

Natural Killer Cell-Related Gene Signature Predicts Malignancy of Glioma and the Survival of Patients

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Abstract

Background: Natural killer (NK) cells-based therapies are one of the most promising strategies against cancer. The aim of this study is to investigate the natural killer cell related genes and its prognostic value in glioma.

Methods: The Chinese Glioma Genome Atlas (CGGA), The Cancer Genome Atlas (TCGA) datasets were used to develop the natural killer cell-related signature. Risk score was built by a multivariate Cox proportional hazards model. A cohort of 326 glioma samples with whole transcriptome expression data from the CGGA database was included for discovery. TCGA dataset was used for validation. GO and KEGG were used to reveal the biological process and function associated with the natural killer cell-related signature. We also collected the clinical pathological features of patients with gliomas to analyze the association with tumor malignancy and patients' survival.

Results: We screened for NK-related genes and built a prognostic signature. Univariate and Multivariate Cox regression analysis were applied to identify and verify the risk score based on the signature. We found that NK-related risk score was independent of various clinical factors. Nature-killer cell gene expression is correlated with clinicopathological features of gliomas. Innovatively, we demonstrated the tight relation between the risk score and immune checkpoints, and found NK-related risk score combined with PD1/PDL1 patients could predict the patient outcome.

Conclusion: Natural killer cell-related gene signature can predict malignancy of glioma and the survival of patients, these results might provide new view for the research of glioma malignancy and individual immunotherapy.

Introduction

Glioma is the most common and malignant primary brain tumor, the prognosis of glioma patients varies greatly and mainly depends on the clinical characteristics[1]. Glioblastoma multiforme (GBM) is the most lethal and malignant brain tumor, the median overall survival of GBM is around 15 months despite surgery and combined radio- and chemo-therapy[1]. Recently, increasing evidence showed that immune infiltration was correlated with the prognosis of the glioma, precise therapies like target therapy and immunotherapy are promising ways to treat GBM. Immune-checkpoint inhibitors, Chimeric antigen receptor (CAR) T cell therapy, Natural killer cell-related therapies, Virotherapy, and Dendritic cells (DC) vaccination were the most encouraging areas in GBM therapy[2].

The molecular classification of central nervous system tumors in 2016 remarkably improved the diagnosis and prognosis prediction by IDH, MGMT methylation, TERT, TP53 et al [3]. However, more precise signatures are needed. Recently, a few gene expression-based risk signature in autophagy, hypoxia, ferroptosis, and glucose has explored their value in predicting prognosis of glioma patients[4–7]. Construction of the immune signature in the survival and malignancy prediction of glioma may lead to a more complete understanding of tumor microenvironment and immunotherapy.

Natural killer(NK) cells are innate cytotoxic lymphocytes encompassing distinct populations based on CD56 intensity in humans and involved in the surveillance and elimination of cancer[8]. As one of the most cutting-edge immunotherapeutic strategies, NK cells related therapies such as adoptive NK cell transfer, chimeric antigen receptor-expressing NK cells (CAR-NKs), bispecific and trispecific killer cell engagers (BiKEs and TriKEs) have emerged as a promising therapeutic target in glioma[9], breast cancer[10], lung cancer[11], colon cancer[12], prostate cancer[13] and hematological malignancies[14]. Although great progress has been made for natural killer cell-based therapy in preclinical and clinical research, there are many things we need to do to advance the research. For instance, we have learned how NK cells employ to recognize and eliminate tumor cells and how cancer cells can also educate and evade NK cell responses[11, 15], little is known about NK cells postsurgical dysfunction and why it works well in hematological malignancies while not good enough for solid tumors [16].

A couple of studies have investigated that the NK cells were one of the least numerous immune cell populations infiltrating the tumour. They represent around 2.11% of the total and the most abundant phenotype is CD56^{dim}CD16^{neg} [17]. Surprisingly, those limited NK cells were potent effectors against brain tumor. Lee SJ et al showed that human NK cells had a strong effect against GBM and could prevent systemic metastasis of GBM [18]. Mukherjee S et al [19] demonstrated that curcumin phytosome induced natural killer cell-dependent repolarization of GBM tumor-associated microglia/macrophages to kill GBM and their stem cells. Scientists also found that virotherapy is limited partially by an antiviral NK cell response involving specific natural cytotoxicity receptors to enhance GBM virotherapy [9].

The prognostic significance of NK cells' activity has been demonstrated in patients with a few solid tumours [20]. However, little is known about NK cells signature in glioma [21]. In the current study, we screened for NK-related genes and built a prognostic signature. Univariate and Multivariate Cox regression analysis was applied to identify and verify the risk score based on the signature. We analyze the NK-related risk score and various clinical factors, including age, sex, IDH1 mutation, and GBM subtype, etc. GO and KEGG were used to reveal the biological process and function associated with the natural killer cell-related signature. Innovatively, we combined the risk score with immune checkpoints to sort out the glioma patients for patient prognosis prediction.

Materials And Methods

Data collection

The mRNA sequencing data of genes encoding calmodulin dependent proteins was downloaded from The Cancer Genome Atlas (TCGA) dataset which was set as the training cohort. The mRNA sequencing data from The Chinese Glioma Genome Atlas (CGGA) dataset was set as the validation cohort. Corresponding clinical information was also downloaded.

Gene Signature Building

We downloaded gene list from import (<https://www.immport.org/resource>) and MSigDB (The Molecular Signatures Database), which were verified to be involved in NK cell-related pathways (Stockwell et al., 2017). Univariate Cox analysis was firstly performed, and genes with P values less than or equal to 0.1 were retained. To assess whether the risk score is independent of other clinical factors, multivariable Cox proportion hazard regression models were performed. We then developed a risk score model on the basis of the expression level of the prognosis-significant genes and the clinical and molecular features of patients, including age, sex, MGMT promoter methylation, IDH1 mutation stage, 1p19q co-deletion, glioma subtype and tumor WHO grade., and their regression coefficients(Coeffs) were derived from the univariate Cox regression analysis. The risk score was expressed as $(\text{exprgene1} \times \text{coefficientgene1}) + (\text{exprgene2} \times \text{coefficientgene2}) + (\text{exprgene3} \times \text{coefficientgene3})$.

Go And Kegg Pathway Analyses Of Degs

The DEGs between NK cell-related high-risk and low-risk groups were screened with a $\log_2\text{FC} > 1$ and adjusted $p < 0.05$. GO analysis with functions including molecular function (MF), biological pathways (BP), cellular component (CC), and KEGG pathway analyses were performed to the DEGs by using R software at the functional level. $P < 0.05$ and $q < 0.05$ were considered to have a significance.

Statistical analysis

The log-rank test was applied to compare overall survival difference between different groups. The Student's t-test was employed to compare two groups and ANOVA analysis was performed to compare multiple groups. $P < 0.05$ was considered as a statistical difference.

Results

Construction of Natural Killer Cell-Related Gene Signature

To characterize the Natural killer cell-related gene expression in gliomas, we examined the RNA-seq data of glioma patients from CGGA and TCGA datasets. We found 134 Natural Killer cell-related genes in glioma from immport, subsequently, 18 GO pathway (GOBP_NATURAL_KILLER_CELL_ACTIVATION, GOBP_NATURAL_KILLER_CELL_ACTIVATION_INVOLVED_IN_IMMUNE_RESPONSE, GOBP_NATURAL_KILLER_CELL_CHEMOTAXIS, et al) was analyzed to be related with natural killer cell from MSigDB, from which 397 genes were collected, after removing the replication genes, 244 genes were left. Details on the Natural Killer Cell-Related genes are presented in Supplement Table 1 and Table 2. A gene-based prognostic model was then established to evaluate the risk of each patient as described in the methods. Consequently, a three-gene signature was generated, and signature risk score was calculated as: $\text{risk score} = (\text{ULBP1} \times 0.048) + (\text{CD70} \times 0.016) + (\text{BID} \times -0.002)$. To validate this gene set, we also calculated patients' risk scores of the CGGA cohort with the same regression Coeffs.

Overall survival status of glioma patient based on gene signature risk score in CGGA and TCGA datasets

Patients with different kinds of glioma were divided into two groups based on their median risk scores. Their percentage of alive patients was 51.6% in the low-risk group versus 21.3% in the high-risk group in the CGGA dataset (Fig. 1A). Similarly, alive patients were 83.3% in low-risk group versus 52.5% in high-risk group in the TCGA dataset (Fig. 1B). The Kaplan–Meier curve for the CGGA dataset showed that the high-risk patients had significantly shorter OS than low-risk patients in the glioma (Fig. 1C), WHO low-grade glioma, and GBM (Fig. 1C). The consistency of results were validated for the TCGA (Figs. 1D).

Nature-killer Cell Gene Signature Is Associated With Clinicopathological Features

The heatmap showed that Nature-killer cell gene expression was correlated with WHO grade, IDH1 mutation, MGMT promoter methylation, 1p19q co-deletion, tumor subtype in glioma patients in CGGA(Fig. 2A) and TCGA datasets (Fig. 2B). Those data were also confirmed by Cox regression analysis (Table 1 and Table 2). After mining the CGGA dataset, the RNA expression of Natural-killer cell gene was higher in high grade glioma than WHO grade II patients (Fig. 3A), higher in the IDH wild type than IDH mutant glioma (Fig. 3B). It was the highest expressed in the Mesenchymal group when compared with Classical, Neural and Proneural glioma types (Fig. 3C). These consistent results were also validated in the TCGA datasets (Fig. 3D-F). Moreover, univariate Cox regression and multivariate Cox regression of the signature of the natural killer-related genes were performed in the CGGA dataset ($p < 0.001$, univariate Cox regression; $p < 0.05$, multivariate Cox regression, Table 1). The independence of the clinical prognostic significance of the signature in glioma. The risk score showed significance in both univariate and multivariate Cox regression. Similar results were also validated in the TCGA dataset (Table 2). The patients with a high-risk score had a markedly higher mortality rate than those with a low-risk score in these two datasets. Meanwhile, with an increase in glioma grade, the risk score increased.

GO and KEGG Pathway Analyses (Gene functional characteristics related to risk scores)

To investigate the function of NK cell-related genes in GBM cells, we analyzed different functional enrichment between low and high-risk cases. TOP 20 pathway type, biological process, cellular component, and molecular function were demonstrated by GO/KEGG enrichment respectively (Fig. 4A and B). The GO enrichment analysis showed the most enrichment pathway was phagosome; the most activated biological processes were neutrophil activation involved in immune response, neutrophil degranulation, neutrophil activation, neutrophil-mediated immunity; the cellular components were largely enriched in extracellular matrix, endosome membrane, and adherens junction; the most enriched molecular function was cell adhesion molecule binding (Fig. 4A). Similarly, the KEGG enrichment analysis confirmed that focal adhesion and Human T-cell leukemia virus 1 infection were the major activated pathways which were also enriched by GO analysis. The major biological process in KEGG were neutrophil-mediated immunity and neutrophil activation; the most enriched cellular component and molecular function were adherens junction, cell adhesion molecule binding and transcription coregulator activity, all of which was similar to the GO analysis (Fig. 4B). Taken together, these results indicated that

the difference between low and high-risk score of NK cell-related gene signature were lines in immune-related adhesion, neutrophil activation and T cell leukemia virus 1 infection.

Correlation analysis between risk score (RS) and immune checkpoints/NK marker genes

To understand the correlation between risk score (RS) and immune checkpoints/NK marker genes, we analyzed the data by circus plots and found that gene signature risk score was related with PDL1, TIM3 and STAT3 in CGGA and TCGA datasets (Fig. 5A). As with the NK markers evaluated with the risk score, we demonstrated that risk score was correlated with CD16, CD226, CD96 and CD112. In summary, the Nature-killer cell gene signature expression is closely related to immune-related pathways and cancer immunotherapy process (Fig. 5B).

Prediction of patient outcome based on the RS and immune checkpoint gene expression

To deeply figure out the status between the risk score, checkpoint gene expression and patient survival. We initially made a Pearson analysis between PD1, PDL1 and risk score. As showed in Fig. 6A, the correlation coefficient of Pearson analysis of PD1 and RS was 0.23 and 0.18 in CGGA and TCGA datasets respectively, the correlation coefficient of PDL1 and RS was 0.10 and 0.38 in CGGA and TCGA. These data indicated that RS has a tight relation with PD1/PDL1 expression (Fig. 6A). Then, we stratified the RS to low and high group and found that both PD1 and PDL1 were higher expressed in high-risk score group than the low-score group by mining both CGGA and TCGA datasets (Fig. 6B). Lastly, we sorted out the data to four groups to better understand the RS, PD1/PDL1 and patient prognosis, Kaplan–Meier survival curves of OS among four patient groups showed that low-RS-low PD1 group as well as low-RS-low PDL1 had a better overall survival than other three groups (Fig. 6C). Those data was verified by TCGA dataset (Fig. 6D). In summary, NK cell-related gene signature combined with PD1/PDL1 can be applied to predict patient's prognosis, and low-RS-low-PD1/PDL1 patients showed better survival outcomes.

Discussion

In this work, we investigated the association between NK cell-related gene signature and glioma. Firstly, we developed an NK cell-related signature and confirmed that it was closely associated with the overall survival of patients in the CGGA and TCGA datasets. Subsequently, we found that this signature could distinguish the clinical and molecular features of gliomas, including WHO grade, TCGA subtype, IDH mutational status, 1p19q co-deletion and MGMT promoter methylation. Based on the differentially expressed genes of the risk score, GO enrichment results indicated that the major difference between high-risk and low-risk score focused on virus infection, immune system, neuropil mediated immunity, cell adhesion. KEGG enrichment results also confirmed that the above immune-related functional and signaling pathways. These results suggested that low-risk score NK cells related signature activated immune system through immune cells infiltration and cell adhesion.

Immunotherapy has shown encouraging benefits for many cancer types. In the current study, we found RS was tightly related to NK CD markers, CD96 and TIGIT together with the co-stimulatory receptor CD226

form a pathway which could enhance the immune response [22]. In addition, Sun H et al[23] found that human intratumoral CD96 + NK cells are functionally exhausted and patients with higher intratumoral CD96 expression exhibit poorer clinical outcomes. CD 16, also known as FcγRIII, is a differentiation molecule found on the surface of natural killer cells, which antibodies, such as cetuximab could mediate apoptosis by CD16 receptors after the recognition [24]. NK cells are large granular lymphocytes of the innate immune system which can directly lyse infected or tumor cells [25, 26]. Increasing evidence demonstrated that NK cells played a vital role in killing GBM by different approaches like KIR, CD16, IFN-γ, TNF-α, NIKG2D, TGF-β, CAR-NK and NK-exosomes [17, 26]. Clearly, NK cell-based immunotherapy is more and more attractive for GBM treatment [2]. Our study showed RS was interconnected with CD16, CD226, CD96 and CD112, which could activate NK cells to kill glioma cells to achieve prolonged survival.

Immune checkpoints were the most promising treatment targets against cancer. Thus, we investigated the correlation between RS with PDL1 and TIM3, which showed low RS linked high PDL1, high RS linked with low TIM3. Laterly, we combined checkpoints and RS to predict the overall survival of glioma patients, as expected, low-RS-low-PD1/PDL1 group gained the most prolonged OS. Therefore, high RS is recognized as an unfavorable feature of glioma.

In summary, our research provided important prognostic resources based on NK cell-related gene signature in glioma. It will contribute to the exploration of NK cell research to promote novel immunotherapy against glioma.

Declarations

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Author contributions Yu Zeng and Fangkun Liu designed and performed the research and wrote the manuscript. All authors contributed to writing and critically revising the manuscript.

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Availability of data and materials The datasets generated and analyzed during the current study are available from the corresponding author on reasonable request.

Conflict of interest none

Consent for publication All listed authors have actively participated in the study and have read and approved the submitted manuscript.

Ethical approval and ethical standards The study was approved by the ethics committee of the Xiangya Hospital, Central South University.

References

1. Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, Belanger K, Brandes AA, Marosi C, Bogdahn U, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med*. 2005;352(10):987–96.
2. Lim M, Xia Y, Bettegowda C, Weller M. Current state of immunotherapy for glioblastoma. *Nat Rev Clin Oncol*. 2018;15(7):422–42.
3. Louis DN, Perry A, Reifenberger G, von Deimling A, Figarella-Branger D, Cavenee WK, Ohgaki H, Wiestler OD, Kleihues P, Ellison DW. The 2016 World Health Organization Classification of Tumors of the Central Nervous System: a summary. *Acta Neuropathol*. 2016;131(6):803–20.
4. Liu HJ, Hu HM, Li GZ, Zhang Y, Wu F, Liu X, Wang KY, Zhang CB, Jiang T. Ferroptosis-Related Gene Signature Predicts Glioma Cell Death and Glioma Patient Progression. *Front Cell Dev Biol*. 2020;8:538.
5. Lin W, Wu S, Chen X, Ye Y, Weng Y, Pan Y, Chen Z, Chen L, Qiu X, Qiu S. Characterization of Hypoxia Signature to Evaluate the Tumor Immune Microenvironment and Predict Prognosis in Glioma Groups. *Front Oncol*. 2020;10:796.
6. Zhao S, Cai J, Li J, Bao G, Li D, Li Y, Zhai X, Jiang C, Fan L. Bioinformatic Profiling Identifies a Glucose-Related Risk Signature for the Malignancy of Glioma and the Survival of Patients. *Mol Neurobiol*. 2017;54(10):8203–10.
7. Wang QW, Liu HJ, Zhao Z, Zhang Y, Wang Z, Jiang T, Bao ZS. Prognostic Correlation of Autophagy-Related Gene Expression-Based Risk Signature in Patients with Glioblastoma. *Onco Targets Ther*. 2020;13:95–107.
8. Demaria O, Cornen S, Daeron M, Morel Y, Medzhitov R, Vivier E. Harnessing innate immunity in cancer therapy. *Nature*. 2019;574(7776):45–56.
9. Alvarez-Breckenridge CA, Yu J, Price R, Wojton J, Pradarelli J, Mao H, Wei M, Wang Y, He S, Hardcastle J, et al. NK cells impede glioblastoma virotherapy through NKp30 and NKp46 natural cytotoxicity receptors. *Nat Med*. 2012;18(12):1827–34.
10. Al Absi A, Wurzer H, Guerin C, Hoffmann C, Moreau F, Mao X, Brown-Clay J, Petrolli R, Casellas CP, Dieterle M, et al. Actin Cytoskeleton Remodeling Drives Breast Cancer Cell Escape from Natural Killer-Mediated Cytotoxicity. *Cancer Res*. 2018;78(19):5631–43.
11. Cong J, Wang X, Zheng X, Wang D, Fu B, Sun R, Tian Z, Wei H. Dysfunction of Natural Killer Cells by FBP1-Induced Inhibition of Glycolysis during Lung Cancer Progression. *Cell Metab*. 2018;28(2):243–55 e245.
12. Otegbeye F, Ojo E, Moreton S, Mackowski N, Lee DA, de Lima M, Wald DN. Inhibiting TGF-beta signaling preserves the function of highly activated, in vitro expanded natural killer cells in AML and

- colon cancer models. *PLoS One*. 2018;13(1):e0191358.
13. Tang M, Gao S, Zhang L, Liu B, Li J, Wang Z, Zhang W. Docetaxel suppresses immunotherapy efficacy of natural killer cells toward castration-resistant prostate cancer cells via altering androgen receptor-lectin-like transcript 1 signals. *Prostate*. 2020;80(10):742–52.
 14. Crinier A, Narni-Mancinelli E, Ugolini S, Vivier E. SnapShot: Natural Killer Cells. *Cell*. 2020;180(6):1280–0 e1281.
 15. Chan IS, Knutsdottir H, Ramakrishnan G, Padmanaban V, Warriar M, Ramirez JC, Dunworth M, Zhang H, Jaffee EM, Bader JS, et al: **Cancer cells educate natural killer cells to a metastasis-promoting cell state**. *J Cell Biol* 2020, 219(9).
 16. Hodgins JJ, Khan ST, Park MM, Auer RC, Ardolino M. Killers 2.0: NK cell therapies at the forefront of cancer control. *J Clin Invest*. 2019;129(9):3499–510.
 17. Kmiecik J, Zimmer J, Chekenya M. Natural killer cells in intracranial neoplasms: presence and therapeutic efficacy against brain tumours. *J Neurooncol*. 2014;116(1):1–9.
 18. Lee SJ, Kang WY, Yoon Y, Jin JY, Song HJ, Her JH, Kang SM, Hwang YK, Kang KJ, Joo KM, et al. Natural killer (NK) cells inhibit systemic metastasis of glioblastoma cells and have therapeutic effects against glioblastomas in the brain. *BMC Cancer*. 2015;15:1011.
 19. Mukherjee S, Fried A, Hussaini R, White R, Baidoo J, Yalamanchi S, Banerjee P. Phytosomal curcumin causes natural killer cell-dependent repolarization of glioblastoma (GBM) tumor-associated microglia/macrophages and elimination of GBM and GBM stem cells. *J Exp Clin Cancer Res*. 2018;37(1):168.
 20. Zhang S, Liu W, Hu B, Wang P, Lv X, Chen S, Shao Z. Prognostic Significance of Tumor-Infiltrating Natural Killer Cells in Solid Tumors: A Systematic Review and Meta-Analysis. *Front Immunol*. 2020;11:1242.
 21. Ren F, Zhao Q, Huang L, Zheng Y, Li L, He Q, Zhang C, Li F, Maimela NR, Sun Z, et al. The R132H mutation in IDH1 promotes the recruitment of NK cells through CX3CL1/CX3CR1 chemotaxis and is correlated with a better prognosis in gliomas. *Immunol Cell Biol*. 2019;97(5):457–69.
 22. Dougall WC, Kurtulus S, Smyth MJ, Anderson AC. TIGIT and CD96: new checkpoint receptor targets for cancer immunotherapy. *Immunol Rev*. 2017;276(1):112–20.
 23. Sun H, Huang Q, Huang M, Wen H, Lin R, Zheng M, Qu K, Li K, Wei H, Xiao W, et al. Human CD96 Correlates to Natural Killer Cell Exhaustion and Predicts the Prognosis of Human Hepatocellular Carcinoma. *Hepatology*. 2019;70(1):168–83.
 24. Veluchamy JP, Spanholtz J, Tordoir M, Thijssen VL, Heideman DA, Verheul HM, de Gruijl TD, van der Vliet HJ. Combination of NK Cells and Cetuximab to Enhance Anti-Tumor Responses in RAS Mutant Metastatic Colorectal Cancer. *PLoS One*. 2016;11(6):e0157830.
 25. Golan I, Rodriguez de la Fuente L, Costoya JA. **NK Cell-Based Glioblastoma Immunotherapy**. *Cancers (Basel)* 2018, 10(12).
 26. Pellegatta S, Eoli M, Frigerio S, Antozzi C, Bruzzone MG, Cantini G, Nava S, Anghileri E, Cuppini L, Cuccarini V, et al. The natural killer cell response and tumor debulking are associated with prolonged

survival in recurrent glioblastoma patients receiving dendritic cells loaded with autologous tumor lysates. *Oncoimmunology*. 2013;2(3):e23401.

Tables

Due to technical limitations, table 1 and 2 is only available as a download in the Supplemental Files section.

Figures

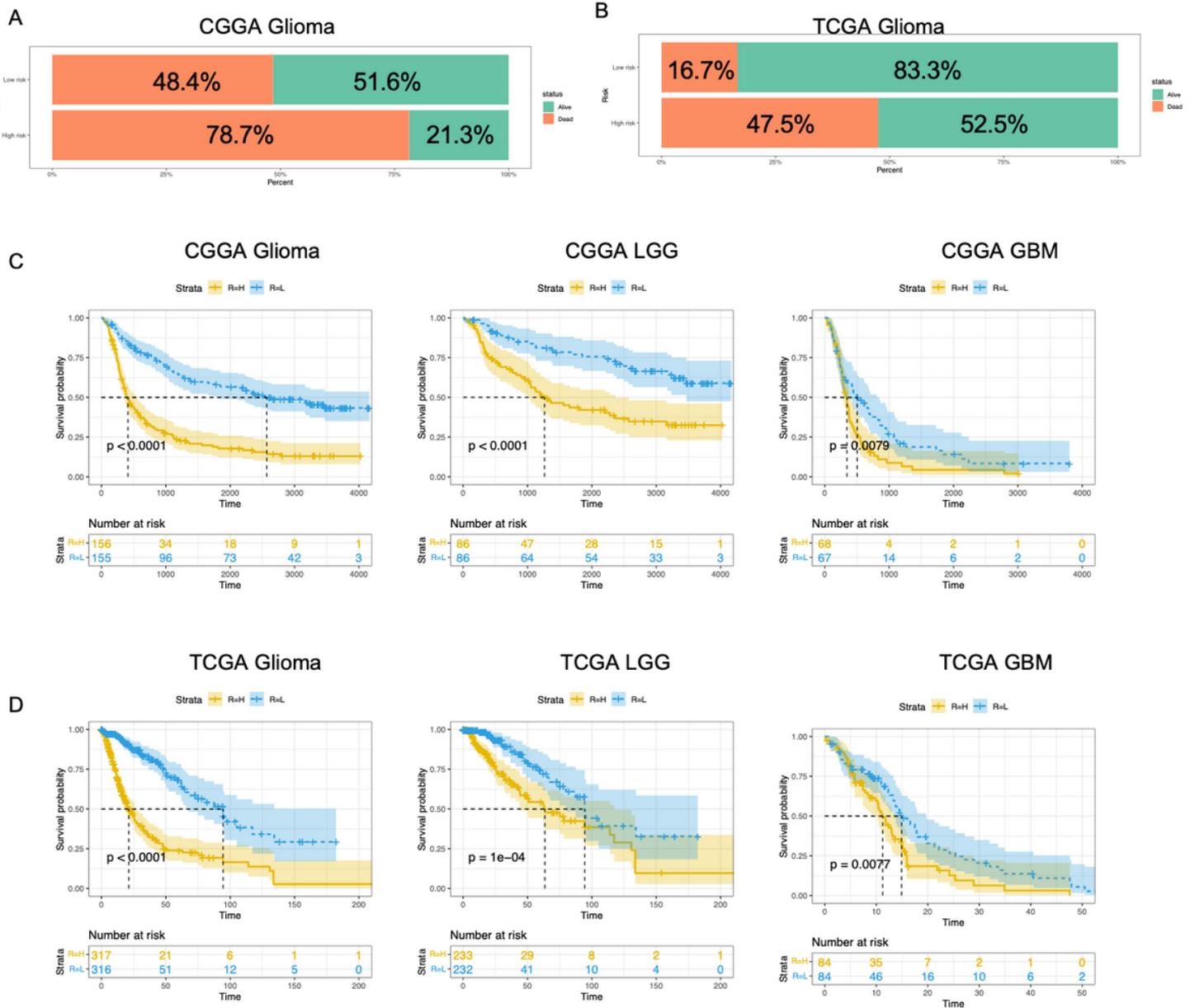


Figure 1

Overall survival status of glioma patient based on gene signature risk score in CGGA and TCGA datasets. (A-B) Mortality rates of the high- and low-risk patient groups; (C-D) Kaplan–Meier overall survival curve among high- and low-risk groups.

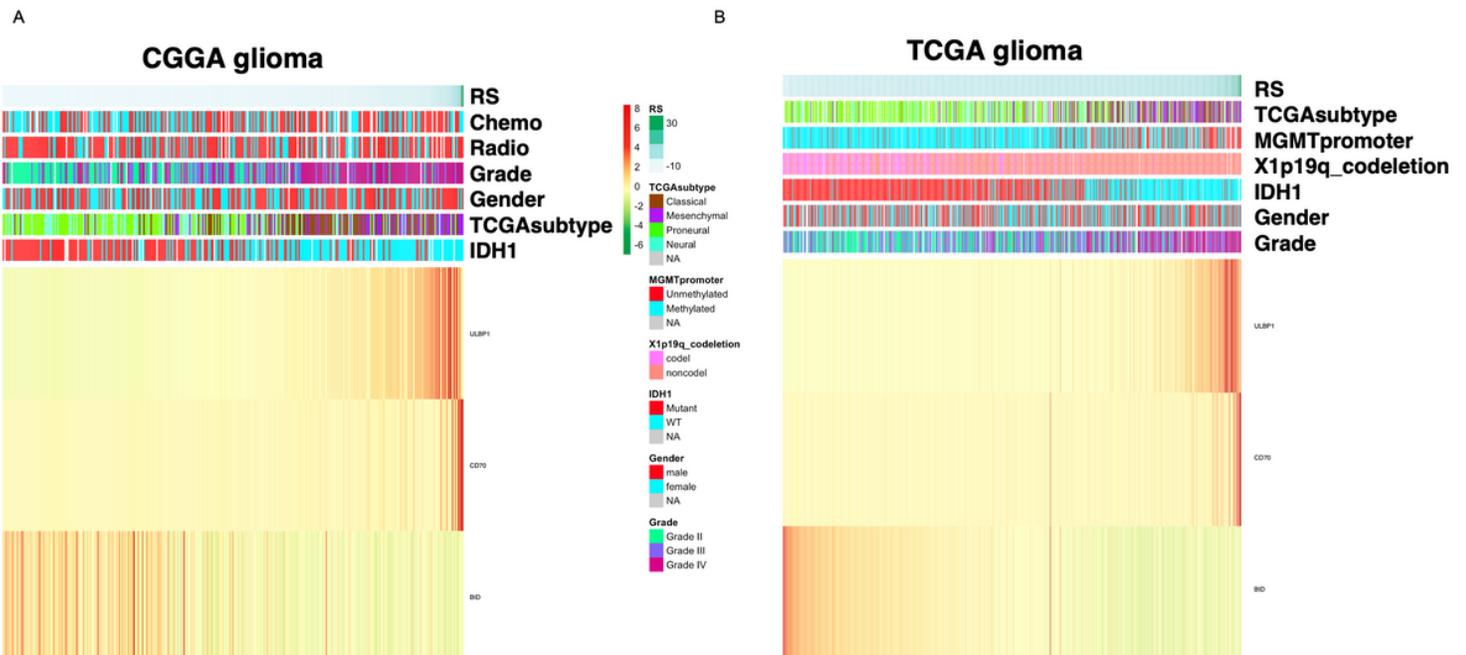


Figure 2

Nature-killer cell gene signature is associated with clinicopathological features in CGGA and TCGA datasets(A and B).

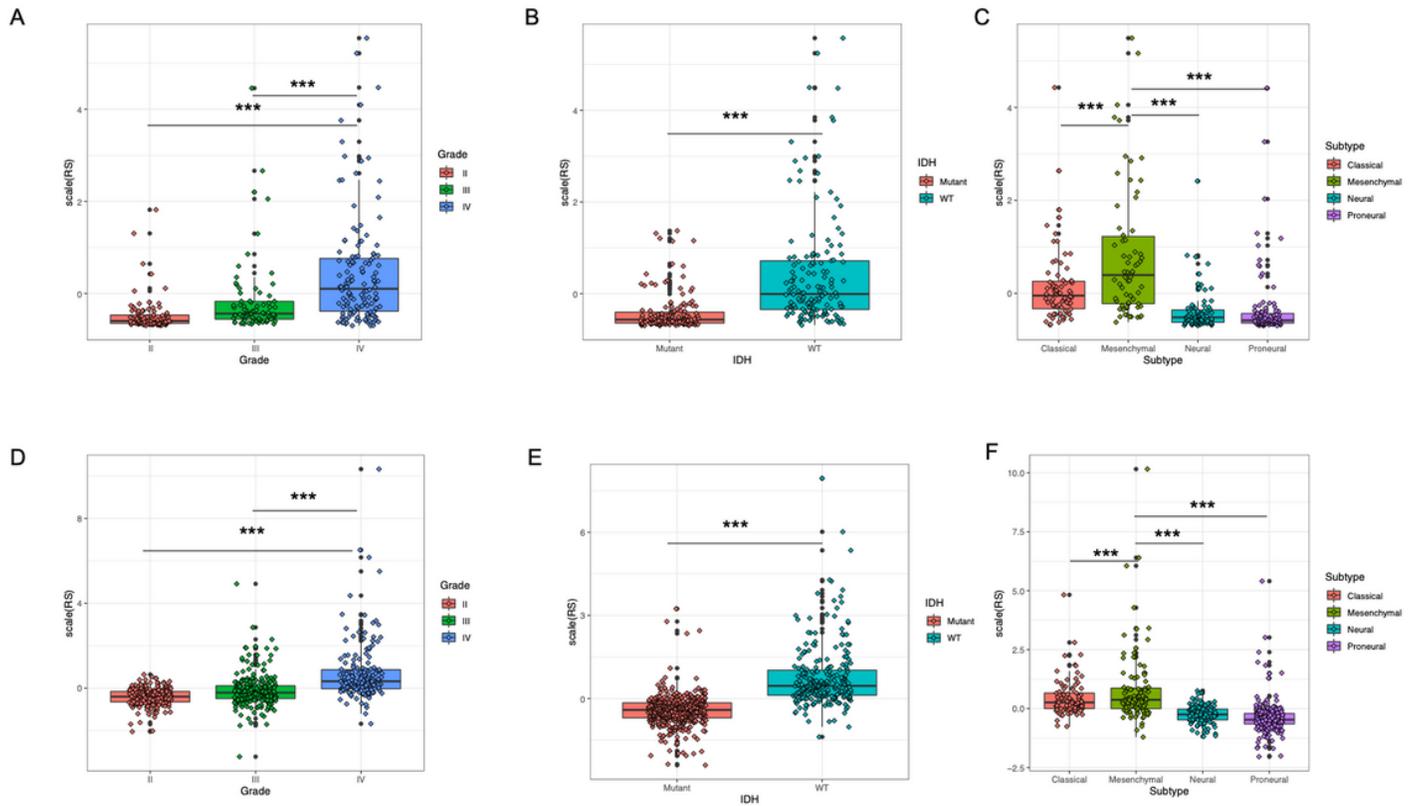


Figure 3

The association of Nature-killer cell gene signature and molecular features of gliomas(A-F).

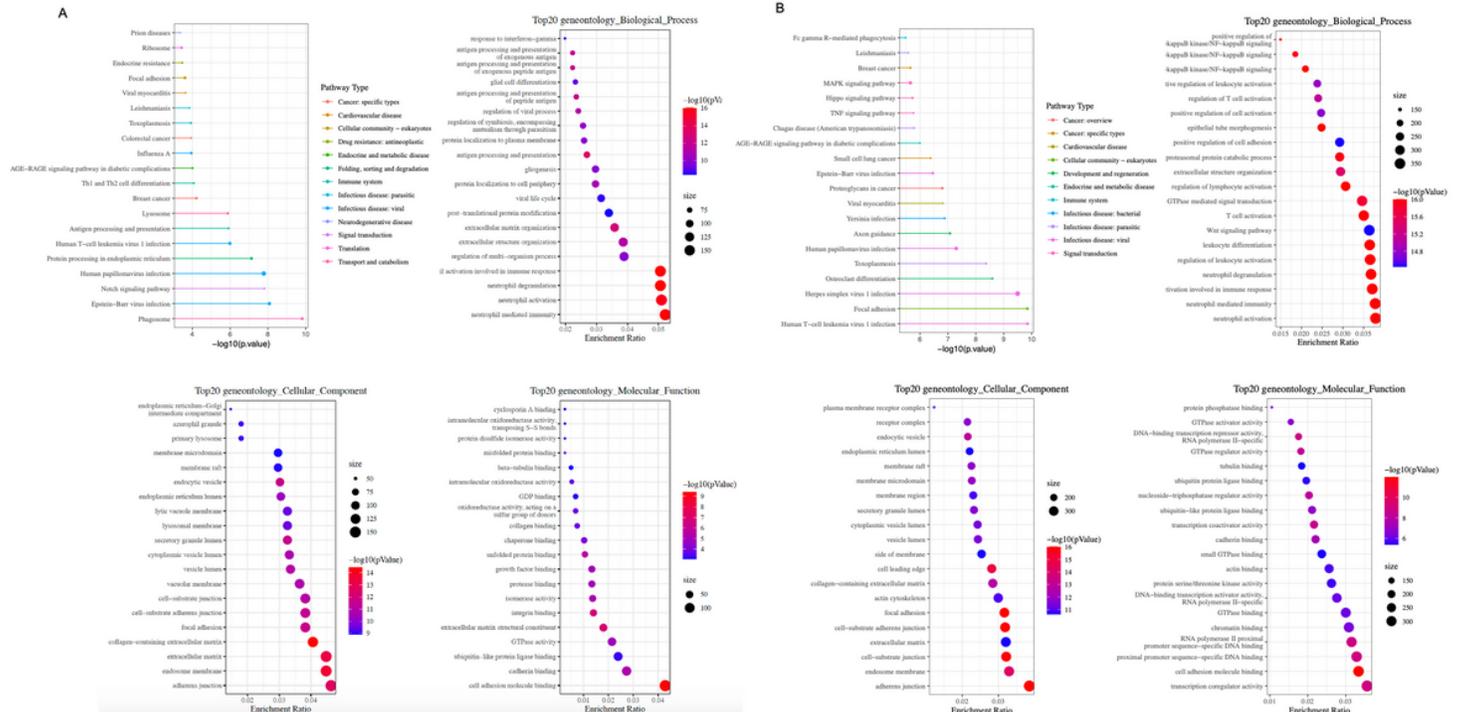


Figure 4

Gene functional characteristics related to risk scores. GO and KEGG analysis of differential genes between low- and high risk cases in two cohorts(A and B).

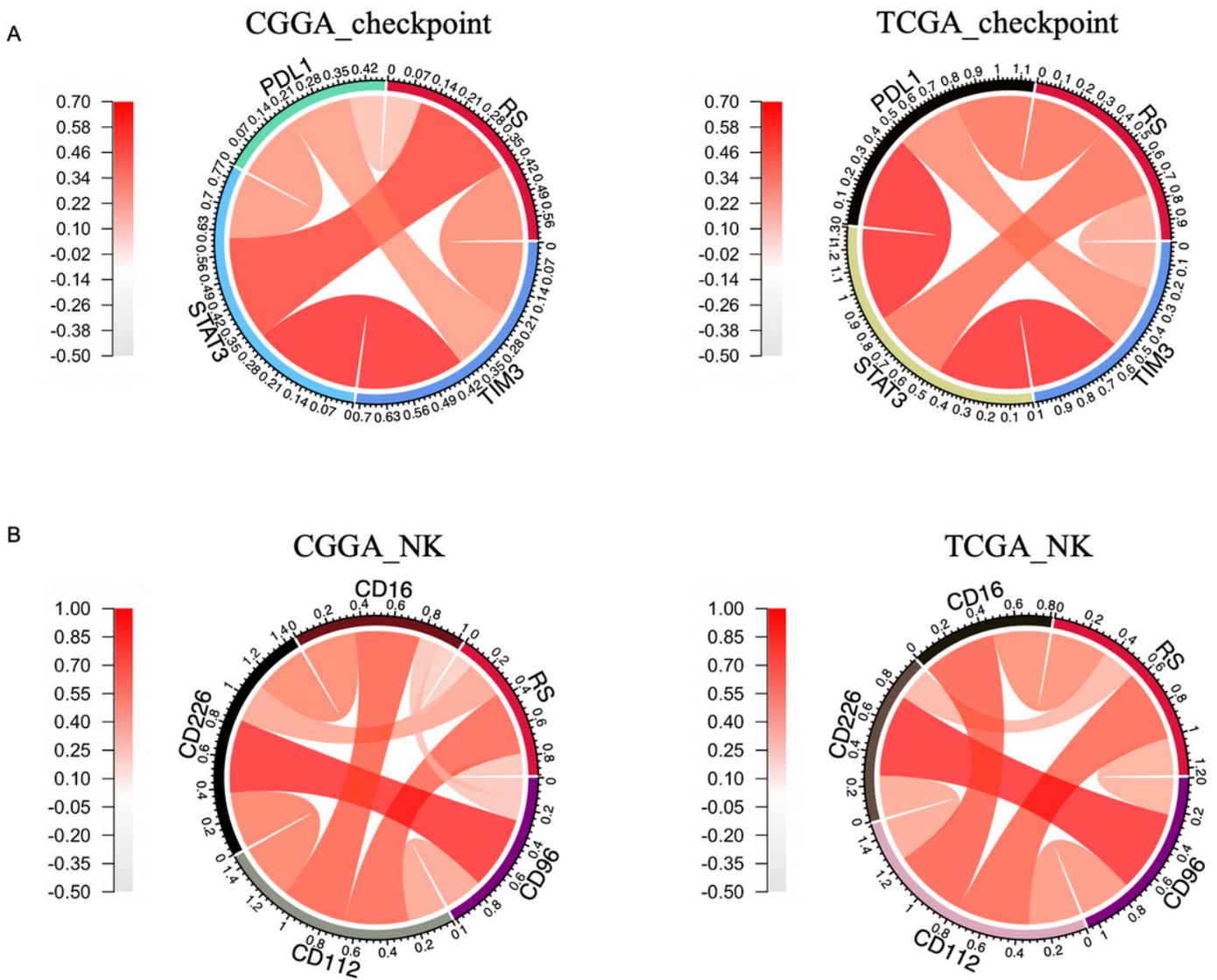


Figure 5

Circos plots showed correlation analysis between risk score (RS) and immune checkpoints/NK marker genes in CGGA and TCGA dataset (A and B).

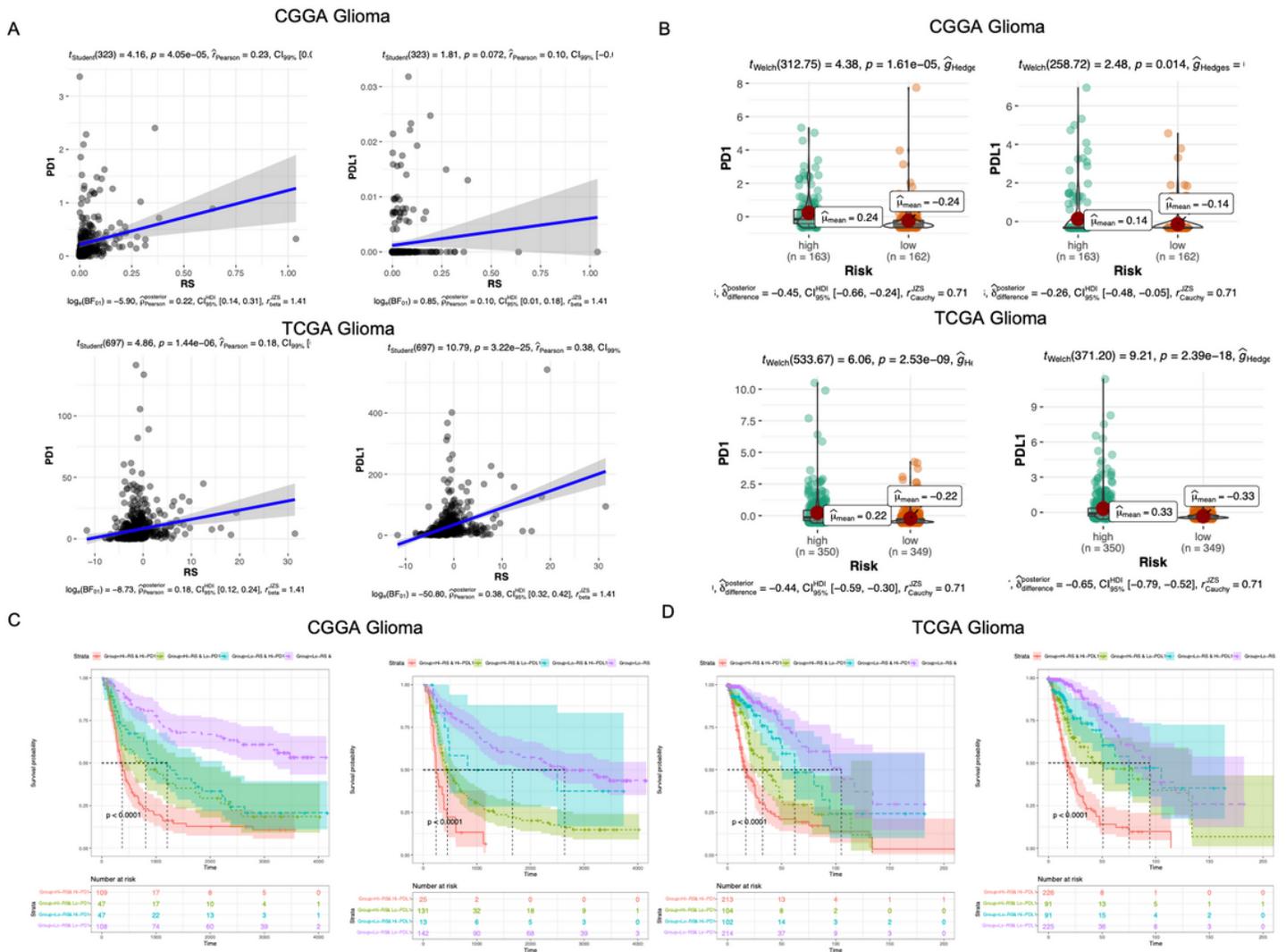


Figure 6

Prediction of patient outcome based on the RS and immune checkpoint gene expression. (A) Correlation coefficient of the RS and PD1/PDL1 gene expression. (B) The expression distribution of PD1/PDL1 in the high- and low-risk groups. (C) Overall survival curves of glioma patient stratified by the RS and immune checkpoint gene expression.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [Table1CGGA.xlsx](#)
- [Table2TCGA.xlsx](#)
- [SupplementTable1NKIMMPORT.xls](#)
- [SupplementTable2MolecularSignaturesDatabase.xlsx](#)