Pathogens isolated in respiratory tract samples of recently hospitalized patients: Implications for primary care physicians

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Research article

Keywords: Lower respiratory tract infection, Pneumonia, Microbiology, Pathogens, Community, Antibiotics, Drug resistance, Bacterial resistance, Epidemiology

Posted Date: September 25th, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-46002/v2>

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Abstract

Background. Little is known about the etiology of community-acquired lower respiratory tract infections (CA-LRTI), and treatment is largely empirical. We aimed to provide clinicians with microbiologic data of the more severe cases, i.e. those that result in hospital admission.

Methods. A retrospective cohort study was conducted at a tertiary care hospital in Haifa, Israel. Consecutive respiratory tract samples obtained from admitted patients older than 12 years between 2014 and 2020 were included. Pathogen distribution and drug susceptibility were described, and factors associated with 14 day mortality rates were analyzed using a multivariable logistic regression with a stepwise model reduction.

Results. A total of 1,395; 2,212; and 2,760 samples were included in the community-acquired LRTI with no healthcare exposure, community-acquired LRTI with recent healthcare exposure, and LRTI diagnosed 48 hours to 7 days within hospital admission LRTI groups, respectively. Gram negative bacteria were the most common bacteria isolated. *Streptococcus pneumoniae* was over represented in patients admitted to the intensive care unit in the first two study groups (accounting for 20% and 10.1% of patients admitted to the ICU versus 12.6% and 6.4% of patients in the entire cohort, $p < 0.001$). *Streptococcus pneumoniae* was susceptible to penicillin in around 50% of cases; to erythromycin in 65% of cases; and to fluoroquinolones and 3rd generation cephalosporins in more than 95% of cases. Twenty-five percent of other typical community pathogens were resistant to ampicillin. Susceptibility of Gram negative bacteria to penicillins with and without a β -lactamase inhibitor and to second generation cephalosporins was lower than 50%. In the multivariable analysis, factors that were associated with increased mortality in the entire cohort were: age, admission to an ICU or surgical department, healthcare-associated infections, and infections with gram negative bacteria, *Staphylococcus aureus*, and *Stenotrophomonas* or *Acinetobacter baumannii*.

Conclusions. Our data shows that gram negative bacteria are the most common pathogens causing LRTI. Typical community acquired pathogens are proportionally more common in younger patients, although the vast majority occur in elderly patients. The susceptibility pattern of these pathogens suggests that for high-risk patients the choice of oral antibiotics is limited.

Background

A community-acquired lower respiratory tract infection (CA-LRTI) is one of the most common infections attended by primary care physicians and results in considerable use of antibiotics [1,2]. As microbiological analysis of sputum cultures is rarely performed in community settings, little is known about the etiology of CA-LRTI, and bacterial susceptibilities of causative pathogens are unknown in most areas. Treatment of LRTI by primary care physicians is thus largely empirical with penicillins, 2nd and 3rd generation cephalosporins, fluoroquinolones, doxycycline and macrolides often used. On the one hand, such empirical treatment may be superfluous resulting in more antibiotic resistance in the community [3]. On the other hand, for the few patients whose treatment fails and results in eventual hospital admission, antibiotic treatment may be inadequate.

In the few studies that have examined the microbiology of community acquired pneumonia among relatively young patients, the only culturable pathogens that were found in a significant proportion of cases (i.e. > 5%) are *Streptococcus pneumoniae* and *Hemophilus influenza* [4]. Other pathogens, such as *Moraxella* spp., *Bordetella* spp. and *Staphylococcus aureus* are uncommon [5]. Whereas, common pathogens such as *Legionella pneumophila*, *Mycoplasma pneumoniae* or *Chlamydia pneumoniae* are only diagnosed through molecular methods or serological tests [6]. Gram negative bacteria are detected in 2.8%-14% of community acquired pneumonia cases, particularly in the elderly and patients who are admitted from a long-term care facility (LTCF) [7,8].

By analyzing the results of respiratory tract cultures of recently admitted patients, we aimed to provide physicians with the relevant microbiologic data of the more severe CA-LRTI cases, i.e. those that resulted in hospital admission. Such microbiological data would provide primary care and emergency room physicians with additional insights as to the causative agents of severe CA-LRTI, thereby improving the process of selecting empirical antibiotic therapy for patients with high-risk LRTI in the community.

Methods

Study Setting and Cohort Description

A retrospective observational cohort study was performed at the Rambam Health Care Campus (RHCC), a 970-bed primary and tertiary care hospital in Haifa, Israel.

The cohort included all consecutive respiratory tract samples obtained from patients older than 12 years who were admitted to the RHCC between January 2014 and January 2020. We assumed these patients to have an active infection, although we did not go through each individual file. The Institutional Review Board of the RHCC approved the study for waiver of informed consent.

Data Collection

For each patient's admission throughout the study period, all unique bacterial species were included (i.e. the first bacteria of each species for each admission). Data were retrieved with the use of the institution's electronic medical records. Data pertaining to demographics, source of admission (home, LTCF), hospitalization at the RHCC within the past 90 days, Charlson Comorbidity Index (CCI), and admission unit were collected for each admission. Microbiology data included dates of sample collection, dates of results, name of bacteria species, and antimicrobial susceptibility.

Study Definitions

Bacterial pathogens were divided into seven groups: 1) Enterobacteriaceae; 2) Gram negative bacteria associated with community-acquired infections - *Hemophilus influenza*, *Moraxella* and *Bordetella* spp.; 3) non-fermenting gram negative bacteria - *Pseudomonas* and *Burkholderia* spp.; 4) Other gram

negative bacteria – *Serratia*, *Providencia*, and *Proteus* spp.; 5) *Staphylococcus aureus*; 6) Opportunistic bacteria common among intensive care unit (ICU) patients – *Stenotrophomonas* and *Acinetobacter* spp.; and 7) *Streptococcus pneumoniae*.

We defined a group of pathogens as ‘typical community pathogens’ based on previous large studies that showed *Streptococcus pneumoniae* and *Hemophilus influenza* to be by far the most common bacterial pathogens to cause CA-LRTI [4]. *Moraxella* and *Bordatella* spp., although uncommon, were also included in this group as they rarely cause hospital-acquired pulmonary infections [9,10].

Drug resistance was defined according to the Clinical and Laboratory Standards Institute (CLSI) breakpoints for susceptibility. Bacteria with intermediate drug susceptibility were defined as resistant. Finally, fourteen-day mortality was assessed in relation to the date the index culture was obtained.

Variable Definitions

The place of acquisition of each culture was classified with the use of three factors: 1) the time elapsed from date of admission to the date the sample was collected; 2) whether the admission was from a LTCF; and 3) if the patient had been hospitalized within the past 3 months. Accordingly, the study consisted of three cohorts as follows:

i) Community-acquired LRTI with no healthcare exposure: cultures which were obtained within less than 48 hours of admission of patients who were not residing in an LTCF and were not hospitalized in the RHCC within the past 3 months; ii) Community-acquired LRTI with recent hospital admission or residence in a LTCF: cultures which were collected within 48 hours of admission of patients who were either admitted from an LTCF or hospitalized in the RHCC within the past 3 months; iii) LRTI diagnosed 48 hours to 7 days within admission: cultures which were collected from patients who were hospitalized for no less than 3 days but for no more than 7 days.

The patients’ ages at the time of culture collection were stratified into 3 groups as follows: 12-18 years; >18 years to 40 years; and older than 40 years. The CCI was stratified into 3 groups: 1) a CCI of 0; 2) a CCI between 1 and 4; and 3) a CCI of >4.

Referring wards were stratified into 5 groups: 1) Internal medicine and Emergency; 2) Pediatrics; 3) ICU; 4) Oncology and Hemato-oncology; and 5) Surgery and Orthopedics.

The main outcome measures were the microbiologic diagnoses of LRTI in the three study groups. The secondary outcome measures were factors associated with 14 day mortality in the entire cohort, and in the pre-specified cohort of patients infected with typical community pathogens.

Statistical Analysis

Continuous variables were described with the use of median and 25-75% interquartile ranges (IQR). We analyzed factors associated with a 14-day mortality in the entire cohort, and in the cohort of patients with typical community pathogens (*Streptococcus pneumoniae*, *Hemophilus influenza*, *Bordatella* spp., *Moraxella* spp.) using a multivariate logistic regression with a stepwise model reduction, and accounting for age, gender, referring ward, and place of acquisition. A p-value of 0.05 was considered statistically significant.

Results

A total of 1,395; 2,212; and 2,760 samples were included in the community-acquired (with and without healthcare exposure), and community-acquired LRTI diagnosed 48 hours to 7 days after admission, respectively. Of the 2,212 community-acquired LRTI with recent hospital admission or residence in a LTCF, 295 were obtained from patients coming from LTCFs, while 1917 were obtained from patients with previous hospital admissions. Samples taken from female patients accounted for 1884 / 6367 (29.6%) of the entire cohort. The mean age was 60.6 ± 19.9 years (median 65, IQR = 50 - 75), and the mean CCI was 5.7 ± 4.1 (median 5, IQR = 2 - 8.5). Samples were obtained in the pediatric ward (267, 4.2%); ICU (1823, 28.6%); emergency room and internal medicine wards (3190, 50.1%); oncology and hemato-oncology wards (190, 2.5%); and surgery and orthopedics wards (897, 14.1%). Sputum samples, tracheal aspirations or broncho-alveolar lavage were used in 3912 (61.4%), 1879 (29.5%), and 576 (9%) of all specimens, respectively. These demographic data are presented in Table 1.

Table 1: Characteristics of patients with LRTI

	Entire cohort (N=6367 samples)
Age (Median, IQR)	65 (50,75)
Female (N (%))	1884 (30)
Charlson comorbidity index (Median, IQR)	5 (2, 8.5)
Place and timing of acquisition	
Community with no healthcare exposure (N (%))	1395 (21.9)
Community with healthcare exposure (N (%))	2212 (34.7)
Cultures obtained 48 hours - 2 days within admission (N (%))	2760 (43.3)
Department	
Pediatric (N (%))	267 (4.2)
Internal medicine (N (%))	3190 (50.1)
Surgery-orthopedic (N (%))	897 (14.1)
Hemato-oncology (N (%))	190 (2.5)
ICU (N (%))	1823 (28.6)
Source of specimen	
Sputum (N (%))	3912 (61.4)
Tracheal aspiration (N (%))	1879 (29.5)
Broncho-alveolar lavage (N (%))	576 (9)

LRTI, lower respiratory tract infection

Pathogen distribution among the three study cohorts is shown in Table 2. The probability of isolating bacteria which we initially expected to be “typical” community-acquired pathogens, namely *Streptococcus pneumoniae*, *Hemophilus influenza*, *Bordatella* spp., *Moraxella* spp., was higher among patients with community-acquired LRTI and with no healthcare exposure ($p < 0.0001$ for all comparisons). Enterobacteriaceae, non-fermenting gram negative bacteria, and other gram negative bacteria, were more frequently isolated from older patients with multiple comorbidities with community-acquired LRTI with recent hospital admission or residence in a LTCF and community-acquired LRTI diagnosed 48 hours to 7 days after admission, but were by far the most common pathogens isolated in all cohorts ($p < 0.0001$ for all comparisons ; Figures 1,2), while *Staphylococcus aureus* was relatively equally distributed among the three study cohorts. These observations were also true when patients were divided into age groups (12-18 years, > 18-40 years, > 40 years; Figure 1), or according to the number of comorbid conditions included in the CCI (Figure 2).

Table 2: Pathogen distribution among patients with community-acquired, healthcare-associated and hospital-acquired LRTI

Bacteria	Community-acquired LRTI with no healthcare exposure Absolute number (%) (N=1,395)	Community-acquired LRTI with healthcare exposure Absolute number (%) (N=2,212)	LRTI diagnosed 3-7 days after admission. Absolute number (%) (N=2,760)
<i>Citrobacter</i> spp., <i>enterobacter</i> spp., <i>Escherichia coli</i> , <i>Klebsiella</i> spp.	371 (26.6%)	564 (25.5%)	860 (31.2%)
<i>Hemophilus</i> spp., <i>Bordatella</i> spp., <i>Moraxella</i> spp.	276 (19.8%)	280 (12.7%)	302 (10.9%)
<i>Pseudomonas</i> and <i>Burkholderia</i> spp.	210 (15.1%)	539 (24.4%)	556 (22.1%)
<i>Serratia</i> , <i>Providencia</i> and <i>Proteus</i> spp.	66 (4.7%)	173 (7.8%)	225 (8.2%)
<i>Staphylococcus aureus</i>	227 (16.3%)	354 (16.0%)	407 (14.8%)
<i>Stenotrophomonas</i> and <i>Acinetobacter</i> spp.	69 (4.9%)	161 (7.3%)	315 (11.4%)
<i>Streptococcus pneumoniae</i>	176 (12.6%)	141 (6.4%)	95 (3.4%)

LRTI, lower respiratory tract infection. Community-acquired LRTI consist of samples taken within 0-2 days of hospital admission. Healthcare-associated LRTI consist of samples taken within 0-2 days of hospital admission and with recent healthcare exposure. Hospital-acquired LRTI consist of samples taken within 3-7 days of hospital admission.

Overall, the number of respiratory cultures was higher among patients older than 40 years compared to other age groups. The relative proportion of samples with a growth of *Streptococcus pneumoniae* was significantly higher ($p < 0.001$) among patients aged 18 to 40 years (9.8% of all samples) when compared to younger patients (7.1%), or to adults aged more than 40 years (5.9%), although the absolute number of samples positive for this bacterium was much higher among older patients. Similarly, *Hemophilus influenza*, *Moraxella* spp., and *Bordatella* spp., were isolated in 15.1%, 18.1% and 12.6% of samples from patients aged 12 to 18 years, > 18 to 40, and > 40 years, respectively ($p < 0.001$ for all comparisons), but the vast majority of samples positive for these bacteria were taken from patients older than 40 years.

Pathogen distribution in samples taken from patients in the ICU is shown in Table 3. Among patients with community-acquired with or without healthcare exposure LRTI, *Streptococcus pneumoniae* was over-represented among patients with CA-LRTI and healthcare-associated LRTI admitted to the ICU (accounting for 20% and 10.1% among patients admitted to the ICU versus 12.6% and 6.4% among patients in these cohort, $p < 0.001$ for both comparisons). Other typical community pathogens (*Hemophilus influenza*, *Moraxella* spp. and *Bordatella* spp.), gram negative bacteria and *Staphylococcus aureus* were not more prevalent among patients in the ICU when compared to their relative frequency in the entire cohort.

Table 3: Frequency of typical community pathogens and *Staphylococcus aureus* in respiratory tract cultures from hospitalized patients

Entire cohort N=6367				ICU N=1823			
	Community acquired LRTI (no healthcare exposure) N=1395	Community acquired LRTI (with healthcare exposure) N=2212	LRTI diagnosed 3-7 days after admission N=2760		Community acquired LRTI (no healthcare exposure) N= 454	Community acquired LRTI (with healthcare exposure) N=426	LRTI diagnosed 3-7 days after admission N=943
<i>Streptococcus pneumoniae</i> N (%)	176 (12.6)	141 (6.4)	95 (3.4)	<i>Streptococcus pneumoniae</i> N (%)	91 (20)	43 (10.1)	32 (3.4)
<i>Hemophilus influenza</i> , <i>Bordatella</i> spp., <i>Moraxella</i> spp. N (%)	276 (19.8)	280 (12.7)	302 (10.2)	<i>Hemophilus influenza</i> , <i>Bordatella</i> spp., <i>Moraxella</i> spp. N (%)	89 (19.6)	92 (21.6)	87 (9.2)
<i>Staphylococcus aureus</i> N (%)	227 (16.3)	354 (16)	407 (14.8)	<i>Staphylococcus aureus</i> N (%)	89 (18.9)	70 (16.4)	98 (10.4)
<i>Citrobacter</i> , <i>Enterobacter</i> , <i>K. pneumoniae</i> , <i>E. coli</i> N (%)	371 (26.6)	564 (25.5)	860 (31.2)	<i>Citrobacter</i> , <i>Enterobacter</i> , <i>K. pneumoniae</i> , <i>E. coli</i> N (%)	113 (24.9)	112 (26.3)	324 (34.3)
<i>Serratia</i> , <i>Providencia</i> , <i>Proterus</i> N (%)	66 (4.7)	173 (7.8)	225 (8.2)	<i>Serratia</i> , <i>Providencia</i> , <i>Proterus</i> N (%)	19 (4.2)	27 (6.3)	77 (8.2)
<i>Pseudomonas speciese</i> , <i>Burkholderia</i> N (%)	210 (15.1)	539 (24.4)	556 (22.1)	<i>Pseudomonas speciese</i> , <i>Burkholderia</i> N (%)	32 (7)	51 (12)	174 (18.5)
<i>Stenotrophomonas</i> , <i>Acinetobacter</i> N (%)	69 (4.9)	161 (7.3)	315 (11.4)	<i>Stenotrophomonas</i> , <i>Acinetobacter</i> N (%)	24 (5.3)	31 (7.3)	151 (16)

The table shows the frequency of pathogens taken from patients hospitalized in all departments and in the intensive care unit (ICU) at the time of sample acquisition.

Among the 190 patients with active malignancy, relatively more samples were obtained through the use of broncho-alveolar lavage (21% versus 9% in the entire cohort). When compared to other patients, *Pseudomonas* spp. (25.3%) and gram negative bacteria (30%) were slightly more commonly isolated in these immunocompromised hosts, while *Streptococcus pneumoniae* (4.2%) and other typical community pathogens (7.9%) were less commonly isolated.

Antibiotic susceptibilities of various bacteria are shown in Table 4. *Streptococcus pneumoniae* was susceptible to penicillin in only around 50% of cases, and to erythromycin in 65% of cases. *Streptococcus pneumoniae* bacteria were susceptible in more than 95% of cases to fluoroquinolones (both ciprofloxacin and levofloxacin) and to 3rd generation cephalosporins (ceftriaxone). Other typical community-acquired pathogens (*Hemophilus influenza*, *Moraxella* spp., and

Bordetella spp.) were also nearly universally susceptible to fluoroquinolones and 3rd generation cephalosporins, but about 25% were resistant to ampicillin. Gram negative bacteria were resistant to penicillins, penicillin and β -lactamase combinations, and 2nd generation cephalosporins in more than 50% of cases. Susceptibility rates to fluoroquinolones and 3rd generation cephalosporins were around 80% and 75% respectively (Table 4).

Table 4: Antimicrobial susceptibilities of typical community pathogens and *Staphylococcus aureus* in LRTIs

otic	Penicillin	Ampicillin	Amoxicillin/Clavulonic acid % susceptible	Oxacillin	Cefuroxime	Ceftriaxone	Ciprofloxacin	Levofloxacin	Erythromycin
ptible	% susceptible	% susceptible		% susceptible	% susceptible	% susceptible	% susceptible	% susceptible	% susceptible
	<i>Hemophilus, Bordetella, Moraxella</i>								
unity 1	-	74	-		98.1	100	100	100	-
icare ated 2	-	75.6	-		99.6	99.6	88.9	89.5	-
al 4	-	71.9	97.3		99.6	100	95.5	100	-
	<i>Streptococcus pneumoniae</i>								
unity 5	59.1	-	-		-	97.9	-	98.6	73.4
1	43.4	-	-		-	95.3		97.2	58.5
al 0	54.5	-	-		-	97.2	-	100	62.3
	<i>Citrobacter, Enterobacter, E. coli, Klebsiella</i>								
unity 1	-	14.9	58.4		69.2	84.3	89.9	-	-
4	-	8	47.9		49.4	68.7	77.5	-	-
al 0	-	6.7	49.8		51.7	70.4	82.8	-	-
	<i>Staphylococcus aureus</i>								
unity 7	23.6	-	-	83.5	-	-	81.1	65.6	66.5
4	17.4	-	-	63.9	-	-	66.3	-	56.8
al 3	25.9	-	-	66	-	-	69.4	-	57.6

LRTI, lower respiratory tract infection. Test susceptibility, with Ceftriaxone as a reference.

Table 5 presents the frequency of typical community pathogens, gram negative bacteria, and *Staphylococcus aureus* among patients who died within 14 days of obtaining the index culture. *Streptococcus pneumoniae* and gram negative bacteria were disproportionately represented among patients who died in the community-acquired with no healthcare exposure LRTI group. Other typical community pathogens were disproportionately represented among patients who survived in all cohorts.

Table 5: Fourteen-day mortality among patients with respiratory cultures of typical community pathogens, *Staphylococcus aureus*, and Gram-negative bacillii

Community acquired LRTI with no healthcare exposure N=1395			Community acquired LRTI with healthcare exposure N=2212			LRTI diagnosed 3-7 days after admission N=2760		
Bacterial species	Dead (N=161)	Alive (N=1234)	Bacterial species	Dead (N=342)	Alive (N=1870)	Bacterial species	Dead (N=551)	Alive (N=2209)
<i>Streptococcus pneumoniae</i> N=181 N (%)	29 (18)	152 (12.3)	<i>Streptococcus pneumoniae</i> N=141 N (%)	18 (5.3)	123 (6.6)	<i>Streptococcus pneumoniae</i> N=95 N (%)	9 (1.6)	86 (3.9)
<i>Hemophilus influenza</i> , <i>Bordatella</i> spp., <i>Moraxella</i> spp. 276 N (%)	26 (16.1)	250 (20.3)	<i>Hemophilus influenza</i> , <i>Bordatella</i> spp., <i>Moraxella</i> spp. N=280 N (%)	29 (8.5)	251 (13.4)	<i>Hemophilus influenza</i> , <i>Bordatella</i> spp., <i>Moraxella</i> spp. N=302 N (%)	36 (6.5)	266 (12)
<i>Staphylococcus aureus</i> N=227 N (%)	29 (18)	198 (16)	<i>Staphylococcus aureus</i> N=354 N (%)	62 (18.1)	292 (15.6)	<i>Staphylococcus aureus</i> N=407 N (%)	95 (17.2)	312 (14.1)
<i>Citrobacter</i> , <i>Enterobacter</i> , <i>K. pneumoniae</i> , <i>E. coli</i> N=371 N (%)	51 (31.7)	320 (25.9)	<i>Citrobacter</i> , <i>Enterobacter</i> , <i>K. pneumoniae</i> , <i>E. coli</i> N=564 N (%)	123 (36)	441 (23.6)	<i>Citrobacter</i> , <i>Enterobacter</i> , <i>K. pneumoniae</i> , <i>E. coli</i> N=860 N (%)	165 (29.9)	695 (31.5)
<i>Serratia</i> , <i>Providencia</i> , <i>Proterus</i> N=66 N (%)	8 (5)	58 (4.7)	<i>Serratia</i> , <i>Providencia</i> , <i>Proterus</i> N=173 N (%)	26 (7.6)	147 (7.9)	<i>Serratia</i> , <i>Providencia</i> , <i>Proterus</i> N=225 N (%)	47 (8.5)	178 (8.1)
<i>Pseudomonas species</i> , <i>Burkholderia</i> N=210 N (%)	17 (10.6)	193 (15.6)	<i>Pseudomonas species</i> , <i>Burkholderia</i> N=539 N (%)	54 (15.8)	485 (25.9)	<i>Pseudomonas species</i> , <i>Burkholderia</i> N=556 N (%)	116 (21.1)	440 (19.9)
<i>Stenotrophomonas</i> , <i>Acinetobacter</i> N=69 N (%)	6 (3.7)	63 (5.1)	<i>Stenotrophomonas</i> , <i>Acinetobacter</i> N=161 N (%)	30 (8.8)	131 (7)	<i>Stenotrophomonas</i> , <i>Acinetobacter</i> N=315 N (%)	83 (15.1)	232 (10.5)

Factors associated with a 14 day mortality were analyzed in the entire cohort, including gram negative bacteria and typical community pathogens (*Streptococcus pneumoniae*, *Hemophilus influenza*, *Moraxella* spp., and *Bordatella* spp.). In the multivariable analysis, factors that were associated with

increased mortality in the entire cohort were: age, admission to an ICU, admission to a surgical department, community-acquired LRTI with recent hospital admission or residence in a LTCF and community-acquired LRTI diagnosed 48 hours to 7 days after admission (when compared to “pure” community acquired infections), and infections with *Staphylococcus aureus*, Enterobacteriaceae, or *Stenotrophomonas* and *Acinetobacter baumannii* (Tables 6a and 6b).

Table 6a: Predictors for 14-day mortality among hospitalized patients with LRTI

Predictor	Adjusted OR (95% CI)	P-value
Age*	1.026 (1.02, 1.032)	<0.001
Department		
Internal medicine	1 (reference)	
Pediatric	0.57 (0.77, 4.27)	0.59
ICU	3.37 (2.34, 4.85)	<0.001
Oncology-hematology	0.53 (0.07, 3.97)	0.53
Surgery-orthopedic	2.48 (1.34, 4.60)	0.004
Bacterial species		
<i>Streptococcus pneumoniae</i>	1 (reference)	
Gram negative bacteria associated with community-acquired infections [‡]	0.83 (0.57, 1.21)	0.34
<i>Staphylococcus aureus</i>	1.55 (1.10, 2.19)	0.01
Non-fermenting gram negative bacteria [€]	1.06 (0.75, 1.49)	0.75
Opportunistic bacteria common among ICU patients [¥]	1.69 (1.16, 2.45)	0.006
Enterobacteriaceae	1.39 (1.001, 1.93)	0.048
Other gram negative bacteria [£]	1.23 (0.83, 1.82)	0.30
Timing and place of acquisition		
Community acquired without healthcare exposure	1 (reference)	
Community acquired with healthcare exposure	1.64 (1.21, 2.23)	0.002
LRTI cases diagnosed 48 hours – 7 days after admission	3.08 (2.28, 4.15)	<0.001

OR, odds ratio; CI, confidence interval; ICU, intensive care unit; LRTI, lower respiratory tract infection

[‡] *Hemophilus influenza*, *Moraxella* and *Bordetella* spp.

[€] *Pseudomonas* and *Burkholderia* spp

[¥] *Stenotrophomonas* and *Acinetobacter* spp

[£] *Serratia*, *Providencia*, and *Proteus* spp

*Increase for each year

Table 6b: Predictors for 14-day mortality among patients who had respiratory cultures with typical community pathogens

Predictor	Adjusted OR (95% CI)	P-value
Age*	1.025 (0.99, 1.05)	0.056
Charlson Comorbidity Index [†]		
Charlson Comorbidity Index = 0 (reference)	1	
Charlson Comorbidity Index 1-4	0.40 (0.14, 1.15)	0.09
Charlson Comorbidity Index >4	0.56 (0.17, 1.84)	0.34
Department		
Internal medicine	1 (reference)	
Children	0.56 (0.17, 1.84)	0.34
ICU	2.23 (1.44, 3.55)	<0.001
Oncology-hematology	1.74 (0.63, 4.78)	0.28
Surgery-orthopedic	1.87 (1.07, 3.26)	0.03
Timing and place of acquisition		
Community acquired (no healthcare exposure)	1 (reference)	
Community acquired (with healthcare exposure)	0.88 (0.08, 9.32)	0.92
LRTI cases diagnosed 48 hours – 3 days within admission	0.97 (0.20, 4.82)	0.97

OR, odds ratio; CI, confidence interval; ICU, intensive care unit;

*Increase for each year

[†] An interaction was found between the Charlson Comorbidity Index and the place of acquisition. Subgroup analysis for healthcare-associated infection is described in the text above.

In the multivariable analysis that included only typical community-acquired pathogens (*Streptococcus pneumoniae*, *Hemophilus influenza*, *Moraxella* spp., and *Bordetella* spp.), factors that were associated with an increased risk of mortality included ICU and surgical ward admission. Because of a strong interaction between the place of acquisition and the CCI, the cohort was stratified into subgroups according to the place of acquisition and the multivariable analysis for a 14-day mortality was repeated. In the subgroup of community-acquired LRTI with recent hospital admission or residence in a LTCF, a CCI between 1 and 4, as well as a CCI greater than 4 were independent predictors for a 14-day mortality (OR 12.94, 95% CI 1.65, 101.15, p=0.01; OR 14.84, 95% CI 1.70, 129.15, p=0.01, respectively). The CCI was not associated with a 14-day mortality in the other subgroups.

Discussion

This retrospective cohort study included data on respiratory samples that were collected from patients with community-acquired LRTI, but divided into three groups: those taken within 48 hours of hospital admission from patients without known healthcare exposure; those taken within 48 hours of hospital admission from patients with known healthcare exposure; and samples taken 3-7 days after hospital admission. A unique finding in this cohort was a rather high percentage of LRTI caused by Gram negative bacteria in all study groups. However, susceptibility to 1st line agents (e.g. penicillins with and without β -lactamase inhibitors) used in the community for the treatment of LRTI was alarmingly low.

Although among young and relatively healthy patients, a larger proportion of samples had growth of typical community pathogens (*Streptococcus pneumoniae*, *Hemophilus influenza*, *Bordetella* spp., *Moraxella* spp.), in terms of absolute numbers the majority of these cases occurred in patients older than

40 years. Cases with community-acquired LRTI and no healthcare exposure that were caused by *Streptococcus pneumoniae*, but not other typical community pathogens, were more likely to be admitted to an ICU and to die.

Our results highlight the limited choice of available oral antibiotics for patients with CA-LRTIs. Primary care physicians often use penicillins with or without β lactamase inhibitors, 2nd generation cephalosporins, macrolides, doxycycline, and fluoroquinolones for LRTIs. Typical community pathogens were nearly always susceptible to 3rd generation cephalosporins that are not universally available in an oral formulation, and to fluoroquinolones. Susceptibility to penicillin, ampicillin, and macrolides was consistently much lower among all study groups, and lower than those described in several previous European and North American studies [4,11-13]. Gram negative bacteria, isolated mostly from frail elderly patients, were resistant to 2nd generation cephalosporins, penicillins and β -lactamase inhibitors in more than 50% of cases. Although *Hemophilus influenza*, *Bordatella* spp., *Moraxella* spp. were nearly universally susceptible to amoxicillin and clavulanic acid, such combinations are unlikely to provide additional benefit over penicillins for the treatment of penicillin-resistant *Streptococcus pneumoniae*, as resistance in these bacteria is mediated by mutations in the penicillin-binding protein rather than by the production of β -lactamases [14]. Other antibiotic agents may also not be adequate for the treatment of LRTI. For instance, trimethoprim sulphamethoxazole is not part of treatment guidelines. Furthermore, 2nd generation cephalosporins in this study were active against only about 50% of gram negative bacteria, are probably less efficacious, and are mostly available in oral formulations with relatively low bioavailability [15,16].

The therapeutic implications of these data are not easy to determine, as many patients with CA-LRTI are treated empirically, and microbiologic data is either received late in the course of treatment or is unavailable altogether. However, it is likely that for the vast majority of younger patients with a low number of comorbid conditions, the likelihood of an adverse outcome from a CA-LRTI is very low, while the ecological price of antibiotic overuse is high [17]. Since CA-LRTIs are very common, resistance to administered antibiotics develops quickly [18]. Treatment with macrolides, 2nd generation cephalosporins, and penicillin and β -lactamase inhibitor combinations is likely to result in a good outcome in low-risk patients. Such low-risk patients who are infected with *Streptococcus pneumoniae* with intermediate resistance to penicillin can probably be adequately treated by higher doses of penicillin. Conversely, for patients who are at a higher risk of complications, i.e. older patients with multiple comorbid conditions, failure of a 1st line antibiotic treatment administered in the community setting may result in increased use of 2nd line antibiotics, and increased rates of hospital admission, mechanical ventilation, and death. A similar population includes older patients discharged from hospital to complete antibiotic treatment for an LRTI in the community. These patients nearly always have multiple risk factors for treatment failure and microbiologic data are usually unavailable. In such patients an initial or follow up treatment using penicillins with or without β -lactamase inhibitors, macrolides, or 2nd generation cephalosporins may be inadequate. Fluoroquinolones, and more specifically the so called “respiratory” fluoroquinolones, provide excellent coverage according to our data and those of previous studies [19-21]. The use of fluoroquinolones, however, has several important drawbacks for patients with significant comorbidities. Namely, they are associated with an increased risk of *Clostridioides difficile* associated diarrhea and probably more so than most other antibiotics [22]; they can cause QT prolongation that may lead to arrhythmias; and their use is associated with tendinopathies, peripheral neuropathy, and rupture of aortic aneurysms [22]. Treatment with fluoroquinolones quickly leads to colonization with resistant pathogens in the treated patients [18,23], and increases the risk of fluoroquinolones resistance in certain geographic areas even in the absence of personal antibiotic use [24].

Improving diagnostics with the use of molecular tools for identification of respiratory pathogens and / or their resistance genes may facilitate antibiotic choice in the near future. In addition, newer antibiotics such as lefamulin, and oral formulations of 3rd generation cephalosporins (i.e. cefditoren pivoxil or cefpodoxime) may prove useful for high-risk patients with relative contraindications to fluoroquinolones.

There are several important limitations for this study. The data are retrospective, and are based on microbiological sampling results. Occasional patients with no active infections could have been included, although RHCC policy is that respiratory tract samples are taken only for patients with suspected infections. The inclusion of patients without actual infection may have resulted in the inclusion of some pathogens that do not have an actual pathogenic role, but are rather colonizers of the respiratory tract. Conversely, samples may not have been obtained for various reasons from patients with clinically significant LRTI. Different clinical syndromes, i.e. pneumonia, acute exacerbation of chronic lung disease, infected bronchiectasis, and bacterial bronchitis, have been analyzed in this cohort. As the epidemiology and microbiology of these entities differ, our results present the “big picture”, but may miss important data relevant to specific infections. We may have missed some CA-LRTI cases caused by less common pathogens that are difficult to diagnose (e.g. *Legionella*). Furthermore, we have not included viral pathogens in the analysis, and certainly viral infections are important causes of LRTI in the community. Empirical treatment for a presumed bacterial infection, however, should be based on the antibiotic susceptibility profile, safety, ecological cost and economic cost of specific agents, and targeted towards the most common community pathogens. Susceptibility tests were performed according to CLSI requirements. Therefore, there are some gaps in potentially useful data, i.e. *Streptococcus pneumoniae* susceptibility rates to 2nd generation cephalosporins.

Conclusions

Our data shows that gram negative bacteria are very common among patients admitted because of a LRTI, and typical community acquired pathogens (*Streptococcus pneumoniae*, *Hemophilus influenza*, *Bordatella* spp., *Moraxella* spp.) are especially common among older patients with multiple comorbidities. The isolation of these bacteria in respiratory tract samples is associated with appreciable rates of ICU admission and 14-day mortality rates. The susceptibility pattern of these community pathogens suggests that for high-risk patients, the choice of effective oral antibiotic agents is limited. Both improvements in diagnostic procedures aimed at providing targeted therapy and newer oral antibiotic formulations are needed.

List Of Abbreviations

CA-LRTI - community-acquired lower respiratory tract infections

CCI - Charlson Comorbidity Index

CLSI - Clinical and Laboratory Standards Institute

ICU – intensive care unit

IQR - interquartile ranges

LTCF - long-term care facility

LRTI - lower respiratory tract infections

RHCC - Rambam Health Care Campus

Declarations

Competing interests: Neuberger Ami has received writing honoraria from Taro Pharmaceuticals Industries Ltd.

Funding: This work was supported by a research grant from Taro Pharmaceuticals Industries Ltd.

Authors contributions: OH – data analysis, writing of manuscript. ARB – data analysis, review of manuscript, AN – study concepts, writing of manuscript

Ethics approval and consent to participate: This study protocol was exempted for review by the Institutional Review Board of the Rambam Health Care Center according to the exemption criteria. In this study, anonymized and deidentified information was used for the analyses; thus, informed consent was not required.

Consent for publication: Not applicable.

Availability of data and materials: The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Acknowledgement: We would like to thank Dr. Ruth Moont for her assistance in reviewing, editing, and preparing the manuscript.

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Figures

CA, community-acquired; LRTI, lower respiratory tract infection; HCA, healthcare-associated; HA, hospital-acquired. Community-acquired LRTI consist of samples taken within 0-2 days of hospital admission. Healthcare-associated LRTI consist of samples taken within 0-2 days of hospital admission and with recent healthcare exposure. Hospital-acquired LRTI consist of samples taken within 3-7 days of hospital admission.

Figure 1

Frequency of bacteria isolated in respiratory tract cultures taken from hospitalized patients by age groups

CA, community-acquired; LRTI, lower respiratory tract infection; HCA, healthcare-associated; HA, hospital-acquired. Community-acquired LRTI consist of samples taken within 0-2 days of hospital admission. Healthcare-associated LRTI consist of samples taken within 0-2 days of hospital admission and with recent healthcare exposure. Hospital-acquired LRTI consist of samples taken within 3-7 days of hospital admission.

Figure 2

Frequency of bacteria isolated in respiratory tract cultures taken from patients by Charlson Comorbidity Index