

# Degradation of Oxytetracycline by Electrochemical, Fenton and Electro-fenton Processes Using SS316 and SS316/ $\beta$ -PbO<sub>2</sub> Anodes: Process Optimization Using Rsm-ccd, Bioassay Test and Degradation Pathway

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## Research Article

**Keywords:** AOPs, Electrochemical, Electro-Fenton, Oxytetracycline, Stainless steel/ $\beta$ -PbO<sub>2</sub>, Toxicity

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1     **Degradation of oxytetracycline by electrochemical, Fenton and electro-Fenton processes**  
2     **using SS316 and SS316/ $\beta$ -PbO<sub>2</sub> anodes: Process optimization using RSM-CCD, bioassay**  
3     **test and degradation Pathway**

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12  
13             **Highlights:**

- 14             ✓ The bioassay was conducted to assess the biological toxicity the inlet and outlet solution.  
15             ✓ SS316/ $\beta$ -PbO<sub>2</sub> anode was used to improve the biodegradability of diazinon.  
16             ✓ Oxytetracycline antibiotics degradation by electrochemical oxidation, Fenton and electro-  
17             Fenton processes.  
18             ✓ Process optimization by RSM-CCD  
19             ✓ degradation mechanism of Oxytetracycline antibiotics.  
20             ✓ The electrofenton process was more efficient than the oxytetracycline antibiotic compared  
21             to the electrochemical processes and Fenton.  
22             ✓ The removal efficiency of oxytetracycline using SS316/  $\beta$ -PbO<sub>2</sub> anode electrode was  
23             100%.  
24             ✓ The bioassay method with microorganisms effectively showed the reduction of the toxicity  
25             of the effluent treated by the electrofenton process.

## 28 **Abstract**

29 The aim of the present study was to evaluate the efficiency of advanced oxidation processes  
30 (electrochemical, Fenton and electro-Fenton) in the removal of oxytetracycline using SS316 and  
31 SS316/ $\beta$ -PbO<sub>2</sub> anodes. This study was performed experimentally on a laboratory scale in a 250  
32 mL reactor. First, experiments were designed for the electrochemical process using a central  
33 composite design, and the optimal conditions for the variables pH(3.53), electric current  
34 density(3.85mA/cm<sup>2</sup>), initial concentration of oxytetracycline (20mg/L) and electrolysis time  
35 (42.35min) was obtained; then, under these conditions, the efficiency of Fenton process with  
36 FeSO<sub>4</sub> variable without the presence of electrodes was evaluated, and its optimal value was 0.3  
37 g/L, and then in the presence of optimal values of the above 5 variables, the efficiency of electro-  
38 Fenton process with H<sub>2</sub>O<sub>2</sub> changes were investigated and the optimal value of 0.12 was obtained  
39 for H<sub>2</sub>O<sub>2</sub>. The removal efficiencies of oxytetracycline in electrochemical, Fenton, and electro-  
40 Fenton processes were 84.7%, 73.4%, and 98.2%, respectively. Under optimal conditions, the  
41 SS316/ $\beta$ -PbO<sub>2</sub> anode electrode enhanced the oxytetracycline efficiency by electron-Fenton  
42 process to 100%. The results of bioassay with microorganisms showed that the reduction of  
43 toxicity of the effluent treated by electro-Fenton process for *Pseudomonas aeruginosa* and  
44 *Staphylococcus aureus* was 84.5% and 69%, respectively.

45 **Keywords:** AOPs, Electrochemical, Electro-Fenton, Oxytetracycline, Stainless steel/ $\beta$ -PbO<sub>2</sub>,  
46 Toxicity.

## 47 **1. Introduction**

48 Due to the increasing use of medicinal compounds, the study of compounds derived from treated  
49 wastewater is very important <sup>[1, 2]</sup>. Today, there are many concerns about medicinal compounds  
50 because the presence of these compounds in the ecosystem can make bacteria resistant to these  
51 compounds <sup>[3, 4]</sup>, which occurs by causing genetic mutations in bacteria; thus, paying attention to

52 the presence of this contaminant in the environment and especially in aquatic environments are  
53 important <sup>[5, 6]</sup>. On the other hand, pharmaceutical compounds are abundant in the wastewater of  
54 hospitals, livestock and residential centers and have even been observed in surface water samples  
55 and these compounds have very strong chemical and physical properties that make these  
56 substances dangerous for non-target groups. However, it is noteworthy that some pharmaceutical  
57 compounds can be transferred to the natural environment through the waste of the pharmaceutical  
58 industry <sup>[7-9]</sup>. On the other hand, we know that the growth of the pharmaceutical industry in some  
59 countries has increased in recent years and the non-compliance of this industry with environmental  
60 standards has caused the effluents from this industry to be discharged into the domestic sewage  
61 network, which is an important issue because these compounds are only partially removed by  
62 treatment processes and their lack of optimal removal causes contamination of surface water  
63 resources <sup>[1, 10]</sup>. The global estimate for the use of antibiotics is between 100,000 and 200,000 tons  
64 per year, which is a significant number <sup>[11]</sup>. Among the types of antibiotics, oxytetracycline (from  
65 the tetracycline category) is of great importance for its widespread use in medicine and veterinary  
66 medicine <sup>[12]</sup>. Because it has the ability to eliminate various infections and is active against a wide  
67 range of bacteria and is mainly excreted in the urine, so the removal of this contaminant from  
68 sewage seems very necessary <sup>[13, 14]</sup>. In order to treat water and sanitation, various processes have  
69 been used in this field. Advanced oxidation processes have received much attention in recent years  
70 <sup>[15]</sup>. Of course, processes such as removal by UV/H<sub>2</sub>O<sub>2</sub> [16], removal by ozone <sup>[17]</sup>, activated carbon  
71 <sup>[18]</sup> and bio-nanocomposite method <sup>[19]</sup> can be mentioned, but each has advantages and  
72 disadvantages; for example, although activated carbon is very economically suitable and has good  
73 potential for treatment, however the limitations of the adsorption process related to the separation  
74 and regeneration of the adsorbent after treatment are considered <sup>[20]</sup>. Of course, in the meantime,

75 attention has been paid to electrochemical processes because compared to conventional processes,  
76 they have different applications in water and wastewater treatment processes in small systems,  
77 remotely or from the entry point [21, 22]. Moreover, the electrochemical method, electrical energy is  
78 used as the driving force [23]. It is noteworthy that in recent years, advanced oxidation processes in  
79 combination with other processes have received much attention. For example, electro-Fenton [24]  
80 is one of the most effective and environmentally friendly processes of EAOPs [25], which is actually  
81 a new and developed process of Fenton [26] in combination with electrochemical, and in this  
82 process, the contaminant is eliminated with the reagents of Fenton process and anodic oxidation  
83 processes [27, 28].

84 Nowadays, the use of statistical methods to optimize different processes due to their economics  
85 and simplicity is very important that the response surface methodology (RSM) is one of these  
86 widely used statistical models [29]. In this study, the central composite scheme (CCD) has been  
87 used to optimize the operational variables [30]. The present study considers the efficiency of  
88 advanced oxidation processes (electrochemical, Fenton and electro-Fenton) for the removal of  
89 oxytetracycline antibiotics from aqueous solutions using stainless steel and stainless steel/ $\beta$ -PbO<sub>2</sub>  
90 electrodes due to the widespread use of the antibiotic oxytetracycline and its entry into aqueous  
91 solutions. The experiments were designed using the response-surface methodology.

92

## 93 **2. Materials and methods**

### 94 **2. 1. Required materials and sample preparation:**

95 The oxytetracycline antibiotic powder was prepared from Sigma, and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>),  
96 ferrous sulfate (FeSO<sub>4</sub>), lead nitrate, and nitric acid from Merck CO. Also, to adjust the pH of the  
97 solution containing the oxytetracycline antibiotics, H<sub>2</sub>SO<sub>4</sub>, and NaOH, with a purity of 98%, were

98 prepared from the Merck Company, Germany. To test for toxicity, standard strains of  
99 *Pseudomonas aeruginosa* and *Staphylococcus aureus* were obtained from the microbiology  
100 laboratory of Ardabil University of Medical Sciences. Stoke solution (1000 mg/L) of  
101 oxytetracycline antibiotic was prepared by dissolving 1 g of oxytetracycline antibiotic precursor  
102 in 1 liter of double distilled water. For determination of surface morphology of the PbO<sub>2</sub> deposited  
103 onto the stainless steel 316, scanning electron microscopy (SEM) coupled with energy dispersive  
104 X-ray spectroscopy (EDX) (model HITACHI S-4160, Japan) was employed. Analysis of the phase  
105 structure of the PbO<sub>2</sub> layer was done using X-ray diffraction (XRD pattern) using an X'Pert Pro  
106 diffractometer (Rigaku RINT2200, Japan) with a 2θ step width of 0.1° and a scan rate of 1520  
107 s/0.1° at 40 kV and the electron probe current of 40 mA.

## 108 **2-2) Design of experiments for electrochemical process:**

109 In this study, the response surface methodology (RSM) with a central composite design was used  
110 to determine the optimal conditions and the effect of independent variables on response  
111 performance (removal of oxytetracycline antibiotic). The experiments were designed with four  
112 factors, i.e., pH (A), electric current intensity (B), initial antibiotic concentration (C), and  
113 electrolysis time (D). The number of experiments in this study was obtained using Equation No.1.  
114 Therefore, in this study, a total of 30 experiments were performed using 6 replications at the central  
115 point. These central points are for evaluating test error and measuring lack of fit. The efficiency of  
116 the quadratic polynomial model was expressed by correlation coefficient (R<sup>2</sup>) and adjusted  
117 correlation coefficient (adjusted R<sup>2</sup>). The significance of each sentence was expressed by  
118 regression equation, and significant expressions were determined by analysis of variance  
119 (ANOVA) for each answer.

$$120 \quad N = 2K ( K - 1 ) + C \quad (1)$$

121 In this equation, N is the number of experiments, K is the number of independent variables, and C  
122 is the number of central points.

### 123 **2.3. Experimental procedure:**

124 This experimental study was performed on a laboratory scale. In this research, in order to perform  
125 electrochemical, Fenton and Electro-Fenton processes, a cylindrical reactor with a volume of 250  
126 ml was used; it was equipped with two stainless steel electrodes (one as a cathode and the other as  
127 an anode) with dimensions (60×20×1 mm) and a distance of 20 mm from each other and connected  
128 to the DC power supply. The actual schematic of the reactor used in this study were shown in  
129 Figure 1. The volume of each sample was 250 ml. For experiments, a certain amount of NaCl salt  
130 (0.2 g/L) was added to the reactor for improvement of ionic properties. In the electrochemical  
131 process, independent variables including pH (3-11), electric current intensity (1-4 mA/cm<sup>2</sup>), initial  
132 concentration of oxytetracycline (20-100 mg/L), and electrolysis time (10-50). Minutes). After  
133 completion of the determined reaction, the sample was centrifuged at 30,000 rpm for 4 minutes,  
134 and after passing through a 0.45 micron filter, it was prepared to measure the residual concentration  
135 of antibiotics. Then, the residual concentration of oxytetracycline was determined by  
136 spectrophotometric device (model DR-5000, HACH Company, Germany) at 348 nm, and finally,  
137 the removal efficiency of oxytetracycline was obtained by equation No.2 [31]. Changes in ferrous  
138 sulfate (0.1, 0.2, 0.3, and 0.4 gr/L) without the presence of electrodes in the Fenton process were  
139 investigated using the optimal conditions determined in the electrochemical process (previous  
140 step) and the optimal amount of ferrous sulfate was obtained to remove oxytetracycline. Finally,  
141 in the presence of the optimal amount of ferrous sulfate (iron ion source) and the optimal amount  
142 of independent variables, changes in hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) in the electro-Fenton process were

143 obtained in a similar reactor and in the presence of stainless steel electrodes to achieve optimal  
144 removal efficiency.

145 
$$\text{removal efficiency (\%)} = \frac{C_0 - C_t}{C_0} \times 100 \quad (2)$$

146 In the above equation,  $C_0$  and  $C_t$  are the initial and final concentrations of antibiotics in solution  
147 (mg/L), respectively.

#### 148 **2. 4. Preparation of stainless steel electrode/lead dioxide (SS316 / $\beta$ -PbO<sub>2</sub>)**

149 Figure 2 shows the stainless steel (a) and stainless steel/lead dioxide (b) electrodes. To prepare  
150 stainless steel/lead dioxide electrodes, stainless steel electrodes with suitable dimensions were  
151 prepared. It was then mechanically polished with fine-grade sandpaper. Also, for degreasing the  
152 surface of stainless steel electrodes, it was placed in a mixture of diluted nitric acid and caustic  
153 soda for 2 hours in the presence of sound waves at a frequency of 40 kHz. It was thoroughly  
154 washed with distilled water. For the of lead oxide on the stainless steel electrode, a 500 ml  
155 electrolyte solution containing lead nitrate and nitric acid was used. To prepare the stainless  
156 steel/lead dioxide electrode, an electrolysis cell was specially designed in which the stainless steel  
157 electrodes were placed and the lead dioxide nanoparticles were deposited simultaneously and  
158 uniformly on each electrode. For preparing the electrodeposition solution, a 24 g of lead nitrate  
159 and 0.3 M nitric acid were used for on liter of distilled water. Then, for electrochemical coating of  
160 lead dioxide per square centimeter, 25 mA current was applied for 6-9 hours <sup>[32]</sup>. After the  
161 electrochemical coating process, the prepared electrodes were thoroughly washed with distilled  
162 water and their performance was evaluated under optimal conditions.

#### 163 **5-2) Bioassay using microorganisms**

164 In order to test the toxicity of the bioassay method, two microorganisms, Gram-negative  
165 *Pseudomonas aeruginosa* and Gram-positive *Staphylococcus aureus*, were used. MacConkey agar,  
166 nutrient agar and lactose broth were sterilized after preparation according to standard method  
167 instructions in an autoclave (temperature of 121°C, pressure of 15 ppm at 15 minutes). Next,  
168 *Pseudomonas aeruginosa* was cultured on McConkey agar medium and *Staphylococcus aureus* on  
169 nutrient agar medium and incubated (37 ° C for 24 hours). After growth of colonies, toxicity test  
170 was performed using broth lactose culture medium under optimal conditions of electro-Fenton  
171 process for oxytetracycline. For each bacterium (gram-negative and gram-positive) 15 test tubes  
172 (5 tubes for each of the inlet, outlet and control solutions) were considered. Each of the inlet and  
173 outlet solutions of the process consisted of 10 ml of broth lactose culture medium, 1 ml of the  
174 desired solution, and 1 loop of bacteria. The control sample contained lactose broth culture  
175 medium and 1 loop of bacteria. Microbial culture was performed in presence of the flame and one  
176 colony was taken with a loop for each tube, and after inoculation and dissolution in the wall of the  
177 test tube, they were placed in an incubator. Then, the spectrophotometer was set to zero with broth  
178 lactose culture medium at a wavelength of 600 nm and after every 2 hours, the absorption rate of  
179 three samples (inlet solution, outlet solution, and control) was read by spectrophotometer for 5  
180 consecutive periods of 2 hours. The experiments were repeated in two steps to ensure the accuracy  
181 of the work. Finally, the growth inhibition percentage of the inlet and outlet solution for each  
182 bacterium in 5 different time intervals was obtained by Equation (3) and the 10-hour mean growth  
183 inhibition rate was calculated.

$$184 \quad GI (\%) = [ ( 1- OD_{600S}/OD_{600B} ) \times 100] \quad (3)$$

185 In the above equation, GI represents the growth inhibition percentage, and OD600S and OD600B  
186 are the optical density of the sample and the control at a wavelength of 600 nm, respectively.

187 2. 6. Investigation of by-products under optimal conditions:

188 LC-MS (Liquid chromatography-mass spectrometry) with Waters Alliance 2695 HPLC-  
189 Micromass Quattro micro API Mass Spectrometer was used to determine the intermediates  
190 obtained from the degradation of the oxytetracycline antibiotic using the electro-Fenton process  
191 under optimal conditions.

### 192 **3. Results and Discussion**

#### 193 **3.1. Determination of structural characteristics of SS316/ $\beta$ -PbO<sub>2</sub> anode**

194 As mentioned above, evaluation of the surface morphology of PbO<sub>2</sub> electrodeposited on stainless  
195 steel 316 (SS316) substrate was carried out by SEM technique (Fig. 3 (a, b)). An agglomeration  
196 with limited individual particle boundary was observed for all particles. According to provided  
197 images, the crystalline structures of  $\beta$ -PbO<sub>2</sub> in the electrodeposited PbO<sub>2</sub> microparticles on SS316  
198 was identified and the tetragonal structure for the  $\beta$ -PbO<sub>2</sub> was detected. Moreover, according to  
199 EDX results, the weight percentage of main elements existed in  $\beta$ -PbO<sub>2</sub>, i.e., oxygen (O) and lead  
200 (Pb) were 20.1% and 79.9%, respectively.

201 For estimation of the phases and crystallinity of PbO<sub>2</sub> and the deposited film, X-ray diffraction  
202 patterns (XRD) measurements were applied. In Fig. 3c, the XRD of the PbO<sub>2</sub> layer deposited on  
203 the SS316 interlayers was brought, in which the diffraction peaks of  $\beta$  form of PbO<sub>2</sub> is observed.  
204 the tetragonal structure of  $\beta$ -PbO<sub>2</sub> is approved by all XRD results. we could observe the main  
205 peaks at  $2\theta$  of 25.4°, 32.0°, 36.2°, 49.1°, and 62.5° for these SS316/  $\beta$ -PbO<sub>2</sub> electrode; these are  
206 related to the (110), (101), (200), (211), (220) plane of  $\beta$ -PbO<sub>2</sub>, respectively.

#### 207 **3. 2. Statistical analysis of central composite design**

208 CCD is a statistical technique used to collect mathematical and statistical data and to analyze model  
209 in which response performance (antibiotic elimination efficiency) is influenced by independent

210 factors. Table 1 shows the results of the design matrix of 30 experiments (runs) with the effect of  
211 independent variables PH (A), electric current intensity (B), initial concentration of antibiotic (C),  
212 and electrolysis time (D) for removal of the oxytetracycline antibiotic. Table 2 shows the analysis  
213 of variance (ANOVA) for the removal of oxytetracycline by electrochemical process and the  
214 significance of the quadratic model. According to this table, the value of P-Value ( $P < 0.0001$ )  
215 indicates the significance of the model and the interaction of variables on response performance  
216 (removal of oxytetracycline). In addition, the value of F-Value equal to 81.58 indicates the  
217 significance of the variance of each variable against the error variance. The correlation coefficient  
218 of R-squared was 0.9870. In fact, this model explains 98.70% of the changes in response, and for  
219 a suitable model, the  $R^2$  must be at least 0.80, and the significance and usefulness of the model is  
220 greater for 1 [33, 34]. Lack of Fit (0.28) describes the difference between the predicted values with  
221 the observed values, and the repetition of the errors of the real values, and the model is significant  
222 when it does not fit well. The coefficient of variation (CV) is a criterion for describing the range  
223 of changes in the data and the maximum acceptable value for the satisfaction of the results of the  
224 proposed model is 10%, which in this study was 4.10%. Adequate precision expresses the signal  
225 to noise ratio and is desirable when its value is more than 4 and its value in our results was 31/10,  
226 which indicates a high reliability [35]. Figure 4 shows the separate and interaction effects of each  
227 of the study variables. Among the variables, electric current intensity (B) and electrolysis time (D)  
228 had the greatest effect on the removal of oxytetracycline, respectively. These findings show that  
229 the regression model provides a good explanation for the relationship between the four factors and  
230 the response. Finally, after removing the non-significant expressions by the proposed model,  
231 Equation 4 was obtained by the quadratic polynomial model for the removal of oxytetracycline.

232  $R = 20.72 + 7.25A + 35.25B - 1.36C + 0.47D - 1.07AB + 0.03AC + 0.005AD + 0.036BC +$   
233  $0.023BD + 0.009CD - 0.56A^2 - 4.30B^2 + 0.005C^2 - 0.009D^2$  (4)

234 In Equation (4), R represents the removal efficiency of oxytetracycline, and A represents the pH.  
235 B represents the density of electric current, C represents the initial concentration of the antibiotic,  
236 and D represents the time of electrolysis.

### 237 **3. 3. The effect of independent variables**

#### 238 **Effect of PH:**

239 In advanced oxidation processes, including electrochemical methods, pH plays an important role  
240 in the removal of contaminants. According to Figure 5-a, with increasing pH, the removal  
241 efficiency of oxytetracycline decreased, so that at the acidic pH, a higher removal percentage was  
242 observed. According to Table 1, in run No.4 and at pH = 3, the removal efficiency is equal to  
243 82.84%, while in the same conditions in run No.25 and at pH = 11, the removal efficiency has  
244 decreased to 61.55%. The effect of pH on the efficiency of the electrochemical process is  
245 manifested by its effect on the electrical production of metal hydroxyl. According to previous  
246 studies, acidic pH values provide better conditions for increasing the efficiency of the  
247 electrochemical process. Lowering the pH of the solution provides the conditions for increasing  
248 the production of hydroxyl radicals. Since the potential of these radicals to degrade pollutants in  
249 acidic environment is higher, so it leads to increased oxidation efficiency [36, 37].

#### 250 **Effect of current density:**

251 One of the important and effective parameters in electrochemical processes is the electric current  
252 density because it plays an important role in the production of hydroxyl radicals for the degradation  
253 and removal of pollutants from aqueous solutions. In this study, the electric current density in the  
254 range of 1-4 mA/cm<sup>2</sup> was used to determine its efficiency on removal efficiency. As can be seen

255 in Figure 5-a, the removal efficiency has gradually increased with increasing electric current  
256 density. In a period of 30 minutes and under constant conditions of other variables, with increasing  
257 electric current density from 1 mA/cm<sup>2</sup> in run No.12 to 4 mA / cm<sup>2</sup> in run No.5, the removal  
258 efficiency was augmented from 32.41% to 73.17% (Table 1). The results of Jafarzadeh et al.  
259 showed that increasing the electric current density leads to increasing the efficiency of pollutant  
260 removal, which were consistent with the present study [38]. In another study by Martinez and  
261 Bahena, the electric current density exhibited a direct relationship with increasing the removal  
262 efficiency in oxidation processes [39].

#### 263 **Effect of initial concentration of the oxytetracycline antibiotic:**

264 The initial concentration of the antibiotic provides an important driving force for overcoming all  
265 solute mass transfer resistances between the aqueous and solid phases. Figure 5-b shows that the  
266 rate of degradation of oxytetracycline in the electrochemical process is affected by its initial  
267 concentration, and with increasing the initial concentration of the oxytetracycline antibiotic from  
268 20 mg/L to 100 mg/L, its removal efficiency has decreased. By increasing the initial concentration,  
269 the amount of contact and exposure of the contaminant to hydroxyl radicals decreases, and  
270 consequently, leads to more consumption of hydroxyl radicals and a decrease in removal efficiency  
271 [40]. Also, increasing the initial concentration leads to the production of more by-products of  
272 oxidation, which results in more consumption of hydroxyl radicals and ultimately reduction of the  
273 removal efficiency [41].

#### 274 **Effect of electrolysis time:**

275 The findings of Figure 5-c indicate that with increasing electrolysis time, the efficiency of the  
276 electrochemical process for removal of oxytetracycline has increased significantly. By increasing  
277 the electrolysis time from 10 min to 50 min (thighs 3 and 6 in Table 1), the removal efficiency has

278 increased from 45.11% to 69.52%. With increasing electrolysis time, the electrodes have more  
279 opportunity to produce ferrous ions and increasing the ferrous ion solubility leads to the production  
280 of more hydroxyl radicals and finally, enhancement of the removal efficiency <sup>[42]</sup>. The study of  
281 Dargahi et al. showed that increasing the electrolysis time is directly related to the contaminant  
282 removal efficiency in the electrochemical process <sup>[43]</sup>. Findings from studies of other researchers  
283 have also shown that the removal efficiency of contaminants from aqueous solutions increases  
284 with increasing electrolysis time in oxidation processes <sup>[34, 44]</sup>.

### 285 **3.4. Optimization of process and investigation of the quality of the proposed model**

286 To achieve the highest removal efficiency of oxytetracycline antibiotic from various experiments  
287 designed with a central composite design, the optimal conditions of each of the variables, e.g., pH,  
288 electric current density ( $\text{mA}/\text{cm}^2$ ), initial concentration of oxytetracycline ( $\text{mg}/\text{L}$ ), and time  
289 Electrolysis (min) was obtained for the quadratic polynomial model according to Table 3, and  
290 finally, it was estimated that under these optimal conditions, the removal efficiency of  
291 oxytetracycline by electrochemical process was equal to 84.7%. Various studies have also pointed  
292 out the importance and usefulness of the central composite design for optimizing independent  
293 variables and achieving the highest efficiency of pollutant removal <sup>[45, 46]</sup>.

294 The quality of the proposed model was evaluated using graphs of values obtained from  
295 experimental values and predicted by the model and the normal distribution of data. As can be  
296 seen in Figure 6-a, the points are uniformly and homogenously aligned and there is no deviation  
297 in the data distribution, indicating a high correlation and normal distribution among the residual  
298 values. Figure 6 - b and c helps to understand the relationship between actual and predicted values  
299 and evaluates the fit of the model well. Since the residual values are randomly scattered in the  
300 standard deviation range and none of the points are out of range; thus, it provides a good agreement

301 between the actual results and the predicted values. As shown in Figure 6-d, the lowest point in  
302 the Box-Cox design with lambda equal to 1 (best lambda) indicates that there is no need to transfer  
303 the response of the proposed model [47, 48].

### 304 **5-3) Evaluation of efficiency of Fenton and electro-Fenton processes**

305 After determining the optimal conditions for the electrochemical process using a central composite  
306 design, the effect of different concentrations of FeSO<sub>4</sub> (0.1, 0.2, 0.3, and 0.4 gr/L) as a source of  
307 divalent iron ions on removal efficiency was investigated at different times in the Fenton process  
308 under optimal conditions, and the results of which are shown in Figure 7-a. By increasing the  
309 amount of FeSO<sub>4</sub> from 0.1 g/L to 0.4 g/L, the removal efficiency increased and the highest removal  
310 efficiency was obtained in the FeSO<sub>4</sub> amount equal to 0.3 g/L, and this amount was selected as the  
311 optimal value in the Fenton process, and removal efficiency in this case was 73.4%. Increasing the  
312 concentration of FeSO<sub>4</sub> leads to the production of more divalent iron ions in solution, which in  
313 turn increases the contaminant removal efficiency by producing more hydroxyl radicals [49]. The  
314 results of Yang et al. showed that increasing the dose of FeSO<sub>4</sub> to some extent increases the  
315 production of hydroxyl radicals and increases the removal efficiency, which were consistent with  
316 the present study [50].

317 According to Figure 7-b, changes in different concentrations of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) (0.03,  
318 0.06, 0.09, 0.12, and 0.15 mg/L) in the electro-Fenton process were investigated using the optimal  
319 process conditions of the electrochemical and optimal amount of FeSO<sub>4</sub> obtained in the Fenton  
320 process. The results showed that with increasing the amount of hydrogen peroxide, the removal  
321 efficiency in the electro-Fenton process increases consequently. In this study, the optimal amount  
322 of H<sub>2</sub>O<sub>2</sub> equal to 0.12 mg/L was obtained, and in this case, the oxytetracycline removal efficiency  
323 by electro-Fenton process reached 98.2%. A normal increase in concentration of hydrogen

324 peroxide leads to produce hydroxyl radicals and increase the power of oxidizing due to reaction  
325 with the ferrous ion, and radicals generated during access times more pollutants were causing  
326 further degradation of oxytetracycline and increased removal efficiency <sup>[51]</sup>. Ahmadzadeh and  
327 Dolatabadi, in their study, showed that increasing the concentration of hydrogen peroxide  
328 increases the efficiency of the electro-Fenton process to remove antibiotics <sup>[52]</sup>.

### 329 **6-3) Comparison of process efficiency, the effect of SS316/ $\beta$ -PbO<sub>2</sub> anode on removal** 330 **efficiency**

331 As can be seen in Figure 8-a, the efficiency of each of the electrochemical processes, Fenton and  
332 electro-Fenton, has been investigated under optimal conditions at different times. According to the  
333 results in the optimal time of 42.35 minutes, the removal efficiencies of Fenton and  
334 electrochemical processes were 73.4% and 84.7%, respectively, while the efficiency in the electro-  
335 Fenton process reached 98.2%, and since this process could offer greatest effect on oxytetracycline  
336 antibiotic removal, it was selected as the optimal process.

337 Coating of the anode electrode using lead dioxide ( $\beta$ -PbO<sub>2</sub>) is one of the effective and practical  
338 methods to remove contaminants using oxidation processes that can affect the removal efficiency  
339 <sup>[43]</sup>. In this study, the stainless steel anode was coated with lead dioxide and then under the optimal  
340 conditions of the electro-Fenton process, the effect of the SS316/ $\beta$ -PbO<sub>2</sub> anode electrode in  
341 comparison with the SS316 anode was investigated, and the relevant results in Figure 8-b showed  
342 that the SS316/ $\beta$ -PbO<sub>2</sub> anode was only more effective in removing oxytetracycline than SS316, so  
343 that at an optimal time of 42.35 min, the removal efficiency for SS316/ $\beta$ -PbO<sub>2</sub> was 100%.

### 344 **7-3) Calculation of electrical energy consumption**

345 The amount of energy consumed under optimal conditions for the outlet solution treated by the  
346 electro-Fenton process was calculated using Equation (5) <sup>[53,54]</sup>. The results showed that the energy

347 consumption for the three voltages (6.5, 11.5 and 15.5 volts) were equal to 0.91, 3.86, and 8.24  
348 kwh/m<sup>3</sup>, respectively. Based on the results, increasing the amount of voltage was led to an increase  
349 in the removal efficiency of oxytetracycline, and the amount of electrical energy consumed was  
350 enhanced (Eq. 5).

$$351 \quad E_{EC} = \frac{UIt_{EC}}{V_s} \times 10^{-3} \quad (5)$$

352 In the above equation,  $E_{EC}$  represents the electrical energy consumed (kwh/m<sup>3</sup>),  $U$  is the reactor  
353 voltage (volts),  $I$  is the electric current intensity (amperes),  $t_{EC}$  is the electrolysis time (hour), and  
354  $V_s$  is the reactor volume (m<sup>3</sup>).

### 355 **8-3) Results of toxicity test**

356 Figure 9 shows the results of the toxicity test of the inlet solution and the effluent treated by the  
357 electro-Fenton process. The results showed that after 10 hours, the growth rate of both bacteria in  
358 the control sample and the effluent increased and consequently the toxicity decreased, while the  
359 growth of bacteria in the inlet solution was less and its toxicity was high. Results 10-hour mean  
360 growth inhibition under optimal electro-Fenton process conditions (pH = 3.53, current density =  
361 3.85 mA/cm<sup>2</sup>, concentration of oxytetracycline = 20 mg/L, electrolysis time = 42.35 min, FeSO<sub>4</sub>  
362 = 0.3 gr/L, and H<sub>2</sub>O<sub>2</sub> = 0.12 mg/L) showed that the toxicity for *Pseudomonas aeruginosa* decreased  
363 from 75.6% in the reactor inlet solution to 11.7% in the outlet solution (84.5% reduction in  
364 toxicity) and for *Staphylococcus aureus*, was declined from 41.4% in the inlet solution to 12.8%  
365 in the outlet solution (69% reduction in toxicity). Nadafi et al. used standard strains of *Escherichia*  
366 *coli* and *Staphylococcus aureus* to evaluate for toxicity using a bioassay; their results showed that  
367 the reduction of *Escherichia coli* and *Staphylococcus* toxicity in the outlet solution was 86.7% and  
368 72.3%, respectively [55]. Dirany et al., by conduction a study on the sulfomethoxazole antibiotic

369 toxicity by bioassay with microorganisms, noted that the toxicity of the effluent treated by the  
370 electro-Fenton process was reduced by about 75% compared to the inlet solution <sup>[56]</sup>. In the present  
371 study, the electro-Fenton process effectively reduced the toxicity of aqueous solution containing  
372 oxytetracycline after treatment, and the bioassay method using microorganisms could be used as  
373 an efficient and cost-effective technique to evaluate the toxicity of aqueous solutions.

### 374 **3) Intermediates from LC-MS analysis**

375 Intermediates obtained from the degradation of the oxytetracycline antibiotic were identified by  
376 LC-MS (Figure 10). According to the spectral data obtained from the method used, the mechanism  
377 of the oxytetracycline compound removal in the presence of hydroxide ions was presented in Table  
378 4 and as schematic 1. Initially, the oxytetracycline compound undergoes a demethylation reaction  
379 in the presence of active hydroxide ions, and the (4S, 4aR, 5S, 5aR, 6S, 12aR) -4-amino-  
380 1,5,6,10,11,12a-hexahydroxy-6 -methyl-3,12-dioxo-3,4,4a, 5,5a, 6,12,12a-octahydrotetracene-2-  
381 carboxamide compound with a molecular mass of 432 is produced. The second demethylation  
382 process is performed and the compound with IUPAC name of (4S, 4aR, 5S, 5aR, 6S, 12aR) -4-  
383 amino-1,5,6,10,11,12a-hexahydroxy-3,12- produces dioxo-3,4,4a, 5,5a, 6,11,11a, 12,12a-  
384 decahydrotetracene-2-carboxamide and mass of 420. This compound forms compounds with  
385 masses of 404 and 361, in two steps and by losing the functional group of hydroxide and CONH<sub>2</sub>,  
386 respectively. The compound with mass of 361 can produce a compound with mass of 268 by losing  
387 all the functional groups of hydroxide and amine. From this compound, it can produce two  
388 compounds with masses of 240 and 200, which in turn, compounds called 1- (1,2,3,4-  
389 tetrahydronaphthalen-2-yl) ethan-1-one with mass 174 and 1,2,3,4-tetrahydronaphthalene-2-  
390 carbaldehyde with mass 160 and finally 1,2,3,4-tetrahydronaphthalene with mass 132 are  
391 produced. At the end of the process, compounds with lower molecular masses such as toluene,

392 benzene, carbonic acid and acetic acid are produced, the degradation of which leads to water and  
393 carbon dioxide.

### 394 **10-3) Study of process kinetics**

395 Kinetic models were used to evaluate the speed of chemical reactions, in which the determination  
396 of the reaction rate of liquids and gases is expressed in terms of the concentration of the substance  
397 (pollutant) per unit time. The basis of the reaction rate is the decrease of the reactant concentration  
398 per unit time or the increase of the product concentration per unit time <sup>[57]</sup>. In this study, a pseudo-  
399 first-order kinetic model under optimal electro-Fenton process conditions was used to estimate the  
400 process kinetics. Equation (6) was used to calculate the pseudo-first-order kinetic model for the  
401 removal of the oxytetracycline antibiotic.

$$402 \quad \text{Ln} \left( \frac{[\text{OxyTET}]_0}{[\text{OxyTET}]_e} \right) = kt \quad (6)$$

403 In the above equation,  $[\text{OxyTET}]_0$  and  $[\text{OxyTET}]_e$  represent the initial and final concentrations of  
404 the antibiotics oxytetracycline and  $k$ , respectively. The constant  $k$  was calculated by plotting the  
405  $\text{Ln} ([\text{OxyTET}]_0/[\text{OxyTET}]_t)$  versus the electrolysis time. Equation (7) was also used to determine  
406 the half-life of the pollutant.

$$407 \quad t_{1/2} = \frac{0.693}{k} \quad (7)$$

408 The results of the evaluation of the oxytetracycline degradation under optimal conditions with a  
409 pseudo-first-order kinetic model were shown in Table 5 and Figure 11. As can be seen, the value  
410 of regression coefficient ( $R^2$ ) for the pseudo-first order kinetic model was determined to be 0.9965,  
411 which indicates the suitability of the kinetic model to express the data for the degradation of  
412 oxytetracycline <sup>[58, 59]</sup>. According to Table 5, the kinetic parameters represent a good linear

413 relationship, and also the degradation of oxytetracycline by the electro-Fenton process follows  
414 pseudo-first-order kinetics; accordingly, the studied system has a very good potential for the  
415 degradation of oxytetracycline. In the present study, the velocity constant (k) of the pseudo-first-  
416 order model for degradation of oxytetracycline by electro-Fenton process was  $0.073 \text{ min}^{-1}$ , and the  
417 half-life ( $t_{1/2}$ ) of oxytetracycline removal was 9.5 min.

#### 418 **4) Conclusion**

419 The findings of this study showed that the efficiency of electrochemical processes, Fenton and  
420 electro-Fenton for the removal of the antibiotic oxytetracycline under optimal test conditions were  
421 84.7%, 73.4%, and 98.2%, respectively; thus, the electro-Fenton process as an optimal process  
422 was selected for oxytetracycline removal. Optimal conditions for pH, electric current density,  
423 initial oxytetracycline concentration, electrolysis time,  $\text{FeSO}_4$  and  $\text{H}_2\text{O}_2$  concentrations are 3.53,  
424  $3.85 \text{ mA/cm}^2$ , 20 mg/L, 42.35 min, 3 g/L, and 0.12 mg/L, respectively. Under the optimal  
425 conditions of the electro-Fenton process, the oxytetracycline antibiotic removal efficiency by the  
426 SS316/ $\beta\text{-PbO}_2$  anode electrode reached 100%, and this anode had a significant effect on the  
427 removal efficiency compared to SS316 alone. The bioassay method using microorganisms  
428 effectively reduced the toxicity of effluent containing oxytetracycline treated by the electro-Fenton  
429 process; thus this method can be used as an effective tool to investigate the toxicity of pollutants  
430 from aqueous solutions. Intermediates obtained from the degradation of the antibiotic  
431 oxytetracycline were identified and determined by LC-MS. Results from the kinetic studies of  
432 process showed that the removal of oxytetracycline using the electro-Fenton process with  $R^2 =$   
433 0.9965 follows quasi-first-order kinetics.

434

435

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451 **Authorship contribution statement**

452 Abdollah Dargahi: Conceptualization, Methodology, Validation, Formal analysis, Investigation,  
453 Resources, Supervision, Funding acquisition. Mehdi Vosoughi: Methodology, Validation,  
454 Resources, Writing - original draft, Writing - review & editing. Somayeh Biparva Haghighi:  
455 Analyses, writing and text revision. Kamal Hasani: Methodology, Validation, Formal analysis,

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Table 1. Matrix results of experimental design for removal of Oxytetracycline antibiotic via electrochemical process

Run	A	B	C	D	Removal (%)	
	pH	current density (mA/cm <sup>2</sup> )	Concentration (mg/L)	electrolysis time (min)	Actual Value	Predicted Value
1	7	2.5	20	10	74.7	73.98
2	7	2.5	60	30	62.7	66.38
3	3	2.5	60	50	69.52	69.73
4	3	2.5	20	30	82.84	82.69
5	3	4	60	30	73.17	72.77
6	3	2.5	60	10	45.11	47.72
7	7	4	60	10	55.78	54.74
8	11	2.5	60	10	35.78	37.17
9	3	2.5	100	30	60.23	57.89
10	11	2.5	60	50	61.76	60.75
11	7	2.5	60	30	63.24	66.38
12	3	1	60	30	32.41	32.48
13	11	1	60	30	35.7	35.61
14	11	4	60	30	50.68	50.11
15	7	2.5	60	30	67.9	66.38
16	7	2.5	60	30	70.63	66.38
17	7	2.5	100	50	81.9	82.12
18	7	1	60	10	31.2	28.72
19	7	2.5	60	30	69.66	66.38
20	7	1	60	50	50.2	50.14
21	7	2.5	60	30	64.15	66.38
22	7	1	20	30	59.86	60.53
23	7	4	100	30	72.34	73.27
24	7	4	60	50	77.54	78.92
25	11	2.5	20	30	61.55	62.78
26	7	2.5	20	50	83	82.26
27	11	2.5	100	30	59.22	58.26
28	7	2.5	100	10	44.55	44.80
29	7	4	20	30	83.8	83.51
30	7	1	100	30	39.55	41.45

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Table 2. ANOVA results pertaining to Oxytetracycline elimination

Source	Sum of Squares	df	Mean Square	F-value	p-value	Status
<b>Model</b>	7086.49	14	506.18	81.58	< 0.0001	significant
A-pH	286.07	1	286.07	46.11	< 0.0001	significant
B-current density	2252.01	1	2252.01	362.97	< 0.0001	significant
C-Concentration	644.75	1	644.75	103.92	< 0.0001	significant
D-electrolysis time	1559.52	1	1559.52	251.36	< 0.0001	significant
AB	166.15	1	166.15	26.78	0.0001	significant
AC	102.82	1	102.82	16.57	0.0010	significant
AD	0.6162	1	0.6162	0.0993	0.7570	not significant
BC	19.58	1	19.58	3.16	0.0959	not significant
BD	1.90	1	1.90	0.3069	0.5877	not significant
CD	210.98	1	210.98	34.00	< 0.0001	significant
A <sup>2</sup>	550.50	1	550.50	88.73	< 0.0001	significant
B <sup>2</sup>	642.20	1	642.20	103.51	< 0.0001	significant
C <sup>2</sup>	437.35	1	437.35	70.49	< 0.0001	significant
D <sup>2</sup>	87.70	1	87.70	14.14	0.0019	significant
<b>Residual</b>	93.07	15	6.20	-	-	-
Lack of Fit	33.56	10	3.36	0.2820	0.9581	not significant
Pure Error	59.51	5	11.90	-	-	-
<b>Cor Total</b>	7179.56	29	-	-	-	-
<b>R<sup>2</sup> = 0.9870</b>				<b>Std. Dev = 2.49</b>		
<b>Adjusted R<sup>2</sup> = 0.9749</b>				<b>Mean = 60.69</b>		
<b>Predicted R<sup>2</sup> = 0.9611</b>				<b>C.V (%) = 4.10</b>		
<b>Adeq Precision = 31.10</b>						

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Table 3. Optimal condition of independent variables for Oxytetracycline removal via electrochemical process.

Factor	Low	High	Optimum
pH	3.0	11.0	3.53
current density (mA/cm <sup>2</sup> )	1.0	4.0	3.85
Concentration (mg/L)	20.0	100.0	20
electrolysis time (min)	10.0	50.0	42.35
Optimum Value %			84.7 %

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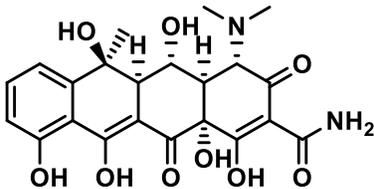
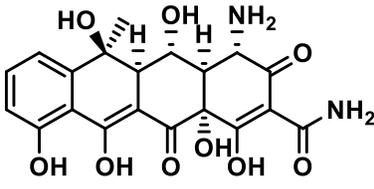
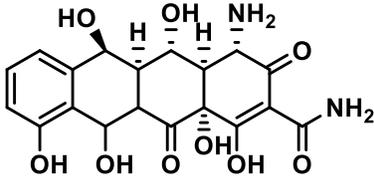
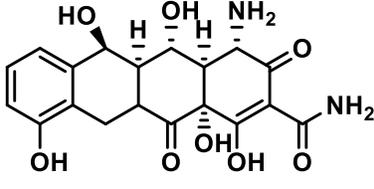
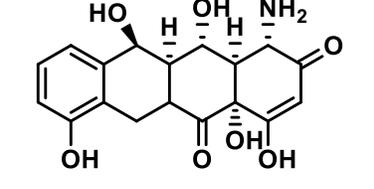
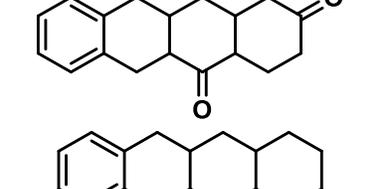
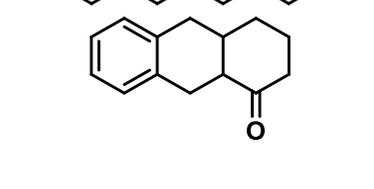
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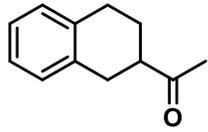
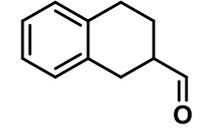
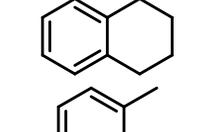
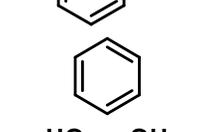
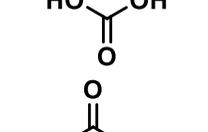
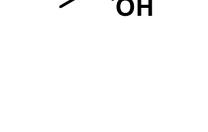
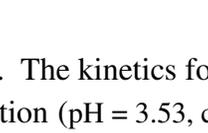
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Table 4. Identified products by LC-MS in the degradation of Oxytetracycline antibiotic by the Electro-Fenton process.

Structure	IUPC name	Molecular Weight
	(4S,4aR,5S,5aR,6S,12aR)-4-(dimethylamino)-1,5,6,10,11,12a-hexahydroxy-6-methyl-3,12-dioxo-3,4,4a,5,5a,6,12,12a-octahydrotetracene-2-carboxamide	460
	(4S,4aR,5S,5aR,6S,12aR)-4-amino-1,5,6,10,11,12a-hexahydroxy-6-methyl-3,12-dioxo-3,4,4a,5,5a,6,12,12a-octahydrotetracene-2-carboxamide	432
	(4S,4aR,5S,5aR,6S,12aR)-4-amino-1,5,6,10,11,12a-hexahydroxy-3,12-dioxo-3,4,4a,5,5a,6,11,11a,12,12a-decahydrotetracene-2-carboxamide	420
	(4S,4aR,5S,5aR,6S,12aR)-4-amino-1,5,6,10,12a-pentahydroxy-3,12-dioxo-3,4,4a,5,5a,6,11,11a,12,12a-decahydrotetracene-2-carboxamide	404
	(1S,4aR,11S,11aR,12S,12aR)-1-amino-4,4a,7,11,12-pentahydroxy-5a,6,11,11a,12,12a-hexahydrotetracene-2,5(1H,4aH)-dione	361
	4,4a,5a,6,11,11a,12,12a-octahydrotetracene-2,5(1H,3H)-dione	268
	1,2,3,4,4a,5,5a,6,11,11a,12,12a-dodecahydrotetracene	240
	3,4,4a,9,9a,10-hexahydroanthracen-1(2H)-one	200

	1-(1,2,3,4-tetrahydronaphthalen-2-yl)ethan-1-one	174
	1,2,3,4-tetrahydronaphthalene-2-carbaldehyde	160
	1,2,3,4-tetrahydronaphthalene	132
	toluene	92
	benzene	78
	carbonic acid	62
	acetic acid	60

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678

679 Table 5. The kinetics for the removal of Oxytetracycline by Electro-Fenton process in optimum  
 680 condition (pH = 3.53, current density = 3.85 mA/cm<sup>2</sup>, concentration of Oxytetracycline = 20 mg/L,  
 681 electrolysis time = 42.35 min, FeSO<sub>4</sub> = 0.3 gr/L, H<sub>2</sub>O<sub>2</sub> = 0.12 mg/L)

kinetics	k (min <sup>-1</sup> )	R <sup>2</sup>	t <sub>1/2</sub> (min)
pseudo-first-order model	0.073	0.9965	9.5

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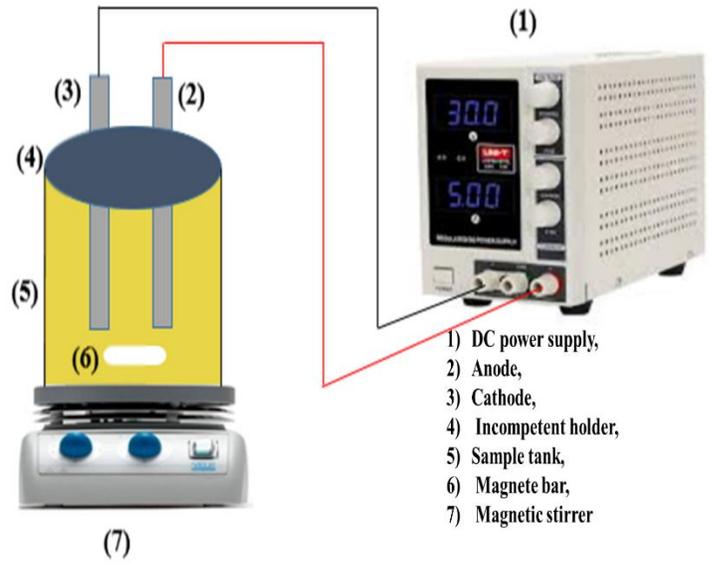
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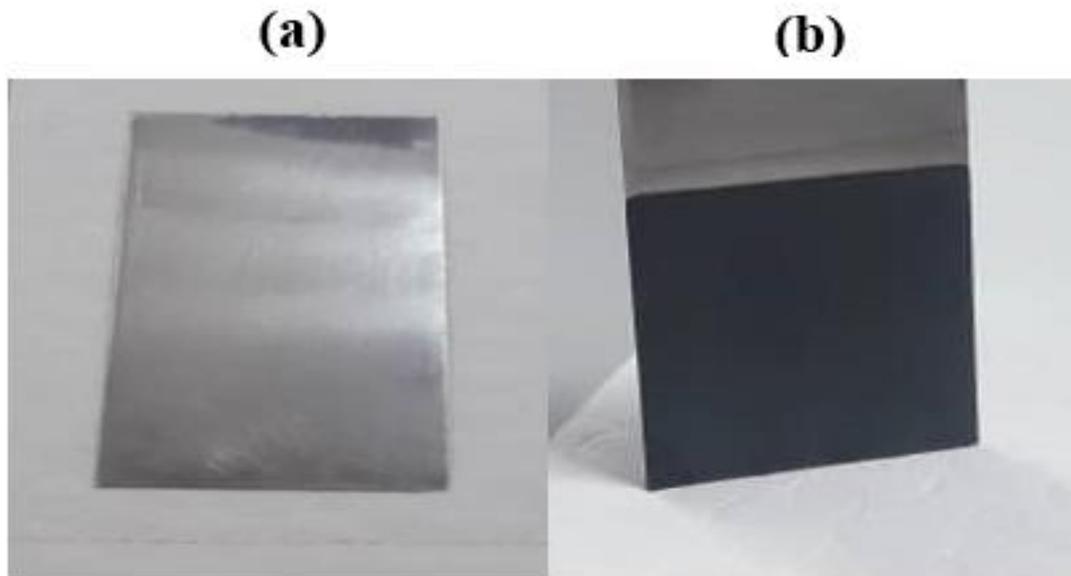
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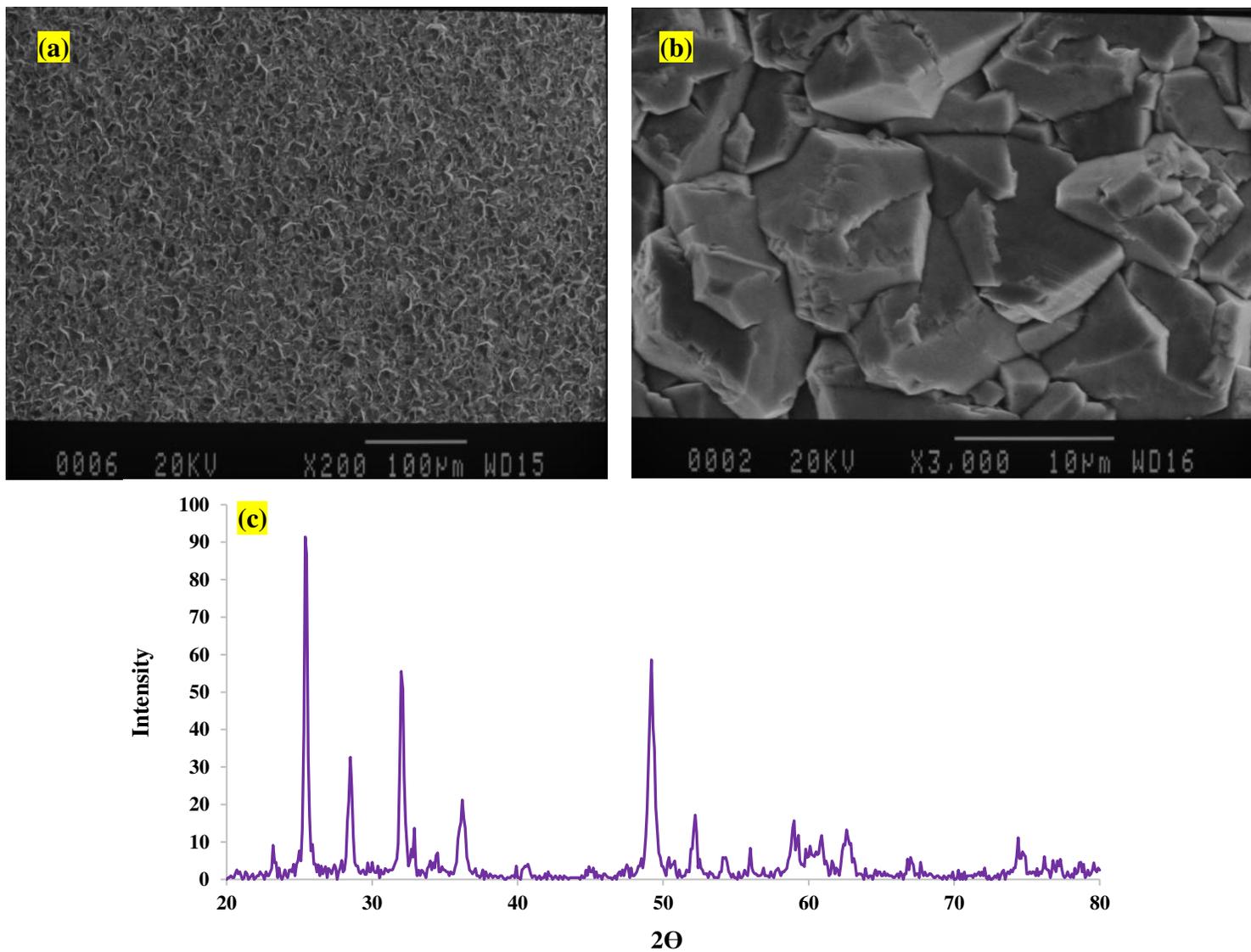
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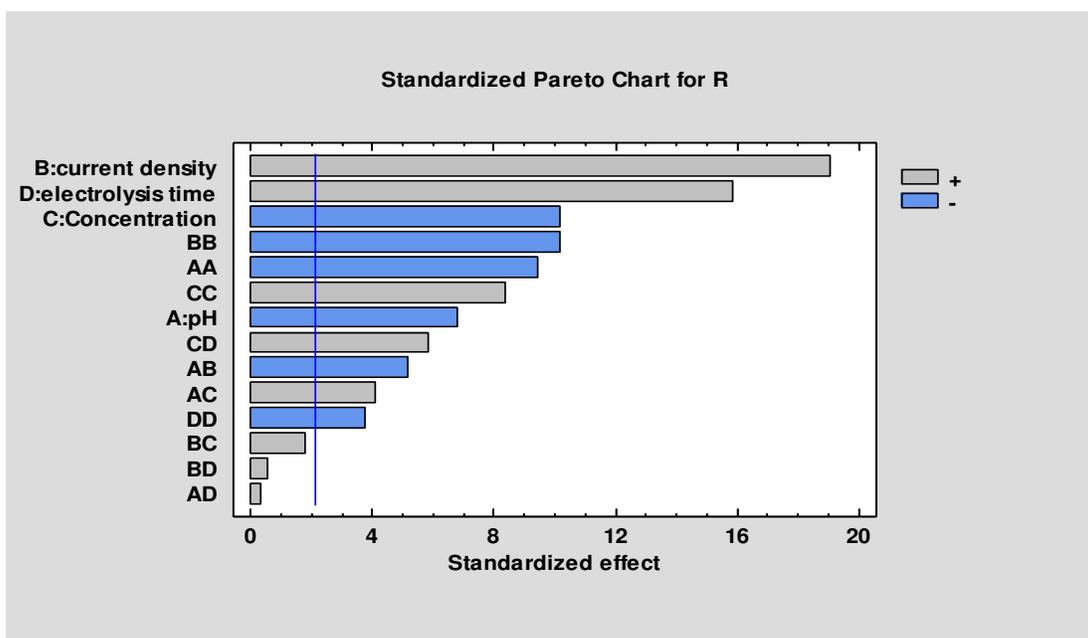
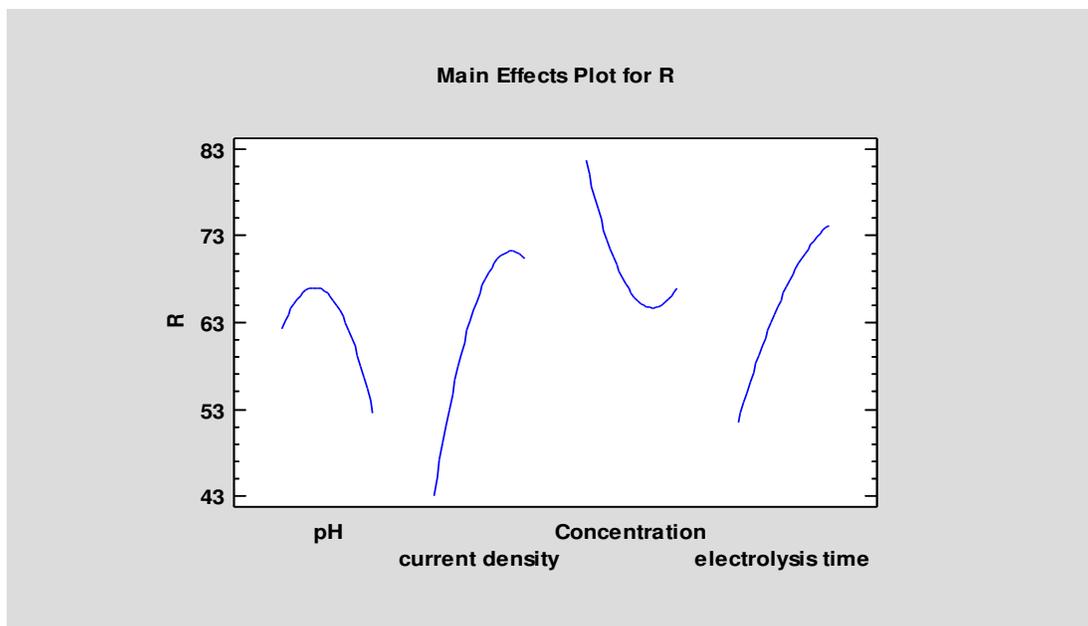
690 **Fig 1:** Schematic of the reactor used for removal of oxytetracycline by oxidation processes



691  
 692 **Fig 2.** (a) Image of stainless steel electrode and (b) Stainless steel electrode coated with lead  
 693 dioxide (SS316/PbO<sub>2</sub>) for removal of the oxytetracycline antibiotic under optimal test conditions



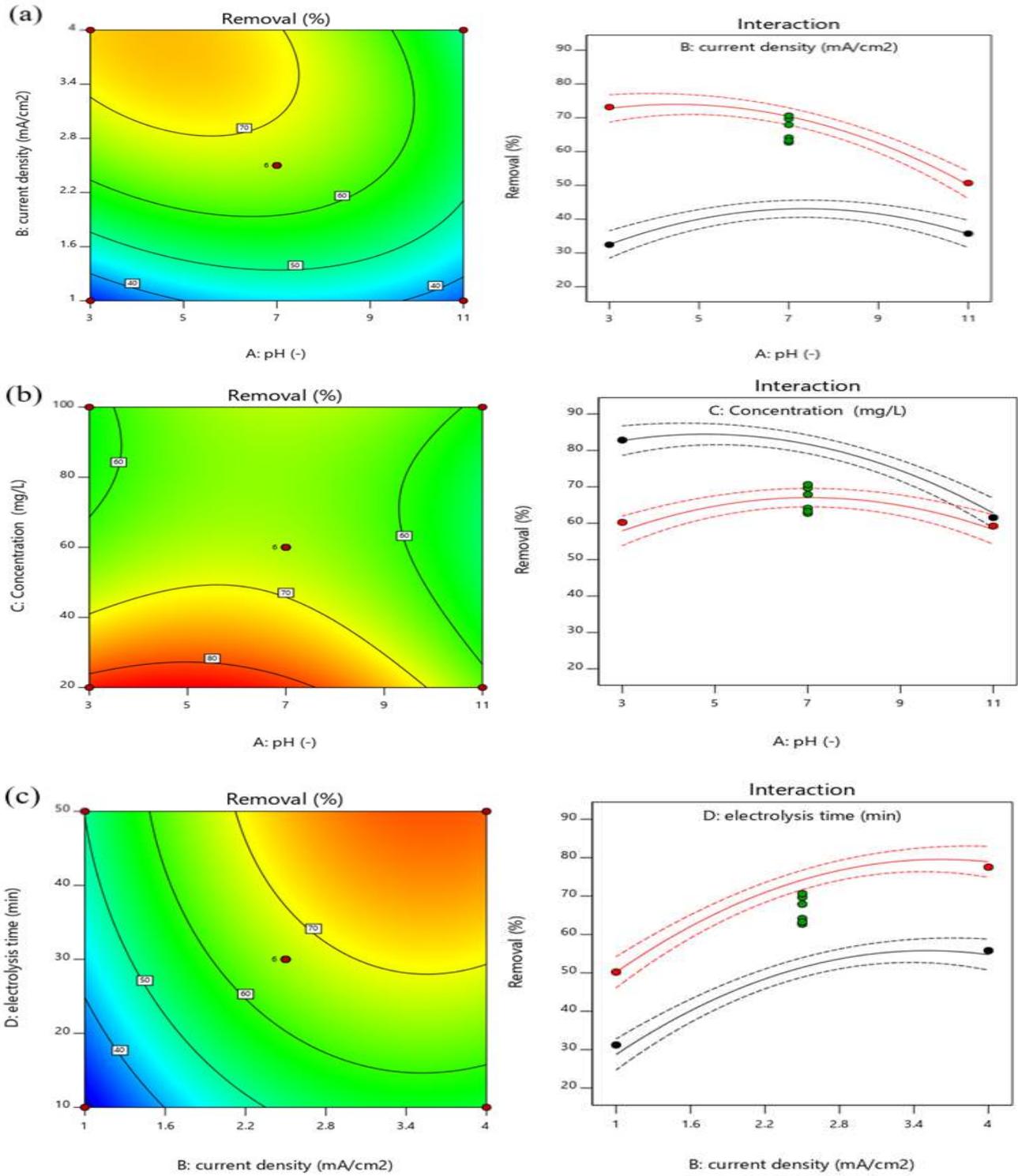
694 **Fig. 3.** (a, b) SEM micrographs of the surface of SS316/ $\beta$ -PbO<sub>2</sub> at different magnifications; (c)  
 695 XRD analysis of SS316/ $\beta$ -PbO<sub>2</sub> anode.



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697 **Fig.4.** Separate effects of variables and the pareto plot for determining the effect of each term on  
 698 the response (removal of Oxytetracycline by electrochemical process).

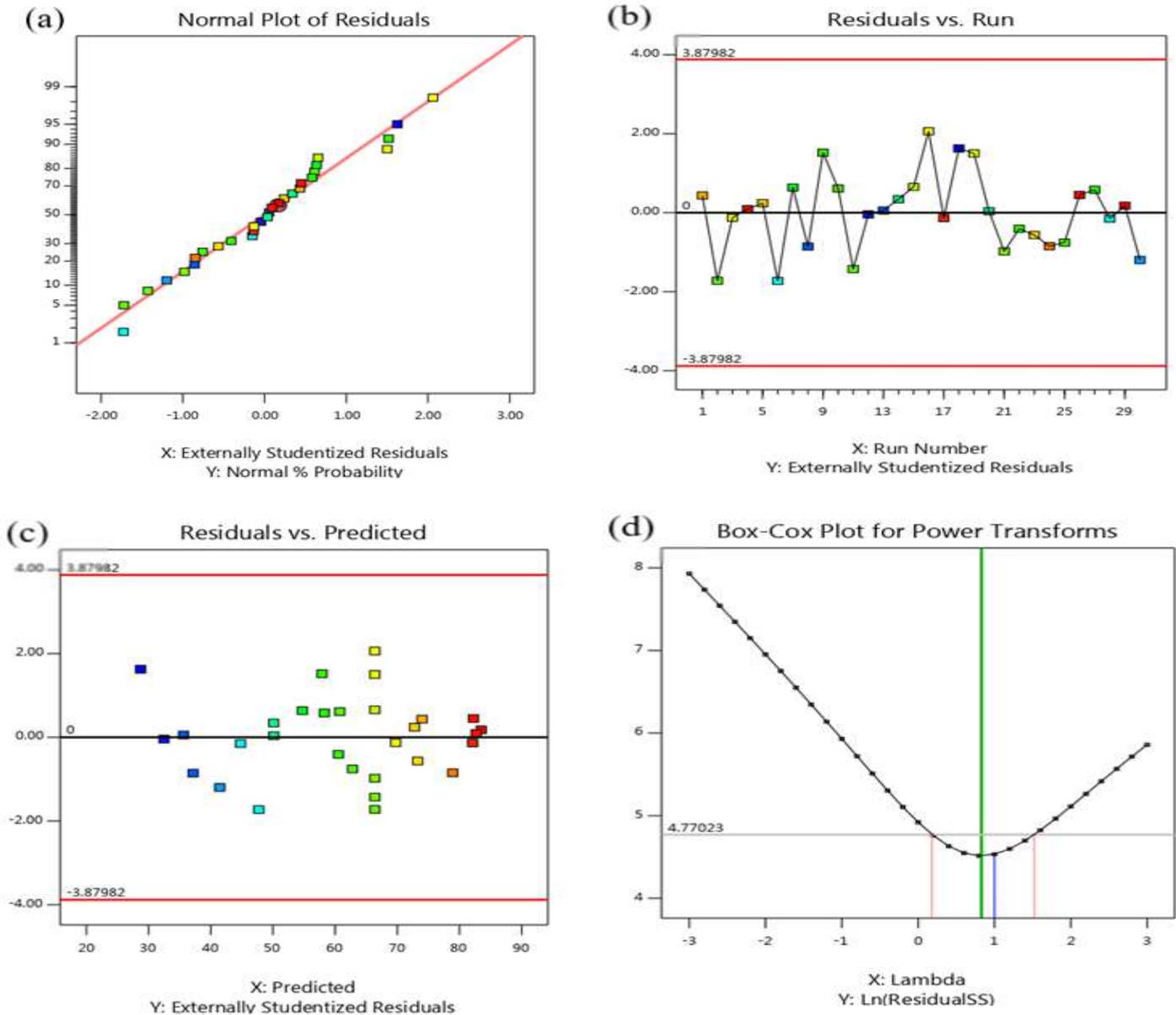
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700 Fig. 5. (a) Contour plots and interaction effects of pH and current density (mA/cm<sup>2</sup>), (b)  
 701 Contour plots and interaction effects of pH and initial concentration of Oxytetracycline (mg/L),  
 702 (c) Contour plots and interaction effects of current density (mA/cm<sup>2</sup>) and electrolysis time (min).

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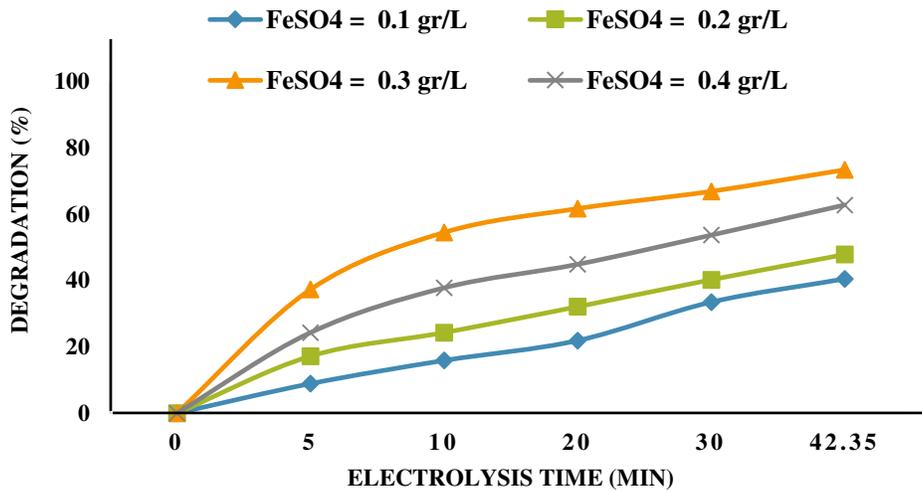
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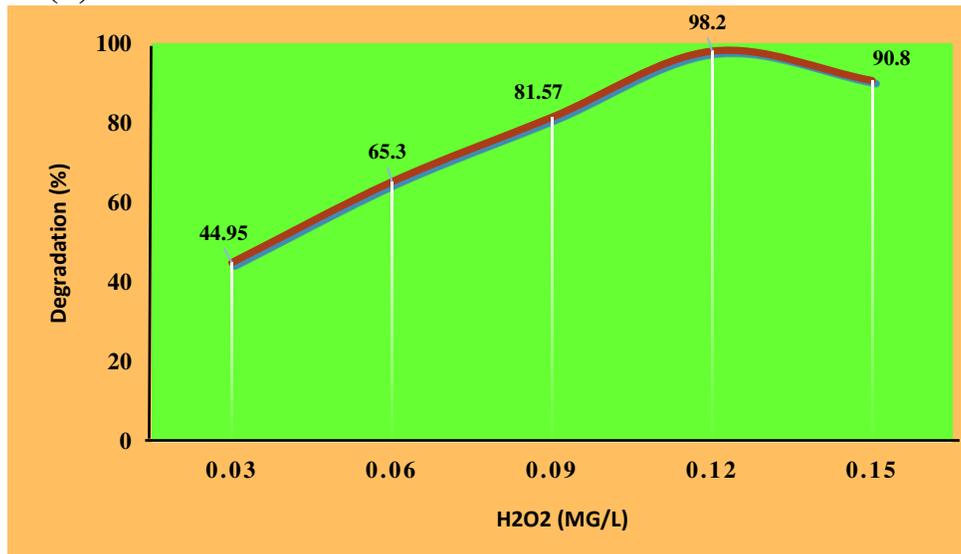
Fig. 6. The diagnostic plots for validation of obtained model: (a) Normal probability distribution of residuals, (b) Externally studentized residuals versus run order, (c) Residuals versus predicted and (d) Box-Cox Plot.

(a)



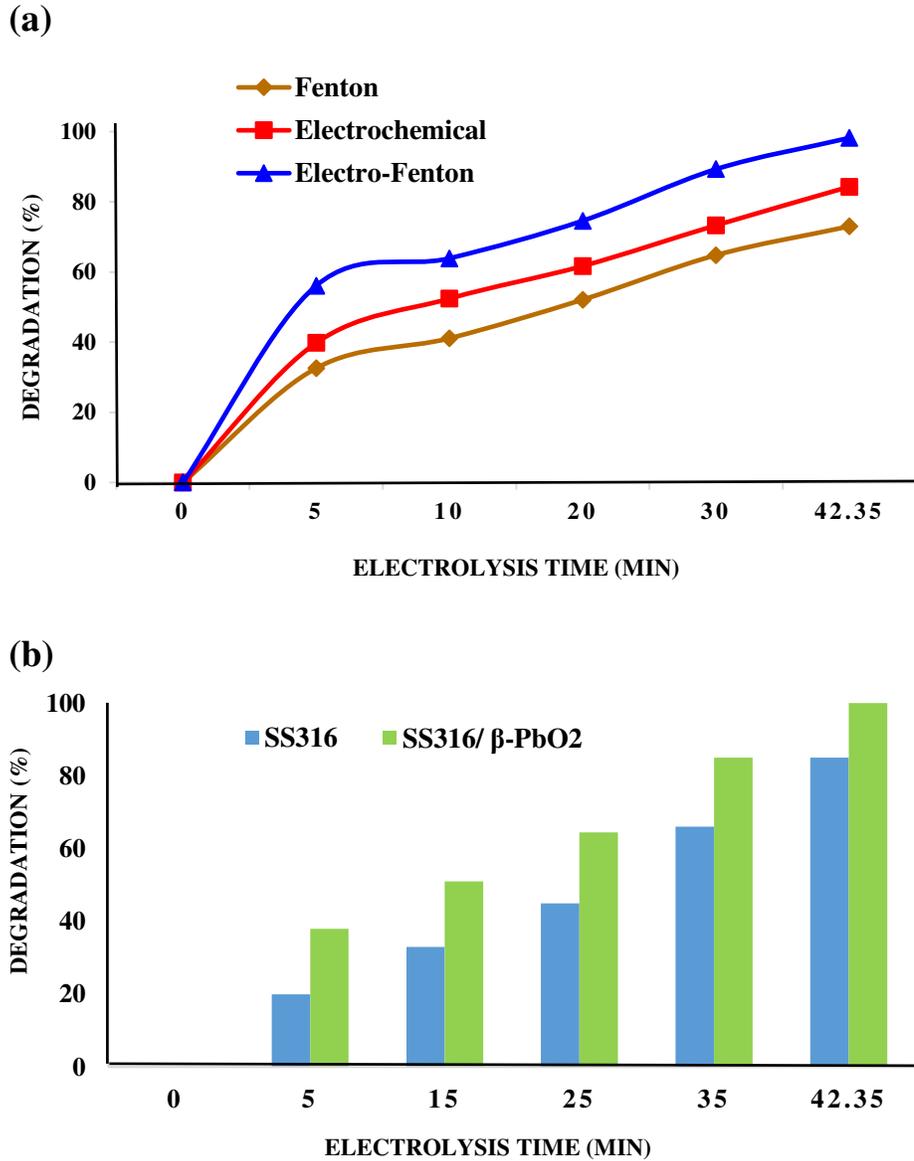
The changes of FeSo4 amounts in optimal condition via Fenton process  
(pH = 3.53, concentration = 20 mg/L)

(b)

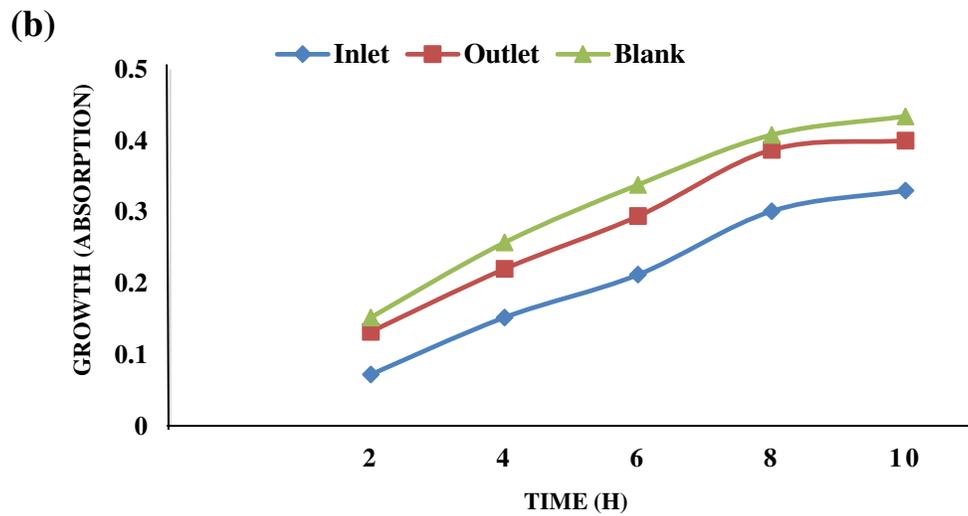
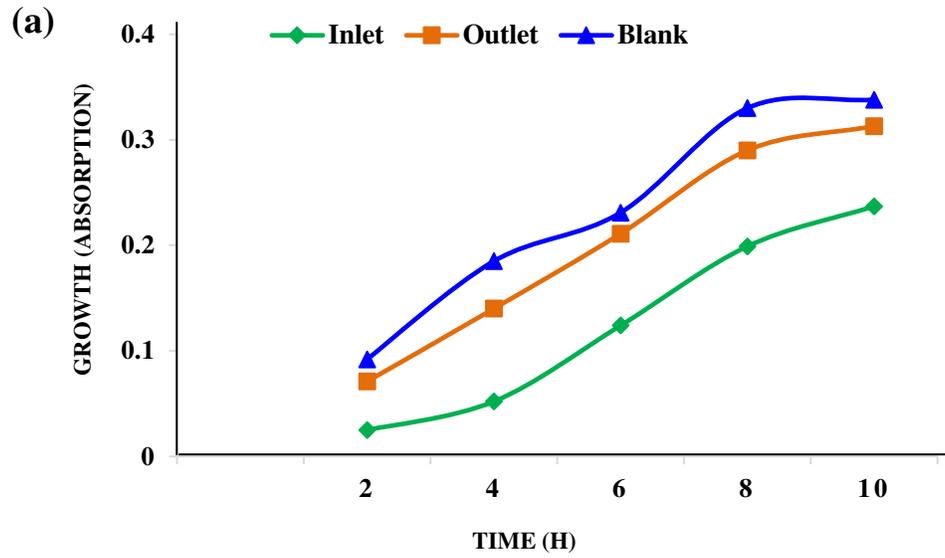


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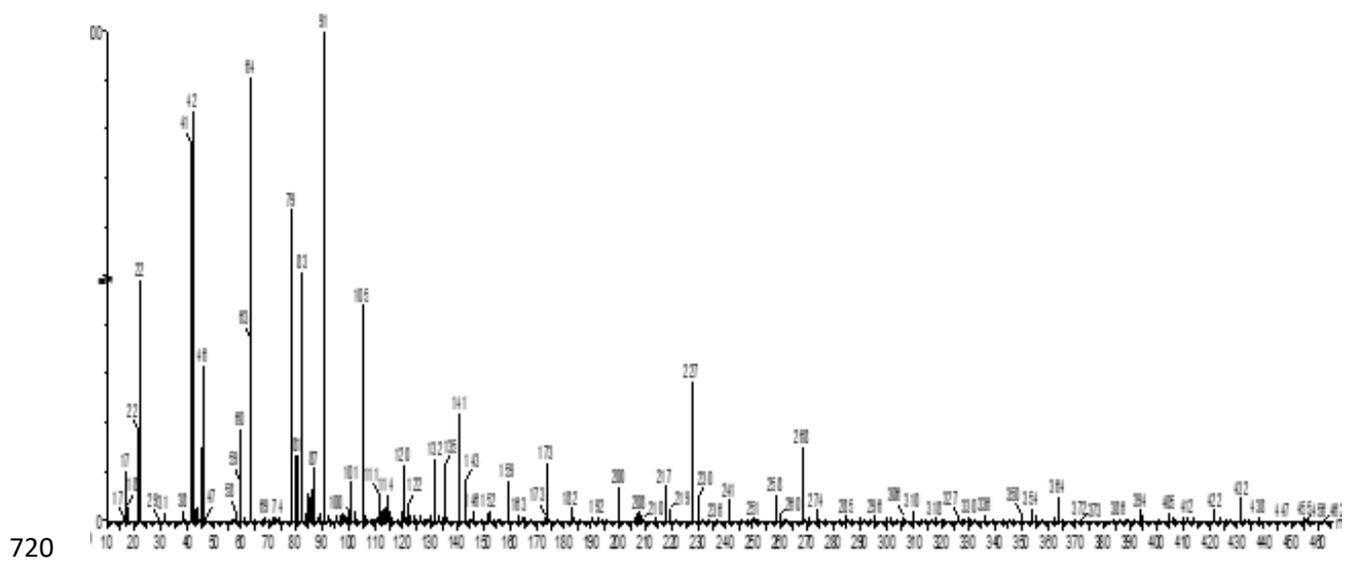
710 Fig. 7. (a) Influence of FeSo4 amounts via Fenton process and (b) H<sub>2</sub>O<sub>2</sub> values in elimination  
711 of Oxytetracycline in electro-Fenton process under optimum condition (pH= 3.53, current  
712 density = 3.85 mA/cm<sup>2</sup>, concentration = 20 mg/L, electrolysis time = 42.35 min and FeSO<sub>4</sub> = 0.3  
713 gr/L). (a: Optimize amount of FeSO<sub>4</sub> = 0.3 gr/L, b: Optimize value of H<sub>2</sub>O<sub>2</sub> = 0.12 mg/L)



714 Fig. 8. (a) The effect of AOPs processes on the degradation of Oxytetracycline antibiotic, (b)  
 715 The elimination of Oxytetracycline antibiotic by the SS316 and SS316/ $\beta$ -PbO<sub>2</sub> anodes (pH =  
 716 3.53, current density = 3.85 mA/cm<sup>2</sup>, concentration of Oxytet = 20 mg/L, electrolysis time = 42.35 min,  
 717 FeSO<sub>4</sub> = 0.3 gr/L, H<sub>2</sub>O<sub>2</sub> = 0.12 mg/L).



718 Fig. 9. (a) Growth trend of *Pseudomonas aeruginosa* (gram-negative) and (b) *Staphylococcus*  
 719 *aureus* (gram-positive) in toxicity test under optimal conditions of Electro-Fenton process.

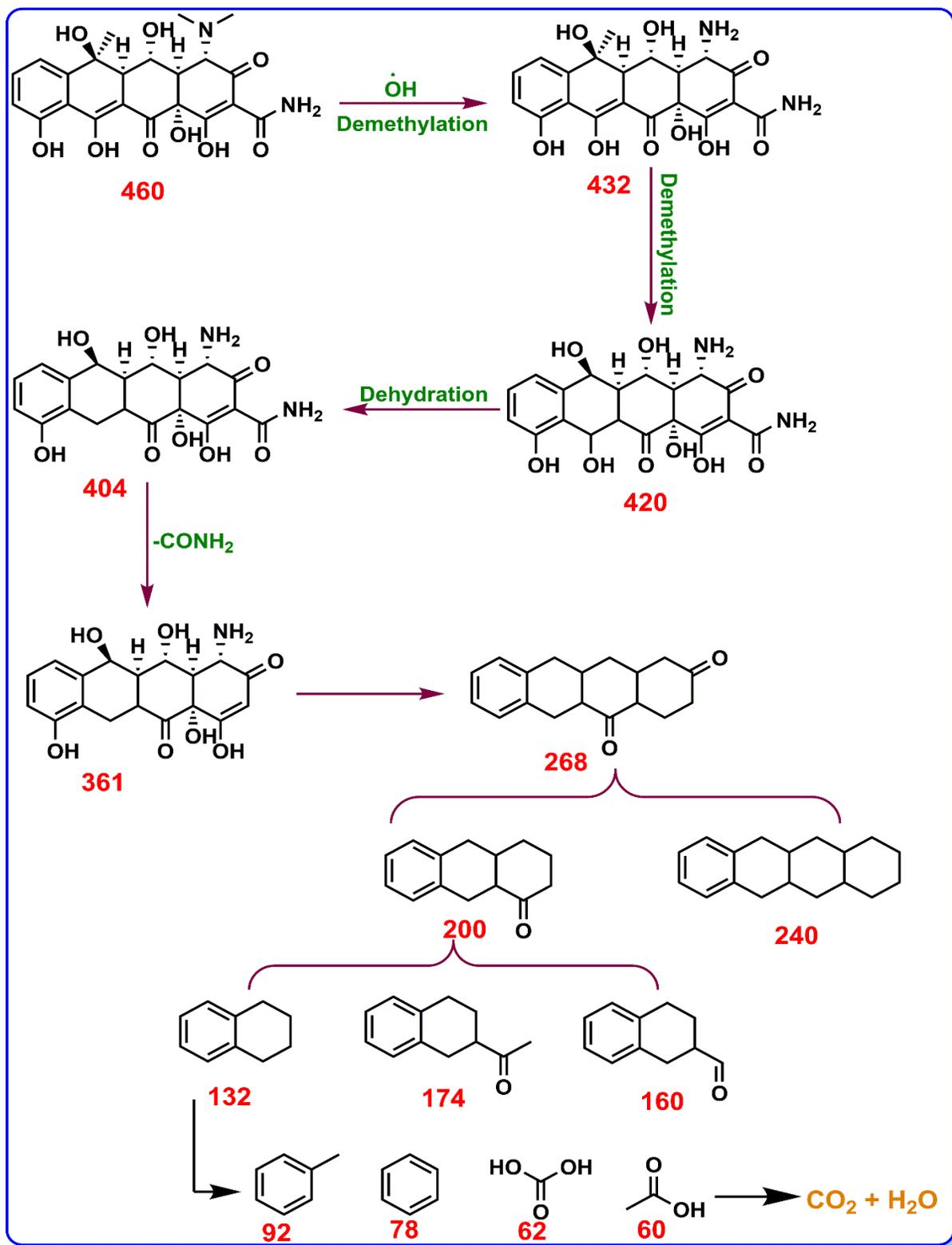


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Fig.10. LC/MS chromatographs after degradation of Oxytetracycline antibiotic via Electro-Fenton process under optimum conditions.

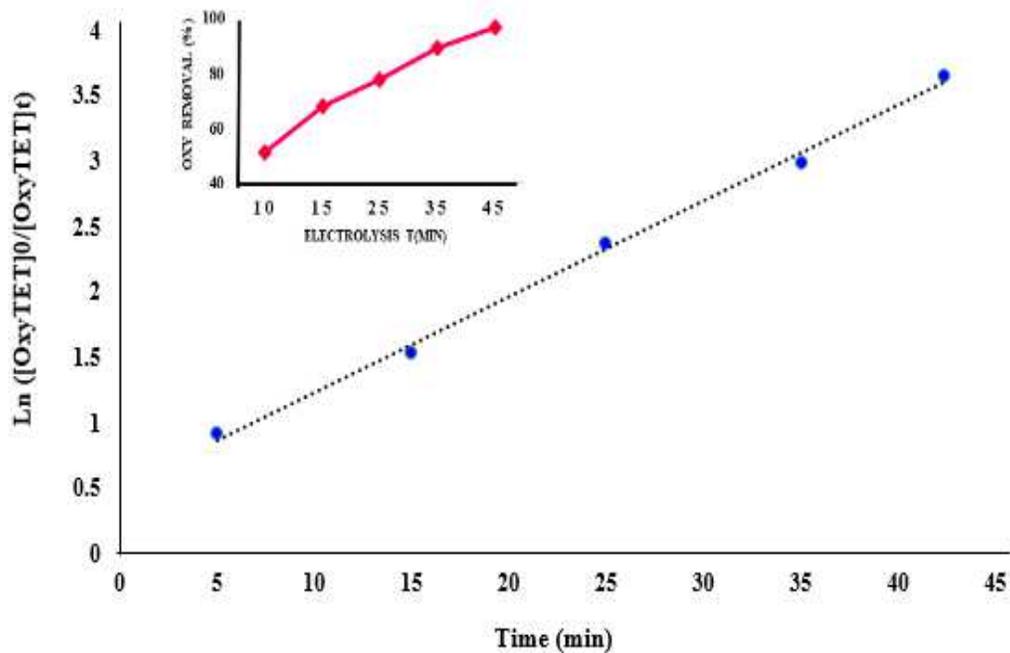


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Scheme 1. Proposed pathway for degradation of Oxytetracycline antibiotic by the Electro-Fenton process.



726

727 **Fig. 11.** Kinetics of Oxytetracycline removal at the optimum conditions (pH = 3.53, current  
 728 density = 3.85 mA/cm<sup>2</sup>, concentration of Oxytetracycline = 20 mg/L, electrolysis time = 42.35 min,  
 729 FeSO<sub>4</sub> = 0.3 gr/L, H<sub>2</sub>O<sub>2</sub> = 0.12 mg/L) with pseudo- order model.

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732

# Figures

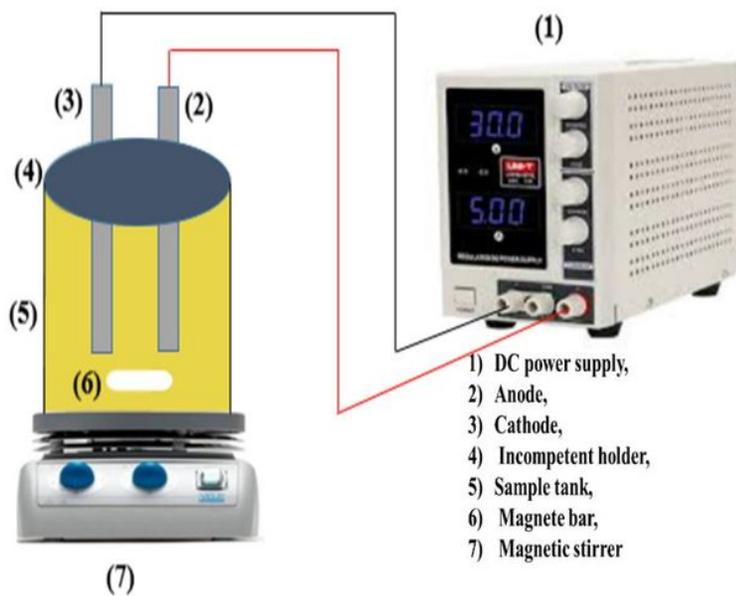


Figure 1

Schematic of the reactor used for removal of oxytetracycline by oxidation processes

**(a)**

**(b)**

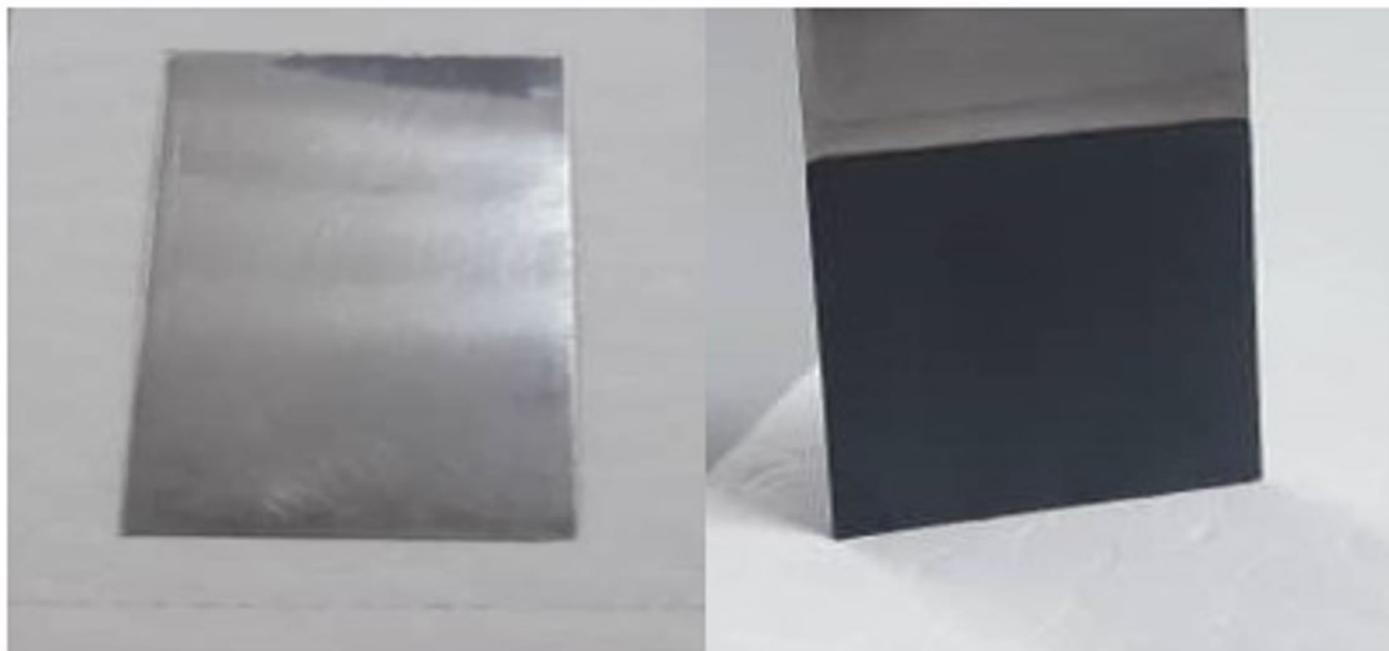
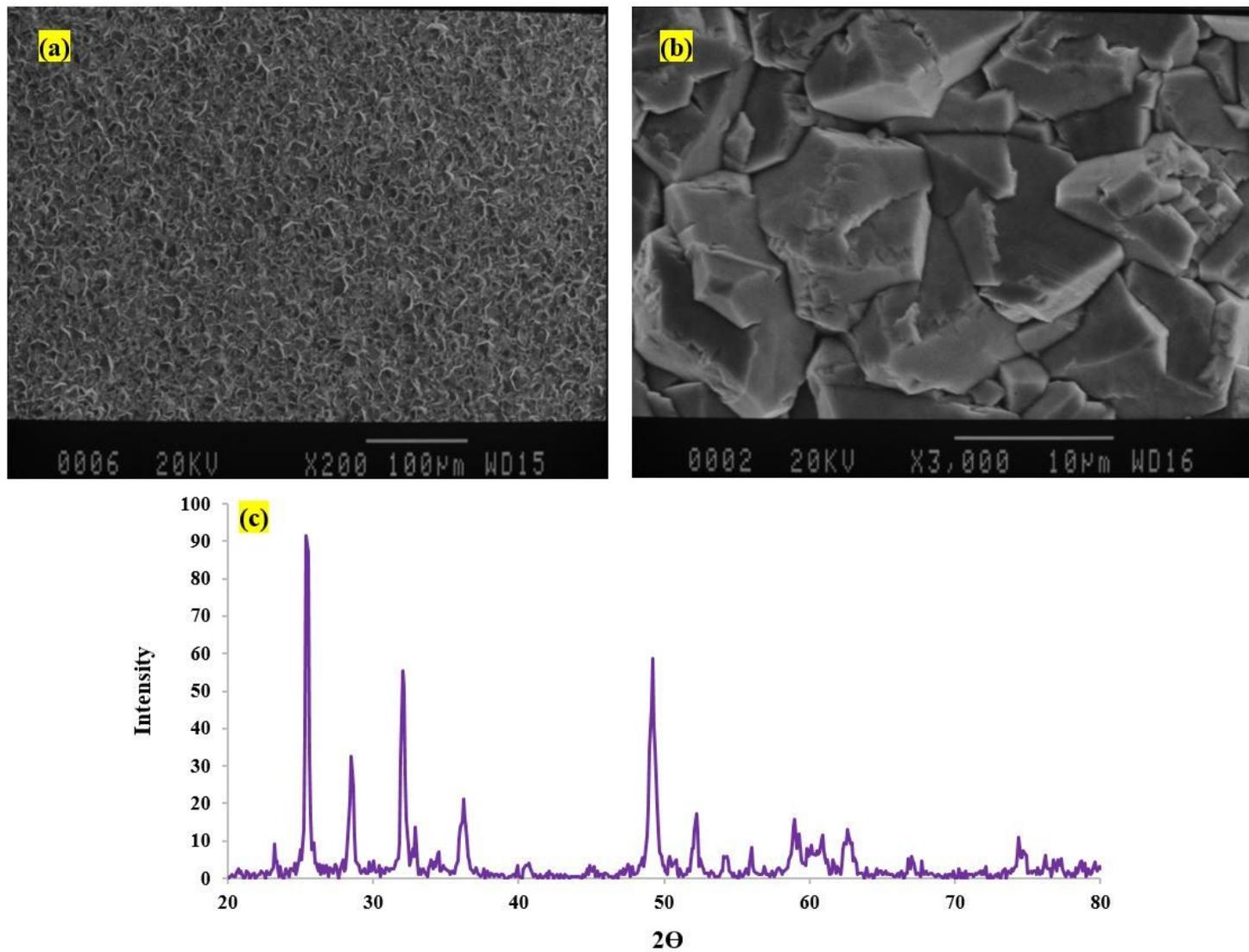


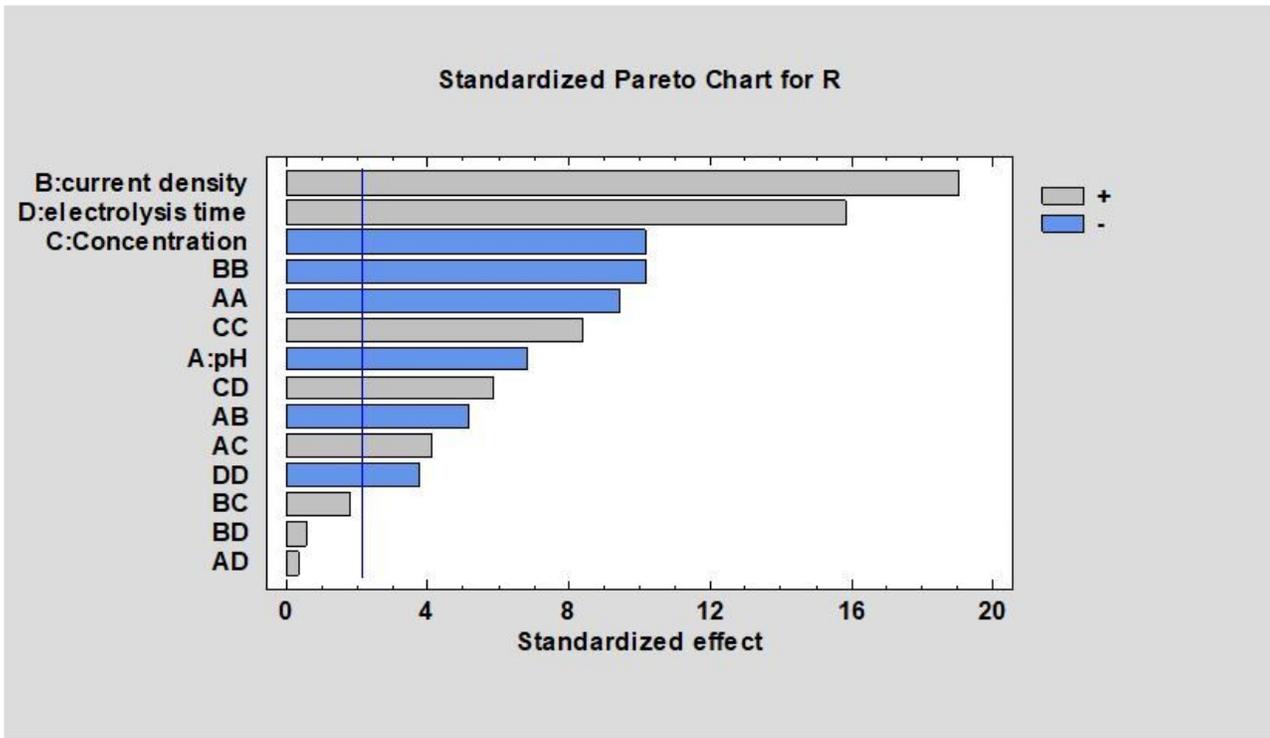
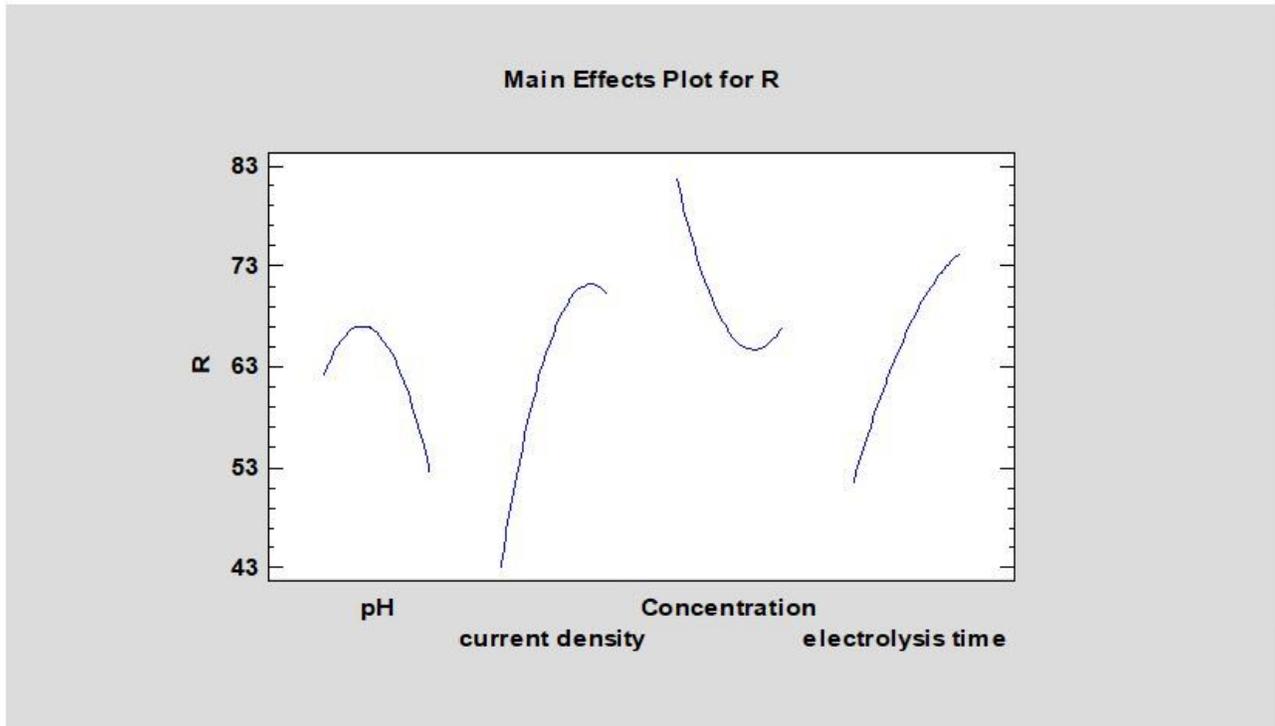
Figure 2

(a) Image of stainless steel electrode and (b) Stainless steel electrode coated with lead dioxide (SS316/PbO<sub>2</sub>) for removal of the oxytetracycline antibiotic under optimal test conditions



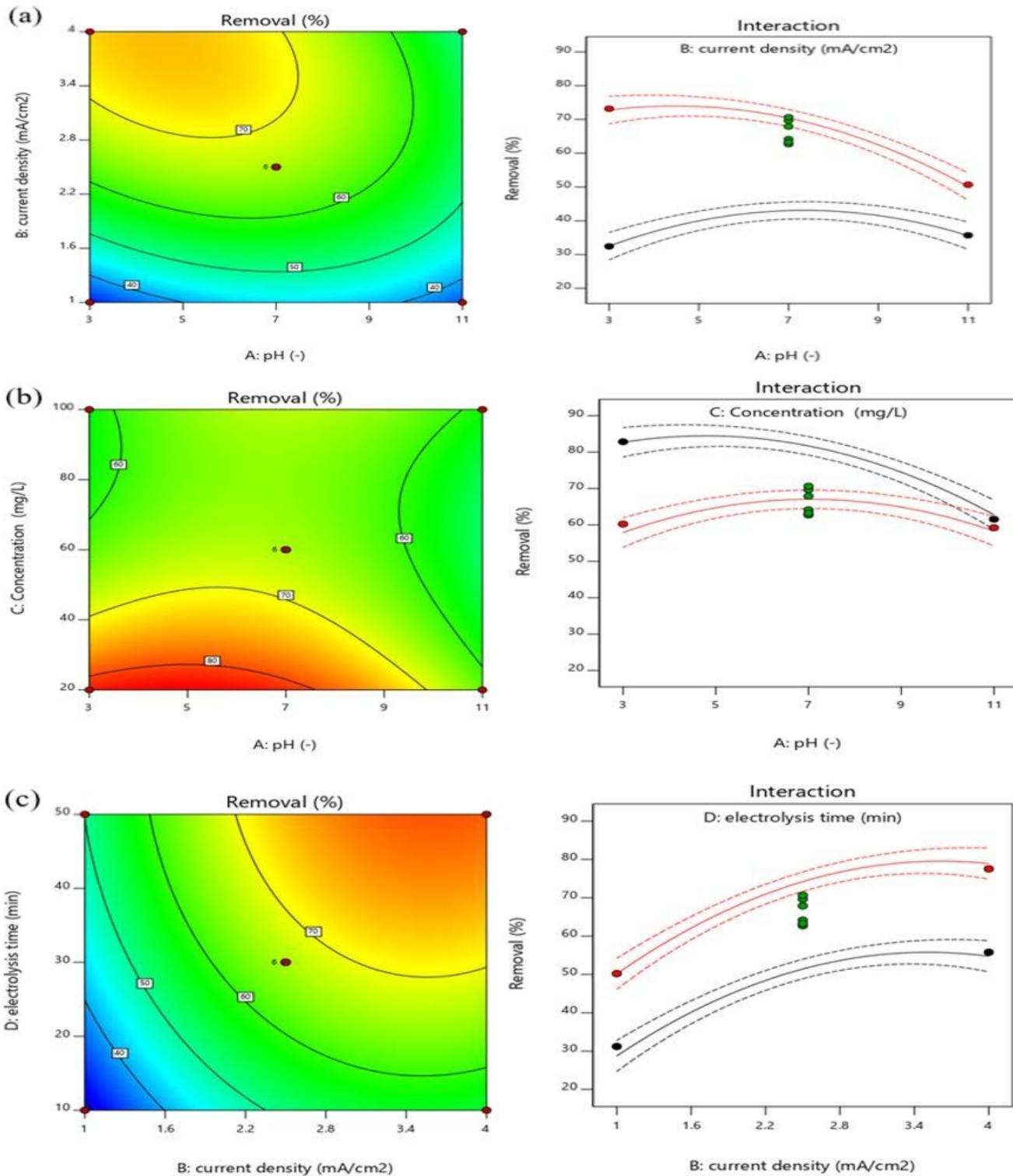
**Figure 3**

(a, b) SEM micrographs of the surface of SS316/ $\beta$ -PbO<sub>2</sub> at different magnifications; (c) XRD analysis of SS316/ $\beta$ -PbO<sub>2</sub> anode.



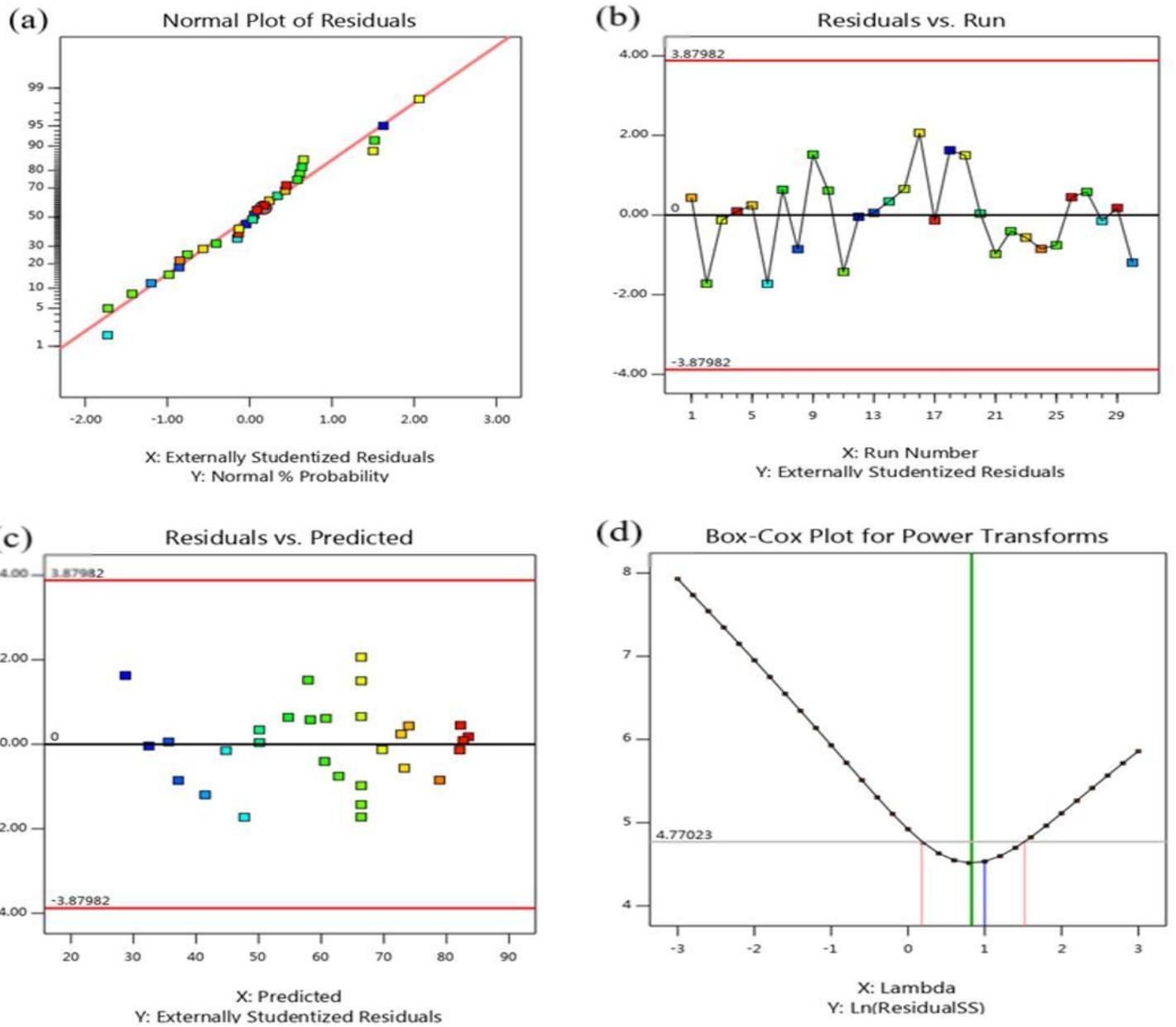
**Figure 4**

Separate effects of variables and the pareto plot for determining the effect of each term on the response (removal of Oxytetracycline by electrochemical process).



**Figure 5**

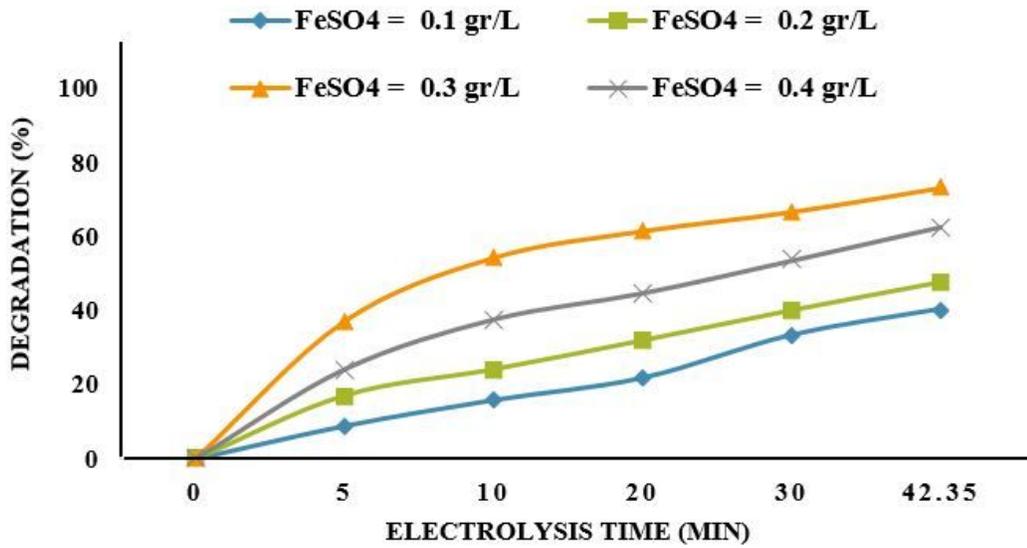
(a) Contour plots and interaction effects of pH and current density (mA/cm<sup>2</sup>), (b) Contour plots and interaction effects of pH and initial concentration of Oxytetracycline (mg/L), (c) Contour plots and interaction effects of current density (mA/cm<sup>2</sup>) and electrolysis time (min).



**Figure 6**

The diagnostic plots for validation of obtained model: (a) Normal probability distribution of residuals, (b) Externally studentized residuals versus run order, (c) Residuals versus predicted and (d) Box-Cox Plot.

(a)



The changes of FeSo4 amounts in optimal condition via Fenton process  
(pH = 3.53, concentration = 20 mg/L)

(b)

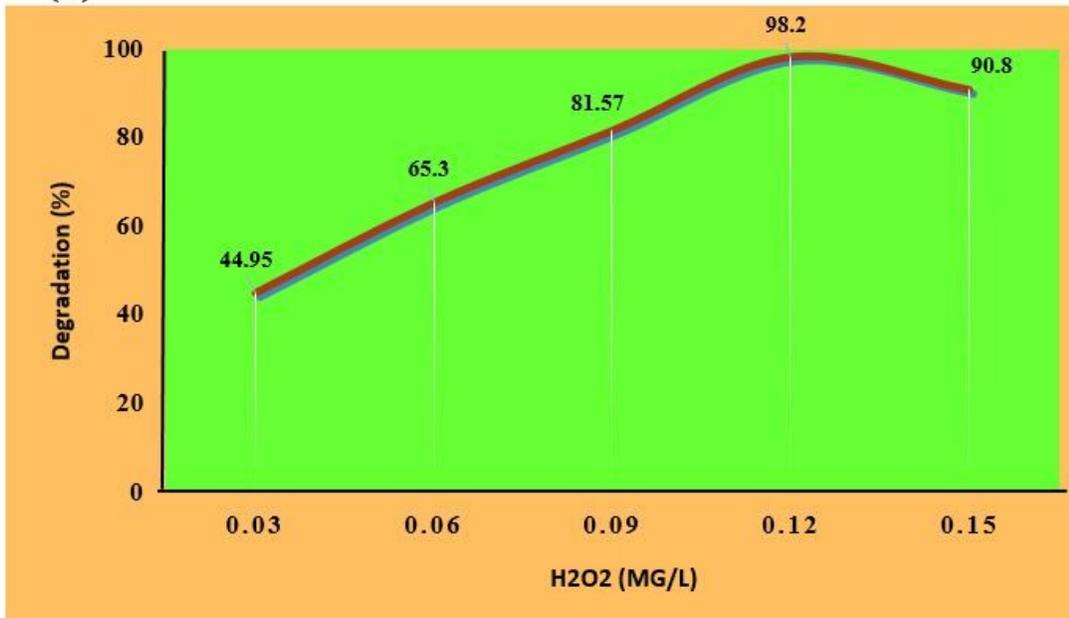
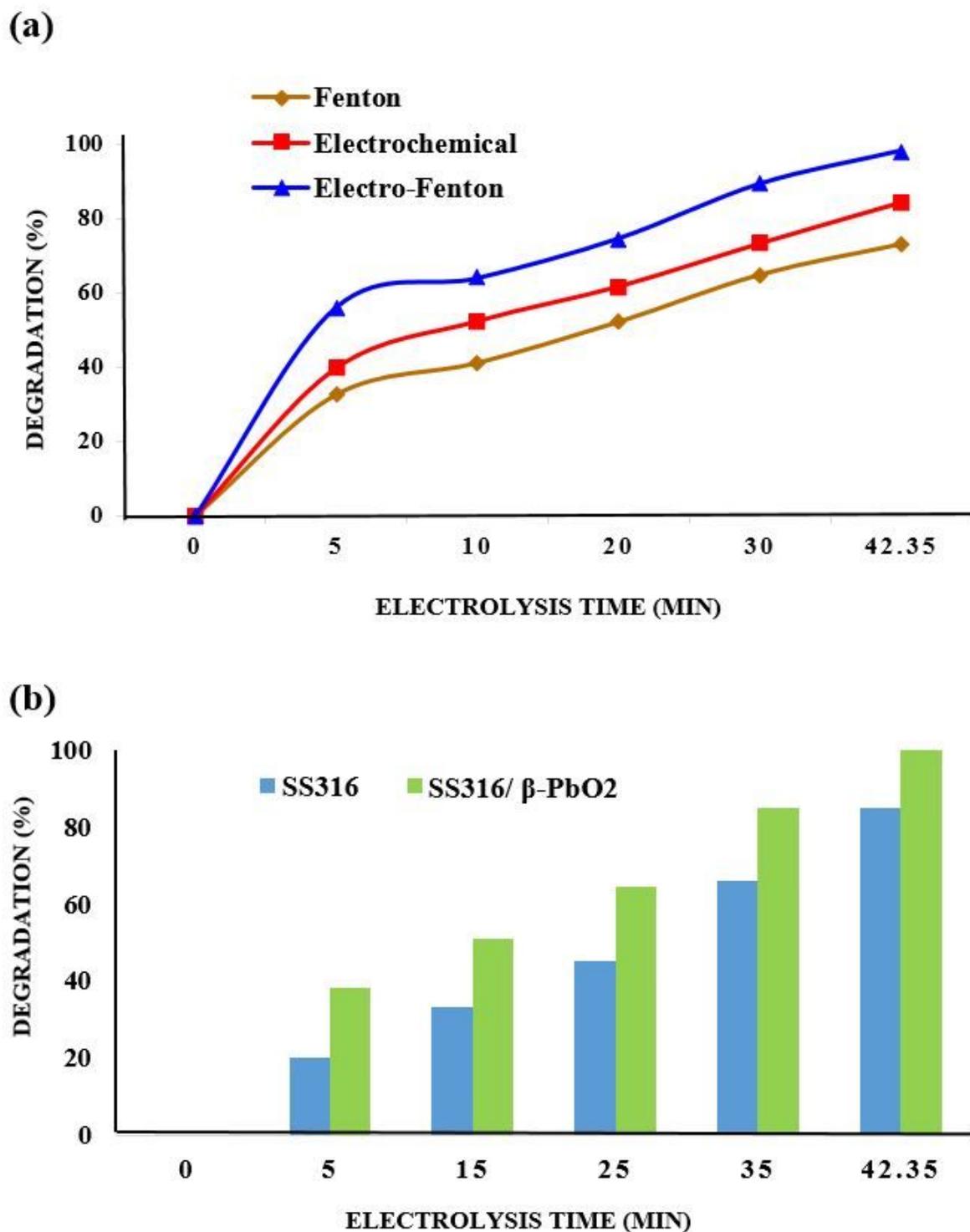


Figure 7

(a) Influence of FeSo4 amounts via Fenton process and (b) H2O2 values in elimination of Oxytetracycline in electro-Fenton process under optimum condition (pH= 3.53, current density = 3.85 mA/cm<sup>2</sup>, concentration = 20 mg/L, electrolysis time = 42.35 min and FeSO<sub>4</sub> = 0.3 gr/L). (a: Optimize amount of FeSO<sub>4</sub> = 0.3 gr/L, b: Optimize value of H<sub>2</sub>O<sub>2</sub> = 0.12 mg/L)



**Figure 8**

(a) The effect of AOPs processes on the degradation of Oxytetracycline antibiotic, (b) The elimination of Oxytetracycline antibiotic by the SS316 and SS316/ $\beta$ -PbO<sub>2</sub> anodes (pH = 3.53, current density = 3.85 mA/cm<sup>2</sup>, concentration of Oxytet = 20 mg/L, electrolysis time = 42.35 min, FeSO<sub>4</sub> = 0.3 gr/L, H<sub>2</sub>O<sub>2</sub> = 0.12 mg/L).

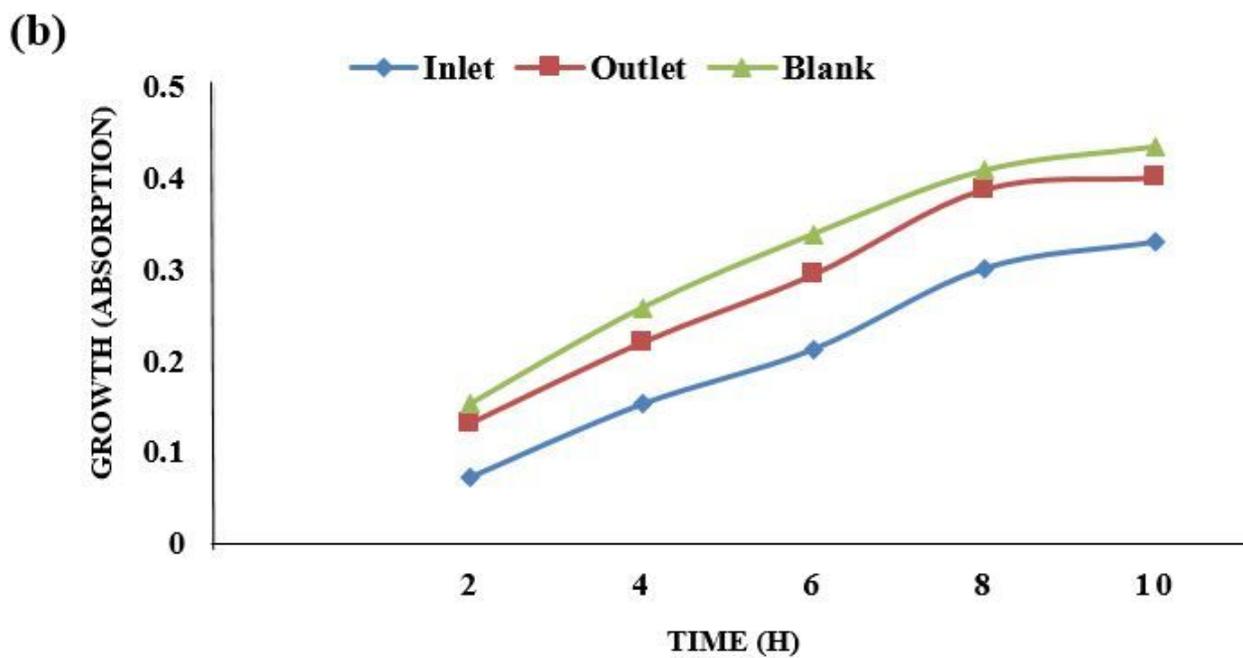
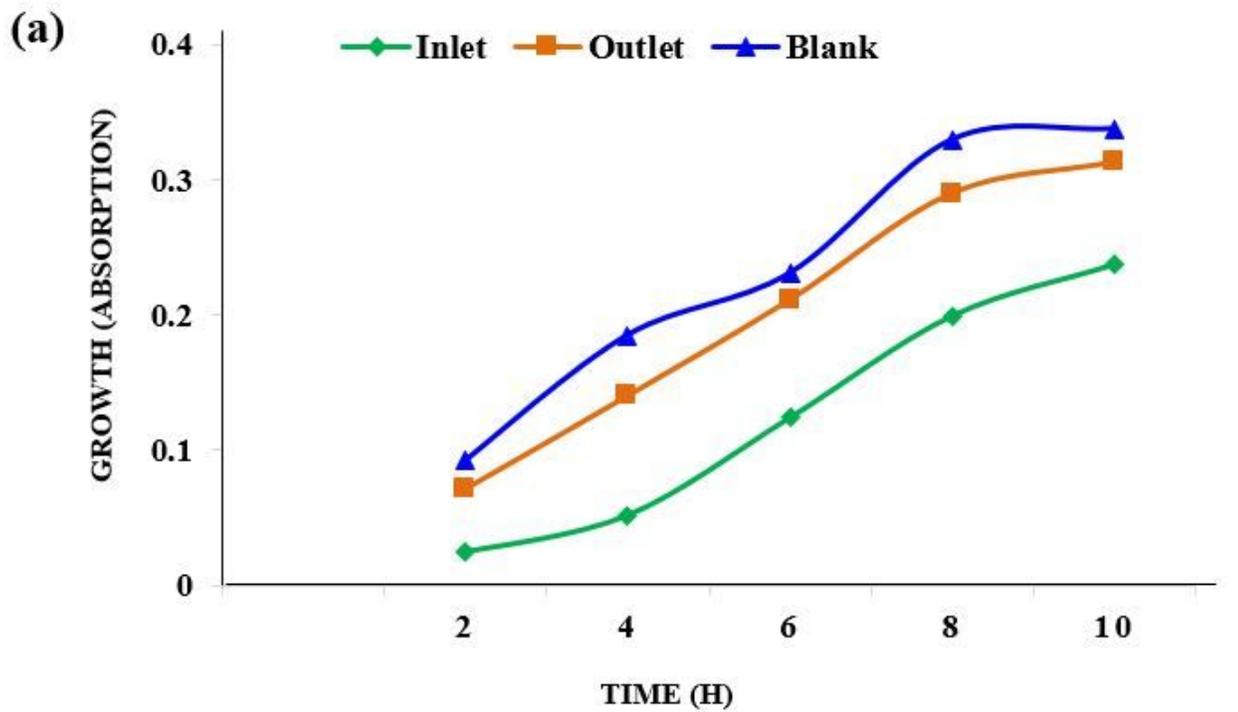


Figure 9

(a) Growth trend of *Pseudomonas aeruginosa* (gram-negative) and (b) *Staphylococcus aureus* (gram-positive) in toxicity test under optimal conditions of Electro-Fenton process.

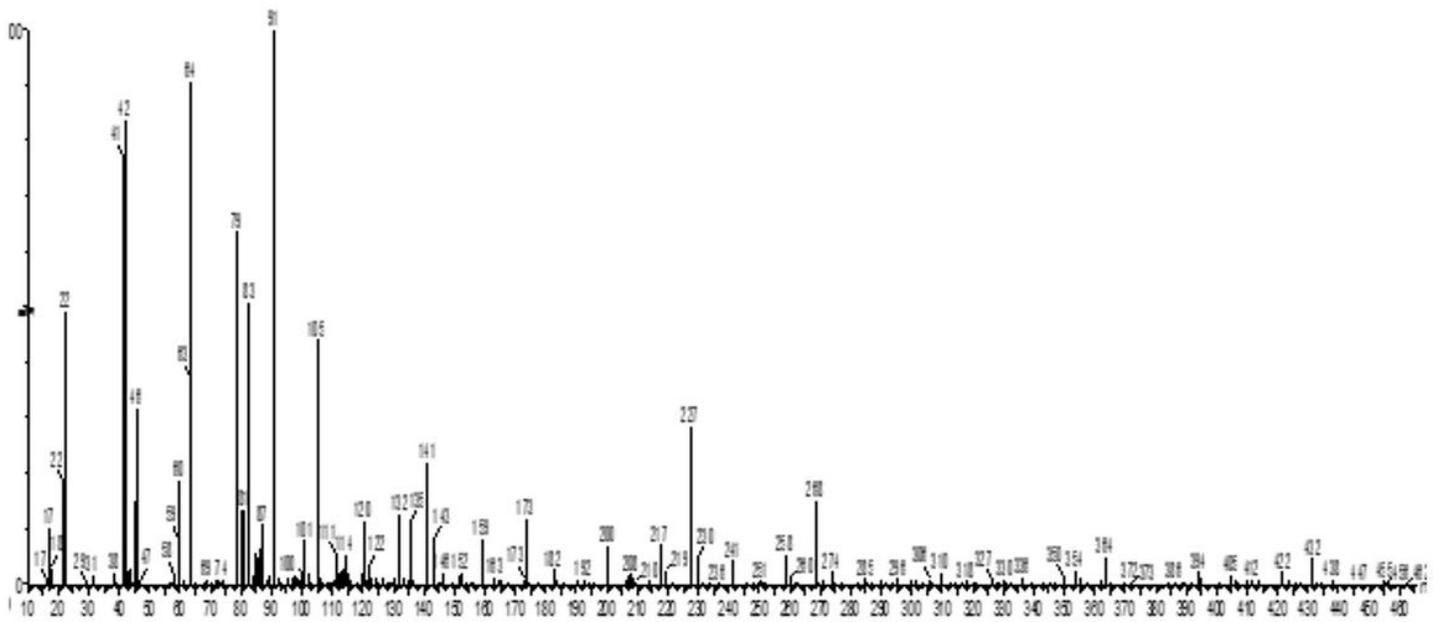
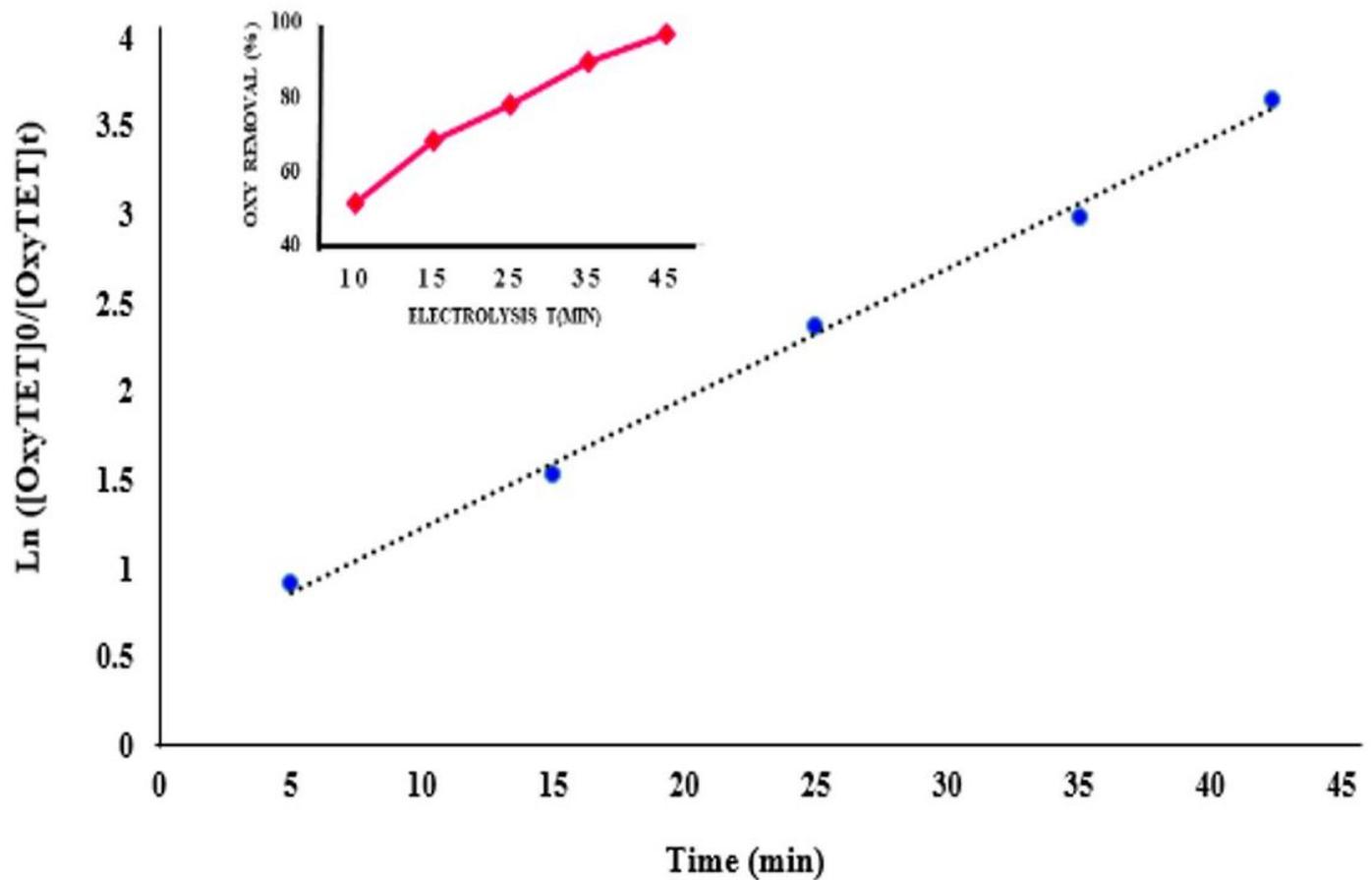


Figure 10

LC/MS chromatographs after degradation of Oxytetracycline antibiotic via Electro-Fenton process under optimum conditions.



## Figure 11

Kinetics of Oxytetracycline removal at the optimum conditions (pH = 3.53, current density = 3.85 mA/cm<sup>2</sup>, concentration of Oxytetracycline = 20 mg/L, electrolysis time = 42.35 min, FeSO<sub>4</sub> = 0.3 gr/L, H<sub>2</sub>O<sub>2</sub> = 0.12 mg/L) with pseudo- order model.

## Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [scheme1.jpg](#)