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Characteristics of the community-structure of A2O processes under different temperature conditions in plateau areas

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Abstract

Background: In this study, laboratory-scale A2O wastewater treatment was performed in Linzhi at an altitude of 3000 meters. Water temperatures were maintained at four operating conditions (25°C, 20°C, 15°C, and 10°C). Sludge in the aerobic tank was assessed by 16SrRNA sequencing and composition analysis. The Phylum, Class, Order, Family, Genus, and Species of the sludge were also confirmed.

Results: The bacteria within the sludge showed significant differences at varying temperatures ($P < 0.05$). A significant correlation between the bacteria numbers in anaerobic tanks and anoxic tanks also occurred. Indicators of community richness, community uniformity, community diversity and other areas showed differences. Significant differences in bacteria abundance were also observed and differed to those of previously reported superior community categories and proportions. T-tests were used to identify temperature-sensitive community at each level. Correlation analysis of environmental factors and colony structure further confirmed the association between temperature and colony structure ($P < 0.05$). The removal rates of TP, TN, NH₃-N, and COD were all affected by the sample community structure. The variety of colony structure include nitrifying bacteria, denitrifying bacteria, phosphorus accumulating bacteria and other bacterial differed, but their proportions were relatively low. Heatmaps were used to identify species sensitive to temperature, TP, TN, NH₃-N, COD at the species level.

Conclusions: Amongst the common dominant bacteria at the species level, Bacteroidetes_bacterium_OLB10 was significantly related to temperature. Other common dominant bacteria that were related to temperature metagenome_g_Gordonia, unclassified_f_Burkholderiaceae, uncultured_bacterium_g_norank_f_Saprosiraceae.

Keywords: plateau area; temperature; 16SrRNA sequencing; anaerobic-anoxic-aerobic (A2O) processes; community structure.

1. Background

As the third pole, the Qinghai-Tibet Plateau has an average altitude of more than 4000 meters. The high altitude creates unique environmental conditions of high cold and low air pressure. In recent years, Tibet has amassed a large number of urban sewage treatment plants, but their operating conditions are unsatisfactory, mainly due to the poor effects of nitrogen and phosphorus removal because of the unique environment conditions (Zong et al., 2019).

According to measurements, the summer water temperature in Changdu City at an altitude of 3500 meters is 12.2 ~ 18.5°C (Zhai et al., 2012) compared to the water temperature of Linzhi City at an altitude of 3000 meters is -1 ~ 15°C (Chen et al., 2018). Studies have shown that the water temperature in Tibet is low, which impacts the microbial structure of the sewage treatment system and the effects of nitrogen and phosphorus removal (Zong et al., 2019). Tian et al. (Tian et al., 2015) identified 51,880 genus microorganisms from the activated sludge ecosystem of municipal sewage

treatment plants using the A2O process, including *Proteus*, *Bacteroides*, *Nitrospira* and *Green Campylobacter* as the dominant bacteria. In view of the large differences in microbial colony structures at different water temperatures, numerous studies on water temperature and microbial structure have been performed. Using biological aerated filters, Dou and colleagues (Dou et al., 2016) reported that when the temperature was lower than 18 °C (low temperature), the removal rate of COD and NH₃-N of the biological aerated filters were less than 60%. The structure of the flora in the tank was simple, and the density of the flora decreased. The activity and quantity of enriched phosphorus accumulating bacteria showed a decreasing trend at increasing temperatures, but when the temperature decreased, the activity of phosphorus accumulating bacteria *Acinetobacter* enhanced its phosphorus removal effect (Fang et al., 2011). Ma and coworkers (Ma et al., 2020) assessed induced temperature periodic changes. They found that the activity and relative abundance of ammonia oxidizing bacteria, nitrite oxidizing bacteria, and anammox bacteria, were dominated by *Candidatus Brocadia* at low temperatures, which changed following periodic temperature shocks. The formation of dissolved organic nitrogen was also influenced by both microbial activity and microbial community structure (Liao et al., 2019). The performance and stabilization of biological wastewater treatment systems are also closely related to the microbial community structure, variations in which are driven by temperature (Chen et al., 2017). Decreased temperatures result in significant reductions in microbiome diversity, and the alpha diversity of the active community (Paul et al., 2020). Upon analysis of the capacity of nitrogen removal and the spatial distribution of microbial communities at low temperatures, it was shown that low temperatures inhibit nitrogen removal (Paul et al., 2020). It is therefore clear that temperature changes impact microbial structure in wastewater treatment processes and the quality of wastewater treatment water.

Microbial composition and changes in A2O processes under different temperature conditions are typical characteristics of the community structure. This must be explored to fully understand microbial response mechanisms under the influence of plateau environmental factors, and provides a theoretical system for sewage biological treatment systems affected by plateau environmental factors. In this study, we selected the A2O system as a typical sewage treatment process to analyze the influence of temperature conditions. We assessed community structure characteristics and their alterations in anoxic and aerobic regions to explore the microbial response mechanisms influenced by plateau environmental factors.

2. Methods

The A2O system was selected as a typical sewage treatment process. Microbial structure characteristics were explored to investigate the influence of low temperature at a range of environmental factors. We further explored the performance of experimental scale A2O processes in response to a range of plateau environmental factors..

2.1 Experimental setup

Laboratory-scale A2O sewage treatment devices were produced from Plexiglas. Each device had an effective volume of 210 L and was divided into 3 compartments; (1) an anaerobic tank; (2) an anoxic tank; (3) an aerobic tank. The volume ratio of the three regions were 35:58:117, and the effective volume of the sedimentation tank was 39 L. Both the anaerobic and anoxic tank were equipped with a stirring device at a stirring speed of 50 rpm. The aerobic tank was equipped with an aeration head for the oxygen supply. A peristaltic pump was used to control the inlet water, return sludge and nitrification liquid. To ensure constant temperatures in the test, a constant temperature circulator was used to control the water temperature. Each tank wall had a sampling port. The specific device processes are shown in Figure 1.

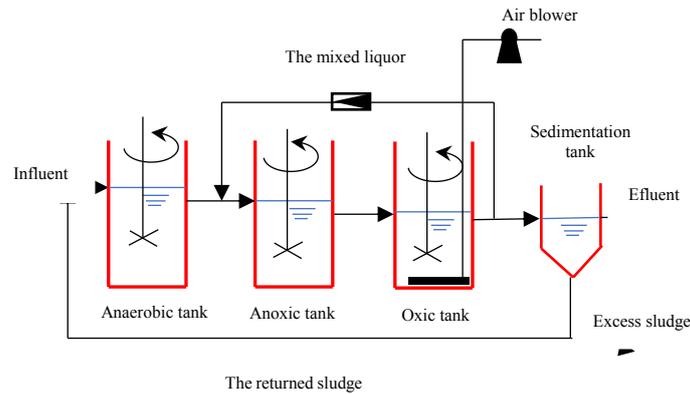


Figure 1. Schematic of Anaerobic-Anoxic-Oxic process

After 35 days of activated sludge culture, the temperature was controlled at 20.0°C, the SV₃₀ was 34%, and the MLSS was 4325 mg/L. We used urban sewage in Linzhi City as the test water, the quality status of which is shown in Table 1.

Table 1. Quality indicators of Sewage

potential of Hydrogen	Chemical Oxygen Demand (mg/L)	Total Nitrogen (mg/L)	Total Phosphorous (mg/L)
8.79-9.04	92.95 ~ 530.62	38.725 ~ 186.775	2.075 ~ 14.325

2.2 A2O Operation

A2O processes at each temperature were investigated. The designed inlet flow rate was 10.0 ± 0.1L/s. The aerobic tank had a dissolved oxygen content of 2 mg/L. The hydraulic retention time was 21.0 ± 0.2h, the anaerobic tank residence time: anoxic tank residence time: and aerobic tank residence time were 35:58:117. The reflux ratio of the mixed solution was RI=200%. The reflux ratio of the sludge was R=100%. Both the mixed solution and sludge were continuously refluxed. Changes in dissolved oxygen were achieved through altering the levels of blown aeration. Controls were set to four levels of 25 °C, 20 °C, 15 °C, and 10 °C. Sampling was performed 72 hours after the temperature reached the designed value.

3. Results and discussion

Microbiological indicators were compared using SPSS 20.0 and are expressed as the mean ± standard deviation (SD). A one-way analysis of variance (ANOVA) to assess significant differences. P < 0.05 (Simon et al., 1989) indicated significant differences between samples.

Anaerobic, anoxic and aerobic tank sludge were sampled at each temperature and 16SrRNA sequencing was performed.

3.1 Basic information

Taxonomic analysis of the domain, kingdom, phylum, class, order, family, genus and species

of each sample were assessed. Operational Taxonomic Units (OTU) were identified using the Silva database. The statistical results of specific samples are shown in Table 2.

Table 2. Sample information

Sample	Domain	Kingdom	Phylum	Class	Order	Family	Genus	Species	OTU
temp_ana_25	1	1	20	35	97	188	376	533	734
temp_ana_20	1	1	27	52	126	234	477	707	969
temp_ana_15	1	1	29	56	134	241	505	757	1067
temp_ana_10	1	1	30	54	131	239	453	664	898
temp_ano_25	1	1	24	41	103	198	413	607	849
temp_ano_20	1	1	27	47	123	235	491	712	970
temp_ano_15	1	1	29	53	131	242	493	745	1028
temp_ano_10	1	1	28	52	129	239	455	675	908
temp_Oxic_25	1	1	27	44	103	194	394	567	772
temp_Oxic_20	1	1	27	50	126	238	471	678	911
temp_Oxic_15	1	1	26	43	102	193	379	560	758
temp_Oxic_10	1	1	27	49	121	225	426	625	837

Only a single Domain and Kingdom were investigated through sequencing, and Phylum \in (20,30), Class \in (35,56), Order \in (97,134), Family \in (188,242), Genus \in (376,505), Species \in (533,757), OTU \in (734, 1067), and the corresponding genealogical values were significantly lower than those stated in the literature. At the Phylum level, values were similar to the 24 reported by Sun and colleagues (sun et al., 2019), but were lower than the value of 51 recorded by Tian and coworkers (Tian et al., 2015). At the Genus level, the values were also lower than the Tian study (Tian et al. ., 2015). The value of the test 800 was low. At the OUT level, the values (924 ~ 1363) were comparable to Wen and colleagues (Wen et al., 2015) in the SBR process, but exceeded those of Jin and coworkers (Jin et al. ., 2017; Zhang et al., 2019; Fan et al., 2018). Upon measurement of the A/O processes, the aerobic tank had low OUT values. Previous studies suggest that the temperature of the Phylum, Class, Order, and Family levels significantly influence the number of species ($P < 0.05$). Compared to previous studies, we observed large differences in the number of microbial species, particularly at the temperature of the plateau, variations in which significantly influenced the number of bacteria at the Phylum, Class, Order, and Family levels. In addition, correlation analysis revealed significant differences in the number of species of phylum, phylum, order, family, genus and species at each temperature ($P < 0.05$). The number of phylum, phylum, order, family, genus and species also differed. A significant correlation between the anaerobic tank and anoxic tank were observed ($P < 0.05$). No significant correlation between the aerobic tank and the above two reactors ($P \geq 0.05$) were observed. There were no obvious correlations between the number of genus, order, family, genus and species ($P \geq 0.05$).

3.2 Alpha diversity analysis

The Alpha diversity comprises the diversity index and the differences between index groups. The commonly used metrics of the index are chao (colony richness), shannoneven (colony uniformity), ace (colony richness), Simpson (colony diversity), and coverage (colony coverage). T tests were performed to detect significant correlations between the two groups. The diversity index of each sample are shown in Table 3.

Table 3. Alpha Diversity Statistics Table

Sample	shannoneven	ace	chao	coverage	simpson
temp_ana_25	0.686311	678.636	672.000	0.996	0.063589
temp_ana_20	0.700519	867.345	904.167	0.996	0.028344
temp_ana_15	0.643979	902.975	912.556	0.996	0.02355
temp_ana_10	0.613907	866.421	892.741	0.994	0.064387
temp_Oxic_25	0.619765	720.442	726.797	0.996	0.045108
temp_Oxic_20	0.656817	839.848	839.318	0.995	0.032669
temp_Oxic_15	0.688048	925.187	882.961	0.995	0.062754
temp_Oxic_10	0.661908	816.326	844.380	0.995	0.08789
temp_ano_25	0.710761	785.327	818.406	0.996	0.042694
temp_ano_20	0.645525	884.870	911.405	0.996	0.029929
temp_ano_15	0.626017	910.082	875.190	0.995	0.032413
temp_ano_10	0.68918	876.520	862.011	0.994	0.063279

The community coverage (Wang et al., 2019) has an average coefficient of 0.994 ~ 0.996, with all exceeding 0.990. The sample sequencing coverage was good, meaning the probability that the sample sequence not being detected was extremely low. The ace coefficient of community richness was 678.6 ~ 925.2 which was comparable to previous studies (Chen et al., 2018; Zhang et al., 2019). Ace values were 1300 ~ 2200 and 1038 ~ 1159. Low levels of community uniformity were observed with a Shannon coefficient of 0.613907 ~ 0.710761, which was lower than previously reported values (Li et al., 2014). The community diversity measured as the Simpson coefficient was 0.02355 ~ 0.08789 in previous studies (Zhang et al., 2019). The Shannon coefficient was 0.935-0.976 which was comparable to Chen and colleagues (Chen et al., 2018) who reported values of 0.835.

Upon assessment of the four operating conditions: excluding operating conditions of 10°C and 20°C, there were significant differences in the ace coefficient of community richness between each of the operating conditions ($p \leq 0.05$). There were significant differences in the number of species except for at 10°C and 20°C. The community diversity measured through the Simpson coefficient differed between 10°C and 20°C, 10°C and 15°C, and 20°C and 25°C ($p < 0.05$); Significance analysis of the community richness chao coefficient revealed significant differences between 10°C and 15°C, 20°C and 25°C, and 15°C and 25°C ($P < 0.05$). The Simpson even coefficient of community uniformity also showed significant differences between 10°C and 20°C, 10°C and 20°C, and 20°C and 25°C ($P < 0.05$). Under the four groups of working conditions, there were one or more significant differences in community richness, community diversity or community uniformity, suggesting that temperature significantly influences colony structure. Differences in community richness from the aerobic to anoxic tank, oxygen tank and anaerobic tank, and anoxic tank and anaerobic tank were observed. The community diversity varied in both the anaerobic and anoxic tank, anaerobic tank and aerobic tank, and anoxic tank and aerobic tank. Differences in community uniformity were observed upon comparison of anaerobic and anoxic, anoxic and aerobic, and anaerobic and aerobic tanks.

3.3 Community composition analysis

The RDPClassifier was used for classification statistics of the three reactors under each of the operating conditions in the order of Domain, Kingdom, Phylum, Class, Order, Family, and Genus. We further focused on the analysis of fungus function at the Genus level, and analyzed each sample. The composition of the middle population when including community with an abundance greater than 1% of different classification levels showed only a single bacteria at the domain analysis level and no rank_d__Bacteria at the Kingdom analysis level.

3.3.1 Analysis of the characteristics of colony structure at the Phylum level

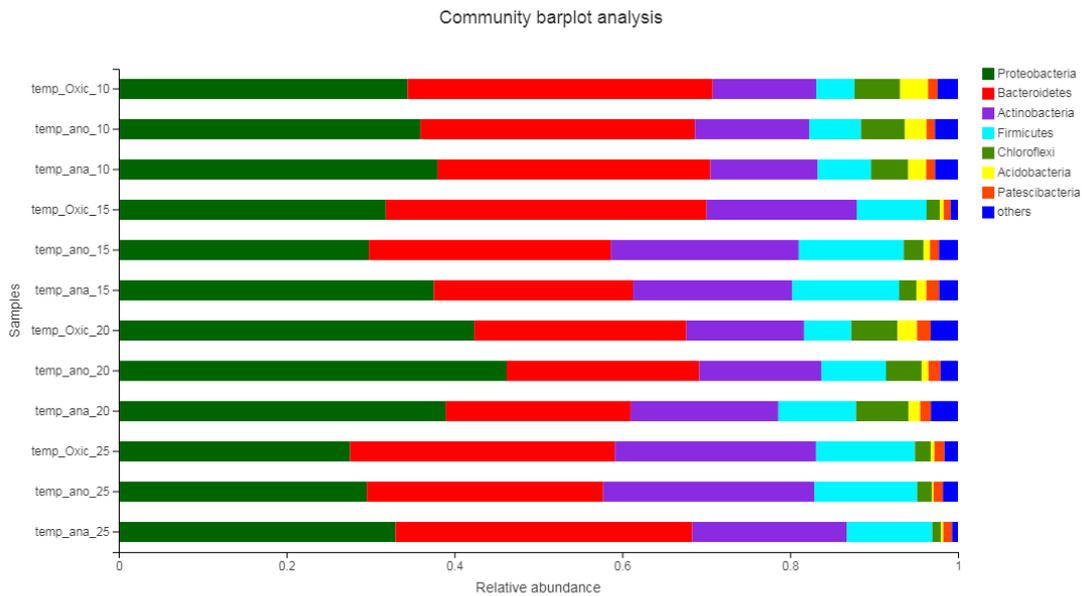


Figure 2. Community composition according to Phylum level

Figure 2 shows that shared dominant communities at the Phylum level were Proteobacteria, Bacteroidetes, Actinobacteria, Firmicutes, and Chloroflexi. A total of 31 Phylums were obtained. The abundance of five of the Phylums were 94.31-99.04% in the anaerobic and anoxic tanks. The aerobic tank showed a decreasing trend as the temperature decreased. More abundant doors and Firmicutes were observed compared to the literature (Cyzdik-Kwiatkowska et al., 2016).

The abundance of Proteobacteria (27.58% ~ 46.22) represented a relatively high proportion of organic matter-degrading bacteria. The abundance of Proteobacteria in anaerobic tanks, anoxic tanks, and aerobic tanks gradually decreased, whilst the abundance of Proteobacteria increased. The most abundant gates are consistent with previous studies (Cyzdik-Kwiatkowska et al., 2016), but the corresponding abundance was lower than previously reported (Muszyński et al., 2015). The abundance of Bacteroidetes was 21.93% ~ 38.25 %. This obligate anaerobic phylum can promote the utilization of nitrogen-containing substances and the hydrolysis of macromolecular organic substances. The aerobic tanks had a significantly higher abundance than anaerobic and anoxic tanks. Bacteroidetes were more abundant than those reported by Juretschko et al. (2002) and Kong et al. (2007). The values obtained were low; the abundance of Actinobacteria were 12.33%-25.14% in heterotrophic aerobic phylum, and its change in abundance suggested that the concentration of the anoxic tank exceeded that of the anaerobic tank and oxygen tank. The abundance of Firmicutes was 4.52% to 12.74%. These are denitrifying bacteria that are mainly present in anoxic conditions. Their abundance in the anaerobic tank was higher, and the abundance of Firmicutes was higher than that reported by Beer et al. (2006), but lower than that reported by Wong et al. (2005). The abundance of Chloroflexi was 1.04% ~ 6.26%. These are aerobic sulphur bacteria the abundance of which decreases in anoxic and aerobic tanks as temperature decreases.

The abundance of Proteobacteria, Actinobacteria, Firmicutes and Chloroflexi all differed according to temperature ($P \leq 0.05$). Of these four dominant bacteria, the Phylum number had a significant effect, but the dominant strains at the gate level showed no significant differences under different reactors.

3.3.2 Analysis of the characteristics of colony structure at the Class level

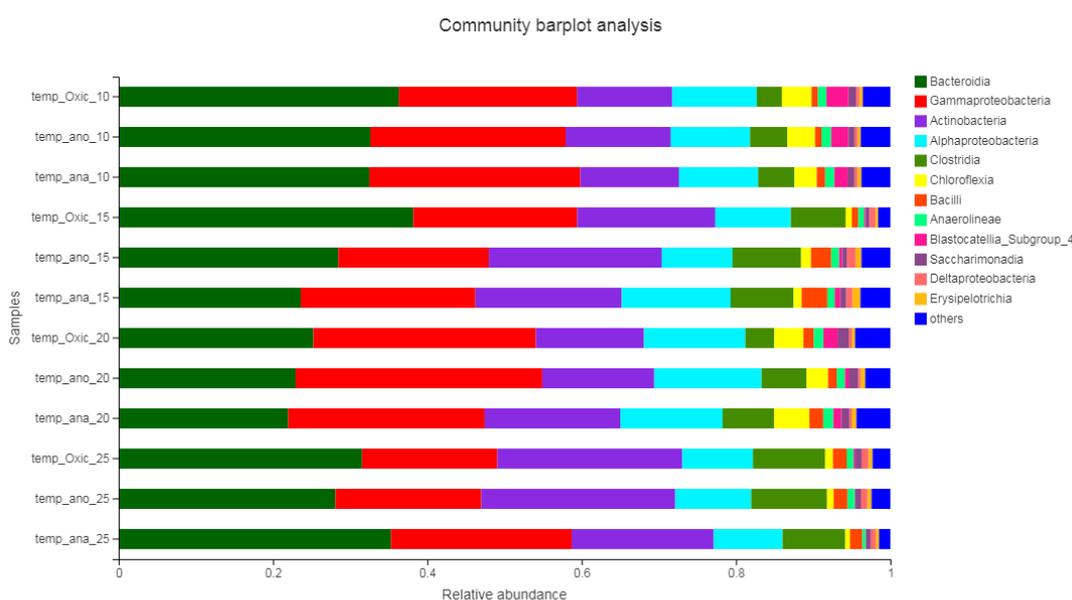


Figure 3. Community composition according to Class level.

Figure 3 shows the five classes of Bacteroidia, Gammaproteobacteria, Actinobacteria, Alphaproteobacteria, Clostridia. A total of 66 Classes were obtained. The abundance of 5 classes were 84.96-94.23% which showed a decreasing trend in the anoxic tank as temperature decreased. These findings were comparable to previous studies (Cyzdik-Kwiatkowska et al., 2016) excluding the β -proteobacteria values. These findings also differed to Kang and colleagues (Kang et al., 2019) in terms of the type and proportion of dominant strains.

Gammaproteobacteria and Alphaproteobacteria belong to the Proteobacteria phylum and represent denitrifying bacteria. The abundance of Alphaproteobacteria and Gammaproteobacteria were higher than the literature (Muszyński et al., 2015), whilst the abundance of Bataproteobacteria were higher than reported by Nielsen et al. (2012), Wong et al. (2005) and Schmid et al. (2003) Clostridia belongs to the Chloroflexi door, which varies greatly in different reactors under different working conditions. The abundance of Actinobacteria (12.33-25.14%) was higher than Zhang et al. (2018). The change interval was smaller than Chen et al. (2017) and the abundance of Bacteroidia was larger than Zhang et al. (2018).

Gammaproteobacteria, Actinobacteria, Clostridia were the three dominant bacteria that showed significant differences ($P \leq 0.05$) under different temperature conditions. In addition, Chloroflexia (0.61-4.61%) and Anaerolineae (0.42 -1.30%) showed changes in abundance at different temperatures ($P \leq 0.05$). Whilst temperature has a significant effect on the number of the above five bacterial community, there no significant differences observed as a result of the different reactors ($P > 0.05$).

3.3.3 Analysis of the characteristics of colony structure at the Order level

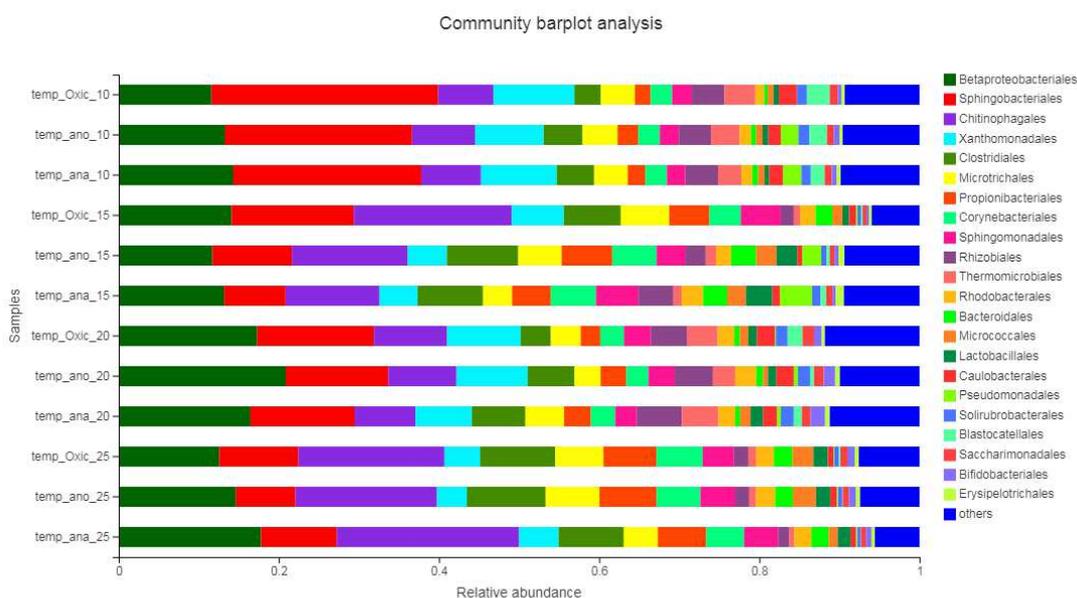


Figure 4. Community composition according to order.

Figure 4 shows that under the above working conditions, the predominant bacterium order were Betaproteobacteriales, Sphingobacteriales, Chitinophagales, Xanthomonadales, Clostridiales, Microtrichales, Propionibacteriales, Corynebacteriales, Sphingomonadales, Rhizobiales, and Rhodobacteriales. A total of 168 Orders were obtained. The abundance of 11 Orders were 71.93-86.24%. In addition, the abundance of seven orders of Thermomicrobiales, Bacteroidales, Micrococcales, Lactobacillales, Caulobacteriales, Pseudomonadales, and Solirubrobacteriales varied greatly. This accounted for 6.77-13.36% of all bacteria that were sensitive to temperature changes.

The analysis showed that: Microtrichales, Propionibacteriales, Corynebacteriales, Micrococcales, Solirubrobacteriales belong to the Actinobacteria phylum. The abundance of Microtrichales and Corynebacteriales were relatively stable, the abundance of Propionibacteriales increased at increasing temperature, and Micrococcales and Solirubrobacteriales were relatively unstable at different temperature conditions. Sphingobacteriales, Chitinophagales, and Bacteroidales belong to the Bacteroidetes order. Sphingobacteriales decreased with increasing temperature and were more abundant in the aerobic tank compared to other reactors. Chitinophagales, Bacteroidales are unstable with changes in temperature changes and showed a correlation. Clostridiales, Lactobacillales and Erysipelotrichales belong to the Firmicutes phylum; Betaproteobacteriales, Xanthomonadales, Sphingomonadales, Rhizobiales, Rhodobacteriales, Caulobacteriales, Pseudomonadales belong to the Proteobacteria phylum. Amongst them, Betaproteobacteriales were more abundant than those of other well-characterised anaerobic bacteria. The proportions of Caulobacteriales and Pseudomonadales in anaerobic and anoxic tanks were higher. Betaproteobacteriales tended to increase with increasing temperatures in each reactor and were higher in anaerobic tanks. Xanthomonadales were higher in aerobic tanks and increased with temperature Sphingomonadales and Rhizobiales did not change with temperature or according to the reactor. Rhodobacteriales showed an increasing trend with increasing temperature in each reactor. Thermomicrobiales belong to the Chloroflexi door, which was greatly affected by temperature, including at 25°C. All three reactors were lower (less than 1%), the aerobic tank was slightly higher

than 1% in the anoxic tank and the anaerobic tank at 15°C.

A total of 8 common dominant bacterial orders of Sphingobacteriales, Chitinophagales, Xanthomonadales, Clostridiales, Propionibacteriales, Corynebacteriales, Sphingomonadales, Rhizobiales showed significant differences under temperature conditions ($P \leq 0.05$), and Thermomicrobiales (0.61-4.60%), Bacteroidales (0.47-3.14%), Caulobacterales (0.63-2.25%), Solirubrobacterales (0.41-1.58%) showed an altered abundance under temperature conditions ($P \leq 0.05$). Temperature therefore had a significant effect on these four bacteria. The predominant strains at the order level showed no significant differences in each reactor ($P > 0.05$).

3.3.4 Analysis of the characteristics of colony structure at the Family level

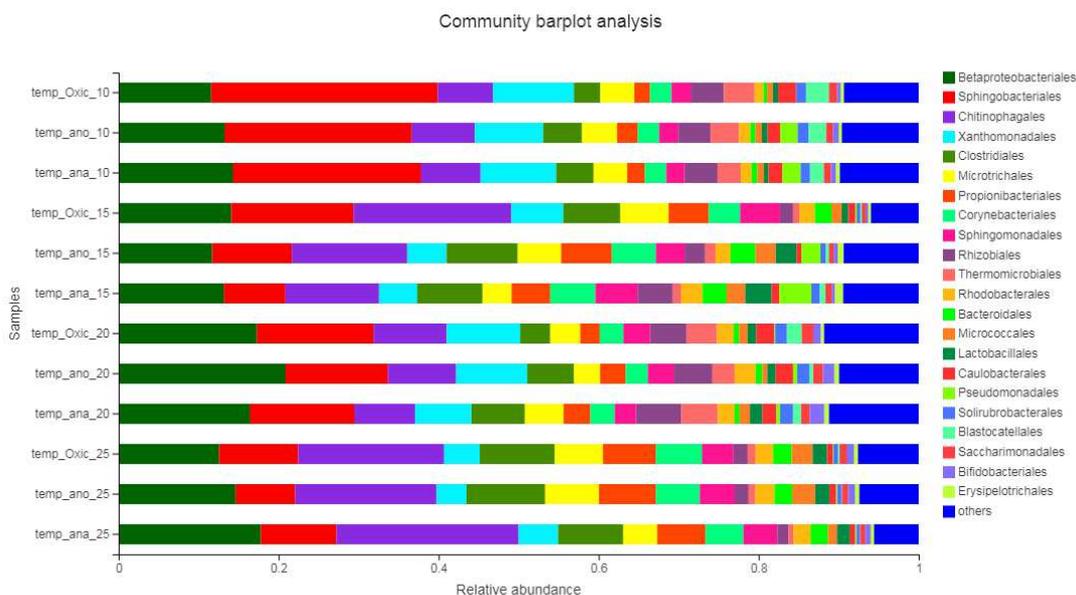


Figure 5. Community composition according to family level.

Figure 5 shows that the dominant fungi were AKYH767, Burkholderiaceae, Saprospiraceae, Microtrichaceae, Propionibacteriaceae, Sphingomonadaceae, Xanthomonadaceae, Rhodanobacteraceae, Chitinophagaceae, Nocardiaceae, Rhodobacteraceae, and Peptostreptococaceae. A total of 325 Families were obtained. The abundance of 5 Families were 54.98-72.41%, which were the dominant fungi in the plateau environment. The composition of the dominant bacteria in previous studies (Yang et al., 2017) lack Comamonadaceae, Nitrospiraceae, Sinobacteraceae, Hyphomicrobiaceae, and Chromatiaceae. It was also important to note that the abundance of nine Families of JG30-KF-CM45, Rhizobiaceae, Clostridiaceae_1, Christensenellaceae, Carnobacteriaceae, Caulobacteraceae, Hydrogenophilaceae, Microbacteriaceae, and Moraxellaceae varied greatly and were more sensitive to temperature.

In addition, Microtrichaceae, Propionibacteriaceae, Nocardiaceae, and Microbacteriaceae belong to Actinobacteria, the abundance of which increased with temperature and were higher in the aerobic tank. AKYH767, Saprospiraceae, and Chitinophagaceae belong to Bacteroidetes, and the abundance of AKYH767 increased with temperature. The aerobic tank showed higher levels of these bacteria than other reactors. The abundance of Saprospiraceae showed an increasing trend with increasing temperature and the aerobic tank was superior to other reactors. The abundance of Chitinophagaceae showed a downward trend with increasing temperature. The reactor was relatively balanced; Peptostreptococaceae, Clostridiaceae_1, Christensenellaceae, and

Carnobacteriaceae belong to the Firmicutes family. The abundance of anaerobic tanks of

Peptostreptococcaceae were significantly higher than other reactors. Clostridiaceae_1, Christensenellaceae, Carnobacteriaceae, ceca Xanthomonadaceae, Rhodanobacteraceae, Rhodobacteraceae, Rhizobiaceae, Caulobacteraceae, Hydrogenophilaceae, and Moraxellaceae belong to the Proteobacteria phylum, amongst which Burkholderiaceae and Hydrogenophilaceae belong to Betaproteobacteriales, which are polyphosphate bacteria. JG30-KF-i45 belongs to the Chloroflex and Chloroflexi.

AKYH767, Saprospiraceae, Microtrichaceae, Propionibacteriaceae, Sphingomonadaceae, Xanthomonadaceae, Rhodanobacteraceae, Chitinophagaceae, Nocardiaceae, Rhodobacteraceae were then the 10 common dominant bacteria under temperature conditions $P \leq 0.05$). JG30-KF-CM45 (0.42-3.56%), Rhizobiaceae (0.62-2.84%), Clostridiaceae_1 (0.77-2.27%), Christensenellaceae (0.26-3.03%), Caulobacteraceae (0.63-2.13%), and Hydrogenophilaceae (0.52-2.89%) showed changes in abundance according to temperature ($P \leq 0.05$). The abundance of Peptostreptococcaceae significantly differed between the reactors ($P \leq 0.05$). No other significant differences between bacteria in different reactors were observed ($P > 0.05$).

3.3.5 Analysis of the characteristics of colony structure at the Genus level

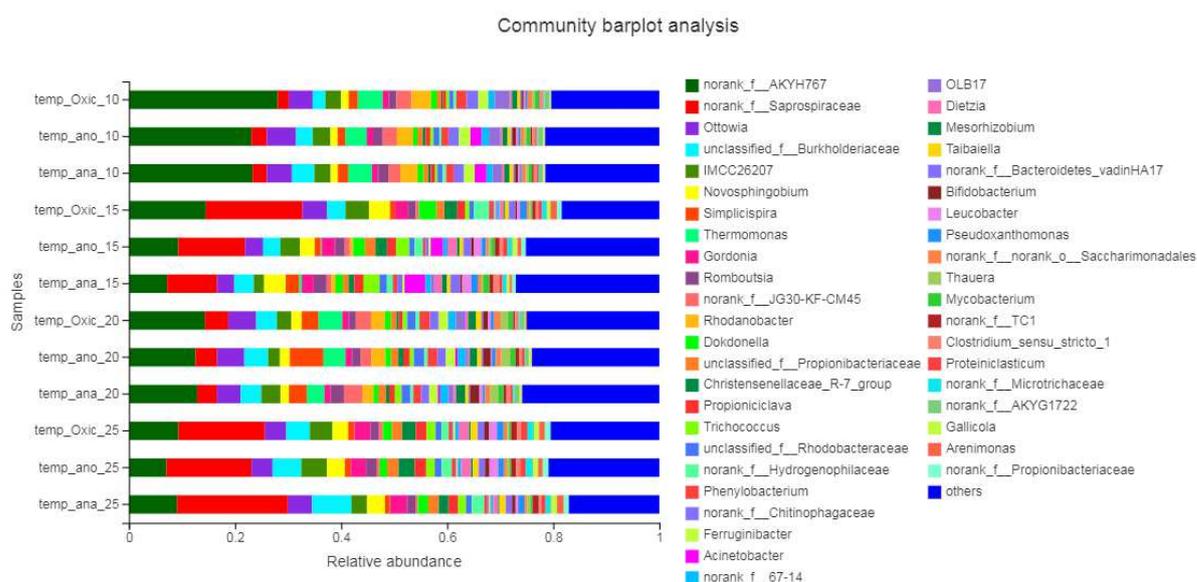


Figure 6. Community composition at the genus level.

Figure 6 shows that the common dominant bacterial genus were norank_f__AKYH767, norank_f__Saprospiraceae, Ottowia, unclassified_f__Burkholderiaceae, IMCC26207, Novosphingobium, Gordonia, Romboutsia octabacterium. A total of 719 Genus were obtained. The abundance of 8 Genus were 33.33-53.11%, which represent the dominant genus in the plateau environment. The dominant bacteria at the genus level reported in previous studies (Yang et al. 2017, Chen et al. 2017) were relatively different, excluding Pseudomonas that were similar (Tian et al. 2015, Kang et al. 2019). The dominant bacterial community differed to these studies.

Thermomonas, Simplicispira, norank_f__JG30-KF-CM45, Rhodanobacter, Dokdonella, unclassified_f__Propionibacteriaceae, Christensenellaceae_R-7_group, Propioniceclava, Trichococcus, norank_f__Hydrogenophilaceae, unclassified_f__ Large (total abundance 15.33-28.97%) were more sensitive to temperature changes.

Norank_f__AKYH767, norank_f__Saprospiraceae, Ferruginibacter,

norank_f__Chitinophagaceae belong to the Bacteroidetes phylum, and these four genera of bacteria vary greatly under temperature conditions; Romboutsia, Christensenellaceae_R-7_group, Trichococcus belong to the Firmicutes phylum. Romboutsia did not significantly change between temperature and reactor. Christensenellaceae_R-7_group increased with increasing temperature, and the abundance of Trichococcus decreased with increasing temperature. Ottowia, unclassified_f__Burkholderiaceae, Novosphingobium, Thermomonas, Simplicispira, Rhodanobacter, Dokdonella, norank_f__Hydrogenophilaceae, unclassified_f__Ottowia, unclassified_f__Burkholderiaceae, Simplicispira, norank_f__Hydrogenophilaceae belong to Betaproteobacteriales as phosphorus-accumulating bacteria. The corresponding proportion in the aerobic tank was significantly higher than other reactors. Norank_f__JG30-KF-CM45 belongs to the Chloroflexi door, which was greatly affected by the temperature conditions, the anoxic tank and the anaerobic tank.

The abundance of six common dominant genera of norank_f__AKYH767, norank_f__Saprospiraceae, Novosphingobium, Thermomonas, Simplicispira, and Gordonia significantly differed under different temperature conditions ($P \leq 0.05$). These included Norank_f__JG30-KF-CM45 (0.42-3.56%), Rhodanobacter (0.03-4.70%), Dokdonella (0.82-3.27%), unclassified_f__Propionibacteriaceae (0.79-2.11%), Christensenellaceae_R-7_group (0.25-3.01%), Propioniciclava (0.71-2.17%), Trichococcus (0.47-3.04%), norank_f__Hydrogenophilaceae (0.43-2.86%), Phenyllobacterium (0.52-1.93%), Ferruginibacter (0.28-2.16%), norank_f__Chitinophagaceae (0.17-2.22%). There were significant differences at each temperature ($P \leq 0.05$). The abundance of the genus-level dominant Genus Romboutsia significantly differed between the reactors ($P \leq 0.05$). There were no significant differences between the other bacteria in the different reactors ($P > 0.05$).

3.3.6 Analysis of the characteristics of colony structure at the Species level

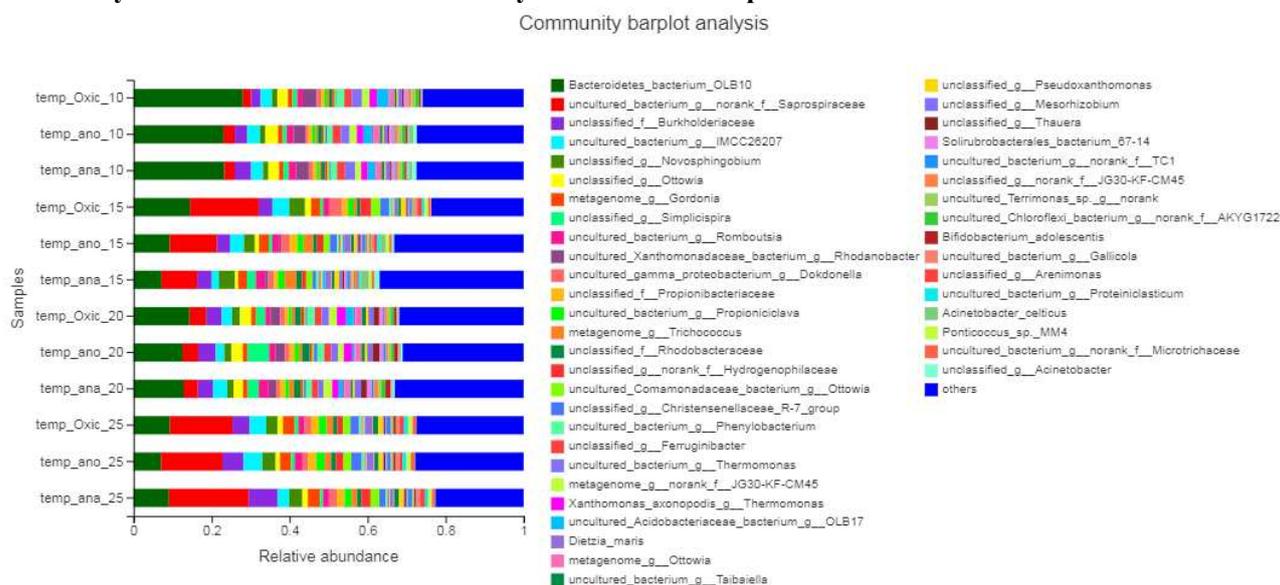


Figure 7. Community composition according to species.

From Figure 7, the common dominant bacteria genus under the above working conditions were Bacteroidetes_bacterium_OLB10, unclassified_bacterium_g_norank_f_Saprospiraceae, unclassified_f__Burkholderiaceae, unclassified_bacterium_g_IMCC26207, unclassified_g__Novosphingobium, metagenium_g__Gordordium, and bacteria. A total of 1178

Species were obtained. The abundance of 8 Species were 28.11-47.98%, which were the dominant strain in the plateau environment. *Unclassified_g_Ottowia*, *unclassified_g_Simplicispira*, *uncultured_Xanthomonadaceae_bacterium_g_Rhodanobacter*, *uncultured_gamma_proteobacterium_g_Dokdonella*, *unclassified_f_Propionibacteriaceae*, *uncultured_bacterium_g_Propioniciclava*, *metagenome_g_Trichococcus*, *unclassified_g_norank_f_Hydrogenophilaceae*, *unclassified_f_Rhodobacteraceae*, *uncultured_Comamonadaceae_bacterium_g_Ottowia*, *unclassified_g_Christensenellaceae_R-7_group*, *uncultured_bacterium_g_Phenylobacterium*, *unclassified_g_Ferruginibacter*, *uncultured_bacterium_g_Thermomonas* also showed changes in abundance. The *metagenome_g_norank_f_JG30-KF-CM45* Species were highly abundant (17.43-25.99%) and more sensitive to temperature changes.

Uncultured_bacterium_g_IMCC26207, *metagenome_g_Gordonia*, *unclassified_f_Propionibacteriaceae*, *uncultured_bacterium_g_Propioniciclava* belong to the Actinobacteria phylum. The abundance of *uncultured_bacterium_g_IMCC26207* increased as temperature increased. *Unclassified_f_Propionibacteriaceae*, *uncultured_bacterium_g_Propioniciclava* also increased at increasing temperature and were significantly superior to the other two reactors in the anoxic tank. *Bacteroidetes_bacterium_OLB10*, *uncultured_bacterium_g_norank_f_Saprospiraceae_unknown_Broderobacterium_genotype_B* Increased showed an upward trend and were most abundant in the aerobic tank. The abundance of *unclassified_g_Ferruginibacter* showed a downward trend at increasing temperatures. *Uncultured_bacterium_g_Romboutsia*, *metagenome_g_Trichococcus*, *unclassified_g_Christensenellaceae_R-7_group* belong to Firmicutes_g, *culture_rich_g-7* and decreased as temperature increased, at were highest in the anaerobic tank. The abundance of *unclassified_g_Christensenellaceae_R-7_group* increased with temperature. *Unclassified_f_Burkholderiaceae*, *unclassified_g_Novosphingobium*, *unclassified_g_Ottowia*, *unclassified_g_Simplicispira*, *uncultured_Xanthomonadaceae_bacterium_g_Rhodanobacter*, *uncultured_gamma_proteobacterium_g_Dokdonella*, *unclassified_g_norank_f_Hydrogenophilaceae*, *unclassified_f_Rhodobacteraceae*, *uncultured_Comamonadaceae_bacterium_g_Ottowia*, *uncultured_bacterium_g_Phenylobacterium*, *uncultured_bacterium_g_Thermomonas* belongs Proteobacteria door, wherein *unclassified_f_Burkholderiaceae*, *unclassified_g_Ottowia*, *unclassified_g_Simplicispira*, *unclassified_g_norank_f_Hydrogenophilaceae*, *uncultured_Comamonadaceae_bacterium_g_Ottowia* belonging to Betaproteobacteriales of PAOs; *metagenome_g_norank_f_JG30-KF-CM45*, *unclassified_g_norank_f_JG30-KF-CM45*, *uncultured_Chloroflexi_bacterium_g_norank_f_AKYG1722*, *metagenome_g_norank_f_JG30-KF-CM45* belong to Chloroflexi door and showed fluctuations at different temperatures.

Bacteroidetes_bacterium_OLB10, *uncultured_bacterium_g_norank_f_Saprospiraceae*, *unclassified_g_Novosphingobium*, *unclassified_g_Ottowia*, *metagenome_g_Gordonia* showed significant changes in abundance (5.52%), *uncultured_Xanthomonadaceae_bacterium_g_Rhodanobacter* (0.03-3.62%), *uncultured_gamma_proteobacterium_g_Dokdonella* (0.79-3.27%), *unclassified_f_Propionibacteriaceae* (0.79-2.11%), *uncultured_bacterium_g_Propioniciclava* (0.70-2.14%), *unclassified_g_norank_f_Hydrogenophilaceae* (0.43-2.86%),

uncultured_Comamonadaceae_bacterium_g__Ottowia (0.36-2.32%),
 unclassified_g__Christensenellaceae_R-7_group (0.22-2.68%),
 uncultured_bacterium_g__Phenylobacterium (0.51-1.92%),
 uncultured_bacterium_g__Thermomonas (0.00-2.67%), metagenome_g__norank_f__JG30-KF-CM45 (0.17-2.25%). These data suggest that temperature has a significant effect on all these bacteria. The abundance of the dominant bacteria uncultured_bacterium_g__Romboutsia at the species level significantly differed between the reactors ($P \leq 0.05$). There were no significant difference between other dominant bacteria in different reactors ($P > 0.05$).

3.4 Relationship between colony structure of the sewage treatment system and environmental factors

Correlation analysis of the environmental factors and colony structure were performed through RDA/CCA analysis, which carries out regression analysis on the community structure and environmental factors at different temperature to reflect the relationship between the community structure and environmental factors. The microbial classification level is Species, environment. The factors corresponding to temperature, TN, TP, $\text{NH}_3\text{-N}$ and COD were assessed. Given the purpose of the experiment design, CCA analysis was performed for the anaerobic tank, anoxic tank and aerobic tank.

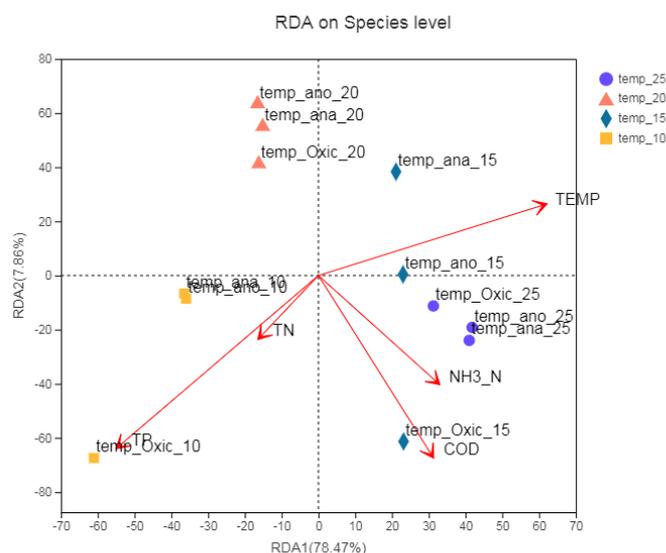


Figure 8. CCA analysis.

CCA analysis showed that the correlation coefficients r of temperature, TN, TP, $\text{NH}_3\text{-N}$, and COD were 0.9555, -0.7292, -0.7916, 0.77882, and 0.5565, respectively. Significance analysis showed that the correlation between temperature and the samples was significant ($p < 0.05$), suggesting that environmental factors show a significant relationship to sample colony structure. The removal rate of TP, TN, $\text{NH}_3\text{-N}$, and COD were affected by the sample community structure. The degree of impact was sequentially weakened. The interpretation degree of RDA1 and RDA2 were 78.47% and 7.86% respectively.

We used correlation Heatmaps to analyze the relationship between different species and environmental variables. To assess the correlation between microorganisms and environmental variables (temperature, TN, TP, $\text{NH}_3\text{-N}$, and COD) samples were classified according to the top 50 species.

Spearman Correlation Heatmap

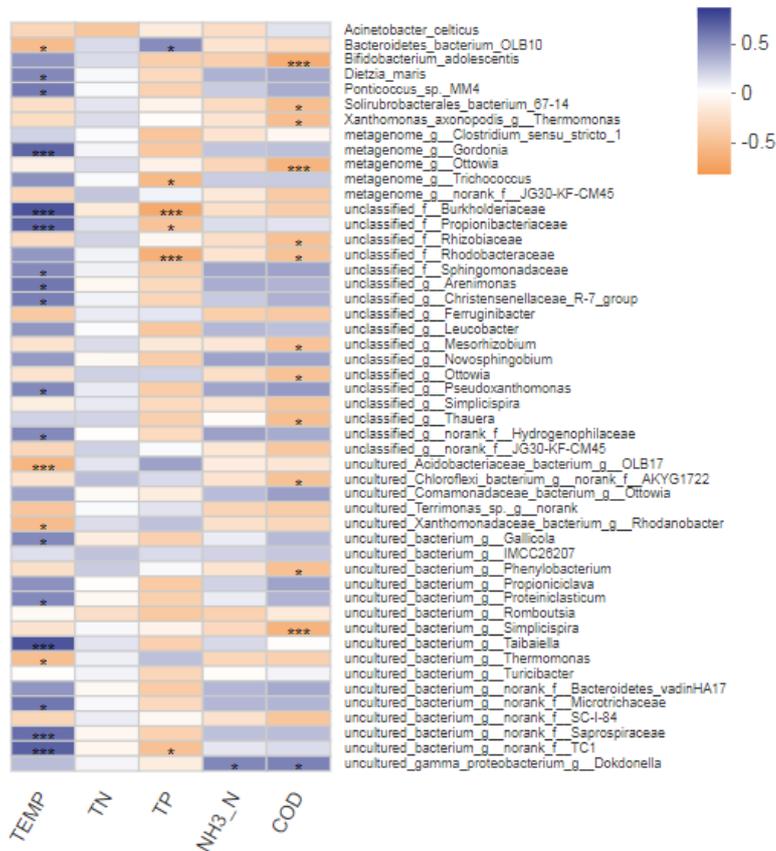


Figure 9. Correlation Heatmap.

Figure 9 shows that: 1) at the species level, temperature is significantly related to the species of Bacteroidetes_bacterium_OLB10, Dietzia_maris, Ponticoccus_sp_MM4, unclassified_f_Sphingomonadaceae, unclassified_g_Arenimonas, unclassified_g_Christensenellaceae_R-7_group, unclassified_g_Pseudoxanthomonas, unclassified_g_norank_f_Hydrogenophilaceae, uncultured_Xanthomonadaceae_bacterium_g_Rhodanobacter, uncultured_bacterium_g_Gallicola, uncultured_bacterium_g_Proteiniclasticum, uncultured_bacterium_g_Thermomonas, uncultured_bacterium_g_norank_f_Microtrichaceae. Temperature also significantly influenced related species including metagenome_g_Gordonia, unclassified_f_Burkholderiaceae, unclassified_f_Propionibacteriaceae, uncultured_Acidobacteriaceae_bacterium_g_OLB17, uncultured_bacterium_g-Taibaiella, uncultured_bacterium_g_norank_f_Saprospiraceae, and uncultured_bacterium_g_norank_f_TC1. TN correlated to the absence of certain bacterial species. TP was significantly associated with species of Bacteroidetes_bacterium_OLB10, metagenome_g_Trichococcus, unclassified_f_Propionibacteriaceae, uncultured_bacterium_g_norank_f_TC1. TP was significantly related to unclassified_f_Burkholderiaceae, unclassified_f_Rhodobacteraceae E. NH₃-N was significantly related to uncultured_gamma_proteobacterium_g_Dokdonella. COD was significantly associated with Solirubrobacterales_bacterium_67-14, Xanthomonas_axonopodis_g_Thermomonas, unclassified_f_Rhizobiaceae, unclassified_f_Rhodobacteraceae,

unclassified_g_Mesorhizobium, unclassified_g_Ottowia, unclassified_g_Thauera, uncultured_Chloroflexi_bacterium_g_norank_f_AKYG1722, uncultured_bacterium_g_Phenylobacterium, uncultured_gamma_proteobacterium_g_Dokdonella. Extremely relevant bacteria and COD species included Bifidobacterium_adolescentis, metagenome_g_Ottowia, and uncultured_bacterium_g_Simplicispira.

4. Conclusions

Using 16SrRNA gene sequencing and correlation analysis, the differences in bacterial communities in anaerobic, anoxic and aerobic tanks under four temperature conditions were analyzed based on species number, diversity, composition, and Heatmaps of the sequenced samples.

1) Sequencing analysis revealed that the number of species in the phylum, Class, order, family, genus and species level in the high-in situ samples were lower than those previously reported and varied according to the working conditions of the three reactors. There were significant differences in community number at the phylum, class, order, family, genus and species level. A significant correlation was observed between the number of community at the phylum, Class, order, family, genus and species level ($P \leq 0.05$). However, no obvious correlation between temperature and the number of phylum, class, order, family, genus and species of each tank were observed.

2) Alpha diversity analysis showed that the community richness, community uniformity, and community diversity under high in situ temperatures differed to those reported in the literature. T tests showed that, excluding 10 °C and 20 °C, significant differences exist in the number of community according to working conditions. Community diversity coefficients were also significantly different between 10 °C and 20 °C, 10 °C and 15 °C, and 20 °C and 25 °C. The community richness chao coefficient was significantly different between the three groups of operating conditions at 10°C and 15°C, 20°C and 25°C, and 15°C and 25°C. The Simpson even coefficient of community uniformity showed that at 10°C, there were significant differences between the three groups of operating conditions at 20 °C, 10 °C and 20 °C, 20 °C and 25 °C. Significance analysis of the three reactors showed no significant differences in the community numbers in the different reactors.

3) We assessed the community composition of the three reactors under different temperature conditions according to Phylum, Class, Order, Family, Genus, and species. The common dominant community at each level were analyzed and their response to temperature were assessed.

4) Using the CCA chart to further verify that temperature is an important factor that influences the composition of the colony structure, and using the correlation heatmap to analyze the species that are significantly related to temperature, our analysis showed that the abundance of 13 species were related to temperature. In total, 7 species showed a significant correlation; TN significantly correlated were not observed; 4 species correlated with TP, 2 of which showed a high correlation; 1 species significantly correlated with NH₃-N; a single species significantly correlated with COD.

Based on these analysis, the A2O species structure in high-in situ correlated with temperature in terms of species number, Alpha diversity, and dominant species composition. The dominant bacteria that were significantly related to temperature at the species level were Bacteroidetes_bacterium_OLB10. The dominant bacteria with an extremely significant correlation were metagenome_g_Gordonia, unclassified_f_Burkholderiaceae, uncultured_bacterium_g_norank_f_Saprospiraceae. The dominant bacteria with significant TP removal rates were Bacteroidetes_bacterium_OLB10. The dominant bacteria with extremely

significant TP removal rates were unclassified_fae. bacteria.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Written informed consent for publication was obtained from all participants..

Availability of data and materials

Due to the confidentiality of the experimental data and the experimental research is in the basic mechanism research stage, the experimental data will not be disclosed for the time being. The data set generated and/or analyzed during the current research period cannot be publicly obtained, but can be obtained from the corresponding author according to reasonable requirements Obtained there. The data in this article can be used after obtaining the consent of all authors. The email address for obtaining data: 1916619203@qq.com.

Competing interests

The authors declare that they have no competing interests.

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Authors' contributions

ZONG Yongchen and LU Guanghua conceived and designed this study. LI Yuanwei, HUANG Decai and HAO Kaiyue conducted experiments. ZONG Yongchen analyzed the data and wrote the manuscript. All authors reviewed the manuscript. The authors read and approved the final manuscript.

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Figures

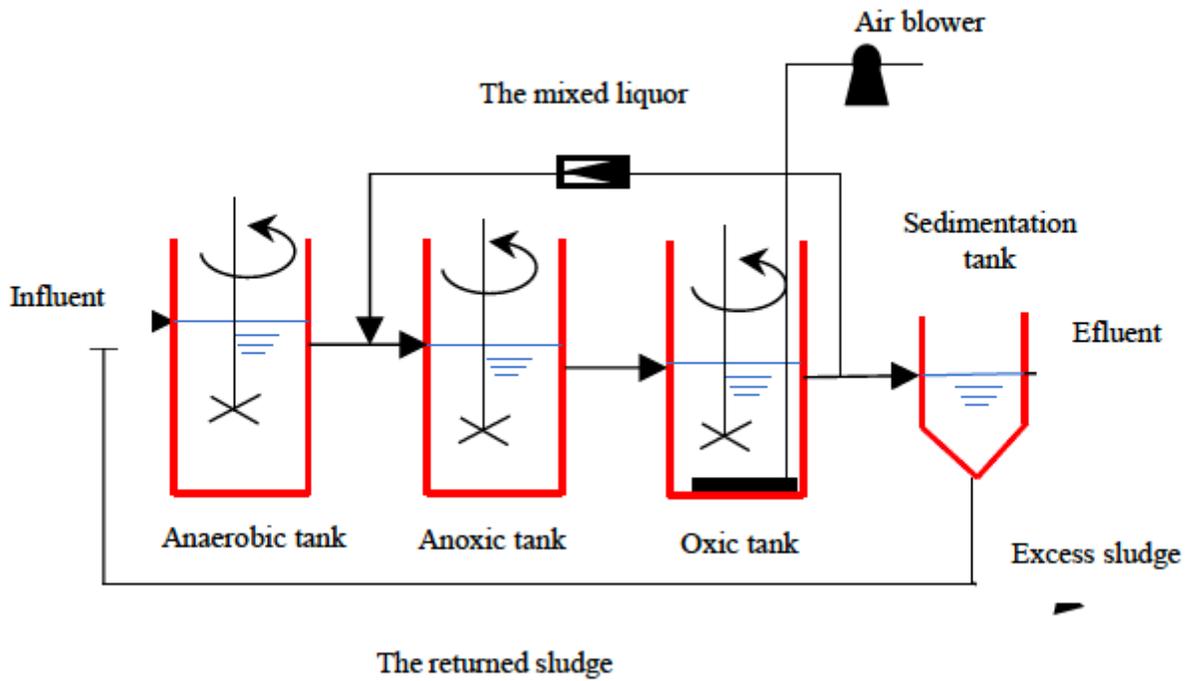


Figure 1

Schematic of Anaerobic-Anoxic-Oxic process

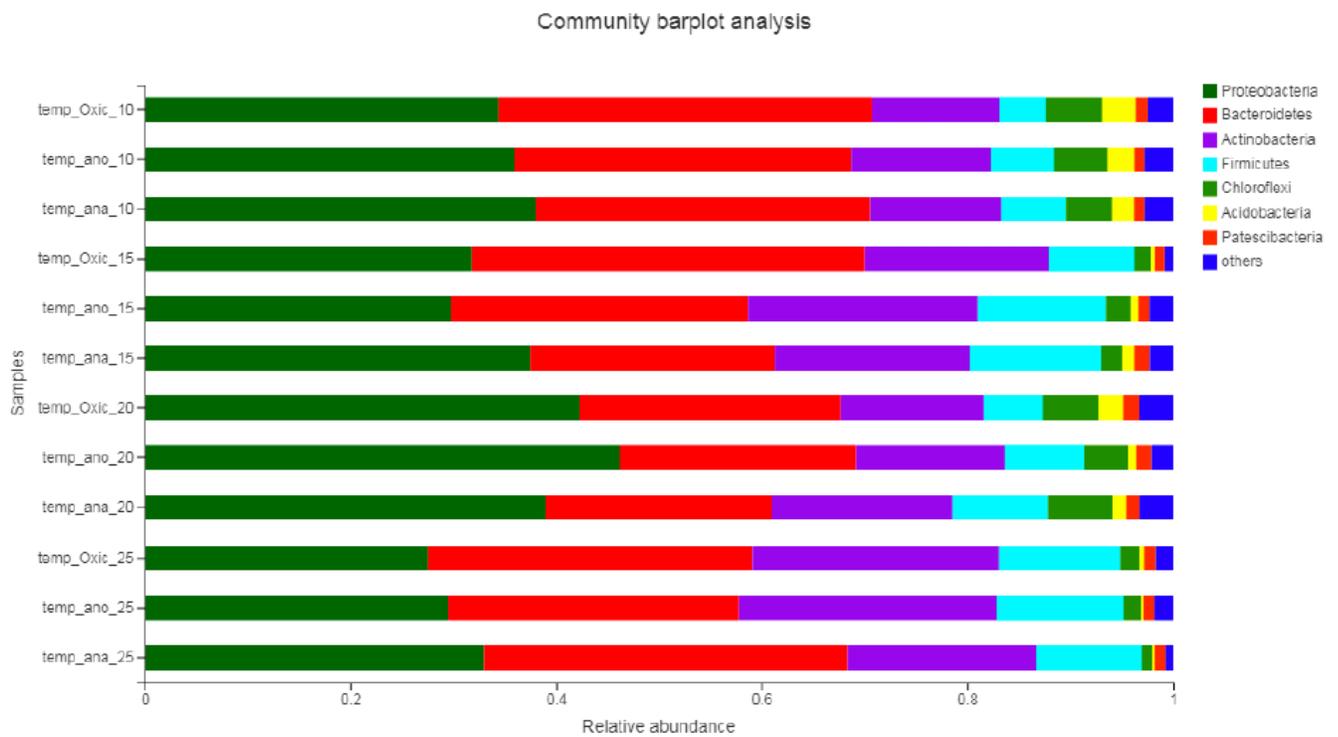


Figure 2

Community composition according to Phylum level

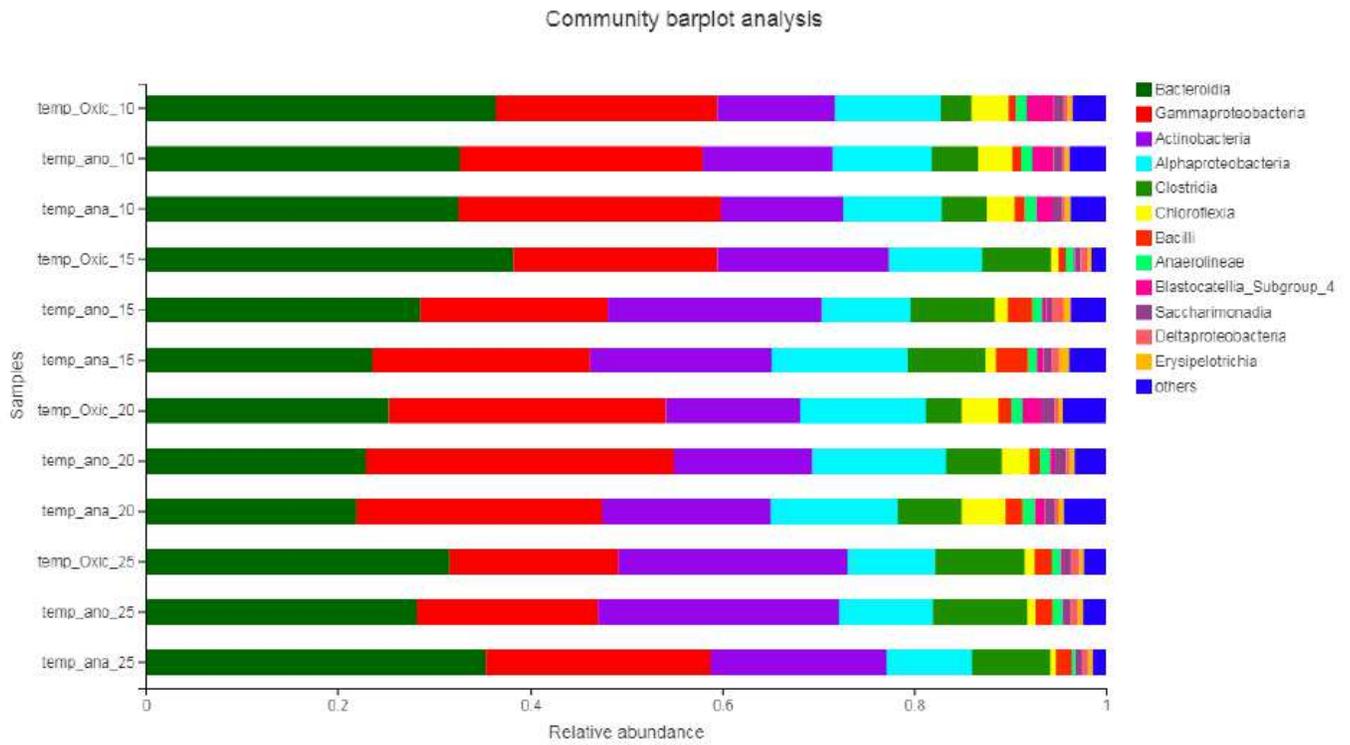


Figure 3

Community composition according to Class level.

Community barplot analysis

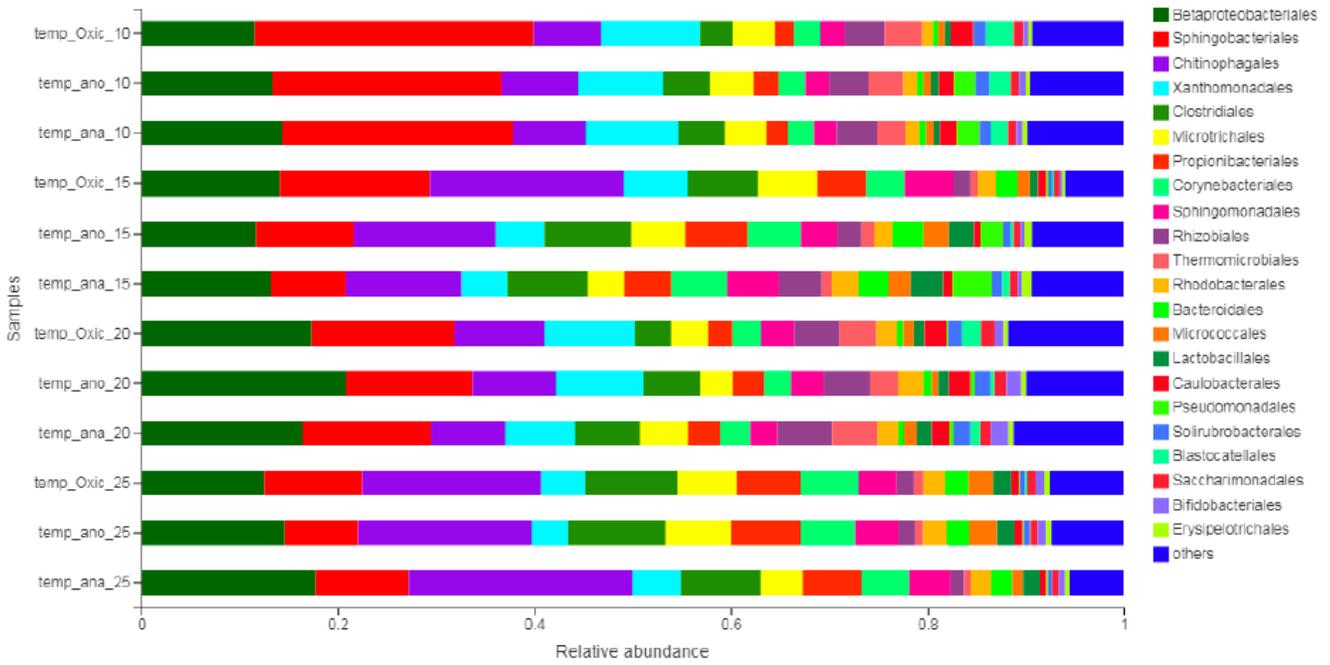


Figure 4

Community composition according to order.

Community barplot analysis

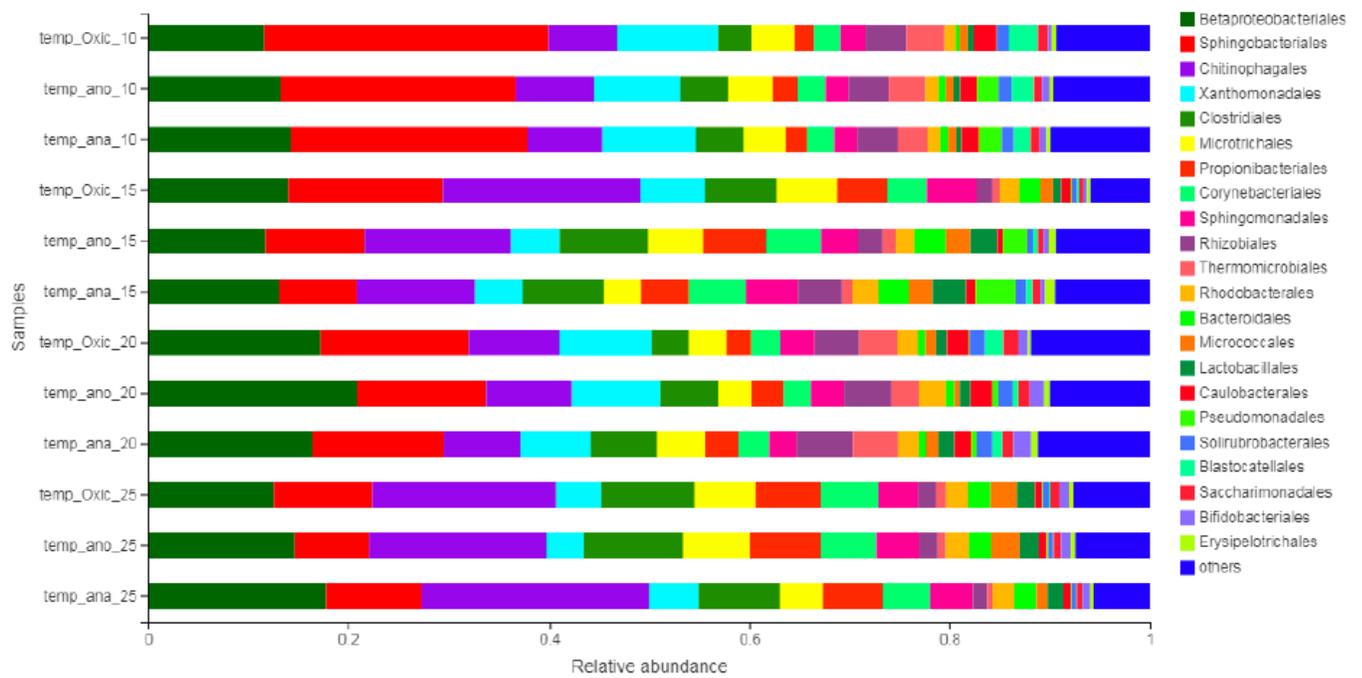


Figure 5

Community composition according to family level.

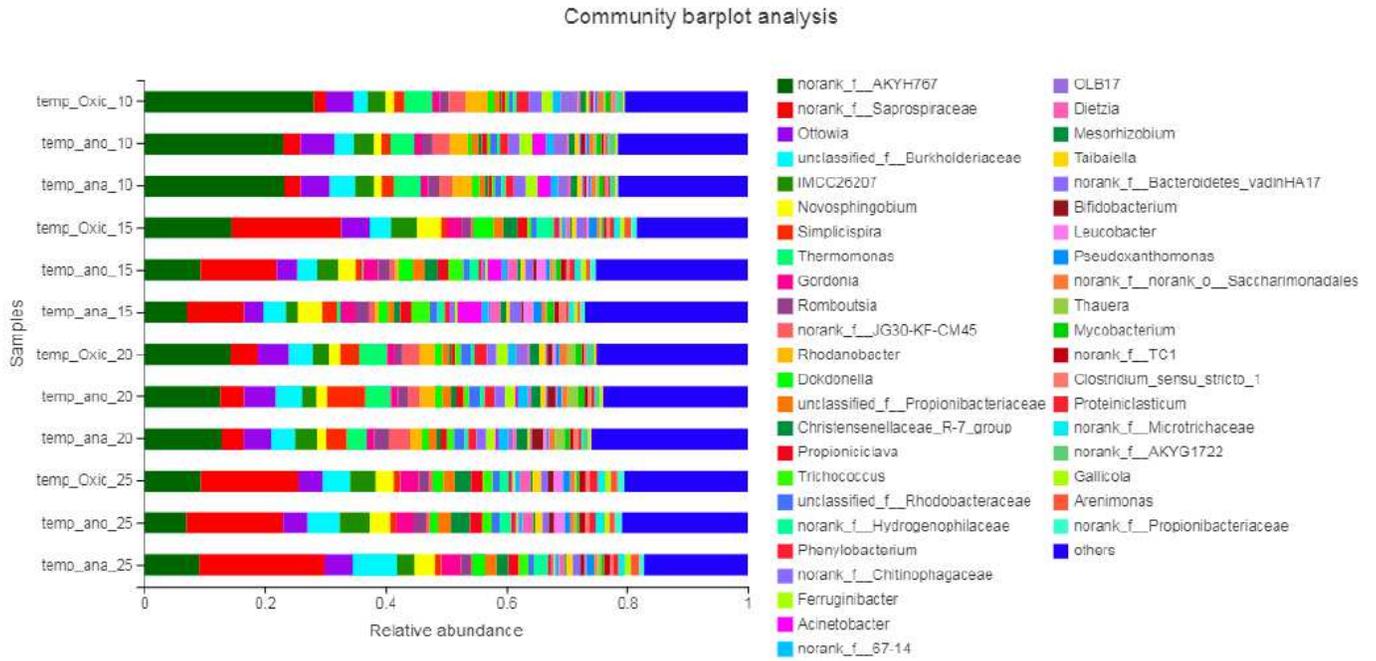


Figure 6

Community composition at the genus level.

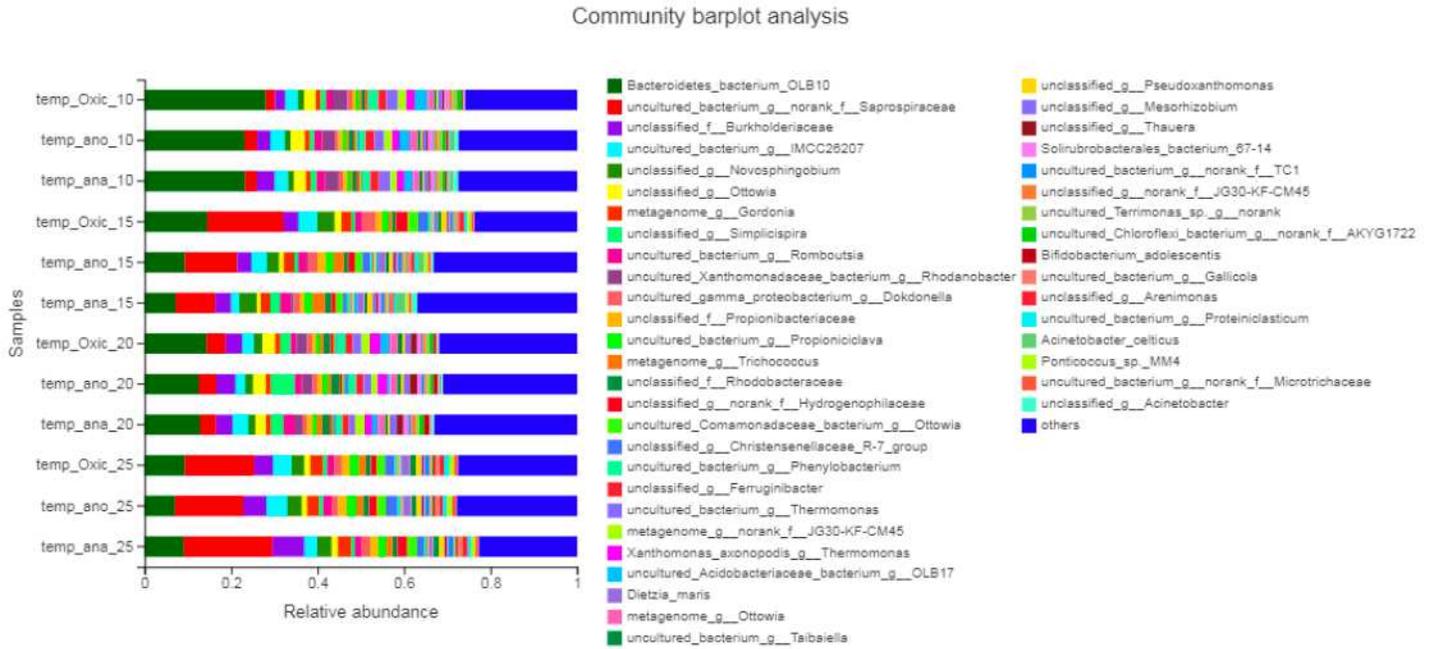


Figure 7

Community composition according to species.

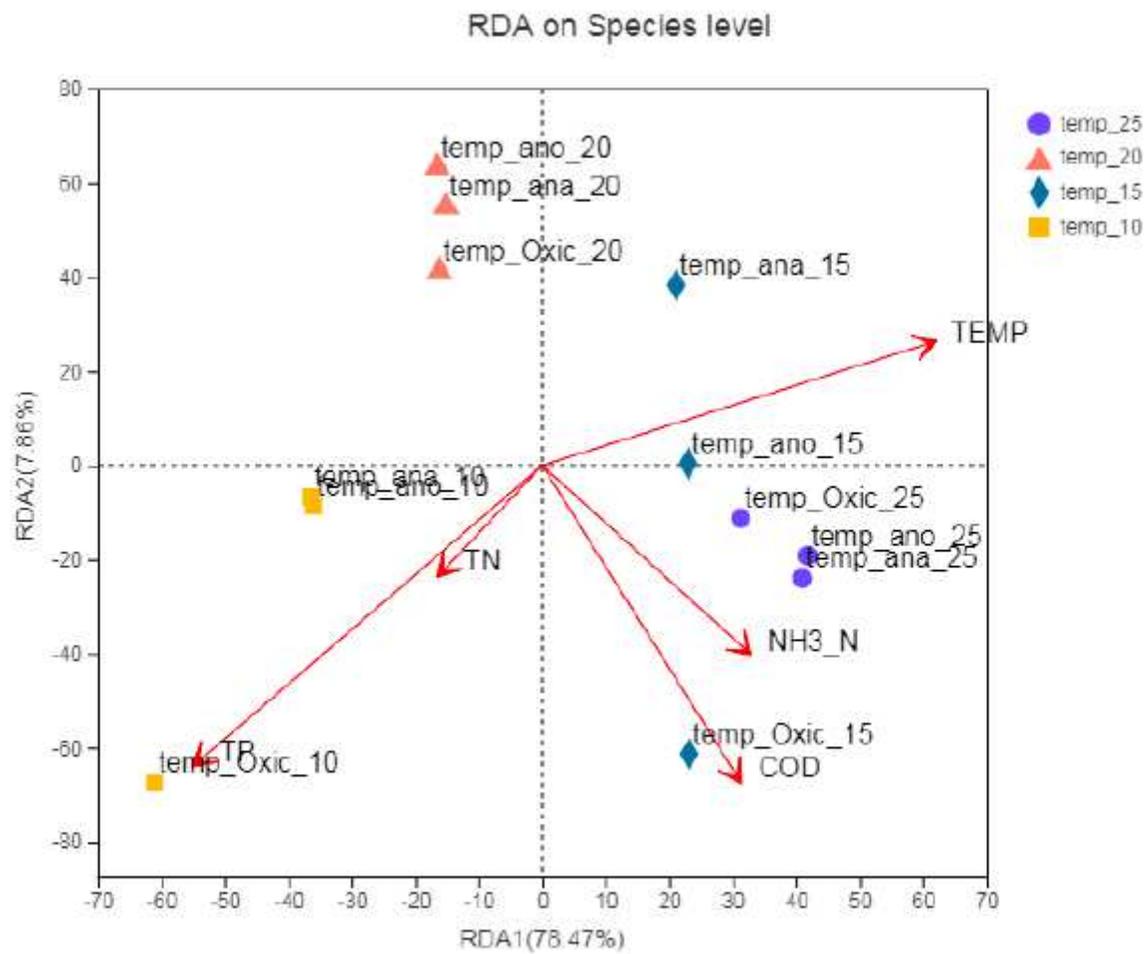


Figure 8

CCA analysis.

Spearman Correlation Heatmap

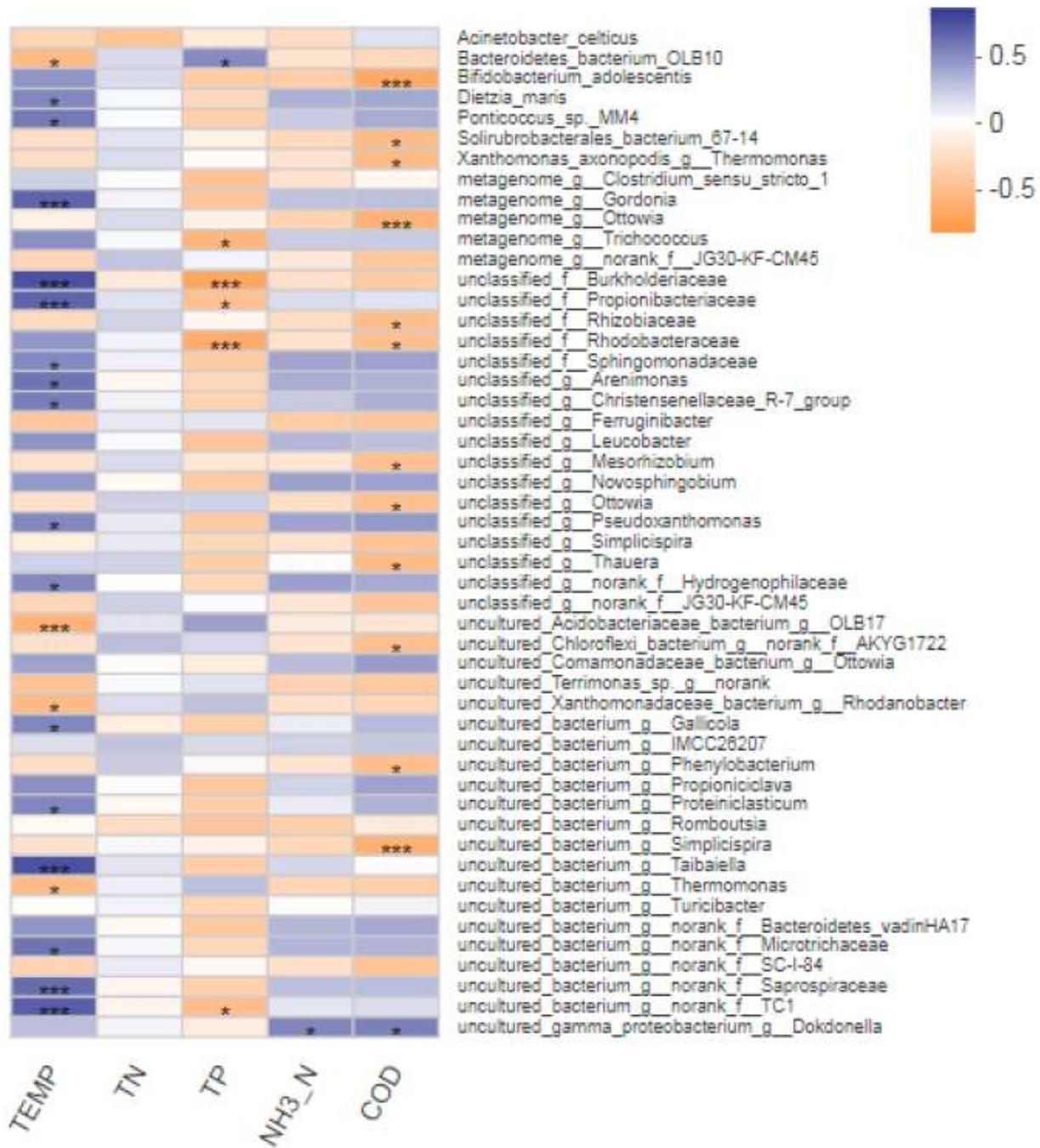


Figure 9

Correlation Heatmap.