

# Treatment of COVID-19 pneumonia and acute respiratory distress with ramatroban, a thromboxane A<sub>2</sub> and prostaglandin D<sub>2</sub> receptor antagonist: A 4-Patient Case Series Report

Martin L. Ogletree (✉ [martin.ogletree@vanderbilt.edu](mailto:martin.ogletree@vanderbilt.edu))

Department of Pharmacology, Vanderbilt University School of Medicine, Nashville, TN

Kate Chander Chiang

Charak Foundation, Orange, CA

Rashmi Kulshreshta

Regulatory Wisdom, Delhi, India

Aditya Agarwal

EyeSight Eye Hospital and Retina Centre, Madhya Pradesh, India

Ashutosh Agarwal

EyeSight Eye Hospital and Retina Centre, Madhya Pradesh, India

Ajay Gupta (✉ [ajayg1@hs.uci.edu](mailto:ajayg1@hs.uci.edu))

Division of Nephrology, Hypertension and Kidney Transplantation, University of California Irvine, Orange, CA

---

## Case Report

**Keywords:** COVID-19, SARS-CoV-2, thromboxane A<sub>2</sub>, prostaglandin D<sub>2</sub>, lipid storm, ramatroban, DPr2, TPr, thromboinflammation, thrombosis, platelet activation, NETs, immunomodulator, antithrombotic

**Posted Date:** November 18th, 2021

**DOI:** <https://doi.org/10.21203/rs.3.rs-474882/v2>

**License:** © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

---

# Abstract

COVID-19 associated pneumonia and acute respiratory distress syndrome are characterized by a lipid mediator storm with massive increases in lung and systemic thromboxane  $A_2$  >> prostaglandin  $D_2$ . Thromboxane  $A_2$  is a potent vasoconstrictor of pulmonary veins >> arteries, and thereby promotes an increase in pulmonary capillary pressures, transudation of fluid into the alveolar space, pulmonary edema and ARDS. Thromboxane  $A_2$  also increases vascular permeability, contracts bronchial smooth muscle, triggers and amplifies platelet activation, and promotes a prothrombotic state.  $PGD_2$  promotes a Th2 immune response that is atypical for viral infections and inhibits antiviral defense by suppressing interferon  $\lambda$  expression. D-dimers, urinary 11-dehydro-TxB<sub>2</sub>, and IL-13, a Th2 cytokine, have emerged as key biomarkers of severity and organ failure in COVID-19. Ramatroban is an orally bioavailable, potent, dual antagonist of the thromboxane  $A_2$  (TPr) and  $PGD_2$  (DPr2) receptors. We report use of ramatroban in 4 COVID-19 outpatients, 22 to 87 years of age, with acute onset / worsening of respiratory distress and hypoxemia. All four patients experienced decrease in respiratory distress and increase in SpO<sub>2</sub> within hours of the first dose and thereby avoided hospitalization. By the 5<sup>th</sup> day all 4 patients had complete resolution of respiratory distress and hypoxemia. Ramatroban (Baynas®, Bayer Yakuhin Ltd., Japan) has an established safety profile, having been indicated in Japan for the treatment of allergic rhinitis for over 20 years. As a broncho-relaxant, anti-vasospastic, anti-thrombotic and immunomodulator, ramatroban addresses the fundamental pathophysiologic mechanisms underlying respiratory and critical organ failure in COVID-19, and therefore merits urgent clinical trials that might impact the ongoing pandemic.

## Background

After symptomatic SARS-CoV-2 infection, 10-20% of patients require hospitalization for respiratory distress and hypoxemia.<sup>1</sup> Currently, only anti-SARS-CoV-2 monoclonal antibodies are approved for treatment of ambulatory patients with COVID-19.<sup>2</sup> Additional antiviral treatments are nearing approval, but they are expensive and not completely effective. There is an unmet medical need for an inexpensive, orally bioavailable drug with an excellent safety profile that can provide symptomatic relief, reduce hypoxemia and prevent hospitalization in outpatients with COVID-19. Identifying the correct therapeutic target is critical to discovering such a drug.

Lungs in COVID-19 patients with acute respiratory distress syndrome (ARDS) exhibit an abundance of proinflammatory lipid mediators with predominance of cyclooxygenase metabolites in bronchoalveolar lavage fluid (BALF), notably thromboxane  $B_2$  (TxB<sub>2</sub>) >> prostaglandin  $E_2$  (PGE<sub>2</sub>) > prostaglandin  $D_2$  ( $PGD_2$ ).<sup>3</sup> The massive increase in TxA<sub>2</sub> metabolites in BALF from COVID-19 associated ARDS,<sup>3</sup> and systemically in hospitalized COVID-19 patients,<sup>4 5</sup> led us to propose a critical role for TxA<sub>2</sub> - TxA<sub>2</sub> prostanoid receptors (TPr) signaling in COVID-19 associated respiratory distress. We hypothesized that TxA<sub>2</sub>/TPr induced contraction of pulmonary veins elevates pulmonary capillary pressure and contributes

to pulmonary edema and hypoxemia in COVID-19 pneumonia (Fig. 1). TPr signaling leads to constriction of intrapulmonary veins and small airways with 10-fold higher potency and greater reduction in luminal area than intrapulmonary arteries.<sup>6</sup> High local concentrations of TxA<sub>2</sub> can effectively shut down pulmonary venous blood flow, increase microvascular pressure and permeability, and force plasma into alveoli.<sup>6</sup> A selective TPr antagonist was previously reported to decrease pulmonary capillary pressure by selectively reducing post-capillary resistance in patients with acute lung injury.<sup>7</sup> Thromboxane A<sub>2</sub> and isoprostanes stimulate TPr-mediated activation of the TGFβ pathway,<sup>8</sup> and early, untimely TGFβ responses in SARS-CoV-2 infection limit antiviral function of natural killer (NK) cells and promote progression to severe COVID-19 disease.<sup>9</sup>

Theken and FitzGerald have proposed early administration of a TxA<sub>2</sub> antagonist as an antithrombotic agent to limit progression of disease in SARS-CoV-2 infection, and administration of an antagonist to block PGD<sub>2</sub> / D-prostanoid receptor 2 (DPr2, formerly referred to as CRTH2) in order to boost interferon lambda (IFN-λ) response in the upper respiratory tract, thereby limiting SARS-CoV-2 replication and transmission.<sup>10 11</sup> Ramatroban is the only dual TxA<sub>2</sub>/TPr and PGD<sub>2</sub>/DPr2 receptor antagonist available for clinical study and has been proposed as an antithrombotic and immunomodulator agent in COVID-19.<sup>12 13</sup> Archambault and colleagues also recently supported the use of ramatroban to block the deleterious effects of PGD<sub>2</sub> and TxA<sub>2</sub> in COVID-19.<sup>3</sup> Ramatroban has an established safety profile, having been used for over 20 years in Japan for the treatment of allergic rhinitis.<sup>14 15</sup> We report here a small case series of four consecutive COVID-19 patients with worsening respiratory distress and hypoxemia who were treated with ramatroban leading to rapid improvement in both respiratory distress and hypoxemia, thereby avoiding hospitalization and promoting recovery from acute disease.

### **The 1<sup>st</sup> case of severe COVID-19 pneumonia treated with ramatroban**

S.D., an 87-year-old Indian lady, experienced sudden onset of fever, cough, diarrhea, anorexia, profound weakness, and slight shortness of breath, 10 days after a 2-hour flight from New Delhi to Indore, Madhya Pradesh, India. Patient had received the first dose of COVAXIN, a whole virion inactivated vaccine against SARS-CoV-2, 30 days prior to beginning of symptoms. On examination the patient was fully alert, oriented, and able to make intelligent conversation but lay listlessly in bed unable to ambulate. Patient weighed 42 kg and exhibited severe pre-existing muscle wasting and marked kyphosis. Vital signs revealed temperature, 102° Fahrenheit; heart rate, 100 per minute; blood pressure, 90/60 mm of Hg; and respiratory rate, 22 per minute. Mucosa were moist, and mild pallor was present. There was no jugular venous distention or pedal edema. Chest examination revealed bilateral coarse rales especially prominent

at both lung bases but no wheezes. Abdomen, cardiovascular, and neurological examinations were unremarkable. Patient was not taking any medications.

*Past medical history* included hypertension for over 40 years; thyrotoxicosis for over 30 years treated with radioiodine therapy in 1999; severe osteoporosis with kyphosis; bladder suspension surgery in 1999; coronary artery disease leading to acute myocardial infarction and cardiac arrest in 2015 which required coronary angioplasty and stent placement; chronic kidney disease with estimated glomerular filtration rate of about 20 mL/min (Figure 2).

*Investigations:* Nasopharyngeal and oropharyngeal swabs were positive for SARS-CoV-2 infection by RNA PCR with cycle threshold (Ct range < 20 cycles). Pulse oximetry revealed oxygen saturation of about 85-88%. Patient was admitted on April 9, 2021 to Medanta Hospital, Indore. CT scan revealed moderate multifocal, patchy ground glass opacities, and consolidation. There was septal thickening in the central and peripheral subpleural aspect of both lung parenchyma. Serial laboratory examinations during the course of the illness are listed in Table 1.

*Hospital course:* During the hospital stay, the patient was treated with high-flow nasal oxygen, prophylactic low-molecular weight heparin, intravenous remdesivir, antibiotics, and methylprednisolone. Patient continued to have fever, cough, shortness of breath, diarrhea, and profound weakness during the hospital stay. SpO<sub>2</sub> on room air ranged between 82-86% (Table 2). After a hospital stay of 5 days, the patient was discharged upon her request on April 14, 2021. Discharge medications included oral oseltamivir, doxycycline, vitamin C, aspirin 75 mg once a day, 5 mg prednisolone, vitamin D<sub>3</sub>, and nebulization with budesonide and salbutamol twice daily. Continued supportive management with betadine gargles, steam inhalation, and breathing exercises was advised.

*Post-discharge course:* On April 15, the day after discharge from the hospital, the patient had fever with a temperature of 101° Fahrenheit. Pulse oximetry revealed an oxygen saturation (SpO<sub>2</sub>) of 82-84% on room air, and patient was continued on oxygen. Patient was profoundly weak and unable to get out of bed without assistance. At this time all drugs including low-dose aspirin were discontinued, and the patient was started on ramatroban (Baynas®, 75 mg tablet) in a dose of one-half tablet (37.5 mg) orally twice daily. The patient was continued on oxygen using a nasal cannula and SpO<sub>2</sub> was not checked on room air. After about 36 hours, having received three one-half doses of ramatroban, there was noticeable improvement in her general condition, and SpO<sub>2</sub> increased to 90% on room air. The dose of ramatroban was increased to 37.5 mg in the morning and 75 mg at bedtime. Patient had complete resolution of

cough and diarrhea over the next 3 days and started ambulating independently without assistance. Ramatroban was discontinued after 2 weeks due to non-availability, and the patient was switched to 75 mg aspirin daily. Patient had recovered almost completely by April 22, 2021, and gradually recovered fully over a period of next 3-4 weeks back to her baseline status. On October 10, 2021, 6 months after the acute COVID-19, a high-resolution, non-contrast CT scan demonstrated non-homogenous ground glass pattern with normal lung volumes and absence of lung fibrosis. Patient continues to be asymptomatic.

Table 1. Serial laboratory values

Analytes	Before admission	During hospital stay	After discharge	Reference Value
	April 8, 2021	April 13, 2021	April 16, 2021	
Hemoglobin (g/dl)	11.7	12.1	12.0	13.0-17.0
Platelet count (per mm <sup>3</sup> )	214,000	285,000	402,000	150,000-410,000
WBC count (per mm <sup>3</sup> )	5040	12100	9010	4000-10000
Neutrophils (%)	77	80	86	38-70
Lymphocytes (%)	18	11	07	21-49
NLR (Neutrophil-Lymphocyte Ratio)	4.3	7.3	12.3	1.1-3.5
Serum CRP (mg/L)	7.86	35.9	15.3	0-5.0
D-dimer (ng FEU/mL)	600	650	659	<500

## Case 2

A.K., a 33-year-old business manager in New Delhi developed sore throat, cough, loss of smell, altered taste, loss of appetite, high grade fever (104 -106° Fahrenheit), profound weakness and severe body aches around April 17, 2021. A.K. had not received the COVID-19 vaccine. Patient has past medical history of mild hypertension, psoriasis and psoriatic arthropathy treated with homeopathy, nasal polyposis and recurrent upper respiratory infections every winter for past several years. Nasopharyngeal

and oropharyngeal swabs taken the next day were positive for SARS-CoV-2 infection by RNA PCR with cycle threshold (Ct range) of 21 cycles.

On April 18, 2021 the patient developed progressive shortness of breath and was started on oral favipiravir, hydroxychloroquine, doxycycline and multivitamins. SpO<sub>2</sub> checked in the morning was about 90%, declining to 82-85% by the evening. The shortness of breath worsened around midnight and during the early morning hours of April 19<sup>th</sup>, patient “could not catch his breath, was unable to speak, and was very anxious and restless.” The SpO<sub>2</sub> was 73% (Table 2). Patient could not be transferred to a COVID hospital because of nonavailability of hospital beds. Desperate attempts to secure an oxygen cylinder failed. Ramatroban was rushed to patient’s home by Uber and first dose of 75mg was taken at 1:30 AM on the morning of April 19<sup>th</sup>. The “breathing improved in 25-30 minutes”, the patient calmed down and fell asleep at 3 AM. Pulse oximetry remained disconnected while the patient was sleeping so as not to disturb him. Patient woke up at 11 AM at which time SpO<sub>2</sub> on room air was 88-90%. On April 20<sup>th</sup>, oral temperature was 101° Fahrenheit, and SpO<sub>2</sub> was 90-92%. Ramatroban was administered in a dose of 75 mg twice daily for a total of 5 days. Patient continued to improve over the next 5 days (Table 2). On the 25<sup>th</sup> of April, patient noticed that the sputum was streaked with blood and oral acetylcysteine was started. A chest CT on April 27<sup>th</sup> revealed ground glass opacities involving bilateral lung fields with mild interstitial thickening giving the appearance of crazy-paving pattern. There were scattered areas of bronchopneumonic changes and consolidation involving both lungs. A few small fibrotic bands were noted in both lower lobes. The patient had made a near complete recovery by May 5<sup>th</sup>, and resumed work on May 10<sup>th</sup>. Patient continues to have altered taste and smell 7 months after the acute illness.

### **Case 3**

S.B., a 22-year-old, healthy lady in New Delhi developed fever, cough, loss of smell and taste and body aches due to COVID-19. S.B. had not received COVID vaccination. S.B. was treated with favipiravir, steroids and multivitamins. Patient experienced progressively worsening shortness of breath and SpO<sub>2</sub> dropped to 85% on room air. Patient was prescribed Ramatroban 75 mg twice daily. Within about 6-8 hours after taking the first dose of ramatroban, respiratory distress improved and the SpO<sub>2</sub> increased to 89%. The next day SpO<sub>2</sub> increased to 90-91%. There was progressive improvement with complete resolution of respiratory symptoms over the next 5 days. On day 5, the SpO<sub>2</sub> was noted to be 94% on room air (Table 2). Patient has made a complete recovery from COVID-19.

## Case 4

B.C., a 70-year-old man living in a rural area of Bihar, India developed high grade fever and cough presumably secondary to SARS-CoV-2 infection. Patient has a history of diabetes mellitus controlled with diet. B.C. was not taking any medications and had only received one dose of COVAXIN vaccine for COVID-19. Patient developed shortness of breath with SpO<sub>2</sub> measuring about 80% on room air. Two to three hours after taking 75 mg ramatroban, respiratory distress and cough improved, and the SpO<sub>2</sub> increased to 85%. After a total of 10 tablets taken over 5 days, dyspnea had resolved, and SpO<sub>2</sub> increased to 96% on room air (Table 2). Patient has made a complete recovery from COVID-19.

**Table 2. Clinical course of COVID-19 patients with acute respiratory distress treated with Ramatroban**

Patient Initials	Ramatroban (Baynas®)	Clinical course; Blood oxygen saturation by pulse oximetry (SpO <sub>2</sub> )		
		Time '0'	Time to partial relief of dyspnea; and the first SpO <sub>2</sub> recorded on room air after initiating ramatroban treatment	Day '5' after taking 10 tablets of ramatroban <sup>^</sup>
Gender	75 mg tab	Ramatroban started		
Age (years)				
Comorbidity				
S.D.; female, 87 yrs.	37.5 mg	Dyspnea ++	24-36 hours;	No dyspnea
Hypertension; Stage 4 CKD; CAD, MI and cardiac arrest 4 years ago	(½ tab) twice daily	SpO <sub>2</sub> : 82%	SpO <sub>2</sub> > 90%, 36 hours after 1 <sup>st</sup> dose*	SpO <sub>2</sub> ≥ 95%
A.K., male, 30 yrs	75 mg	Dyspnea +++	1-2 hours;	No dyspnea
Hypertension, psoriasis, recurrent URTI	twice daily	SpO <sub>2</sub> : 73%	SpO <sub>2</sub> 90%, 9 hours after 1 <sup>st</sup> dose*	SpO <sub>2</sub> 96%
S.B., female, 22 yrs	75 mg	Dyspnea ++	4-6 hours;	No dyspnea
	twice daily	SpO <sub>2</sub> : 85%	SpO <sub>2</sub> 89%, 6-8 hours after 1 <sup>st</sup> dose	SpO <sub>2</sub> 94%
B.C., male, 70 yrs	75 mg	Dyspnea ++	2-3 hours;	No dyspnea
Diabetes mellitus	twice daily	SpO <sub>2</sub> : 80%	SpO <sub>2</sub> 85%, 2-3 hours after 1 <sup>st</sup> dose	SpO <sub>2</sub> 96%

\*SpO<sub>2</sub> on room air was not checked at earlier time points

^ For patients 2, 3, and 4, ramatroban could be administered only for a total of 5 days due to limited supplies.

## Discussion

We present the first reported cases of COVID-19 treated with ramatroban (Baynas®), a dual antagonist of the TxA<sub>2</sub>/TPr and PGD<sub>2</sub>/DPr2 receptors. All four COVID-19 patients were characterized by respiratory distress that was new in onset or had worsened (Table 2). Despite severe hypoxemia, all patients were able to avoid hospitalization and recovered without any further need for steroids.

The rapidity of improvement following treatment with oral ramatroban is consistent with an acute hemodynamic effect. We hypothesize that this involves primarily blocking TxA<sub>2</sub> / TPr-mediated selective pulmonary venous constriction and pulmonary capillary hypertension. A consequent increased transcapillary pressure gradient across the pulmonary microvasculature leads to transudation of fluid from the vascular compartment into the alveoli and small airways<sup>6</sup> (Fig. 1). Notably, U-46619, a TxA<sub>2</sub> mimetic in a concentration of 1 nM is sufficient to reduce guinea-pig pulmonary venous luminal area by 50%.<sup>6</sup> A 50% reduction in luminal area increases vascular resistance by 4-fold, indicating that sub-nanomolar concentrations of thromboxane A<sub>2</sub> could produce meaningful increases in pulmonary venous resistance.<sup>6</sup> This is consistent with the measured effect of ifetroban, a selective TPr antagonist which reduced pulmonary venous resistance and capillary pressure in patients with acute lung injury.<sup>16</sup> Moreover, TPr antagonism has been shown to attenuate airway mucus hyperproduction induced by cigarette smoke<sup>17</sup> and reduce tissue edema in mouse models of acute lung injury.<sup>18</sup> In the cases presented here, we hypothesize that TPr blockade with ramatroban rapidly reduced pulmonary capillary pressures, improved ventilation-perfusion matching, promoted resolution of edema, reduced bronchoconstriction and airway mucus hyperproduction, improved lung compliance and gas exchange, and thereby mitigated respiratory distress and hypoxemia (Table 3 and Fig. 1).

Lung TxA<sub>2</sub> generation is sufficiently elevated in symptomatic COVID-19 that TPr activation may affect other critical organ functions. For example, coronary vascular effects might include vasospasm and thrombosis resulting in angina, arrhythmias and/or myocardial infarction.<sup>19</sup> In the cerebral circulation, TPr activation can increase blood-brain barrier permeability,<sup>20</sup> which may contribute to brain fog in COVID-19. The potential of TPr blockade to affect function of these and other critical organs merits focused COVID-19 research.

In COVID-19, TPr activation by massively elevated levels of TxA<sub>2</sub> and isoprostanes may be further compounded by increased expression of TPr resulting from suppressed expression of microRNA-31.<sup>21</sup> MicroRNA-31 suppression in endothelial progenitor cells, as found in coronary artery disease patients, leads to higher TPr expression,<sup>22</sup> suggesting potential for exacerbation of TxA<sub>2</sub> mediated effects in COVID-19 patients with underlying cardiovascular disease.

PGD<sub>2</sub> / DPr2 signaling also promotes allergic inflammation by stimulating Th2 and innate lymphocyte class 2 (ILC2) cells as in asthma (Fig. 1).<sup>23 24</sup> The maladaptive immune response in COVID-19 is characterized by a shift from Th1 to Th2 with basophilia, eosinophilia, lymphopenia and an increase in plasma levels of type 2 cytokines produced by Th2 cells, including IL-4 and IL-13.<sup>25-27</sup> IL-4 is known to impair the barrier function of endothelial cells, leading to microvascular leakage and edema formation (Fig. 1).<sup>28</sup> IL-13 increases hyaluronan accumulation in mouse lungs,<sup>29</sup> and mucus overproduction in cultured human bronchial epithelial cells,<sup>30</sup> and is correlated with ARDS, need for mechanical ventilation, acute kidney injury (AKI), and mortality in COVID-19.<sup>31</sup> The IC<sub>50</sub> of ramatroban for inhibiting IL-4 and IL-13 production induced by 100 nM PGD<sub>2</sub> is 103 and 118 nM, respectively.<sup>23</sup> Whether ramatroban inhibits hyaluronan accumulation in ARDS remains to be investigated.

The early beneficial effects of ramatroban may be additionally attributed to an enhanced antiviral activity due to TxA<sub>2</sub> / TPr and PGD<sub>2</sub> / DPr2 antagonism. First, TxA<sub>2</sub> / TPr activation stimulates activation of the TGFβ pathway,<sup>8</sup> and early, untimely TGFβ responses in SARS-CoV-2 infection limit antiviral function of natural killer (NK) cells.<sup>9</sup> Second, PGD<sub>2</sub>/DPr2 signaling suppresses innate mucosal antiviral responses by inhibiting expression of IFN-λ, the first line of defense against viruses at mucosal surfaces. Notably IFN-λ is markedly suppressed in the upper respiratory tract in COVID-19.<sup>32</sup> An increased expression of phospholipase A<sub>2</sub> group IID and PGD<sub>2</sub> in the elderly may further suppress IFN-λ expression,<sup>33</sup> thereby impairing their antiviral responses and contributing to the increased morbidity and mortality observed consistently in the elderly with COVID-19.<sup>11</sup> Surprisingly, expression of nasal and pharyngeal PGD<sub>2</sub> and DPr2 in SARS-COV-2 infection remain to be investigated even though there is significant elevation of PGD<sub>2</sub> levels in alveolar lavage fluid,<sup>3 34</sup> and expression of PGD<sub>2</sub> synthase and DPr2 in COVID-19 kidneys.<sup>35</sup> Interestingly, 11-dehydro-TxB<sub>2</sub> (11dhTxB<sub>2</sub>), a major stable metabolite of thromboxane A<sub>2</sub>, serves as a full agonist of DPr2 receptors, and urinary 11dhTxB<sub>2</sub> levels are markedly increased in COVID-19 and correlate with length of hospitalization, mechanical ventilation and mortality.<sup>36</sup> In rabbits infused with TxB<sub>2</sub>, 11dhTxB<sub>2</sub> was the first major metabolite to appear and remained a prominent product in blood for the remainder of the infusion. Enzymatic conversion of TxB<sub>2</sub> to 11dhTxB<sub>2</sub> was not detected in blood cells or plasma.<sup>37</sup> The dehydrogenase catalyzing formation of 11dhTxB<sub>2</sub> was tissue bound and

widespread with the highest activity in lung, kidney, stomach and liver.<sup>37</sup> The above suggests that elevated lung TxA<sub>2</sub> is rapidly converted to 11dhTxB<sub>2</sub> which may exert effects in the lungs via DPr2. In a neonatal mouse model of severe respiratory syncytial virus-induced bronchiolitis, treatment with a DPr2 antagonist decreased viral load and improved morbidity associated with upregulating interferon (IFN)-λ expression.<sup>10 33</sup> Whether ramatroban enhances innate NK cell responses and IFN-λ responses by TPr and DPr2 antagonism, respectively, and reduces SARS-CoV-2 viral load remains to be investigated.

Currently, there is no treatment for the persisting symptoms following recovery from acute illness, referred to as long-haul COVID. Long-haul COVID is often characterized by neuropsychiatric manifestations including “brain fog,” anxiety or depression, fatigue and problems with mobility, dyspnea due to lung fibrosis and lung diffusion impairment, and microvascular thrombosis persisting for > 4 months in about 25% of patients.<sup>38 39</sup> Despite persistence of ground glass opacities 6 months later in patient 1, lung fibrosis was not detected. This is consistent with inhibition of the process triggering lung fibrosis by ramatroban in an animal model of silicosis that is associated with markedly increased pulmonary thromboxane A<sub>2</sub> and PGD<sub>2</sub>.<sup>40</sup> Moreover, in well-established animal models of depression, elevation in PGD<sub>2</sub> mediates depression-like behavior, while ramatroban restores object exploration and social interaction.<sup>41</sup> The above suggests that ramatroban may help prevent and/or treat certain long-haul COVID symptoms (Table 3).

This report has several limitations. Only 4 patients could be treated with ramatroban, and the duration of treatment was brief due to very limited availability of the drug in India. Only the first patient had laboratory studies performed. Patients 2, 3 and 4 were not examined by a physician and the clinical course was reported by patients or their relatives.

During the ongoing pandemic, there is an unmet need for a drug that can provide rapid relief of respiratory symptoms, respiratory distress and hypoxemia; halt progression of disease and avoid hospitalization, since the latter is associated with poor outcomes for the patient and added burden on the healthcare system. Ramatroban (Baynas®, Bayer Yakuhin, Ltd., Japan) has been safely used for the treatment of allergic rhinitis in Japan since 2000.<sup>15</sup> The usual adult oral dose of 75 mg twice daily achieves an average plasma concentration of about 0.1 mg/L or 240 nM which is sufficient to inhibit pulmonary venous constriction, platelet activation, and release of type 2 cytokines (Table 3).

The rapid and salutary responses to ramatroban reported here, its diverse actions targeting the major pathobiologic mechanisms underlying COVID-19 (Table 1 and Fig. 1), coupled with its oral bioavailability

and an excellent safety profile make ramatroban an attractive therapeutic agent to test in randomized controlled clinical trials.

**Table 3. Proposed effect of antagonizing Thromboxane A<sub>2</sub>/TPr and Prostaglandin D<sub>2</sub>/DPr2 signaling by ramatroban in patients with COVID-19**

COVID-19	Thromboxane A <sub>2</sub> /TPr	Prostaglandin D <sub>2</sub> /DPr2
<b>Endogenous agonists for the receptors</b>	Thromboxane A <sub>2</sub> F2-Isoprostanes	Prostaglandin D <sub>2</sub> 11-dehydro-thomboxane B <sub>2</sub>
<b>Acute effects of antagonism (minutes-hours)</b>	Hypoxemia ↓ , V/Q mismatch ↓ Bronchoconstriction ↓ Pulmonary edema ↓ Ý Pulmonary microvascular permeability ↓ Pulmonary capillary pressure ↓ Pulmonary venous constriction ↓  NK cell SARS-CoV-2 killing ↑ Ý TGFβ ↓	Hyaluronan accumulation ↓ Ý IL-13 ↓  Antiviral Ý IFN-λ ↑
<b>Subacute short-term effect of antagonism (days-weeks)</b>	Microvascular thrombosis ↓  Anti-inflammatory (thromboinflammation ↓)	Antiviral activity Ý Th1 Response ↑ Th2 Response ↓
<b>Long-term effect of antagonism (weeks-months)</b>	Brain fog ↓, Brain edema ↓ Ý Blood-brain barrier ↑  Lung fibrosis ↓ Ý TGFβ ↓	Depression ↓ Activity ↑

## References

1. Trust for American's Health. CDC Data Show High Hospitalization Rates for Diagnosed COVID-19 Patients with Underlying Conditions in the United States 2021 [Available from: <https://www.tfah.org/wp-content/uploads/2020/04/COVIDunderlyingconditions040320.pdf> accessed October 30 2021.
2. NIH. What's New in the Guidelines 2021 [Available from: <https://www.covid19treatmentguidelines.nih.gov/about-the-guidelines/whats-new/> accessed October 30 2021.
3. Archambault AS, Zaid Y, Rakotoarivelo V, et al. High levels of eicosanoids and docosanoids in the lungs of intubated COVID-19 patients. *The FASEB Journal* 2021;35(6) doi: 10.1096/fj.202100540r
4. Al-Hakeim HK, Al-Hamami SA, Almulla AF, et al. Increased Serum Thromboxane A2 and Prostacyclin but Lower Complement C3 and C4 Levels in COVID-19: Associations with Chest CT Scan Anomalies and Lowered Peripheral Oxygen Saturation. *COVID* 2021;1(2):489-502. doi: 10.3390/covid1020042
5. Hottz ED, Azevedo-Quintanilha IG, Palhinha L, et al. Platelet activation and platelet-monocyte aggregates formation trigger tissue factor expression in severe COVID-19 patients. *Blood* 2020 doi: 10.1182/blood.2020007252 [published Online First: 2020/07/18]
6. Larsson AK, Hagfjård A, Dahlén SE, et al. Prostaglandin D<sub>2</sub> induces contractions through activation of TP receptors in peripheral lung tissue from the guinea pig. *Eur J Pharmacol* 2011;669(1-3):136-42. doi: 10.1016/j.ejphar.2011.07.046 [published Online First: 2011/08/30]
7. Walch L, De Montpreville V, Brink C, et al. Prostanoid EP1- and TP-receptors involved in the contraction of human pulmonary veins. *British Journal of Pharmacology* 2001;134(8):1671-78. doi: 10.1038/sj.bjp.0704423
8. Craven PA, Studer RK, DeRubertis FR. Thromboxane/Prostaglandin Endoperoxide-Induced Hypertrophy of Rat Vascular Smooth Muscle Cells Is Signaled by Protein Kinase C-Dependent Increases in Transforming Growth Factor- $\beta$ . *Hypertension* 1996;28(2):169-76. doi: doi:10.1161/01.HYP.28.2.169
9. Witkowski M, Tizian C, Ferreira-Gomes M, et al. Untimely TGF $\beta$  responses in COVID-19 limit antiviral functions of NK cells. *Nature* 2021 doi: 10.1038/s41586-021-04142-6
10. Theken KN, Fitzgerald GA. Bioactive lipids in antiviral immunity. *Science* 2021;371(6526):237-38. doi: 10.1126/science.abf3192
11. Sposito B, Broggi A, Pandolfi L, et al. The interferon landscape along the respiratory tract impacts the severity of COVID-19. *Cell* 2021 doi: <https://doi.org/10.1016/j.cell.2021.08.016>
12. Gupta A, Kalantar-Zadeh K, Srinivasa RT. Ramatroban as a Novel Immunotherapy for COVID-19. *Molecular and Genetic Medicine* 2020;14(3) doi: 10.37421/jmglm.2020.14.457

13. Gupta A, Chiang KC. Prostaglandin D2 as a mediator of lymphopenia and a therapeutic target in COVID-19 disease. *Medical Hypotheses* 2020;143:110122. doi: 10.1016/j.mehy.2020.110122
14. Uller L, Mathiesen JM, Alenmyr L, et al. Antagonism of the prostaglandin D2 receptor CRTH2 attenuates asthma pathology in mouse eosinophilic airway inflammation. *Respiratory Research* 2007;8(1) doi: 10.1186/1465-9921-8-16
15. Ishizuka T, Matsui T, Okamoto Y, et al. Ramatroban (BAY u 3405): a novel dual antagonist of TXA2 receptor and CRTH2, a newly identified prostaglandin D2 receptor. *Cardiovasc Drug Rev* 2004;22(2):71-90. doi: 10.1111/j.1527-3466.2004.tb00132.x
16. Schuster DP, Kozlowski J, Brimiouelle S. Effect of thromboxane receptor blockade on pulmonary capillary hypertension in acute lung injury. 2001 Meeting of the American Thoracic Society. San Francisco, CA, 2001.
17. An J, Li JQ, Wang T, et al. Blocking of thromboxane A(2) receptor attenuates airway mucus hyperproduction induced by cigarette smoke. *Eur J Pharmacol* 2013;703(1-3):11-7. doi: 10.1016/j.ejphar.2013.01.042 [published Online First: 2013/02/13]
18. Kobayashi K, Horikami D, Omori K, et al. Thromboxane A2 exacerbates acute lung injury via promoting edema formation. *Scientific Reports* 2016;6(1):32109. doi: 10.1038/srep32109
19. Bauer J, Ripperger A, Frantz S, et al. Pathophysiology of isoprostanes in the cardiovascular system: implications of isoprostane-mediated thromboxane A2receptor activation. *British Journal of Pharmacology* 2014;171(13):3115-31. doi: 10.1111/bph.12677
20. Zhao Z, Hu J, Gao X, et al. Hyperglycemia via activation of thromboxane A2 receptor impairs the integrity and function of blood-brain barrier in microvascular endothelial cells. *Oncotarget* 2017;8(18):30030-38. doi: 10.18632/oncotarget.16273
21. Keikha R, Hashemi-Shahri SM, Jebali A. The relative expression of miR-31, miR-29, miR-126, and miR-17 and their mRNA targets in the serum of COVID-19 patients with different grades during hospitalization. *Eur J Med Res* 2021;26(1):75. doi: 10.1186/s40001-021-00544-4 [published Online First: 2021/07/15]
22. Wang H-W, Huang T-S, Lo H-H, et al. Deficiency of the MicroRNA-31&#x2013;MicroRNA-720 Pathway in the Plasma and Endothelial Progenitor Cells From Patients With Coronary Artery Disease. *Arteriosclerosis, Thrombosis, and Vascular Biology* 2014;34(4):857-69. doi: doi:10.1161/ATVBAHA.113.303001
23. Xue L, Gyles SL, Wetley FR, et al. Prostaglandin D2 Causes Preferential Induction of Proinflammatory Th2 Cytokine Production through an Action on Chemoattractant Receptor-Like Molecule Expressed on Th2 Cells. *The Journal of Immunology* 2005;175(10):6531-36. doi: 10.4049/jimmunol.175.10.6531

24. Xue L, Salimi M, Panse I, et al. Prostaglandin D2 activates group 2 innate lymphoid cells through chemoattractant receptor-homologous molecule expressed on TH2 cells. *Journal of Allergy and Clinical Immunology* 2014;133(4):1184-94.e7. doi: 10.1016/j.jaci.2013.10.056
25. Yang L, Liu S, Liu J, et al. COVID-19: immunopathogenesis and Immunotherapeutics. *Signal Transduction and Targeted Therapy* 2020;5(1) doi: 10.1038/s41392-020-00243-2
26. Lucas C, Wong P, Klein J, et al. Longitudinal analyses reveal immunological misfiring in severe COVID-19. *Nature* 2020;584(7821):463-69. doi: 10.1038/s41586-020-2588-y
27. Perlman S. COVID-19 poses a riddle for the immune system. *Nature* 2020;584(7821):345-46. doi: 10.1038/d41586-020-02379-1 [published Online First: 2020/08/19]
28. Skaria T, Burgener J, Bachli E, et al. IL-4 Causes Hyperpermeability of Vascular Endothelial Cells through Wnt5A Signaling. *PLOS ONE* 2016;11(5):e0156002. doi: 10.1371/journal.pone.0156002
29. Donlan AN, Sutherland TE, Marie C, et al. IL-13 is a driver of COVID-19 severity. *JCI Insight* 2021 doi: 10.1172/jci.insight.150107
30. Tanabe T, Fujimoto K, Yasuo M, et al. Modulation of mucus production by interleukin-13 receptor alpha2 in the human airway epithelium. *Clin Exp Allergy* 2008;38(1):122-34. doi: 10.1111/j.1365-2222.2007.02871.x [published Online First: 2007/11/22]
31. Gómez-Escobar LG, Hoffman KL, Choi JJ, et al. Cytokine signatures of end organ injury in COVID-19. *Scientific Reports* 2021;11(1):12606. doi: 10.1038/s41598-021-91859-z
32. Broggi A, Ghosh S, Sposito B, et al. Type III interferons disrupt the lung epithelial barrier upon viral recognition. *Science* 2020;369(6504):706-12. doi: 10.1126/science.abc3545
33. Werder RB, Lynch JP, Simpson JC, et al. PGD2/DP2 receptor activation promotes severe viral bronchiolitis by suppressing IFN-lambda production. *Sci Transl Med* 2018;10(440) doi: 10.1126/scitranslmed.aao0052
34. Ricke-Hoch M, Stelling E, Lasswitz L, et al. Impaired immune response mediated by prostaglandin E2 promotes severe COVID-19 disease. *PLOS ONE* 2021;16(8):e0255335. doi: 10.1371/journal.pone.0255335
35. Diao B, Wang C, Wang R, et al. Human kidney is a target for novel severe acute respiratory syndrome coronavirus 2 infection. *Nature Communications* 2021;12(1):2506. doi: 10.1038/s41467-021-22781-1
36. Tantry US, Bliden KP, Cho A, et al. First Experience Addressing the Prognostic Utility of Novel Urinary Biomarkers in Patients With COVID-19. *Open Forum Infectious Diseases* 2021;8(7) doi: 10.1093/ofid/ofab274

37. Westlund P, Kumlin M, Nordenström A, et al. Circulating and urinary thromboxane B2 metabolites in the rabbit: 11-dehydro-thromboxane B2 as parameter of thromboxane production. *Prostaglandins* 1986;31(3):413-43. doi: 10.1016/0090-6980(86)90106-1 [published Online First: 1986/03/01]
38. Townsend L, Fogarty H, Dyer A, et al. Prolonged elevation of D-dimer levels in convalescent COVID-19 patients is independent of the acute phase response. *J Thromb Haemost* 2021;19(4):1064-70. doi: 10.1111/jth.15267 [published Online First: 2021/02/16]
39. Huang L, Yao Q, Gu X, et al. 1-year outcomes in hospital survivors with COVID-19: a longitudinal cohort study. *The Lancet* 2021;398(10302):747-58. doi: 10.1016/s0140-6736(21)01755-4
40. Pang J, Qi X, Luo Y, et al. Multi-omics study of silicosis reveals the potential therapeutic targets PGD(2) and TXA(2). *Theranostics* 2021;11(5):2381-94. doi: 10.7150/thno.47627 [published Online First: 2021/01/28]
41. Onaka Y, Shintani N, Nakazawa T, et al. CRTH2, a prostaglandin D2 receptor, mediates depression-related behavior in mice. *Behavioural Brain Research* 2015;284:131-37.  
doi: <https://doi.org/10.1016/j.bbr.2015.02.013>

## Figures



increase in transcapillary pressure in pulmonary microvasculature, and transudation of fluid into the alveoli, thereby causing impaired gas exchange and ARDS. TxA<sub>2</sub>/TPr axis also induces bronchoconstriction and mucus secretion. TxA<sub>2</sub> is rapidly converted to 11-dehydro-TxB<sub>2</sub> in the lungs. PGD<sub>2</sub> and 11-dehydro-TxB<sub>2</sub> stimulate the DPr<sub>2</sub> receptor on Th<sub>2</sub> and ILC<sub>2</sub> cells leading to release of type 2 cytokines, IL-4 and IL-13. IL-4 promotes vascular permeability thereby exacerbating fluid transudation while IL-13 induces hyaluronic acid accumulation and mucus hypersecretion. Ramatroban inhibits the DPr<sub>2</sub> and TPr receptors thereby promoting pulmonary vasorelaxation, bronchorelaxation and improving capillary barrier function, while attenuating the maladaptive type 2 immune response and mucus secretion, thereby alleviating pulmonary edema and ARDS. Tx, thromboxane; PG, prostaglandin; TPr, thromboxane prostanoid receptor; DPr<sub>2</sub>; D-prostanoid receptor 2; Th<sub>2</sub>; T helper 2; ILC<sub>2</sub>; innate lymphoid class 2