proportions of the mutations were low (Table 3).

Table 3
The multi-base mutations combined with rtM204I/V

Types of mutation patterns	The rate of the patients with mutations, n (%) (n = 73)
rtM204I	
rtM204I alone	25 (34.3)
rtM204I + rtL80I/V	5 (6.8)
rtM204I + rtA181T/V	2 (2.7)
rtM204I + rtL180M	0
rtM204I + rtN236T	12 (16.4)
rtM204I + rtI233V	12 (16.4)
rtM204I + rtA194T	0
rtM204V	
rtM204V alone	20 (27.4)
rtM204V + rtL80I/V	5 (6.8)
rtM204V + rtA181T/V	0
rtM204V + rtL180M	0
rtM204V + rtN236T	13 (17.8)
rtM204V + rtI233V	9 (12.3)
rtM204V + rtA194T	0
Abbreviations: n, number.	

3.4 Pre-existing HBV mutations and the TBV treatment effect

After receiving TBV treatment during the pregnancy, 71.2% (52/73) of patients still had HBV DNA load $\geq 10^3$ IU/mL at delivery. The complex of HBV quasispecies of these patients were not found being significantly different from that of the patients with HBV DNA load less than 10^3 IU/mL at delivery (0.40 ± 0.09 vs. 0.40 ± 0.09 , $P = 0.353$). The frequency of rtM204I was not significantly higher in patients with HBV DNA load $\geq 10^3$ IU/mL than that of patients with HBV DNA $< 10^3$ IU/mL at delivery (0.13 ± 0.12 vs. 0.15 ± 0.11 , $P = 0.669$), either. Among them, 75.0% patients with rtM204I positive had HBV DNA load $\geq 10^3$ IU/mL at delivery, which was comparable with the subjects without rtM204I (75.0% vs. 70.8%, $P = 0.710$). In addition, the patients were further divided into high mutation group (the frequency of rtM204I

$\geq 20\%$, 10% and 5%) and low mutation group (the frequency $< 20\%$, 10% and 5%) before the TBV treatment. As shown in Fig. 3, the proportion of maternal HBV DNA load $\geq 10^3$ IU/mL at delivery was 71.4% in the rtM204I $\geq 20\%$ group, which was similar with that in the rtM204I $< 20\%$ group (71.4% vs. 72.3%, $P = 0.961$). Similar trend was observed in groups with 10% (72.7% vs. 72.1%, $P = 0.968$) and 5% frequency of rtM204I (62.5% vs. 75.0%, $P = 0.325$).

3.5 HBV mutations after the short-term TBV treatment

The impact of TBV short-time treatment to HBV mutations was analyzed among the 73 pregnant women. No change was found in the frequencies of rtM204I mutation before and after (0.34 ± 0.23 vs. 0.32 ± 0.23 , $P = 0.681$, Fig. 4B) the TBV treatment. Compared with the HBV quasispecies complexity at baseline, there was no significant increase after the TBV treatment (0.40 ± 0.09 vs. 0.41 ± 0.12 , $P = 0.599$, Fig. 4A) as well.

3.6 Safety of TBV treatment

TBV treatment was generally tolerated well by the mothers and their infants, there were no maternal severe adverse effects observed in this study. Mild creatinine kinase (CK) elevation ($< 2 \times$ ULN) was reported for 1 of 73 mothers (1.4%), and CK level normalized after telbivudine withdrawal (Table. S1). Among the 73 infants, there was no preterm, low birth weight, Apgar scores < 10 infant, and none of them had congenital deformities. No infant was found seropositive for HBsAg, HBeAg, and HBV DNA in the follow-up.

4. Discussion

In our study, we found that the overall viral quasispecies complexity was 0.40 ± 0.09 and 30 of 73 patients (41.1%) had rtM204I/V positive, while 71.2% of pregnant women had serum HBV DNA load more than 10^3 IU/mL at delivery. It was reported that the pre-existing primary resistance mutations could reduce the susceptibility of anti-HBV monotherapy or even combined-therapy, such as rtM204I was refractory to LAM and TBV, the efficacy of the corresponding NAs could be affected[26]. We tested the association between the mutation frequency of rtM204I and the HBV DNA load decrease, no significant association was found in either high mutation frequency group or low mutation frequency group (Fig. 3), which indicated that the pre-existing primary resistance might have no influence on the short-term TBV treatment. Besides, rtN236T/A mutation that was related to decreasing sensitivity of tenofovir disoproxil fumarate (TDF) presented in 53.4% of the pregnant women, which had no difference with the proportion of the patients with rtM204I/V mutation. This indicated that TDF with high genetic barrier to HBV resistance might not be superior to TBV for the pregnant women to prevent MTIT from the view of pre-existing primary resistance mutations.

We used UDPS to detect the drug resistance mutations, which is much more sensitive than the methods of many studies used before (5–20% variants detected in NAs-naïve patients)[10–12]. The UDPS can

detect minor HBV variants and reveal the massive genetic heterogeneity by parallel amplification and detection of abundant small size sequences[27]; moreover, it can provide longer reads than other techniques and is suitable for viral resistance studies[28]. As far as we know, this is the first work to evaluate the pre-existing NA resistance mutations by UDPS in a moderate sample of pregnant women with chronic HBV infection. In the present study, 41.1% of patients were rtM204I/V positive, while only 9.6% (7/73) patients had rtM204I/V frequency 20% or more. In addition, the average frequency of the mutation was 0.13 ± 0.11 , both of which were consistent with the previous study findings [10–12]. Two previous studies conducted rtM204I/V mutation testing with sensitive methods. Kirishima et al. reported 22.2% (4/18) NA-naïve patients had rtM204I/V mutation by peptide nucleic acid mediated polymerase chain reaction clamping which could detect mutation rate as low as 0.01 – 0.001% [13], and Ayres et al. detected 12.5% (3/24) pregnant women had the mutation by UDPS [15], which were lower than the rate in our study, the difference may be associated with the very limited sample sizes in the above two studies.

Drug-resistant HBV variants were reported to emerge in the mothers accepted short-term LAM treatment during late pregnancy [15]. Han et al. reported rtM204 mutation arose in two mothers at 22 weeks and 71 weeks of TBV treatment, respectively [12]. In our study, approximately 7 months TBV treatment was administrated in the pregnant women. No increases of the viral quasispecies complexity and the frequency of rtM204I mutation were observed, which supplemented the safety of TBV treatment in late pregnancy. Furthermore, some pregnant women had multi-base mutations combined with rtM204I/V at baseline, including rtM204I + rtA181T/V, rtM204I/V + rtL80I/V, rtM204I/V + rtN236T, and rtM204I/V + rtI233V, which may affect their sensitivity to LAM, TBV, ADV and TDF. The complexity of viral quasispecies and the frequency of rtM204I mutation had no significant increase after TBV treatment in those pregnant women; however, caution has to be taken for them to choose NAs in subsequent long-term therapy due to the drug resistance mutations.

In this study, the prevalence of HBV pre-existing resistant mutants in pregnant women, the influence of the efficacy of short-term TBV treatment and the drug-resistant mutations after TBV therapy were assessed retrospectively. Although the number of subjects was moderate, a prospective cohort study with larger sample size is necessary to evaluate the relationship between the HBV mutations and the antiviral treatment effect.

In conclusion, the prevalence of pre-existing HBV mutation among the pregnant women was as high as 41.1%. However, the pre-existing HBV mutation had limited influence on the efficacy of short-term TBV treatment, and TBV treatment during late pregnancy seemed not to increase the risk of emerging HBV resistant mutants.

Declarations

Conflict of Interest Statements: All the authors declare that they have no competing interests, and all authors confirm its accuracy.

Ethics approval: All procedures performed in this study involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. The study protocol was approved by the ethics committee of the First Affiliated Hospital of Xi'an Jiaotong University 2010-Lunshenkezi-No. 13).

Consent of publication: not applicable.

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Availability of data: The data in the current study are available from the corresponding author on reasonable request.

Authors' contributions: TC, YH, YZ and JZ conceived of the presented idea. JW, JL, LJ, NY and QY acquired data in the study. YY, TY, CH and JW analyzed data. JW drafted the paper, and all authors discussed the results and contributed to the final manuscript.

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References

1. Terrault NA, Lok ASF, McMahon BJ, Chang KM, Hwang JP, Jonas MM, Brown RS, Jr., Bzowej NH, Wong JB: **Update on Prevention, Diagnosis, and Treatment of Chronic Hepatitis B: AASLD 2018 Hepatitis B Guidance.** *Clinical liver disease* 2018, **12**(1):33-34.
2. **EASL 2017 Clinical Practice Guidelines on the management of hepatitis B virus infection.** *J Hepatol* 2017, **67**(2):370-398.
3. Sarin SK, Kumar M, Lau GK, Abbas Z, Chan HL, Chen CJ, Chen DS, Chen HL, Chen PJ, Chien RN *et al.* **Asian-Pacific clinical practice guidelines on the management of hepatitis B: a 2015 update.** *Hepatol Int* 2016, **10**(1):1-98.
4. Wen WH, Chang MH, Zhao LL, Ni YH, Hsu HY, Wu JF, Chen PJ, Chen DS, Chen HL: **Mother-to-infant transmission of hepatitis B virus infection: significance of maternal viral load and strategies for intervention.** *J Hepatol* 2013, **59**(1):24-30.
5. Deng L, Tang H: **Hepatitis B virus drug resistance to current nucleos(t)ide analogs: Mechanisms and mutation sites.** *Hepatol Res* 2011, **41**(11):1017-1024.
6. Seeger C, Mason WS: **Hepatitis B virus biology.** *Microbiology and molecular biology reviews : MMBR* 2000, **64**(1):51-68.
7. Pastor R, Habersetzer F, Fafi-Kremer S, Doffoel M, Baumert TF, Gut JP, Stoll-Keller F, Schvoerer E: **Hepatitis B virus mutations potentially conferring adefovir/tenofovir resistance in treatment-naive**

- patients. *World J Gastroenterol* 2009, **15**(6):753-755.
8. Pollicino T, Isgro G, Di Stefano R, Ferraro D, Maimone S, Brancatelli S, Squadrito G, Di Marco V, Craxi A, Raimondo G: **Variability of reverse transcriptase and overlapping S gene in hepatitis B virus isolates from untreated and lamivudine-resistant chronic hepatitis B patients.** *Antivir Ther* 2009, **14**(5):649-654.
 9. Margeridon-Thermet S, Shulman NS, Ahmed A, Shahriar R, Liu T, Wang C, Holmes SP, Babrzadeh F, Gharizadeh B, Hanczaruk B *et al*: **Ultra-deep pyrosequencing of hepatitis B virus quasispecies from nucleoside and nucleotide reverse-transcriptase inhibitor (NRTI)-treated patients and NRTI-naive patients.** *J Infect Dis* 2009, **199**(9):1275-1285.
 10. Masaadeh HA, Hayajneh WA, Alqudah EA: **Hepatitis B virus genotypes and lamivudine resistance mutations in Jordan.** *World J Gastroenterol* 2008, **14**(47):7231-7234.
 11. Shi M, Yang ZJ, Wang RS, Zhang H, Zhu YF, Xu YP, Lin QY, Jin LJ: **Rapid quantitation of lamivudine-resistant mutants in lamivudine treated and untreated patients with chronic hepatitis B virus infection.** *Clin Chim Acta* 2006, **373**(1-2):172-175.
 12. Akarsu M, Sengonul A, Tankurt E, Sayiner AA, Topalak O, Akpinar H, Abacioglu YH: **YMDD motif variants in inactive hepatitis B carriers detected by Inno-Lipa HBV DR assay.** *J Gastroenterol Hepatol* 2006, **21**(12):1783-1788.
 13. Kirishima T, Okanoue T, Daimon Y, Itoh Y, Nakamura H, Morita A, Toyama T, Minami M: **Detection of YMDD mutant using a novel sensitive method in chronic liver disease type B patients before and during lamivudine treatment.** *J Hepatol* 2002, **37**(2):259-265.
 14. Ghany M, Liang TJ: **Drug targets and molecular mechanisms of drug resistance in chronic hepatitis B.** *Gastroenterology* 2007, **132**(4):1574-1585.
 15. Ayres A, Yuen L, Jackson KM, Manoharan S, Glass A, Maley M, Yoo W, Hong SP, Kim SO, Luciani F *et al*: **Short duration of lamivudine for the prevention of hepatitis B virus transmission in pregnancy: lack of potency and selection of resistance mutations.** *J Viral Hepat* 2014, **21**(11):809-817.
 16. Liu Y, Wang M, Yao S, Yuan J, Lu J, Li H, Zeng W, Deng Y, Zou R, Li J *et al*: **Efficacy and safety of telbivudine in different trimesters of pregnancy with high viremia for interrupting perinatal transmission of hepatitis B virus.** *Hepatol Res* 2016, **46**(3):E181-188.
 17. Han GR, Cao MK, Zhao W, Jiang HX, Wang CM, Bai SF, Yue X, Wang GJ, Tang X, Fang ZX: **A prospective and open-label study for the efficacy and safety of telbivudine in pregnancy for the prevention of perinatal transmission of hepatitis B virus infection.** *J Hepatol* 2011, **55**(6):1215-1221.
 18. Liu J, Wang J, Jin D, Qi C, Yan T, Cao F, Jin L, Tian Z, Guo D, Yuan N *et al*: **Hepatic flare after telbivudine withdrawal and efficacy of postpartum antiviral therapy for pregnancies with chronic hepatitis B virus.** *J Gastroenterol Hepatol* 2017, **32**(1):177-183.
 19. Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen YJ, Chen Z *et al*: **Genome sequencing in microfabricated high-density picolitre reactors.** *Nature* 2005, **437**(7057):376-380.

20. Solmone M, Vincenti D, Prosperi MC, Bruselles A, Ippolito G, Capobianchi MR: **Use of massively parallel ultradeep pyrosequencing to characterize the genetic diversity of hepatitis B virus in drug-resistant and drug-naive patients and to detect minor variants in reverse transcriptase and hepatitis B S antigen.** *J Virol* 2009, **83**(4):1718-1726.
21. Gong L, Han Y, Chen L, Liu F, Hao P, Sheng J, Li XH, Yu DM, Gong QM, Tian F *et al*: **Comparison of next-generation sequencing and clone-based sequencing in analysis of hepatitis B virus reverse transcriptase quasispecies heterogeneity.** *J Clin Microbiol* 2013, **51**(12):4087-4094.
22. Nishijima N, Marusawa H, Ueda Y, Takahashi K, Nasu A, Osaki Y, Kou T, Yazumi S, Fujiwara T, Tsuchiya S *et al*: **Dynamics of hepatitis B virus quasispecies in association with nucleos(t)ide analogue treatment determined by ultra-deep sequencing.** *PLoS One* 2012, **7**(4):e35052.
23. Yin F, Wu Z, Fang W, Wu C, Rayner S, Han M, Deng F, Du R, Liu J, Wang M *et al*: **Resistant mutations and quasispecies complexity of hepatitis B virus during telbivudine treatment.** *J Gen Virol* 2015, **96**(11):3302-3312.
24. Domingo E, Martin V, Perales C, Grande-Perez A, Garcia-Arriaza J, Arias A: **Viruses as quasispecies: biological implications.** *Curr Top Microbiol Immunol* 2006, **299**:51-82.
25. He X, Wang F, Huang B, Chen P, Zhong L: **Detection and analysis of resistance mutations of hepatitis B virus.** *International journal of clinical and experimental medicine* 2015, **8**(6):9630-9639.
26. Liu BM, Li T, Xu J, Li XG, Dong JP, Yan P, Yang JX, Yan L, Gao ZY, Li WP *et al*: **Characterization of potential antiviral resistance mutations in hepatitis B virus reverse transcriptase sequences in treatment-naive Chinese patients.** *Antiviral Res* 2010, **85**(3):512-519.
27. Sede M, Lopez-Ledesma M, Frider B, Pozzati M, Campos RH, Flichman D, Quarleri J: **Hepatitis B virus depicts a high degree of conservation during the immune-tolerant phase in familiarly transmitted chronic hepatitis B infection: deep-sequencing and phylogenetic analysis.** *J Viral Hepat* 2014, **21**(9):650-661.
28. Rodriguez C, Chevaliez S, Bensadoun P, Pawlotsky JM: **Characterization of the dynamics of hepatitis B virus resistance to adefovir by ultra-deep pyrosequencing.** *Hepatology* 2013, **58**(3):890-901.

Table S1

Table S1. Adverse events reported in current study*

Adverse event	Number (%)
Gastrointestinal symptom	5 (6.85%)
Nausea	1 (1.37)
Emesis	1 (1.37)
Diarrhea	1 (1.37)
flatulence	1 (1.37)
Dyspepsia	1 (1.37)
Fatigue	2 (2.74)
Dizziness	1 (1.37)
Insomnia	2 (2.74)
Myalgia	1 (1.37)
CK elevation	1 (1.37)
Cesarean delivery	35 (47.95)
Threatened abortion	11 (15.07)
Prolonged Labor	4 (6.25)

*Number (percentages) of patients with adverse events were reported. Abbreviations: CK, creatinine kinase.

Figures

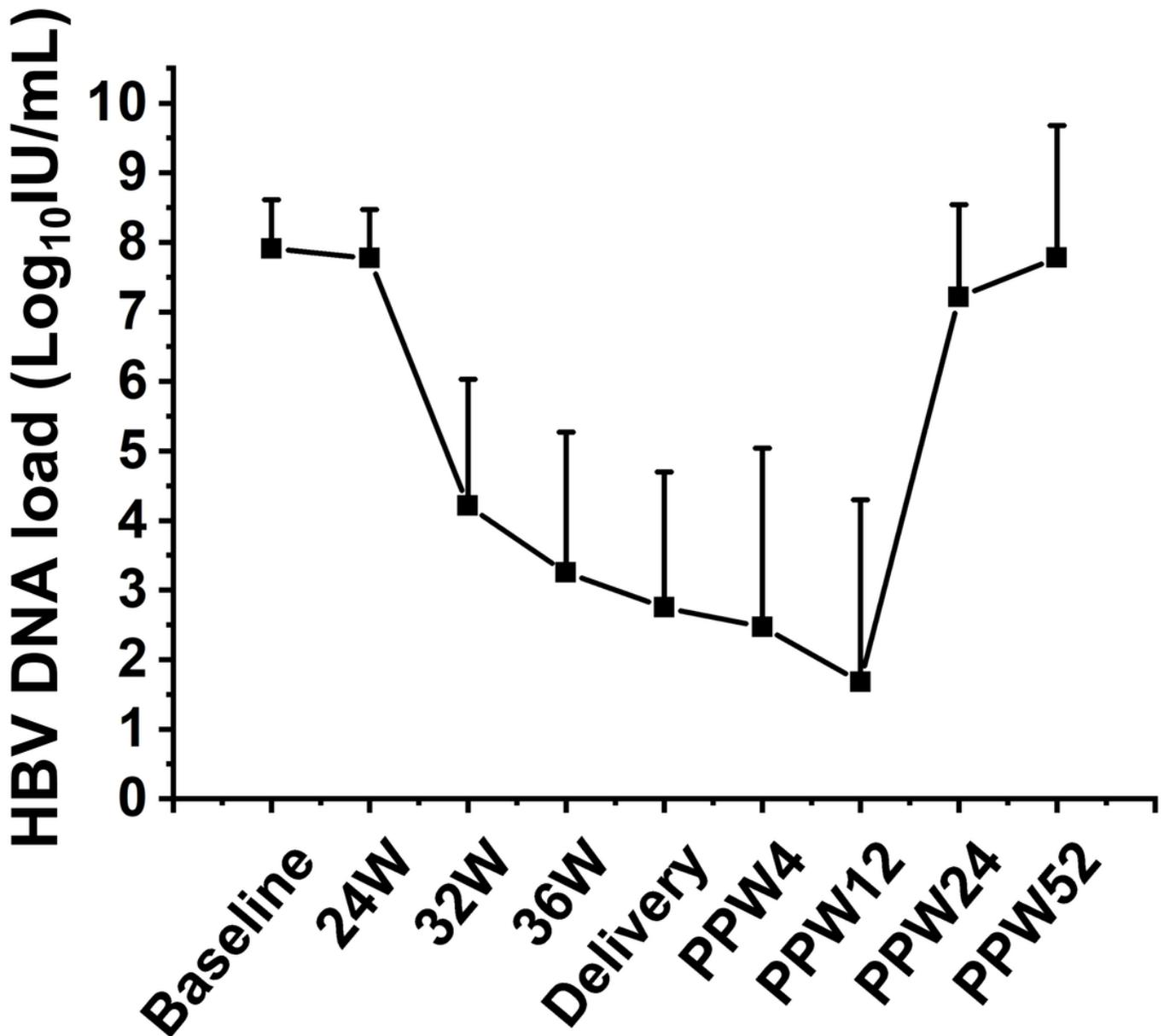


Figure 1

HBV DNA load kinetics in pregnancy and postpartum. Abbreviations: HBV, hepatitis B virus; W, week; PPW, postpartum week.

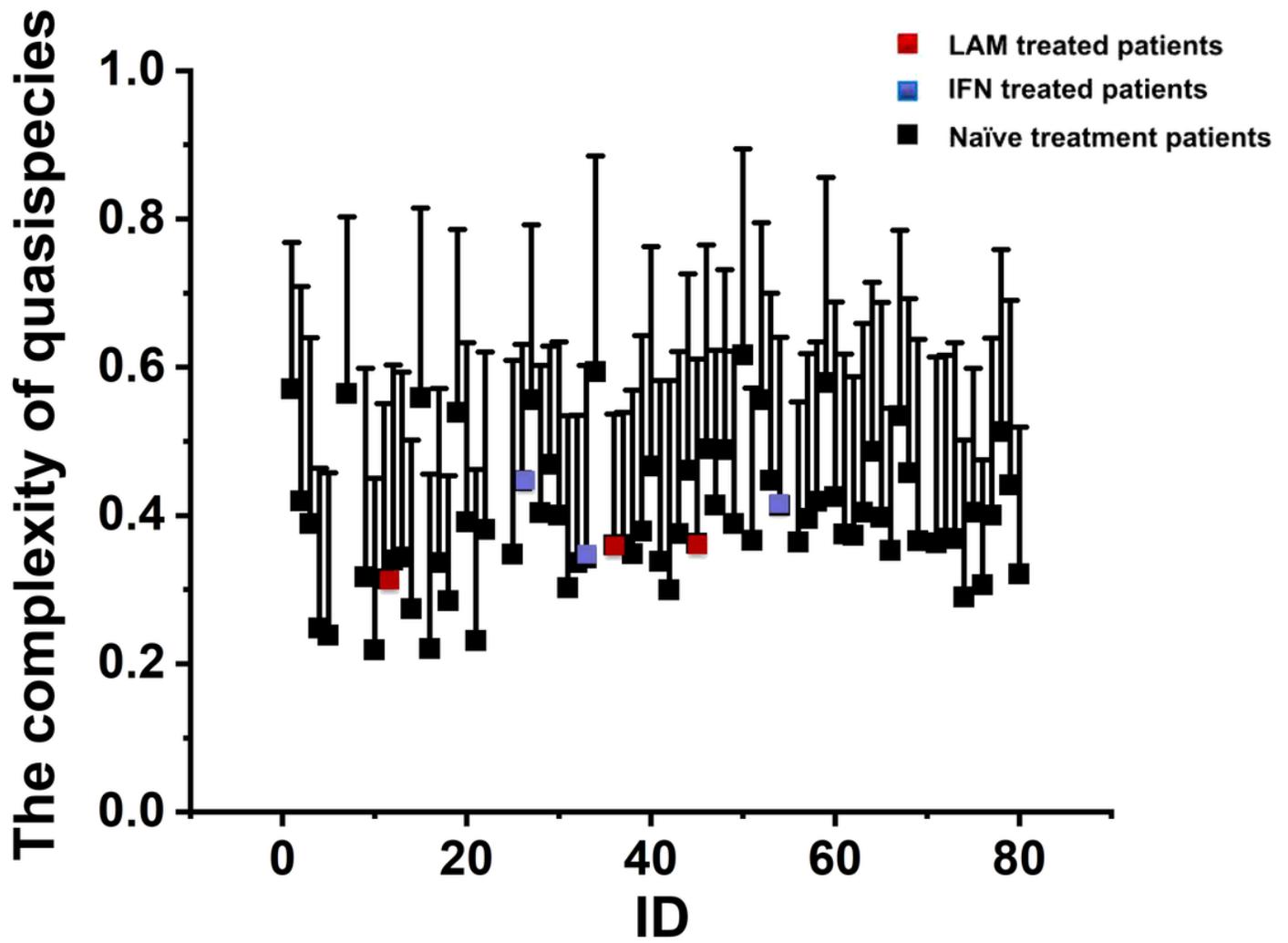


Figure 2

Scatter diagram of HBV quasispecies complexity. Abbreviations: IFN, interferon; LAM, lamivudine.

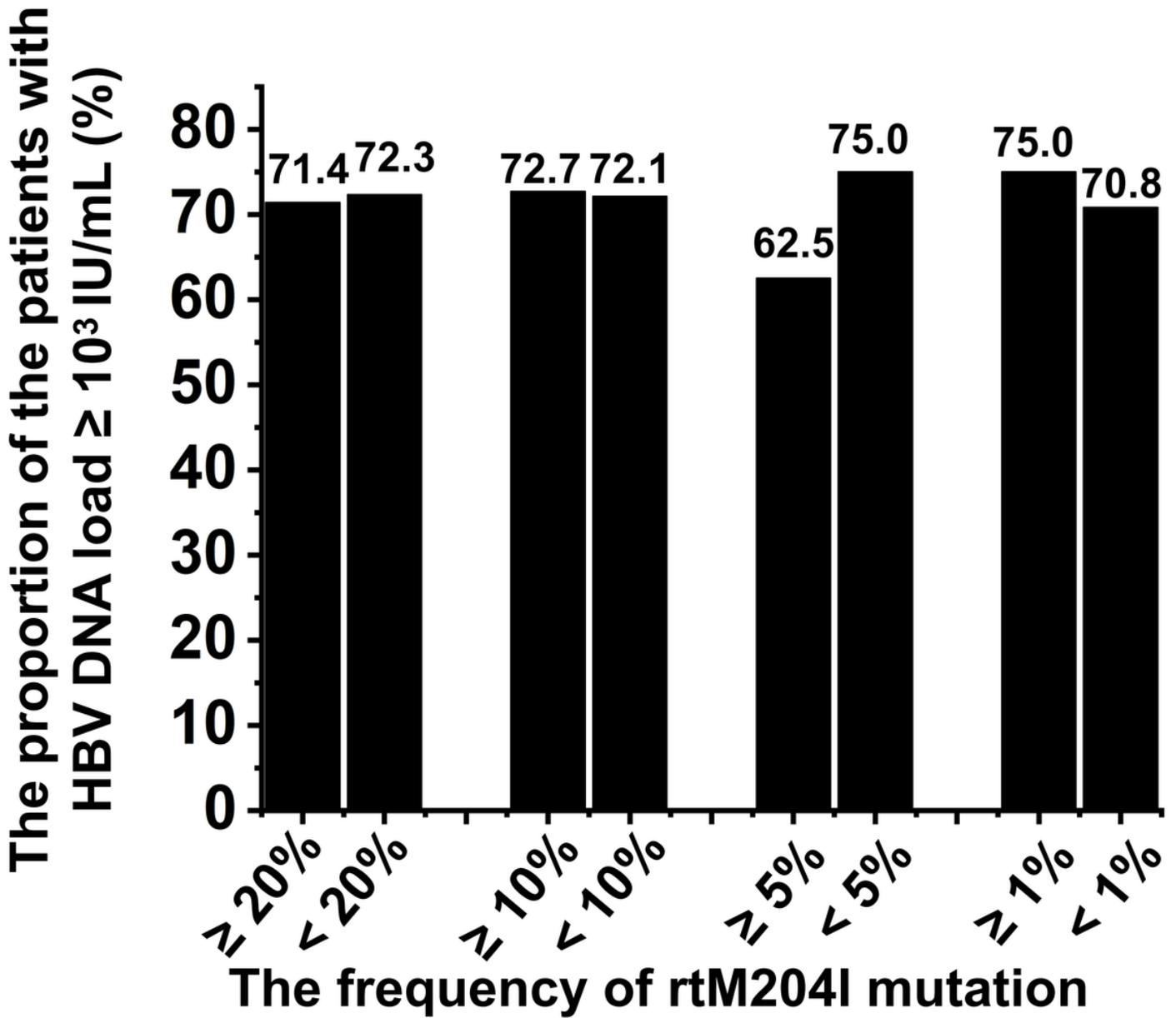


Figure 3

The relation between the frequency of rtM204I and maternal HBV DNA load at delivery. Abbreviations: HBV, hepatitis B virus.

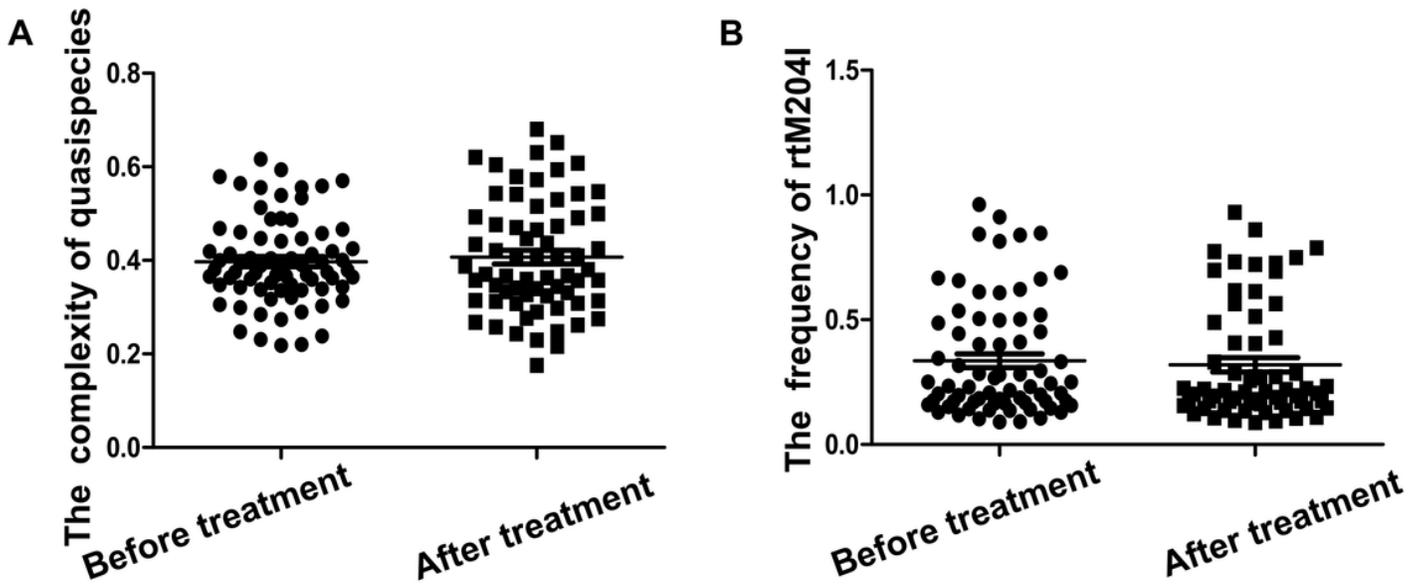


Figure 4

The impact of telbivudine short-time treatment to HBV mutations: (A) The change of HBV quasispecies complexity before and after telbivudine treatment; (B) The change of rtM204I mutation frequency before and after telbivudine treatment.