

Dendrites of Neocortical Pyramidal Neurons: The Key to Understand Intellectual Disability

Alberto Granato (✉ alberto.granato@unito.it)

University of Turin: Universita degli Studi di Torino <https://orcid.org/0000-0003-3755-1036>

Adalberto Merighi

University of Turin: Universita degli Studi di Torino <https://orcid.org/0000-0002-1140-3556>

Research Article

Keywords: cerebral cortex, Down syndrome, fetal alcohol, calcium, dendritic spike, apical dendrite.

Posted Date: May 10th, 2021

DOI: <https://doi.org/10.21203/rs.3.rs-472842/v1>

License:  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Version of Record: A version of this preprint was published at Cellular and Molecular Neurobiology on July 3rd, 2021. See the published version at <https://doi.org/10.1007/s10571-021-01123-1>.

Abstract

Pyramidal neurons (PNs) are the most abundant cells of the neocortex and display a vast dendritic tree, divided into basal and apical compartments. Morphological and functional anomalies of PN dendrites are at the basis of virtually all neurological and mental disorders, including intellectual disability. Here we provide evidence that the cognitive deficits observed in different types of intellectual disability might be sustained by different parts of the PN dendritic tree, or by a dysregulation of their interaction.

Introduction

The father of modern Neuroscience, Santiago Ramón y Cajal, postulated that neocortical pyramidal neurons (PNs) might play an outstanding role for the accomplishment of higher cognitive functions. He defined these cells, representing the vast majority of neocortical neurons, the “psychic cells” (Cajal 1917). Despite his feeling of shame, aimed at counterbalancing the “audacity of language”, after more than a century we have to recognize that, once again, the claim of the great Spanish scientist was substantially right. On the same line of reasoning, it can be assumed that a functional derangement of PNs is the pathophysiological basis of the cognitive deficit observed in intellectual disability (Granato and De Giorgio 2014).

Intellectual disability (ID), previously referred to as mental retardation, is classically defined as a neurodevelopmental disorder with IQ below 70, although more complex definitions, based on poor adaptive functioning and reduced daily life skills, have been provided by the DSM-5 (American Psychiatric Association 2013) and the American Association on Intellectual Developmental Disabilities (Shogren and Turnbull 2010).

The present point of view deals with the anomalies of neocortical PNs, as observed in experimental studies reproducing known causes of ID, as well as in the brains of affected human individuals. Given that dendritic alterations are considered among the most relevant anatomical and functional counterparts of ID (Kaufmann and Moser 2000) and owing to the great extension and geometric complexity of the dendritic arborizations of PNs, we shall focus primarily on dendritic anomalies. Some excellent reviews cover exhaustively the relationships between dendritic alterations and ID (Kaufmann and Moser 2000, Dierssen and Ramakers 2006, Quach et al. 2021). Our purpose is to provide mechanistic insights into how the disruption of PN dendritic function contributes to the genesis of ID, with a special emphasis on the role of the different parts of the dendritic tree.

Dendrites Of Pns

PNs represent the majority of neocortical neurons and are distributed in all cortical layers except layer 1. Among PNs, the thick-tufted cells are the most thoroughly studied, provide the cortical output directed to subcortical structures, and can be found in the deep part of layer 5 (layer 5b (Spruston 2008, Ramaswamy and Markram 2015)). Thick-tufted PNs are characterized by a prominent apical dendrite

spanning all the way to the pial surface and terminating with a branching apical tuft, whose radius often equals (or exceeds) that of the basal dendrites (Fig. 1). Other classes of PNs are those of layer 2/3 (providing cortico-cortical associative and callosal projections) and those bearing a slender apical dendrite, mainly located in the superficial part of layer 5 (Krieger et al. 2017), see also Fig. 1 in (Shepherd 2013). Modified PNs projecting to the thalamus and claustrum reside in layer 6 (Thomson 2010).

The prototypical PNs, the thick-tufted cells of layer 5, display geometric differences among the basal, oblique, and apical dendritic domains that are clearly recognizable at a first glance (see Fig. 1). In a seminal paper published exactly thirty years ago, Alan Larkman provided a rigorous formal description of such branching pattern differences, pointing out, for instance, that basal dendrites branching points are close to the cell body, whereas intermediate branches of the apical tuft are relatively longer than distal ones (Larkman 1991). It is well known that the dendritic geometry impinges upon the functional properties of neurons (Mainen and Sejnowski 1996). Furthermore, different parts of the dendritic tree of layer 5 PNs are potentially involved in the microcircuitry of different cortical layers. Therefore, it is conceivable that basal and apical dendrites contribute differently to the cortical machinery during physiological cognitive tasks.

Regarding the connections, the basal dendrites receive feedforward input from the receptor surfaces through the thalamic relay. The ascending input is thought to be conveyed indirectly to the dendrites of layer 5 through the canonical cortical circuit (thalamus → layer 4 → layer 2/3 → layer 5 (Thomson and Morris 2002, Lübke and Feldmeyer 2007)). However, layer 5 neurons can be also contacted directly by thalamic afferents (Meyer et al. 2010, Constantinople and Bruno 2013). Conversely, most of the apical tuft of PNs lies in layer 1, where it receives feedback connections from higher cortical areas, conveying input related to attention, context, expectations (Coogan and Burkhalter 1990, Caulker 1995, Caulker et al. 1998). Together, the basal and apical dendritic arborization are in the ideal position to integrate bottom-up and top-down streams of information. The refinement of dendritic recording (Davie et al. 2006) made it possible to ascertain that action potentials can back propagate through the apical dendrite of PNs (Stuart and Sakmann 1994) and to shed light on the interplay between basal and apical dendrites. Using multiple patch-clamp recordings from the soma and the apical dendrite of layer 5 PNs, it has been demonstrated that the coincidence of a back propagated action potential generated at the soma and of an apical dendritic input is able to generate a dendritic calcium spike that, in turn, elicits a burst of somatic spikes (Larkum et al. 1999). This mechanism, originally called backpropagation-activated calcium spike firing (BAC firing) has been considered the electrophysiological basis of the top-down / bottom-up integration operated by a single PN (Larkum 2013). This idea was widened by Bill Phillips and Matthew Larkum, leading to the concept of “apical amplification”, the mean by which the information coming from the external world (bottom-up) is modulated by context-sensitive (top-down) information (Phillips et al. 2016, Phillips 2017). A role of the apical dendrite in cognition and consciousness has been also postulated by (LaBerge 2006) . Recently, it has been demonstrated that apical dendritic potentials can gate sensory perception and that such a modulation depends on contextual information (Takahashi et al. 2020). A dysregulation of context-modulated sensory perception and learning abilities can represent a prominent feature of ID (Alevriadou et al. 2004, Carr et al. 2010, Murray et al. 2019).

As to the distribution of ion channels on the membrane of different dendritic compartments, it has been proposed that the apical tuft and basal dendrites are dominated by NMDA receptors and associated potentials, while a calcium initiation zone, located just beneath the apical tuft, is endowed with voltage gated calcium channels (Nevian et al. 2007, Larkum et al. 2009), Fig. 1). Moreover, the correct degree of coupling between basal and apical compartments would be ensured by hyperpolarization-activated HCN channels, responsible for the I_h current and densely distributed on the apical dendrite of PNs (Phillips et al. 2016). Interestingly, these ion channels are developmentally regulated (Atkinson and Williams 2009) and their dysregulation might be involved in the genesis of neurodevelopmental disorders (see below).

In the last years, a conspicuous line of research focused on the dual basal/apical organization of PNs. Changes in the apical amplification process have been implicated in the pathophysiology of several mental disorders, including schizophrenia (Phillips et al. 2016, Mäki-Marttunen et al. 2019). Furthermore, the integration of two different compartments with feedforward and feedback input seems to be ideally suited to bridge the gap between artificial intelligence and neuroscience, since there are similarities between deep learning algorithms and the functional subdivisions observed in PNs (Guergiev et al. 2017).

Dendrites of pyramidal neurons are covered with spines, which receive most of synaptic inputs and are thought to play a central role in several functions, from electrical filter/isolation to synaptic and structural plasticity (reviewed in (Yuste 2011, Sala and Segal 2014)). The density and distribution of dendritic spines appear to be differently regulated in apical and basal dendrites of PNs during learning (Knafo et al. 2001), in response to hormones (Gould et al. 1990), and in experimental models of neurological illness (Perez-Cruz et al. 2011).

Pn Dendritic Domains And Id

From the features outlined above, it is clear that the extensive dendrites of neocortical PNs play a pivotal role in neural computation and higher functions. Therefore, the changes of PN dendrites appear to be central in the genesis of ID. Moreover, a disruption of the interplay between functionally distinct basal and apical compartments might contribute to the pathophysiology of several mental disorders, including ID. Here we focus on PN dendritic alterations in some of the most frequently observed genetic and non-genetic types of ID. Among genetically determined IDs, Down syndrome (DS), caused by trisomy of the human chromosome 21, besides representing the most commonly identified form (Sherman et al. 2007), can be also reproduced by murine models (Dierssen et al. 2001). Notably, dendritic alterations have been reported in both species (reviewed in (Benavides-Piccione et al. 2004)). Interestingly, in the cortex of individuals with DS, the branching of PN dendrites display a higher complexity during the first months of postnatal life, followed by a reduction of dendritic branches thereafter, and dendritic alterations were evident both in the apical and basal dendrites (Becker et al. 1986). In addition to these alterations in branching, a reduction of spine density has been reported in the apical dendrite of humans affected by DS (Suetsugu and Mehraein 1980). It should be noted, however, that PNs of the prefrontal cortex in a murine model of DS display an increased density of spines (Thomazeau et al. 2014). An augmented

number of dendritic spines is a consistent feature of a common inherited cause of ID, the fragile X syndrome (FXS), characterized by mutations of the FMRP, the protein encoded by the *FMR1* gene (Bagni and Greenough 2005).

The most common form of nongenetic ID is the consequence of the exposure to alcohol *in utero* and is nowadays referred to as fetal alcohol spectrum disorders (FASD). Rodent experimental models of FASD allowed to clarify several aspects in the pathogenesis of ID, including those related to PN dendritic anomalies (Valenzuela et al. 2012). Experimental FASD might represent an interesting case of dissociation between dendritic anomalies of basal and apical compartments: after exposure to ethanol during the first week of postnatal life in rats, corresponding to the third trimester of gestation in humans, the basal dendritic branches of PNs are strongly simplified, both in the somatosensory and in the prefrontal cortex (Granato et al. 2003, Granato et al. 2012, Hamilton et al. 2010). Conversely, the spine density of layer 2/3 basal dendrites is not affected (Hamilton et al. 2010, De Giorgio and Granato 2015). Using the same experimental protocols, a specular alteration was observed in the apical dendrites, that featured normal branching properties along with a decreased spine density (Whitcher and Klintsova 2008, Granato et al. 2012, De Giorgio and Granato 2015). It is worth mentioning, however, that in other types of ID, such as congenital/neonatal hypothyroidism, the branching pattern of the apical dendrite is anomalous (Ipiña and Ruiz-Marcos 1986).

Pn Dendrite Physiology And Id

As noted above, active currents generated locally in the dendritic tree can play a fundamental role for the function of PNs. Disruption of dendritic potentials can be the signature of many neurological and mental disorders (Palmer 2014). In the experimental model of FASD, we have demonstrated that the generation of Ca^{2+} spikes in the apical dendrites of layer 5 PNs is strongly impaired (Granato et al. 2012). A derangement of dendritic Ca^{2+} signaling has been also reported in the murine model of FXS (Meredith et al. 2007) and might be part of a more complex set of dendritic channelopathies observed in this condition (Brager and Johnston 2014). Calcium spikes are required to induce synaptic plasticity (Kampa et al. 2006, Cichon and Gan 2015). In addition, they support the apical amplification mechanism described above, that in turn is thought to provide the neurobiological basis for context-sensitive perception and learning (Phillips 2017). Interestingly, shutting down the Ube3a protein leads to a selective defect of growth of PN apical dendrites (Miao et al. 2013). The deficiency of the Ube3a protein in humans is associated to the Angelman syndrome, a condition characterized by ID and whose murine model displays a deficit of contextual learning (Jiang et al. 1998). Besides a direct impairment of dendritic calcium electrogenesis, other factors can contribute to the anomalous function of the apical dendrite and to the genesis of ID. For instance, HCN channels, responsible for the I_h current, play a role in the interaction between the basal and the apical dendites of PNs and some of their variants can be associated to ID (Marini et al. 2018). Another factor ensuring the communication between different dendritic compartments is represented by the backpropagation of axon potential along the apical dendrite. Loss of sodium channels sustaining the backpropagation can also lead to ID (Spratt et al.

2019). Finally, PNs are inserted into a cortical microcircuit to which dendrite-targeting GABAergic interneurons provide a substantial contribution (Palmer et al. 2012). Furthermore, top-down projections from higher cortical areas can engage in a disinhibitory circuit by contacting inhibitory neurons (chiefly VIP-calretinin cells) that, in turn, synapse onto other inhibitory cells, thus activating PN dendrites. Such a disinhibitory circuit might represent part of the neuronal basis for the apical amplification, since it is ideally suited to be involved in recalling past experiences and exploiting contextual cues (Pi et al. 2013, Karnani et al. 2014). Interestingly, an increase in the number of calretinin interneurons has been reported both in a model of FASD and in the Ts65Dn model of DS (Granato 2006, Pérez-Cremades et al. 2010).

Concluding Remarks

Considering that PNs are the most abundant neuronal type of the cerebral cortex, and that they bear a massive dendritic tree, it turns out that the vast majority of neocortical volume is made by PN dendrites. In other words, the neocortex seems to be a “dendritic machine” and the involvement of PN dendrites in ID is quite obvious. Times are not yet mature, however, to classify ID according, for instance, to the PN dendritic domain involved, to the spine density, or to the specific interplay between inhibitory interneurons and PN dendrites. Although such a classification might prove useful to steer the clinical and therapeutic interventions, any effort in this direction appears to be challenging. In fact, ID is a permanent condition that is usually established early during neural development, often through intermediate phases showing transient features (see, for example, the dendritic hypertrophy observed in DS). Furthermore, some of the pathologic features might merely represent a byproduct, or a compensation attempt, of primary alterations. This might be the case for the increased number of potentially disinhibitory interneurons observed in DS and FASD.

Studies on PN dendrites in humans affected by ID are shadowed by technical limitations, in particular by the capricious Golgi staining. The refinement of recording and staining techniques, along with the increased possibility of modeling neurons and their dendrites, can help to improve the results of human investigations (Goriounova et al. 2018). Not to mention the fact that animal models of ID allow detailed *in vitro* and *in vivo* explorations of dendritic and spine function. Therefore, the collaboration among clinical, computational, and experimental neuroscientists will warrant a bright future for the research on ID and dendrites.

Declarations

Authors' contribution

AG and AM conceived and wrote the manuscript

The authors declare that there is no conflict of interest

Uno de los hechos mejor apreciados entonces fue la revelación de la existencia constante en la corteza cerebral de batracios, reptiles, aves y mamíferos, del *corpúsculo piramidal*, que osé llamar, con audacia

de lenguaje de que hoy me avergüenzo un tanto, la *célula psíquica*.

One of the most appreciated facts at that time was the revelation of the constant existence in the cerebral cortex of batrachians, reptiles, birds and mammals, of the pyramidal cell, which I dared to call, with the audacity of language of which today I am a bit ashamed, the psychic cell.

Santiago Ramón y Cajal

References

1. Alevriadou A, Hatzinikolaou K, Tsakiridou H, Grouios G (2004) Field dependence-independence of normally developing and mentally retarded boys of low and upper/middle socioeconomic status. *Percept Mot Skills* 99(3 Pt 1):913–923. doi:10.2466/pms.99.3.913-923
2. American Psychiatric Association (2013) Diagnostic and statistical manual of mental disorders, 5th edn. APA, Washington, DC
3. Atkinson SE, Williams SR (2009) Postnatal Development of Dendritic Synaptic Integration in Rat Neocortical Pyramidal Neurons. *J Neurophysiol* 102(2):735–751. doi:10.1152/jn.00083.2009
4. Bagni C, Greenough WT (2005) From mRNP trafficking to spine dysmorphogenesis: the roots of fragile X syndrome. *Nat Rev Neurosci* 6(5):376–387. doi:10.1038/nrn1667
5. Becker LE, Armstrong DL, Chan F (1986) Dendritic atrophy in children with Down's syndrome. *Ann Neurol* 20(4):520–526. doi:10.1002/ana.410200413
6. Benavides-Piccione R, Ballesteros-Yáñez I, de Lagrán MM, Elston G, Estivill X, Fillat C, Defelipe J, Dierssen M (2004) On dendrites in Down syndrome and DS murine models: a spiny way to learn. *Prog Neurobiol* 74(2):111–126. doi:10.1016/j.pneurobio.2004.08.001
7. Brager DH, Johnston D (2014) Channelopathies and dendritic dysfunction in fragile X syndrome. *Brain Res Bull* 103:11–17. doi:10.1016/j.brainresbull.2014.01.002
8. Cajal SRy (1917) Recuerdos de mi vida - Historia de mi labor científica, vol 2. Moya, Madrid
9. Carr JL, Agnihotri S, Keightley M (2010) Sensory processing and adaptive behavior deficits of children across the fetal alcohol spectrum disorder continuum. *Alcohol Clin Exp Res* 34(6):1022–1032. doi:10.1111/j.1530-0277.2010.01177.x
10. Cauller L (1995) Layer I of primary sensory neocortex: where top-down converges upon bottom-up. *Behav Brain Res* 71(1):163–170. doi:[https://doi.org/10.1016/0166-4328\(95\)00032-1](https://doi.org/10.1016/0166-4328(95)00032-1)
11. Cauller LJ, Clancy B, Connors BW (1998) Backward cortical projections to primary somatosensory cortex in rats extend long horizontal axons in layer I. *J Comp Neurol* 390(2):297–310
12. Cichon J, Gan WB (2015) Branch-specific dendritic Ca(2+) spikes cause persistent synaptic plasticity. *Nature* 520(7546):180–185. doi:10.1038/nature14251
13. Constantinople CM, Bruno RM (2013) Deep cortical layers are activated directly by thalamus. *Science* 340(6140):1591–1594. doi:10.1126/science.1236425

14. Coogan TA, Burkhalter A (1990) Conserved patterns of cortico-cortical connections define areal hierarchy in rat visual cortex. *Exp Brain Res* 80(1):49–53. doi:10.1007/BF00228846
15. Davie JT, Kole MH, Letzkus JJ, Rancz EA, Spruston N, Stuart GJ, Häusser M (2006) Dendritic patch-clamp recording. *Nat Protoc* 1(3):1235–1247. doi:10.1038/nprot.2006.164
16. De Giorgio A, Granato A (2015) Reduced density of dendritic spines in pyramidal neurons of rats exposed to alcohol during early postnatal life. *Int J Dev Neurosci* 41:74–79. doi:S0736-5748(15)00007-6 [pii];10.1016/j.ijdevneu.2015.01.005 [doi]
17. Dierssen M, Fillat C, Crnic L, Arbonés M, Flórez J, Estivill X (2001) Murine models for Down syndrome. *Physiol Behav* 73(5):859–871. doi:10.1016/s0031-9384(01)00523-6
18. Dierssen M, Ramakers GJ (2006) Dendritic pathology in mental retardation: from molecular genetics to neurobiology. *Genes Brain Behav* 5 Suppl 2:48–60. doi:10.1111/j.1601-183X.2006.00224.x
19. Goriounova NA, Heyer DB, Wilbers R, Verhoog MB, Giugliano M, Verbist C, Obermayer J, Kerkhofs A, Smeding H, Verberne M, Idema S, Baayen JC, Pieneman AW, de Kock CP, Klein M, Mansvelder HD (2018) Large and fast human pyramidal neurons associate with intelligence. *eLife* 7. doi:10.7554/eLife.41714
20. Gould E, Allan MD, McEwen BS (1990) Dendritic spine density of adult hippocampal pyramidal cells is sensitive to thyroid hormone. *Brain Res* 525(2):327–329. doi:10.1016/0006-8993(90)90884-e
21. Granato A (2006) Altered organization of cortical interneurons in rats exposed to ethanol during neonatal life. *Brain Res* 1069(1):23–30. doi:10.1016/j.brainres.2005.11.024
22. Granato A De GiorgioA (2014) Alterations of neocortical pyramidal neurons: turning points in the genesis of mental retardation. *Front Pediatr* 2:86. doi:10.3389/fped.2014.00086 [doi]
23. Granato A, Di Rocco F, Zumbo A, Toesca A, Giannetti S (2003) Organization of cortico-cortical associative projections in rats exposed to ethanol during early postnatal life. *Brain Res Bull* 60(4):339–344. doi:[https://doi.org/10.1016/S0361-9230\(03\)00052-2](https://doi.org/10.1016/S0361-9230(03)00052-2)
24. Granato A, Palmer LM, De Giorgio A, Tavian D, Larkum ME (2012) Early exposure to alcohol leads to permanent impairment of dendritic excitability in neocortical pyramidal neurons. *J Neurosci* 32(4):1377–1382. doi:10.1523/JNEUROSCI.5520-11.2012
25. Guergiev J, Lillicrap TP, Richards BA (2017) Towards deep learning with segregated dendrites. *eLife* 6:e22901. doi:10.7554/eLife.22901
26. Hamilton GF, Whitcher LT, Klintsova AY (2010) Postnatal binge-like alcohol exposure decreases dendritic complexity while increasing the density of mature spines in mPFC Layer II/III pyramidal neurons. *Synapse* 64(2):127–135. doi:10.1002/syn.20711
27. Ipiña SL, Ruiz-Marcos A (1986) Dendritic structure alterations induced by hypothyroidism in pyramidal neurons of the rat visual cortex. *Brain Res* 394(1):61–67. doi:10.1016/0165-3806(86)90082-9
28. Jiang YH, Armstrong D, Albrecht U, Atkins CM, Noebels JL, Eichele G, Sweatt JD, Beaudet AL (1998) Mutation of the Angelman ubiquitin ligase in mice causes increased cytoplasmic p53 and deficits of

- contextual learning and long-term potentiation. *Neuron* 21(4):799–811. doi:10.1016/s0896-6273(00)80596-6
29. Kampa BM, Letzkus JJ, Stuart GJ (2006) Requirement of dendritic calcium spikes for induction of spike-timing-dependent synaptic plasticity. *J Physiol* 574(Pt 1):283–290. doi:10.1113/jphysiol.2006.111062
30. Karnani MM, Agetsuma M, Yuste R (2014) A blanket of inhibition: functional inferences from dense inhibitory connectivity. *Curr Opin Neurobiol* 26:96–102. doi:10.1016/j.conb.2013.12.015
31. Kaufmann WE, Moser HW (2000) Dendritic anomalies in disorders associated with mental retardation. *Cereb Cortex* 10(10):981–991. doi:10.1093/cercor/10.10.981
32. Knafo S, Grossman Y, Barkai E, Benshalom G (2001) Olfactory learning is associated with increased spine density along apical dendrites of pyramidal neurons in the rat piriform cortex. *Eur J Neurosci* 13(3):633–638. doi:<https://doi.org/10.1046/j.1460-9568.2001.01422.x>
33. Krieger P, de Kock CPJ, Frick A (2017) Calcium Dynamics in Basal Dendrites of Layer 5A and 5B Pyramidal Neurons Is Tuned to the Cell-Type Specific Physiological Action Potential Discharge. *Front Cell Neurosci* 11 (194). doi:10.3389/fncel.2017.00194
34. LaBerge D (2006) Apical dendrite activity in cognition and consciousness. *Conscious Cogn* 15(2):235–257. doi:<https://doi.org/10.1016/j.concog.2005.09.007>
35. Larkman AU (1991) Dendritic morphology of pyramidal neurones of the visual cortex of the rat: I. Branching patterns. *J Comp Neurol* 306(2):307–319. doi:10.1002/cne.903060207
36. Larkum M (2013) A cellular mechanism for cortical associations: an organizing principle for the cerebral cortex. *Trends Neurosci* 36(3):141–151. doi:10.1016/j.tins.2012.11.006
37. Larkum ME, Nevian T, Sandler M, Polsky A, Schiller J (2009) Synaptic integration in tuft dendrites of layer 5 pyramidal neurons: a new unifying principle. *Science* 325(5941):756–760. doi:10.1126/science.1171958
38. Larkum ME, Zhu JJ, Sakmann B (1999) A new cellular mechanism for coupling inputs arriving at different cortical layers. *Nature* 398(6725):338–341. doi:10.1038/18686
39. Lübke J, Feldmeyer D (2007) Excitatory signal flow and connectivity in a cortical column: focus on barrel cortex. *Brain Structure Function* 212(1):3–17. doi:10.1007/s00429-007-0144-2
40. Mainen ZF, Sejnowski TJ (1996) Influence of dendritic structure on firing pattern in model neocortical neurons. *Nature* 382(6589):363–366. doi:10.1038/382363a0
41. Mäki-Marttunen T, Devor A, Phillips WA, Dale AM, Andreassen OA, Einevoll GT (2019) Computational Modeling of Genetic Contributions to Excitability and Neural Coding in Layer V Pyramidal Cells: Applications to Schizophrenia Pathology. *Front Comput Neurosci* 13:66–66. doi:10.3389/fncom.2019.00066
42. Marini C, Porro A, Rastetter A, Dalle C, Rivolta I, Bauer D, Oegema R, Nava C, Parrini E, Mei D, Mercer C, Dhamija R, Chambers C, Coubes C, Thévenon J, Kuentz P, Julia S, Pasquier L, Dubourg C, Carré W, Rosati A, Melani F, Pisano T, Giardino M, Innes AM, Alembik Y, Scheidecker S, Santos M, Figueiroa S, Garrido C, Fusco C, Frattini D, Spagnoli C, Binda A, Granata T, Ragona F, Freri E, Franceschetti S,

- Canafoglia L, Castellotti B, Gellera C, Milanesi R, Mancardi MM, Clark DR, Kok F, Helbig KL, Ichikawa S, Sadler L, Neupauerová J, Laššuthova P, Šterbová K, Laridon A, Brilstra E, Koeleman B, Lemke JR, Zara F, Striano P, Soblet J, Smits G, Deconinck N, Barbuti A, DiFrancesco D, LeGuern E, Guerrini R, Santoro B, Hamacher K, Thiel G, Moroni A, DiFrancesco JC, Depienne C (2018) HCN1 mutation spectrum: from neonatal epileptic encephalopathy to benign generalized epilepsy and beyond. *Brain* 141(11):3160–3178. doi:10.1093/brain/awy263
43. Meredith RM, Holmgren CD, Weidum M, Burnashev N, Mansvelder HD (2007) Increased threshold for spike-timing-dependent plasticity is caused by unreliable calcium signaling in mice lacking fragile X gene FMR1. *Neuron* 54(4):627–638. doi:10.1016/j.neuron.2007.04.028
44. Meyer HS, Wimmer VC, Hemberger M, Bruno RM, de Kock CPJ, Frick A, Sakmann B, Helmstaedter M (2010) Cell type-specific thalamic innervation in a column of rat vibrissal cortex. *Cerebral cortex (New York, NY: 1991)* 20 (10):2287–2303. doi:10.1093/cercor/bhq069
45. Miao S, Chen R, Ye J, Tan GH, Li S, Zhang J, Jiang YH, Xiong ZQ (2013) The Angelman syndrome protein Ube3a is required for polarized dendrite morphogenesis in pyramidal neurons. *J Neurosci* 33(1):327–333. doi:10.1523/jneurosci.2509-12.2013
46. Murray G, McKenzie K, Murray A, Whelan K, Cossar J, Murray K, Scotland J (2019) The impact of contextual information on the emotion recognition of children with an intellectual disability. *J Appl Res Intellect Disabil* 32(1):152–158. doi:10.1111/jar.12517
47. Nevian T, Larkum ME, Polsky A, Schiller J (2007) Properties of basal dendrites of layer 5 pyramidal neurons: a direct patch-clamp recording study. *Nat Neurosci* 10(2):206–214. doi:10.1038/nn1826
48. Palmer L, Murayama M, Larkum M (2012) Inhibitory Regulation of Dendritic Activity in vivo. *Frontiers in Neural Circuits* 6 (26). doi:10.3389/fncir.2012.00026
49. Palmer LM (2014) Dendritic integration in pyramidal neurons during network activity and disease. *Brain Res Bull* 103:2–10. doi:<https://doi.org/10.1016/j.brainresbull.2013.09.010>
50. Pérez-Cremades D, Hernández S, Blasco-Ibáñez JM, Crespo C, Nacher J, Varea E (2010) Alteration of inhibitory circuits in the somatosensory cortex of Ts65Dn mice, a model for Down's syndrome. *J Neural Transm (Vienna)* 117(4):445–455. doi:10.1007/s00702-010-0376-9
51. Perez-Cruz C, Nolte MW, van Gaalen MM, Rustay NR, Termont A, Tanghe A, Kirchhoff F, Ebert U (2011) Reduced spine density in specific regions of CA1 pyramidal neurons in two transgenic mouse models of Alzheimer's disease. *J Neurosci* 31(10):3926–3934. doi:10.1523/jneurosci.6142-10.2011
52. Phillips WA (2017) Cognitive functions of intracellular mechanisms for contextual amplification. *Brain Cogn* 112:39–53. doi:10.1016/j.bandc.2015.09.005
53. Phillips WA, Larkum ME, Harley CW, Silverstein SM (2016) The effects of arousal on apical amplification and conscious state. *Neurosci Conscious* 2016 (1):niw015-niw015. doi:10.1093/nc/niw015
54. Pi HJ, Hangya B, Kvitsiani D, Sanders JI, Huang ZJ, Kepcs A (2013) Cortical interneurons that specialize in disinhibitory control. *Nature* 503(7477):521–524. doi:10.1038/nature12676

55. Quach TT, Stratton HJ, Khanna R, Kolattukudy PE, Honnorat J, Meyer K, Duchemin AM (2021) Intellectual disability: dendritic anomalies and emerging genetic perspectives. *Acta Neuropathol* 141(2):139–158. doi:10.1007/s00401-020-02244-5
56. Ramaswamy S, Markram H (2015) Anatomy and physiology of the thick-tufted layer 5 pyramidal neuron. *Front Cell Neurosci* 9 (233). doi:10.3389/fncel.2015.00233
57. Sala C, Segal M (2014) Dendritic Spines: The Locus of Structural and Functional Plasticity. *Physiol Rev* 94(1):141–188. doi:10.1152/physrev.00012.2013
58. Shepherd GMG (2013) Corticostriatal connectivity and its role in disease. *Nat Rev Neurosci* 14(4):278–291. doi:10.1038/nrn3469
59. Sherman SL, Allen EG, Bean LH, Freeman SB (2007) Epidemiology of Down syndrome. *Ment Retard Dev Disabil Res Rev* 13(3):221–227. doi:10.1002/mrdd.20157
60. Shogren KA, Turnbull HR (2010) Public Policy and Outcomes for Persons With Intellectual Disability: Extending and Expanding the Public Policy Framework of AAIDD's 11th Edition of Intellectual Disability: Definition, Classification, and Systems of Support. *Intellectual Developmental Disabilities* 48(5):375–386. doi:10.1352/1934-9556-48.5.375
61. Spratt PWE, Ben-Shalom R, Keeshen CM, Burke KJ Jr, Clarkson RL, Sanders SJ, Bender KJ (2019) The Autism-Associated Gene Scn2a Contributes to Dendritic Excitability and Synaptic Function in the Prefrontal Cortex. *Neuron* 103(4):673–685.e675. doi:10.1016/j.neuron.2019.05.037
62. Spruston N (2008) Pyramidal neurons: dendritic structure and synaptic integration. *Nat Rev Neurosci* 9(3):206–221. doi:10.1038/nrn2286
63. Stuart GJ, Sakmann B (1994) Active propagation of somatic action potentials into neocortical pyramidal cell dendrites. *Nature* 367(6458):69–72. doi:10.1038/367069a0
64. Suetsugu M, Mehraein P (1980) Spine distribution along the apical dendrites of the pyramidal neurons in Down's syndrome. A quantitative Golgi study. *Acta Neuropathol* 50(3):207–210. doi:10.1007/bf00688755
65. Takahashi N, Ebner C, Sigl-Glöckner J, Moberg S, Nierwetberg S, Larkum ME (2020) Active dendritic currents gate descending cortical outputs in perception. *Nat Neurosci* 23(10):1277–1285. doi:10.1038/s41593-020-0677-8
66. Thomazeau A, Lassalle O, Iafrati J, Souchet B, Guedj F, Janel N, Chavis P, Delabar J, Manzoni OJ (2014) Prefrontal deficits in a murine model overexpressing the down syndrome candidate gene dyrk1a. *J Neurosci* 34(4):1138–1147. doi:10.1523/jneurosci.2852-13.2014
67. Thomson A (2010) Neocortical layer 6, a review. *Frontiers in Neuroanatomy* 4 (13). doi:10.3389/fnana.2010.00013
68. Thomson AM, Morris OT (2002) Selectivity in the inter-laminar connections made by neocortical neurones. *J Neurocytol* 31(3):239–246. doi:10.1023/A:1024117908539
69. Valenzuela CF, Morton RA, Diaz MR, Topper L (2012) Does moderate drinking harm the fetal brain? Insights from animal models. *Trends Neurosci* 35(5):284–292. doi:10.1016/j.tins.2012.01.006

70. Whitcher LT, Klintsova AY (2008) Postnatal binge-like alcohol exposure reduces spine density without affecting dendritic morphology in rat mPFC. *Synapse* 62(8):566–573. doi:10.1002/syn.20532
71. Yuste R (2011) Dendritic spines and distributed circuits. *Neuron* 71(5):772–781.
doi:10.1016/j.neuron.2011.07.024

Figures

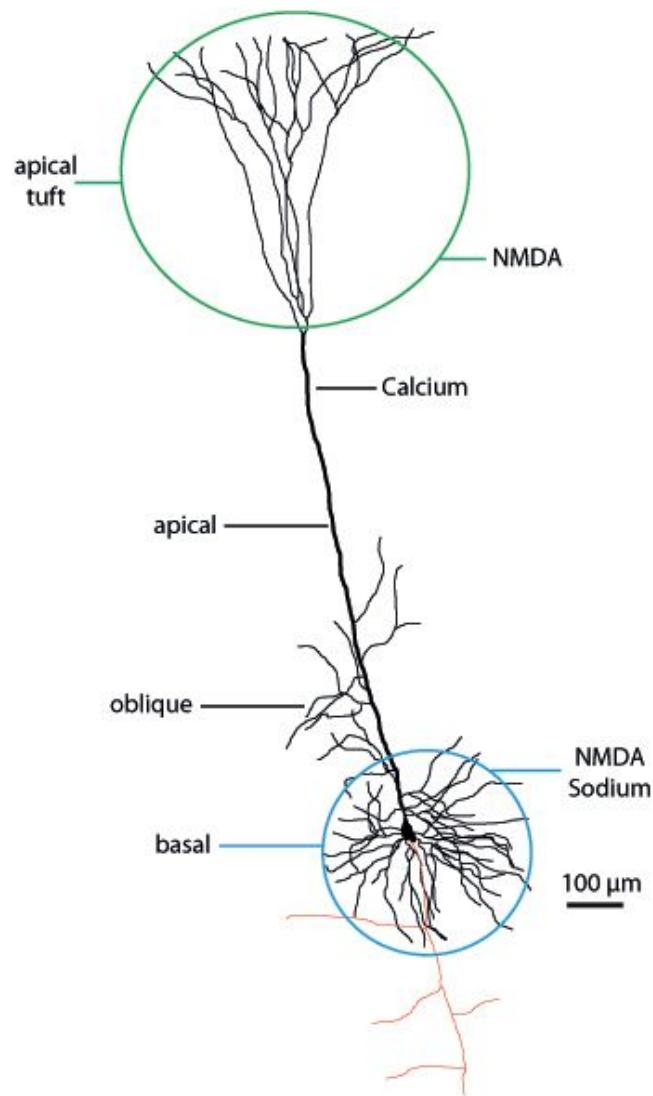


Figure 1

3D reconstruction of a thick-tufted PN of layer 5. On the left of the neuron there is the indication of the different sections of the dendritic tree. On the right the main regenerative events occurring in that dendritic domain, according to Larkum et al., 2009. Red: axon