

iTRAQ-Based Quantitative Proteomic Analysis of Heat Stress-Induced Mechanisms in Pepper Seedlings

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Abstract

Background: As one of the most important vegetable crops, pepper has rich nutritional value and high economic value. Increasing heat stress due to the global warming has a negative impact on the growth and yield of pepper.

Result: In the present study, we investigated the changes of phenotype, physiology, and proteome in heat-tolerant (17CL30) and heat-sensitive (05S180) pepper seedlings in response to heat stress. Phenotypic and physiological changes showed that 17CL30 had a stronger ability to resist heat stress compared with 05S180. In proteomic analysis, a total of 3,874 proteins were identified, and 1,591 proteins were considered to participate in the process of heat stress response. According to bioinformatic analysis of heat-responsive proteins, the heat tolerance of 17CL30 might be related to a higher photosynthesis, signal transduction, carbohydrate metabolism, and stress defense, compared with 05S180.

Conclusion: To understand the heat stress response mechanism of pepper, an iTRAQ-based quantitative proteomic analysis was employed to identify possible heat-responsive proteins and metabolic pathways in 17CL30 and 05S180 pepper seedlings under heat stress. This study provided new insights into the molecular mechanisms involved in heat tolerance of pepper and might offer supportive reference for the breeding of new pepper variety with heat resistance.

Introduction

Pepper (*Capsicum annuum* L.) is one of the most important vegetable crops worldwide, which greatly contributes to the vegetable supply and farmers' income. Capsaicin and capsanthin extracted from pepper or its other components, such as vitamin and trace element, can improve people's appetite and health. It is well known that pepper can well grow in warm place but sensitive to high temperature. Pepper will suffer from heat stress when the ambient temperature exceeds 32 °C [1], and its yield and quality are seriously reduced by the high temperature during the hot summer. Therefore, it is necessary to investigate the molecular mechanism underlying the pepper's response to heat stress in order provide crucial information for the research on complex traits and breeding heat-tolerant pepper cultivars.

Heat stress is one of the adverse environment factors affecting the growth and productivity of plants, resulting in wilting or even death of plants [2, 3]. According to the predication of IPCC (Intergovernmental Panel on Climate Change), the global temperature is likely to increase by about 2–5 °C by the end of 21th century (<https://www.ipcc.ch/>), and the pepper yield will be reduced by about 70%, leading to serious economic loss [4].

Until now, many studies have investigated the molecular mechanism of heat stress responses. To cope with heat stress, a set of defense mechanisms are activated to protect plants from the heat stress-induced damage, including the change of signal transduction and transcriptional regulation, the accumulation of osmolytes, the production of reactive oxygen species (ROS) scavenging enzymes, and the synthesis and accumulation of heat shock proteins (HSPs) [2, 5–8]. The generation and

accumulation of ROS caused by heat stress can lead to the peroxidation of membrane lipids and proteins as well as carbohydrate oxidation and DNA damage [9, 10]. To scavenge and detoxify ROS, plants have evolved both enzyme and non-enzyme systems [9]. The non-enzymatic antioxidants, including glutathione (GSH) and ascorbate, and enzymatic antioxidants, such as peroxidase (POD), glutathione S-transferase (GST), ascorbate peroxidase (APX), dehydroascorbate reductase (DHAR), catalase (CAT), and superoxide dismutase (SOD), play critical roles in the detoxification of ROS [11–13]. The ROS scavenging capacity of may be related to the tolerance to heat stress. Furthermore, HSPs, as molecular chaperones, are involved in folding, transferring and assembly of proteins, and they also can repair or remove denatured proteins in plant cell. HSP90, HSP70 and small HSPs (sHSPs) have been demonstrated to protect plants against the heat stress [14, 15].

In recent years, quantitative proteomic analysis has been applied to investigate the response to abiotic stress in plants [16–18]. Proteomic analysis of plants under heat stress has been widely performed, and some heat-responsive proteins and metabolic pathways have been identified to play important roles in the process of plant response to heat stress [19, 20]. In the present study, heat-tolerant (HT) and heat-sensitive (HS) pepper seedlings were subjected to heat stress (40 °C) to identify the differentially accumulated proteins (DAPs) in different pepper genotypes, and the contents of proteins in pepper leaves collected at four time points were identified and quantified by using the iTRAQ-based quantitative proteomic approach. Moreover, we aimed to identify some key proteins and pathways related to heat stress response in pepper. In addition, we also attempted to provide a new insight into the molecular mechanisms involved in heat tolerance of pepper.

Results

Phenotype and physiological responses of HT and HS under heat stress

After 48 h of heat stress, HS seedlings showed more wilting than HT, suggesting that the effect of heat stress on pepper seedlings was slighter in HT compared with HS (Fig. 1A). After 48 h of heat stress, the contents of proline and soluble sugar were increased in the two pepper genotypes, while their contents in the HT genotype were higher compared with the HS genotype (Fig. 1B, C). Compared with the control samples, the content of MDA was significantly increased both in HT and HS after 48 h of heat stress. However, HS showed a stronger up-regulation (Fig. 1D). RWC values after 48 h of heat stress were decreased in leaves, and such reduction was greater in HS compared with HT (Fig. 1E).

iTRAQ data analysis and responses of DAPs to heat stress

Based on the iTRAQ experiment, a total of 1,628,623 spectra were identified from pepper seedlings. Moreover, 23,706 peptides, 21,335 unique peptides, and 5,098 proteins were detected from 193,696 known spectra (Fig. S1A). The MS data were deposited into the identifier PXD019556. Fig. S1B shows the mass distribution of proteins, and the majority of them were 10-70 kDa in size. In addition, sequence coverage of most proteins was <35% (Fig. S1C). The coefficient of variation of replicates was calculated to evaluate the reliability of proteomic data. The result showed that proteins with 20% coefficient of

variation were more than 80% of identified proteins (Fig. 1D), suggesting that the data were reliable. In a PCA model based on 24 samples, control (0 h) and heat-treatment samples (3, 28 and 48 h) were clearly separated (Fig. S2). PCA1 accounted for 40.30% of the variability, while the PCA2 accounted for 38.30% of the variability (Fig. S2).

Compared with the control samples, a total of 1,591 DAPs were identified under heat stress in HT and HS. Among these DAPs, 157/156, 589/301, and 976/1,025 were significantly accumulated in HT/HS under heat stress at 3, 28, and 48 h, respectively (Fig. 2A). In short, there were more DAPs identified in HT compared with HS (Fig. 2A). Moreover, 457 and 394 specific DAPs were identified in HT and HS, respectively, while 740 common DAPs were found both in HT and HS (Fig. 2B).

Functional analysis of DAPs in response to heat stress

Of the 1,591 DAPs, 997 DAPs were categorized into 24 categories using COGs database. The largest group was general function prediction only, followed by post-translational modification, protein turnover, chaperones, translation, ribosomal structure and biogenesis, energy production and conversion, carbohydrate transport and metabolism, and amino acid transport and metabolism (Fig. 3A and Table S1).

To further characterize all heat-responsive proteins, these DAPs identified in HT and HS were subjected to GO analysis. Of the 1,591 DAPs identified in HT and HS under heat stress, 1,453 proteins were annotated into three groups as cellular component (CC), molecular function (MF), and biological process (BP). GO analysis showed that cell part, cell, and organelle were the major CC terms; catalytic activity, binding, structural molecule activity and transporter activity were the dominant MF terms; and cellular process, signaling, metabolic process, single-organism process and response to stimulus were the most dominant BP terms (Fig. 4).

To further investigate the major metabolic pathways responding to heat stress, KEGG enrichment analysis was carried out with heat-responsive proteins from HT and HS. The result showed that DAPs were matched to 99 KEGG pathways in HT and 106 KEGG pathways in HS. Heat stress could affect protein processing in photosynthesis, endoplasmic reticulum, terpenoid backbone biosynthesis, porphyrin and chlorophyll metabolism, carbon fixation in photosynthetic organisms, and pentose phosphate pathways in both two genotypes (Fig. 5). Furthermore, KEGG metabolic pathways, including pyrimidine metabolism, galactose metabolism and amino sugar and nucleotide sugar metabolism, were highly enriched in HT, whereas other pathways, such as glyoxylate and dicarboxylate metabolism, phenylpropanoid biosynthesis, carbon metabolism, nitrogen metabolism, biosynthesis of amino acids, glutathione metabolism, phenylalanine metabolism, and glycine, serine and threonine metabolism, were considerably enriched in HS ($P < 0.05$, Table S2 and S3).

Antioxidants in response to heat stress

In response to heat stress, antioxidant enzymes (GST, POD, SOD, DHAR, APX and CAT) were differentially accumulated according to proteomic data. Under heat stress, most GSTs were significantly increased with the increase of treatment time (Fig. 6A). Some of them were accumulated in both two genotypes, while three of them were suppressed in response to heat stress. Among six identified PODs, POD 12-like and 3-like were significantly increased in HT, while POD p7-like, 17-like, and 51-like were decreased in HT and HS (Fig. 6A). Under heat stress, two SODs were identified in response to heat stress, and both of them were greatly increased in HT, while there were no obvious changes in HS (Fig. 6A). One DHAR and one CAT were significantly increased in HT, while they were slightly increased in HS (Fig. 6A). In addition, most APXs were increased in HT, while they were only there APXs were increased in HS.

Furthermore, in order to further compare the differences of antioxidant capacity between HT and HS, the content of GSH and the activities of GST, POD, SOD, APX, and CAT were determined at 0 and 48 h after the heat treatment. Fig. 7A shows that the content of GSH was increased in both two genotypes under heat stress. However, its content was higher in HT compared with HS. The activities of GST and POD were higher in HT compared with HS after 48 h of heat stress (Fig. 7B, C). Compared with the control samples, the activities of SOD, APX, and CAT were significantly increased in HT. However, there were no obvious changes in HS (Fig. 7D, E, F).

Specific DAPs of HT in response to heat stress

To further investigate the mechanism underlying the higher heat tolerance in HT, the functions of 457 specific DAPs were studied based on GO analysis (Table S4). In short, these DAPs were mainly enriched in protein autophosphorylation, hexokinase-dependent signaling, and regulation of ROS metabolic process. Among these proteins, 95 proteins were up-regulated at each treatment time point under heat stress, including glutamate decarboxylase (Capana00g003786), 26s protease regulatory subunit 8 homolog a (Capana01g002147), sHSP (18.2 kDa class I heat shock protein), universal stress protein a-like protein, and so on (Table S4). Moreover, some DAPs related to signal transduction were also identified in HT, such as calcium-dependent protein kinase (Capana04g002552), serine threonine-protein kinase ht1 (Capana07g001488), and atp-dependent 6-phosphofructokinase 6 (Capana07g001111).

Discussion

Heat stress seriously restricts the quality and yield of pepper. Therefore, it is necessary to uncover the mechanisms underlying the heat stress response for breeding heat-tolerant pepper varieties. In the present study, we investigated the phenotypic and physiological changes of two pepper seedlings under heat stress. Furthermore, the comparative analysis of heat-induced proteomic changes was carried out to obtain a global view of heat stress responses in pepper at the protein level. As a result, we identified 1,591 heat-responsive proteins in pepper leaves, and the functions of these proteins were annotated using some public databases. The functions of these proteins and main related metabolic processes were further discussed below.

DAPs involved in signal transduction

When plants are subjected to abiotic stress, the complex signal networks are regulated to cope with the unavoidable environmental conditions and keep growth and development [21, 22]. It is well known that plant protein kinases are involved in signal transduction by mediating the response of plant cell to external environment under various abiotic stress [22]. Overexpression of rice serine- threonine protein kinase *SAPK4* resulted in improved the ability of salt stress acclimation [23]. Receptor-like protein kinases are also believed to play an important role in abiotic stress [24], and overexpression of receptor-like kinase *ER* in rice and tomato improves thermotolerance [25]. In the current research, four serine-threonine protein kinases and one receptor-like protein kinase were increased in HT compared with HS (Table S5). Meanwhile, calmodulin plays a critical role in heat shock signal transduction, and the expressions of calmodulin genes are up-regulated after heat treatment [26]. Herein, calmodulin and calmodulin-7 were up-regulated in HT and HS (Table S5). These findings suggest that protein kinase and calmodulin functioned as positive regulators to improve the heat tolerance of HT seedlings. In addition, 14-3-3 proteins were also identified as cold/salt/drought-responsive proteins, respectively [22, 27, 28]. Moreover, 14-3-3 protein 4, 14-3-3 protein 9, and 14-3-3 protein 10 were more accumulated in HT compared with HS under heat stress. This finding indicated that 14-3-3 proteins executed an important function in resistance to heat stress in pepper seedlings. The more accumulation of these proteins was involved in signal transduction, which might contribute to the heat tolerance of HT seedlings. However, the deeper regulatory mechanism remains to be further investigated.

DAPs involved in photosynthesis

Photosynthesis is the most basic physiological process for plant growth and development [20, 29]. Owing to its complex molecular mechanisms and demand for multiple enzymes, photosynthesis is often cited as the most heat-sensitive physiological process [12, 30]. Previous studies have found that the photosynthesis is inhibited by heat stress [29, 31]. It is well documented that PSII is the most heat-sensitive component of photosynthesis [29]. Our study suggested that eight proteins related to PSII were down-regulated, such as two *PsbA*, *PsbB*, *PsbC*, *PsbD*, *PsbS*, *PsbR*, and *Psb28* (Table S5). Meanwhile, nine proteins (three *PsaA*, *PsaB*, two *PsaE*, *PsaF*, *PsaH*, *PsaO*) involved in PSI were differentially accumulated under heat stress (Table S5). However, some of them were up-regulated, and some of them were slightly down-regulated. Furthermore, DAPs related to cytochrome b6/f complex, such as *PetB*, *PetA*, *PetH*, *PetC*, and *PetE*, photosynthetic electron transport including *PetE* and *PetH*, and F-type ATPase enzyme with multiple subunits (a, alpha, two b, beta, delta, epsilon and gamma), were also down-regulated. These finding indicated that heat stress could dramatically decrease PSII compared with PSI. Overall, the photosynthesis of HT and HS was both decreased under heat stress. However, HT could maintain relatively high photosynthesis compared with HS.

DAPs involved in carbohydrate metabolism

Glycolysis, tricarboxylic acid (TCA) cycle, and pentose phosphate pathway are important components of carbohydrate metabolism. The glycolysis and TCA cycle are inhibited with the increase of temperature [13]. Table S5 shows that there were 41 proteins involved in glycolysis, and most of them were down-

regulated after heat stress, such as glucose-6-phosphate isomerase, hexokinase, pyruvate dehydrogenase, and so on. Interestingly, HT could maintain a higher glycolysis under heat stress owing to the relatively high activities of related enzymes compared with HS. In addition, nine proteins were identified to be involved in TCA cycle. The TCA cycle can provide essential energy and precursors for the synthesis of substances, and it can also be affected by abiotic stress [32, 33]. Pyruvate dehydrogenase complex (PDC) is a type of multienzyme complex located in the mitochondrial matrix. PDC, involved in the irreversible oxidation of pyruvate, plays an important role in energy metabolism of mitochondrial respiratory chain [34]. Here, we found that the abundance of pyruvate dehydrogenase e1 component subunit beta (PDHB) was reduced in HT under heat stress, while only little change was found in HS (Table S5). This finding indicated that the activity of TCA cycle was more significantly decreased in HT compared with HS. It has been reported that the reduced consumption of organic acids caused by the decreased activity of TCA cycle may contribute to the synthesis of compounds to cope with stressful condition [35, 36]. Therefore, reducing the activity of TCA cycle in HT might be conducive to the synthesis of osmosis substances to cope with heat stress. The pentose phosphate pathway (PPP) is also a major source of reductant and intermediates for substance biosynthesis [37]. In our present study, the key members of the PPP, including ribose-5-phosphate isomerase, ribulose-phosphate 3, ATP-dependent 6-phosphofructokinase, 6-phosphogluconate decarboxylating 3, and others, were reduced in both HT and HS (Table S5). However, the activities of related enzymes in HS were lower compared with HT, suggesting that HS suffered more from oxidative damage caused by heat stress.

DAPs involved in stress defense

In the present study, 93 DAPs were identified to be involved in the process of heat stress defense, including enzymatic and non-enzymatic antioxidants, and molecular chaperones.

Heat stress can disturb the balance between generation and scavenging of ROS and resulting in damaged membrane system and other cellular components. To cope with the peroxidation damages, plants have developed an efficient enzymatic and non-enzymatic antioxidant system [9]. It is well established that GSH is one of the important metabolites in plant cells and organelles, which functions as the intracellular defense against oxidative damage induced by ROS. In this study, compared with the corresponding control group, the content of GSH was increased in HT, while it was slightly increased in HS (Fig. 6B), suggesting that HT was subjected to minor damage from ROS. It has also been suggested that the up-regulation of antioxidant enzymes can regulate the ROS level and protect the plant cells from oxidative damage when they are under heat stress [13, 20, 38, 39]. In the present study, some antioxidant enzymes were identified, including GST, POD, SOD, APX, and CAT (Fig. 6A). Moreover, we found that some thioredoxins (TRXs) were accumulated under heat stress, which can function in the process of plant cell against oxidative damage (Table S5) [40, 41]. Interestingly, HT showed a higher ability to adapt to high temperature with more accumulation of antioxidants compared with HS, which was consistent with the minor MDA content in HT compared with HS under heat stress.

Molecular chaperones are crucial proteins, which can prevent the synthesis of misfolded proteins and maintain the intracellular homeostasis in plant under suitable or stressful condition [14, 42, 43]. HSPs are one of chaperones, which are induced when plants suffer from heat stress [44]. Over-expression of *CaHSP70a* contributes to the higher heat tolerance, and CaHSP70a protein is also accumulated in pepper under heat stress [45]. In addition, HSP90 [46] and sHSPs [47] are also involved in the response of heat stress. In our present study, 10 HSP70s, five HSP90s, and 11 sHSPs were up-regulated after heat stress (Table S5). Previous studies indicated that ubiquitin-conjugating enzyme and other chaperone protein help plants cope with the adverse environment [48]. The up-regulated of these proteins might indicate that they functioned as the molecular chaperones and played a critical role in response to heat stress in pepper.

Conclusions

In the present study, we studied the physiological and proteomic changes of HT and HS seedlings to uncover the molecular mechanisms in heat tolerance. A total of 1,591 DAPs were identified as heat-responsive proteins in pepper seedlings. The bioinformatic analysis revealed that these DAPs were involved in energy production and conversion; carbohydrate transport and metabolism; catalytic activity; structural molecule activity and transporter activity; and response to stimulus. The KEGG enrichment analysis showed that photosynthesis, endoplasmic reticulum, terpenoid backbone biosynthesis, porphyrin and chlorophyll metabolism, carbon fixation in photosynthetic organisms, and pentose phosphate pathways were significantly related to the response to heat stress. Furthermore, some proteins were confirmed to play a critical role in protein folding, osmoregulation and ROS scavenging. A model of pepper response to heat stress was generated based on physiological and proteomic changes (Fig. 8). Collectively, our findings provided valuable insights into the molecular mechanisms underlying the heat tolerance of pepper and might be useful reference for the breeding of new pepper variety with heat resistance.

Methods

Plant materials and heat treatment

Two pepper cultivars 17CL30 (HT) and 05S180 (HS) were obtained from Vegetable Institution of Hunan Academy of Agricultural Science. The seed germination and seedling management of pepper were carried out as previously described [49]. Six-leaf stage seedlings of HT and HS cultivars were subjected to heat stress at 40°C, and the leaves were harvested at 0, 3, 28 and 48 h for physiological assays and proteomic analysis. Three biological replicates were performed for each treatment.

Measurement of physiological indices

Control (0 h) and 48-h heat-treated samples were used for the measurement of physiological indices. The relative water content (RWC) of leaves was determined as previously described [50]. The contents of

proline, soluble sugar, malondialdehyde (MDA), reduced GSH, POD, GST, SOD and CAT were determined using commercially available kits (Nanjing Jiancheng Bioengineering Institute, China) according to the manufacturer's instructions.

Protein extraction, iTRAQ labeling, and data analysis

Total proteins were extracted from 24 samples using plant total protein extraction kit (Beijing Bio-Fly Bioscience Co., Ltd., China) according to the manufacturer's instructions. The iTRAQ labeling and analysis were performed at Biomarker (Beijing). The iTRAQ-labeled samples were analyzed using Q Exactive HF-X (Thermo Scientific). The raw file was processed using the Mascot engine embedded into the Proteome Discoverer 2.1 against the pepper protein database (Zunla-1 version 2). The principal component analysis (PCA) was used to evaluate the relationship among 24 samples. T-test was used to compare the differential expressions of proteins. Proteins with fold-changes >1.2 or <0.83 and P-value < 0.05 were considered as DAPs.

Bioinformatic analysis

The functional classification of DAPs was performed by the Clusters of Orthologous Groups (COGs) protein database (<https://www.ncbi.nlm.nih.gov/COG/>). Gene Ontology (GO) database (<http://geneontology.org/>) was used for functional annotation of DAPs. The prediction of metabolic pathways was carried out with Kyoto Encyclopedia of Genes and Genomes (KEGG) database (<https://www.genome.jp/kegg>).

Abbreviations

APX, ascorbate peroxidase; BP, biological process; CAT, catalase; CC, cellular component; COGs, Clusters of Orthologous Groups; DAPs, differentially accumulated proteins; DHAR, dehydroascorbate reductase; GO, Gene Ontology; GSH, glutathione; GST, glutathione S-transferase; HS, heat-sensitive; HSPs, heat shock proteins; HT, heat-tolerant; IPCC, Intergovernmental Panel on Climate Change; KEGG, Kyoto Encyclopedia of Genes and Genomes; MDA, malondialdehyde; MF, molecular function; PCA, principal component analysis; PDC, Pyruvate dehydrogenase complex; PDHB, pyruvate dehydrogenase e1 component subunit beta; POD, peroxidase; PPP, pentose phosphate pathway; ROS, reactive oxygen species; RWC, relative water content; sHSPs, small HSPs; SOD, superoxide dismutase; TCA, tricarboxylic acid; TRXs, thioredoxins.

Declarations

Acknowledgement

Not applicable.

Author contributions

Jing Wang, Xuexiao Zou, Wenchao Chen, Chunhai Jiao, and Niran Juntawong conceived the study. Jing Wang, Chengliang Liang, Jingshuang Song, Fangjun, Tan, Xilu Zhang and Fei Wang performed the experiments. Jing Wang, Junheng Lv, Zhoubin Liu, Yuhua Liu, and Jingshuang Song performed the data analysis. Jing Wang wrote the manuscript. All authors have read and approved the final manuscript.

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Availability of data and materials

The MS data in the current study have been deposited to the ProteomeXchange with the dataset identifier PXD019556.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing financial interests.

References

1. Guo M, Yin YX, Ji JJ, Ma BP, Lu MH, Gong ZH: **Cloning and expression analysis of heat-shock transcription factor gene CaHsfA2 from pepper (*Capsicum annuum* L.)**. *Gen Mol Res* 2014, **13**(1):1865-1875.
2. Wahid A, Gelani S, Ashraf M, Foolad M: **Heat tolerance in plants: An overview**. *Environ Exp Bot* 2007, **61**(3):199-223.
3. Lesk C, Rowhani P, Ramankutty N: **Influence of extreme weather disasters on global crop production**. *Nature* 2016, **529**(7584):84-87.
4. Gajanayake B, Trader BW, Reddy KR, Harkess RL: **Screening Ornamental Pepper Cultivars for Temperature Tolerance Using Pollen and Physiological Parameters**. *HortScience* 2011, **46**(6):878-884.
5. Ohama N, Sato H, Shinozaki K, Yamaguchi-Shinozaki K: **Transcriptional Regulatory Network of Plant Heat Stress Response**. *Trends Plant Sci* 2017, **22**(1):53-65.

6. Vu LD, Gevaert K, De Smet I: **Feeling the Heat: Searching for Plant Thermosensors.** *Trends Plant Sci* 2019, **24**(3):210-219.
7. Ding Y, Shi Y, Yang S: **Molecular Regulation of Plant Responses to Environmental Temperatures.** *Molecular Plant* 2020, **13**(4):544-564.
8. Gong Z, Xiong L, Shi H, Yang S, Herrera-Estrella LR, Xu G, Chao D-Y, Li J, Wang P-Y, Qin F *et al.* **Plant abiotic stress response and nutrient use efficiency.** *Science China Life Sciences* 2020.
9. Gill SS, Tuteja N: **Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants.** *Plant Physiol Biochem* 2010, **48**(12):909-930.
10. Zhang H, Yang J, Li W, Chen Y, Lu H, Zhao S, Li D, Wei M, Li C: **PuHSA4a Enhances Tolerance To Excess Zinc by Regulating Reactive Oxygen Species Production and Root Development in Populus.** *Plant Physiol* 2019, **180**(4):2254-2271.
11. Smirnoff N: **Ascorbic acid: metabolism and functions of a multi-faceted molecule.** *Curr Opin Plant Biol* 2003, **3**:229-235.
12. Shi J, Chen Y, Xu Y, Ji D, Chen C, Xie C: **Differential Proteomic Analysis by iTRAQ Reveals the Mechanism of *Pyropia haitanensis* Responding to High Temperature Stress.** *Sci Rep* 2017, **7**:44734.
13. Wang L, Ma KB, Lu ZG, Ren SX, Jiang HR, Cui JW, Chen G, Teng NJ, Lam HM, Jin B: **Differential physiological, transcriptomic and metabolomic responses of *Arabidopsis* leaves under prolonged warming and heat shock.** *BMC Plant Biol* 2020, **20**(1):86.
14. Wang W, Vinocur B, Shoseyov O, Altman A: **Role of plant heat-shock proteins and molecular chaperones in the abiotic stress response.** *Trends Plant Sci* 2004, **9**(5):244-252.
15. Wang J, Gao X, Dong J, Tian X, Wang J, Palta JA, Xu S, Fang Y, Wang Z: **Over-Expression of the Heat-Responsive Wheat Gene TaHSP23.9 in Transgenic *Arabidopsis* Conferred Tolerance to Heat and Salt Stress.** *Front Plant Sci* 2020, **11**:243.
16. Tian X, Liu Y, Huang Z, Duan H, Tong J, He X, Gu W, Ma H, Xiao L: **Comparative proteomic analysis of seedling leaves of cold-tolerant and -sensitive spring soybean cultivars.** *Mol Biol Rep* 2015, **42**(3):581-601.
17. Cao Y, Luo Q, Tian Y, Meng F: **Physiological and proteomic analyses of the drought stress response in *Amygdalus Mira* (Koehne) Yu et Lu roots.** *BMC Plant Biol* 2017, **17**(1):53.
18. Zhan Y, Wu Q, Chen Y, Tang M, Sun C, Sun J, Yu C: **Comparative proteomic analysis of okra (*Abelmoschus esculentus* L.) seedlings under salt stress.** *BMC Genomics* 2019, **20**(1):381.
19. Han F, Chen H, Li XJ, Yang MF, Liu GS, Shen SH: **A comparative proteomic analysis of rice seedlings under various high-temperature stresses.** *Biochim Biophys Acta* 2009, **1794**(11):1625-1634.
20. Lu Y, Li R, Wang R, Wang X, Zheng W, Sun Q, Tong S, Dai S, Xu S: **Comparative Proteomic Analysis of Flag Leaves Reveals New Insight into Wheat Heat Adaptation.** *Frontiers in Plant Science* 2017, **8**:1086.
21. Kaur N, Gupta AK: **Signal transduction pathways under abiotic stress.** *Curr Sci* 2005, **88**(11).

22. Abreu IA, Farinha AP, Negrao S, Goncalves N, Fonseca C, Rodrigues M, Batista R, Saibo NJ, Oliveira MM: **Coping with abiotic stress: proteome changes for crop improvement.** *J Proteomics* 2013, **93**:145-168.
23. Diedhiou CJ, Popova OV, Dietz KJ, Golldack D: **The SNF1-type serine-threonine protein kinase SAPK4 regulates stress-responsive gene expression in rice.** *BMC Plant Biol* 2008, **8**:49.
24. Jung CG, Hwang SG, Park YC, Park HM, Kim DS, Park DH, Jang CS: **Molecular characterization of the cold- and heat-induced Arabidopsis PXL1 gene and its potential role in transduction pathways under temperature fluctuations.** *J Plant Physiol* 2015, **176**:138-146.
25. Shen H, Zhong X, Zhao F, Wang Y, Yan B, Li Q, Chen G, Mao B, Wang J, Li Y *et al*: **Overexpression of receptor-like kinase ERECTA improves thermotolerance in rice and tomato.** *Nat Biotechnol* 2015, **33**(9):996-1003.
26. Al-Quraan NA, Locy RD, Singh NK: **Expression of calmodulin genes in wild type and calmodulin mutants of Arabidopsis thaliana under heat stress.** *Plant Physiol Biochem* 2010, **48**(8):697-702.
27. Zhang S, Zhang L, Chai Y, Wang F, Li Y, Su L, Zhao Z: **Physiology and proteomics research on the leaves of ancient Platycladus orientalis (L.) during winter.** *J Proteomics* 2015, **126**:263-278.
28. Yang Z, Wang C, Xue Y, Liu X, Chen S, Song C, Yang Y, Guo Y: **Calcium-activated 14-3-3 proteins as a molecular switch in salt stress tolerance.** *Nat Commun* 2019, **10**(1):1199.
29. Ashraf M, Harris PJC: **Photosynthesis under stressful environments: An overview.** *Photosynthetica* 2013, **51**(2):163-190.
30. Centritto M, Brillì F, Fodale R, Loreto F: **Different sensitivity of isoprene emission, respiration and photosynthesis to high growth temperature coupled with drought stress in black poplar (Populus nigra) saplings.** *Tree Physiol* 2011, **31**(3):275-286.
31. Li T, Xu X, Li Y, Wang H, Li Z, Li Z: **Comparative transcriptome analysis reveals differential transcription in heat-susceptible and heat-tolerant pepper (Capsicum annum L.) cultivars under heat stress.** *J Plant Biol* 2015, **58**(6):411-424.
32. Williams TC, Poolman MG, Howden AJ, Schwarzlander M, Fell DA, Ratcliffe RG, Sweetlove LJ: **A genome-scale metabolic model accurately predicts fluxes in central carbon metabolism under stress conditions.** *Plant Physiol* 2010, **154**(1):311-323.
33. Zhang Y, Beard KFM, Swart C, Bergmann S, Krahnert I, Nikoloski Z, Graf A, Ratcliffe RG, Sweetlove LJ, Fernie AR *et al*: **Protein-protein interactions and metabolite channelling in the plant tricarboxylic acid cycle.** *Nat Commun* 2017, **8**:15212.
34. Yu H, Du X, Zhang F, Zhang F, Hu Y, Liu S, Jiang X, Wang G, Liu D: **A mutation in the E2 subunit of the mitochondrial pyruvate dehydrogenase complex in Arabidopsis reduces plant organ size and enhances the accumulation of amino acids and intermediate products of the TCA cycle.** *Planta* 2012, **236**(2):387-399.
35. Widodo, Patterson JH, Newbigin E, Tester M, Bacic A, Roessner U: **Metabolic responses to salt stress of barley (Hordeum vulgare L.) cultivars, Sahara and Clipper, which differ in salinity tolerance.** *J Exp Bot* 2009, **60**(14):4089-4103.

36. Kiani-Pouya A, Roessner U, Jayasinghe NS, Lutz A, Rupasinghe T, Bazihizina N, Bohm J, Alharbi S, Hedrich R, Shabala S: **Epidermal bladder cells confer salinity stress tolerance in the halophyte quinoa and *Atriplex* species.** *Plant Cell Environ* 2017, **40**(9):1900-1915.
37. Kruger NJ, von Schaewen A: **The oxidative pentose phosphate pathway: structure and organisation.** *Curr Opin Plant Biol* 2003, **6**(3):236-246.
38. Das K, Roychoudhury A: **Reactive oxygen species (ROS) and response of antioxidants as ROS-scavengers during environmental stress in plants.** *Frontiers in Environmental Science* 2014, **2**.
39. Xu Y, Yuan Y, Du N, Wang Y, Shu S, Sun J, Guo S: **Proteomic analysis of heat stress resistance of cucumber leaves when grafted onto *Momordica* rootstock.** *Hortic Res* 2018, **5**:53.
40. Laloi C, Mestres-Ortega D, Marco Y, Meyer Y, Reichheld J-P: **The *Arabidopsis* Cytosolic Thioredoxin h5 Gene Induction by Oxidative Stress and Its W-Box-Mediated Response to Pathogen Elicitor.** *Plant Physiol* 2004, **134**(3):1006-1016.
41. Vieira Dos Santos C, Rey P: **Plant thioredoxins are key actors in the oxidative stress response.** *Trends Plant Sci* 2006, **11**(7):329-334.
42. Kang H, Park SJ, Kwak KJ: **Plant RNA chaperones in stress response.** *Trends Plant Sci* 2013, **18**(2):100-106.
43. Voth W, Jakob U: **Stress-Activated Chaperones: A First Line of Defense.** *Trends Biochem Sci* 2017, **42**(11):899-913.
44. Merret R, Carpentier MC, Favory JJ, Picart C, Descombin J, Bousquet-Antonelli C, Tillard P, Lejay L, Deragon JM, Charng YY: **Heat Shock Protein HSP101 Affects the Release of Ribosomal Protein mRNAs for Recovery after Heat Shock.** *Plant Physiol* 2017, **174**(2):1216-1225.
45. Kim NH, Hwang BK: **Pepper Heat Shock Protein 70a Interacts with the Type III Effector AvrBsT and Triggers Plant Cell Death and Immunity.** *Plant Physiol* 2015, **167**(2):307-322.
46. Guo X, Ronhovde K, Yuan L, Yao B, Soundararajan MP, Elthon T, Zhang C, Holding DR: **Pyrophosphate-dependent fructose-6-phosphate 1-phosphotransferase induction and attenuation of Hsp gene expression during endosperm modification in quality protein maize.** *Plant Physiol* 2012, **158**(2):917-929.
47. Waters ER, Vierling E: **Plant small heat shock proteins - evolutionary and functional diversity.** *New Phytol* 2020.
48. Sharma B, Bhatt TK: **Genome-wide identification and expression analysis of E2 ubiquitin-conjugating enzymes in tomato.** *Sci Rep* 2017, **7**(1):8613.
49. Wang J, Lv J, Liu Z, Liu Y, Song J, Ma Y, Ou L, Zhang X, Liang C, Wang F *et al*: **Integration of Transcriptomics and Metabolomics for Pepper (*Capsicum annuum* L.) in Response to Heat Stress.** *Int J Mol Sci* 2019, **20**(20).
50. Zegaoui Z, Planchais S, Cabassa C, Djebbar R, Belbachir OA, Carol P: **Variation in relative water content, proline accumulation and stress gene expression in two cowpea landraces under drought.** *J Plant Physiol* 2017, **218**:26-34.

Figures

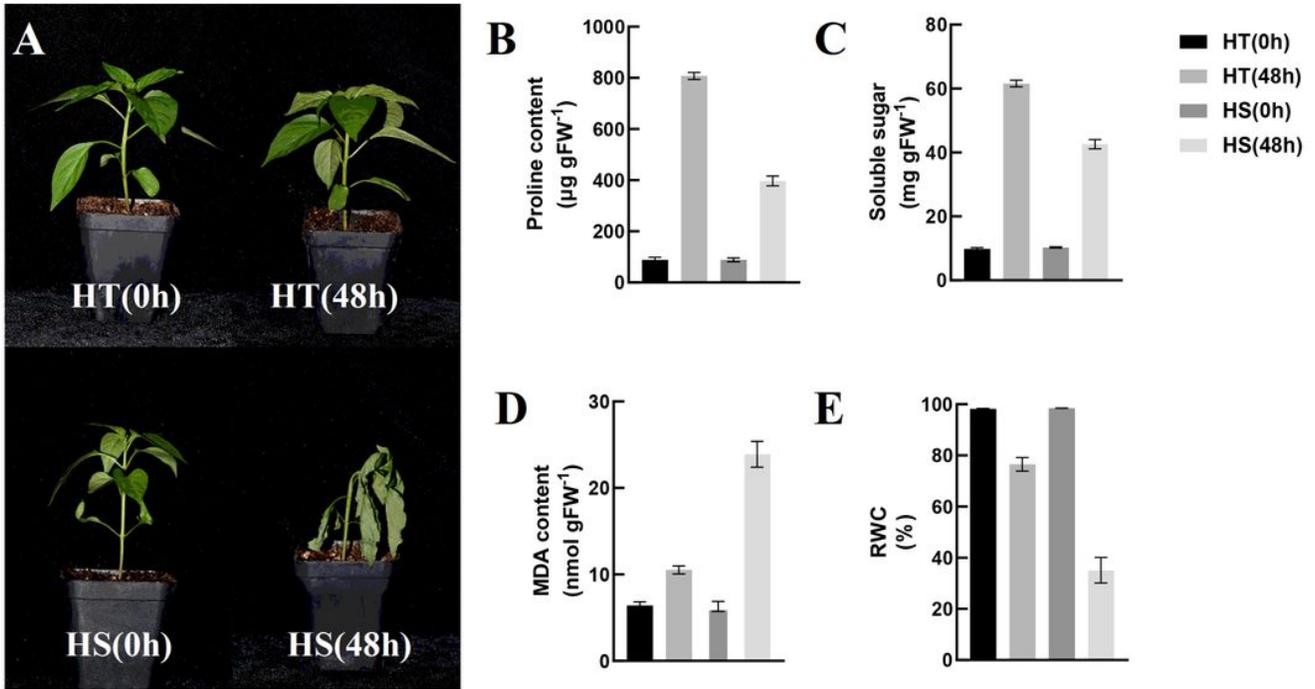


Figure 1

Phenotype and physiology responses of heat-tolerant (HT) and heat-sensitive (HS) pepper genotypes under heat stress. (A) The phenotype of HT and HS after 48 h heat stress. (B) Proline (C) soluble sugar (D) MDA and (E) RWC of HT and HS under 0, and 48 h heat stress.

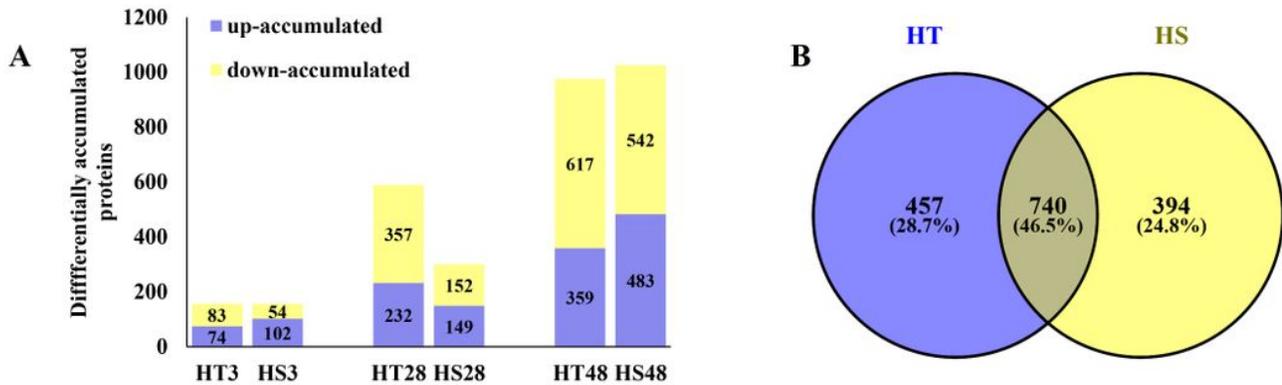


Figure 2

The differentially accumulated proteins (DAPs) in HT and HS genotypes under heat stress. (A) The number of the DAPs identified in HT and HS at different time point of heat stress. (B) Venn graph of the DAPs of HT and HS.

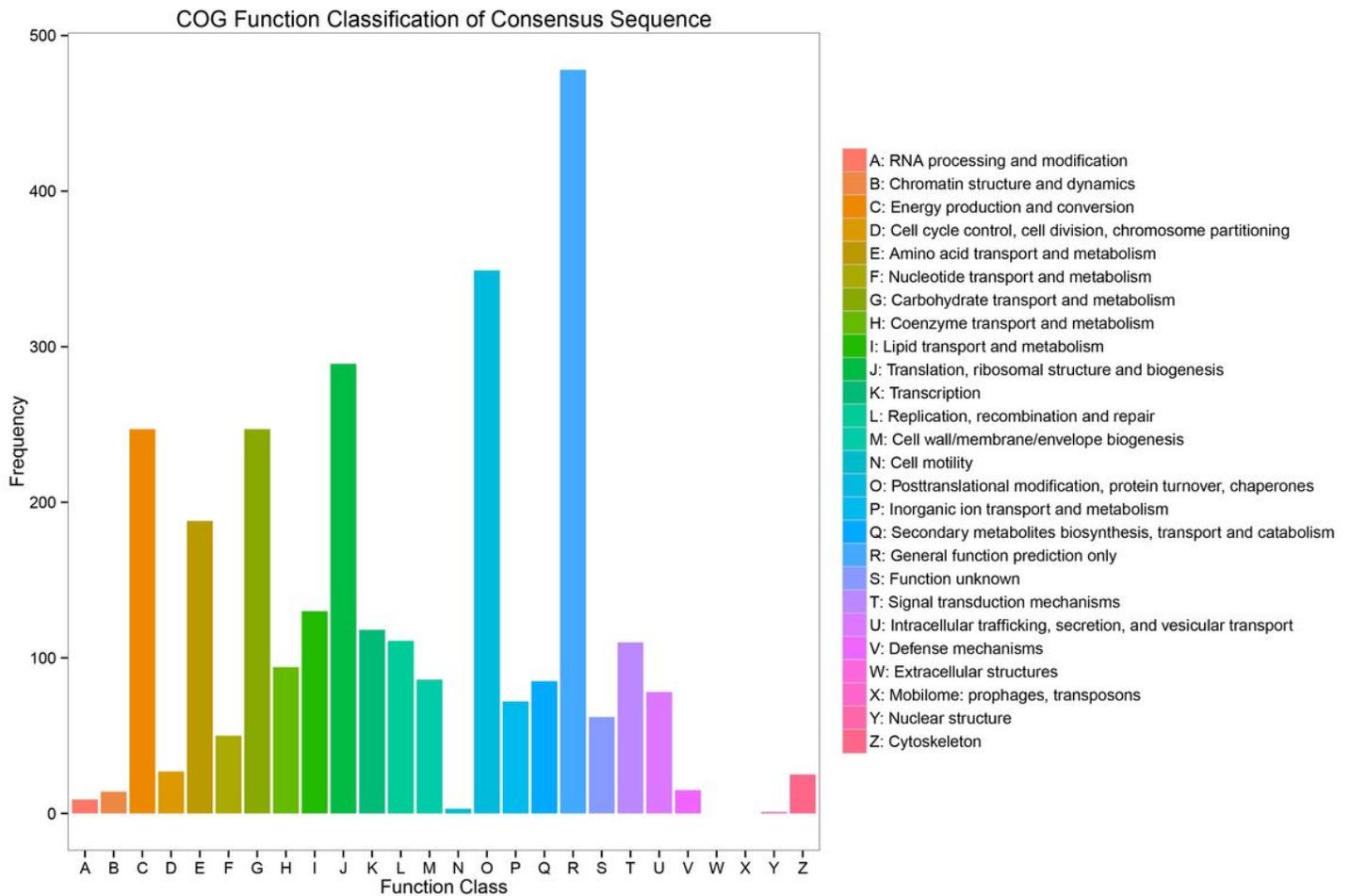


Figure 3

Clusters of Orthologous Groups (COG) annotation analysis of all heat responsive proteins in HT and HS.

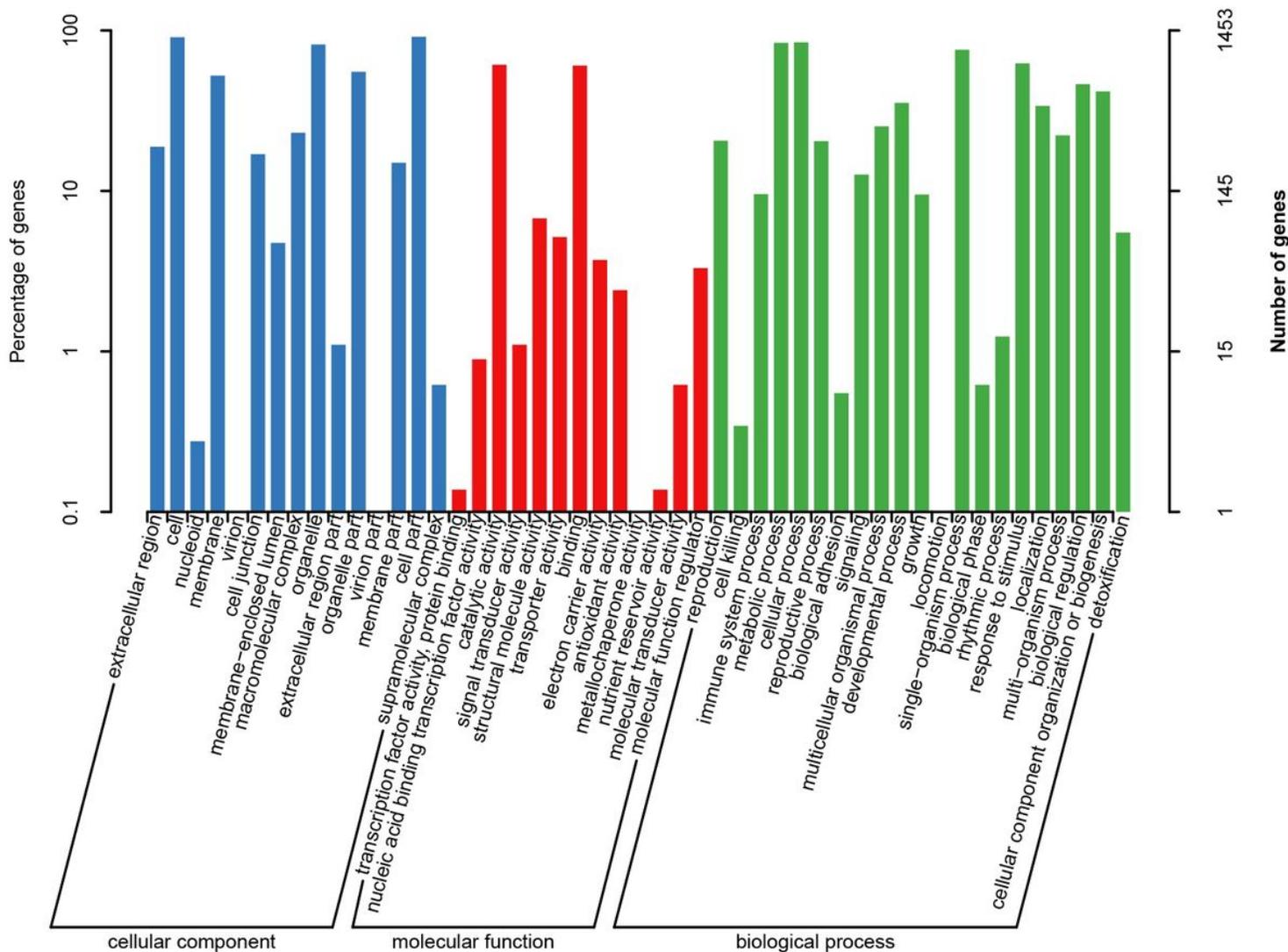


Figure 4

Gene Ontology (GO) annotation of all heat responsive proteins in HT and HS.

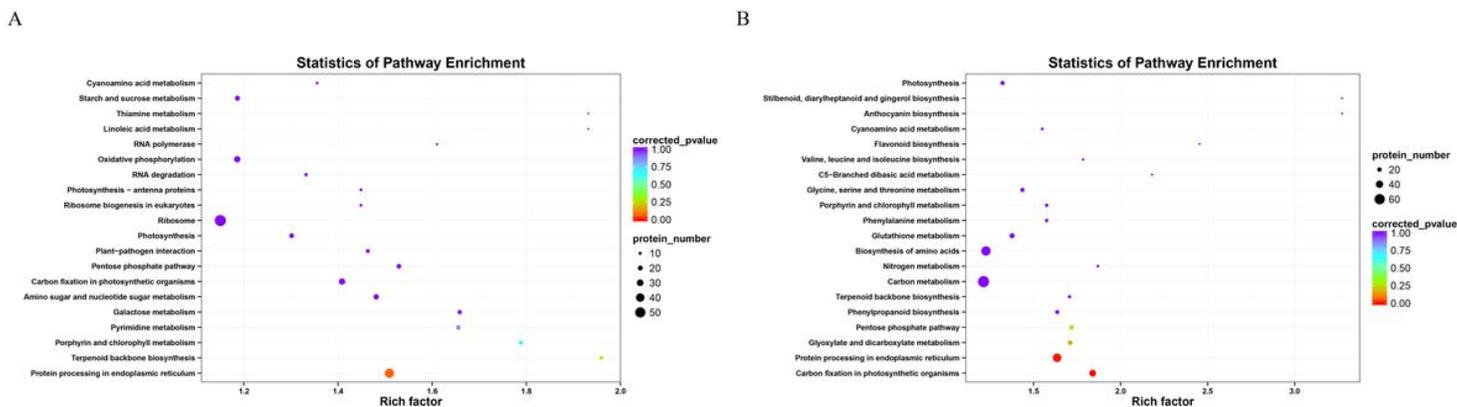


Figure 5

Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment of DAPs in HT (A) and HS (B).

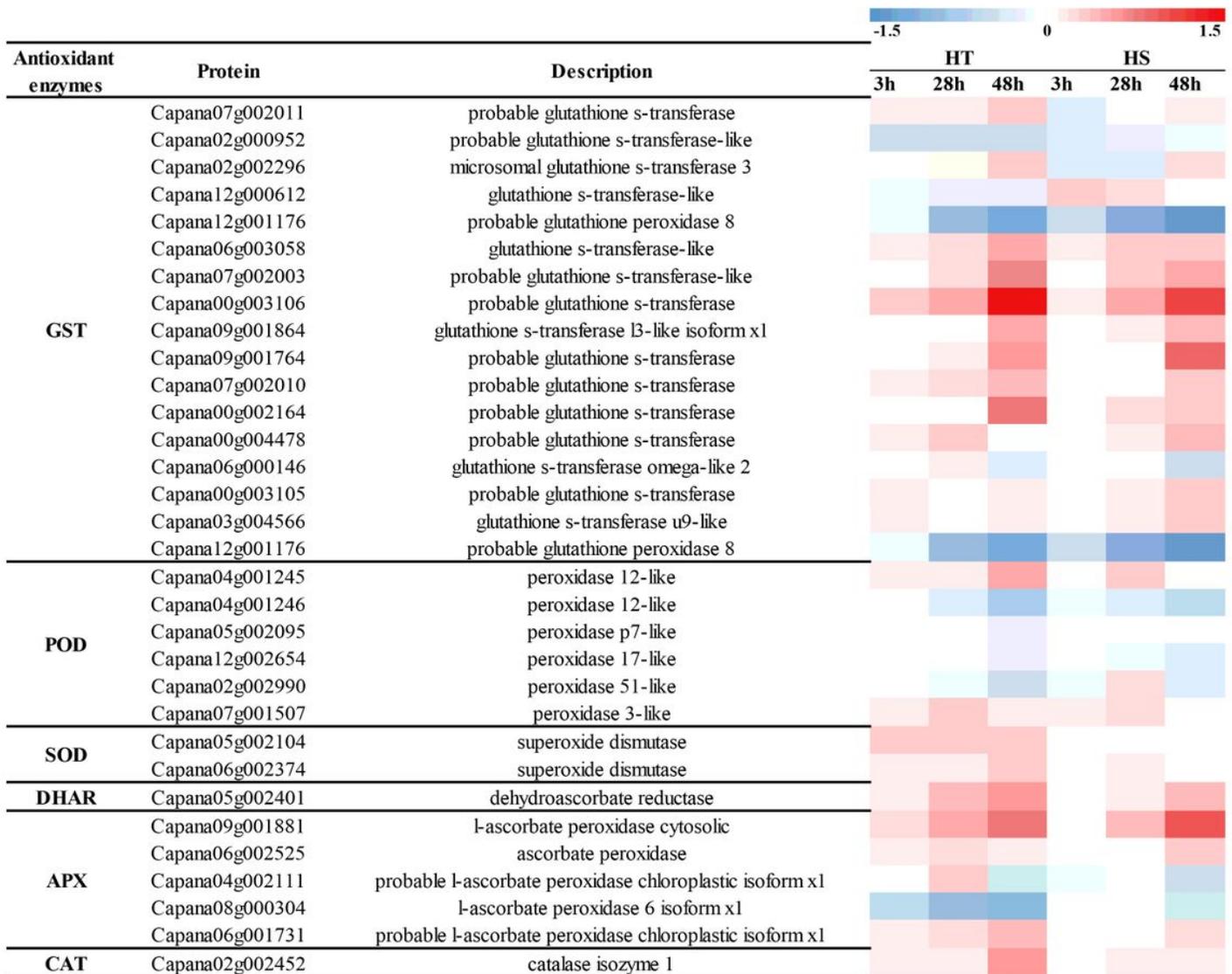


Figure 6

The change of enzymatic antioxidants of HT and HS in response to heat stress based on the proteomics data.

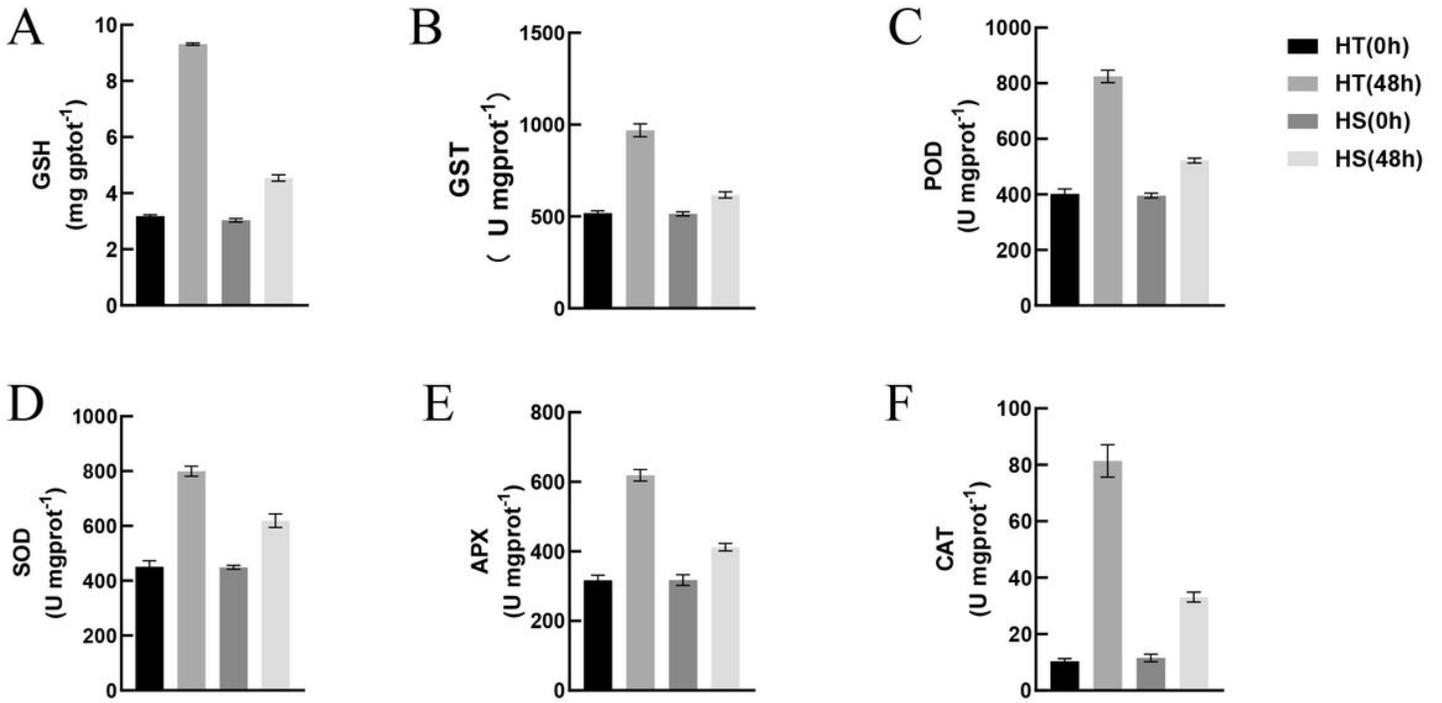


Figure 7

The content of GSH (A) and the activities of GST (B), POD (C), SOD (D), APX(E), and CAT(F) in HT and HS under 0, and 48 h heat stress using commercially available kits.

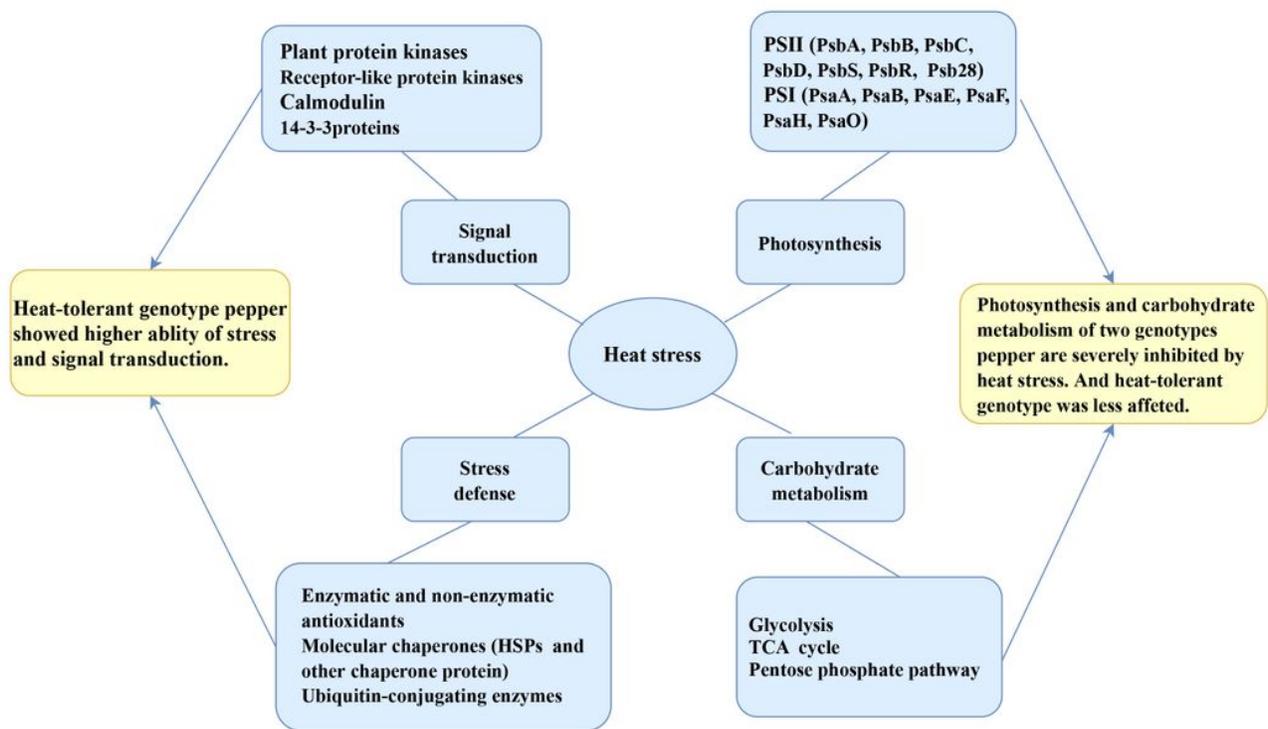


Figure 8

The model of pepper response to heat stress based on physiological and proteomic changes.

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