

DNA methylation abnormalities of imprinted genes in congenital heart disease: A pilot study

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Abstract

Background: Congenital heart disease (CHD) is resulted from the interaction of genetic aberration and environmental factors. Imprinted genes, which are regulated by epigenetic modifications, are essential for the normal embryonic development. However, the role of imprinted genes in the pathogenesis of CHD remains unclear.

Results: We investigated the effect of imprinted gene methylation on the pathogenesis of CHD. Eighteen imprinted genes that are known to affect early embryonic development were selected and the methylation modification genes were detected by massarray in 27 CHD children and 28 healthy children. Altered methylation of 8 imprinted genes was found, including 2 imprinted genes with hypermethylation of *GRB10* and *MEST* and 6 genes with hypomethylation of *PEG10*, *NAP1L5*, *INPP5F*, *PLAGL1*, *NESP* and *MEG3*. Stratified analysis showed that the methylation degree of imprinted genes was different in different types of CHD. Risk analysis showed that 6 imprinted genes within a specific methylation level range were the risk factor for CHD except *MEST* and *NAP1L5*.

Conclusion: Altered methylation of imprinted genes is associated with CHD and varies in different types of CHD. Further experiments are warranted to identify the methylation characteristics of imprinted genes in different types of CHD and clarify the pathogenesis mechanism of imprinted genes in CHD.

Background

Congenital heart disease (CHD) is the most common type of congenital malformations. Despite the diversity of the CHD category, there appear to be a shared epigenetic factor in the etiology of CHD for the following reasons. First, the genetic changes associated with many types of CHD, such as point mutation and chromatin abnormality (trisomy 21), remain unclear [1]. Second, environmental factors have been found to be related to CHD [2, 3]. Third, maternal folic acid supplementation significantly reduces the incidence of CHD through epigenetic programming [4].

Among the different epigenetic mechanisms, the potential relationship between abnormal DNA methylation and CHD has been increasingly recognized. DNA methylation has been found to be highly dynamic with the feature of demethylation in cardiomyocyte-associated gene sets during cardiomyocyte development [5]. Studies have shown that there are hypermethylation of myocardial-related genes in myocardial tissues of CHD, which is closely related to gene downregulation [6, 7]. In CHD patients, decreased transcriptional activity of *CITE2*, *ZIC3*, *NR2F2* and *BRG1* is associated with abnormal methylation [8–11].

DNA methylation has a major influence on the establishment of imprinting markers [12]. Several human disorders have been found to be associated with varied methylation modifications at imprinting control regions. For example, Silver-Russell syndrome is characterized by hypomethylation and Prader-Willi Syndrome by hypermethylation of the related imprinted genes [13]. Epigenetic imprinting is particularly

vulnerable in early embryonic development. Exploring the epigenetic modification of imprinted genes may provide pathogenesis mechanism of congenital disorders including CHD.

In placental mammals, the imprinted genes regulate embryonic development. At present, about 94 human imprinted genes have been identified [14]. It has been proposed that *IGF2* and *GRB10* and co-regulated genes of *PEG1/MEST*, *GTL2/MEG3*, *CDKN1C*, *PLAGL1* and *DLK1* regulates embryonic growth and development by forming a proposed "imprinted gene network" [15]. The most well-studied example of involving in cardiac development is *DLK1-DIO3*, encoded ncRNAs, which participate in the commitment of the mesoderm to different subsets of a specific cardiac cell lineage through many *DLK1-DIO3* ncRNAs [16]. However, there has been no study on the epigenetic modification of imprinted genes in CHD.

Therefore, we aimed to investigate the effect of imprinted genes on the pathogenesis of CHD through methylation modification. We selected 18 imprinted genes that are known to play an important role in early embryonic developments.

Results

Comparison of the altered methylation levels of the imprinted genes between CHD and control groups

Table 1 showed the values and statistical results. The methylation of 18 imprinted genes in 28 controls showed low heterogeneity except for NNST, which may reflect truly the methylation level of these imprinted genes in normal human. Compared with control group, 8 of the 18 selected genes were found to be significantly different in CHD group; two of them were hypermethylated and 6 hypomethylated. The two hypermethylated-imprinted genes were *GRB10* and *MEST*, in which *GRB10* increased from 43.42% in the normal group to 51.12% ($P < 0.01$) in the CHD group and *MEST* increased from 53.22–56.6% ($P < 0.05$). The 6 hypomethylation-imprinted genes were *PEG10*, *NAP1L5*, *INPP5F*, *PLAGL1*, *NESP* and *MEG3*, in which *PEG10* decreased from 50.92% in control to 45.17% in CHD group ($P < 0.01$), *NAP1L5* decreased from 68.86–62.12% ($P < 0.01$), *INPP5F* decreased from 73.17–67.02% ($P < 0.01$), *PLAGL1* decreased from 42.80–40.97%, ($P < 0.05$), *NESP* decreased from 41.12–31.31% ($P < 0.01$), and *MEG3* decreased from 45.31–39.53% ($P < 0.01$) (Fig. 1; Additional file 1–8, Fig. S1-S8).

Table 1
Global methylation levels of the imprinted genes between CHD and control groups

Imprinted genes	Control group (n = 28)		CHD group (n = 27)		P value
	n	methylation level (%, $\bar{x} \pm SD$)	n	methylation level (%, $\bar{x} \pm SD$)	
<i>GRB10</i>	23	43.42 ± 7.43	25	51.12 ± 7.18	0.001
<i>PEG10</i>	22	50.92 ± 5.87	25	45.17 ± 6.07	0.002
<i>MEST</i>	22	53.22 ± 4.79	20	56.60 ± 4.62	0.025
<i>NAP1L5</i>	23	68.86 ± 4.35	20	62.12 ± 10.29	0.007
<i>INPP5F</i>	17	73.17 ± 3.17	22	67.02 ± 4.33	0.000
<i>PLAGL1</i>	25	42.80 ± 2.97	26	40.97 ± 2.33	0.018
<i>NESP</i>	23	41.12 ± 3.38	7	31.31 ± 2.70	0.000
<i>MEG3</i>	24	45.31 ± 3.88	20	39.53 ± 3.41	0.000
<i>MCST2</i>	25	32.43 ± 2.80	25	33.27 ± 3.37	0.339
<i>NNAT</i>	20	88.60 ± 10.76	24	88.99 ± 13.83	0.919
<i>NESPAS</i>	19	37.00 ± 2.68	14	34.78 ± 4.86	0.095
<i>GNAS</i>	23	41.72 ± 3.46	24	41.11 ± 6.09	0.677
<i>IGF2</i>	23	53.25 ± 3.77	22	53.24 ± 4.31	0.991
<i>H19</i>	20	47.01 ± 5.22	14	42.35 ± 8.38	0.051
<i>IG</i>	18	53.65 ± 3.84	18	51.69 ± 4.13	0.149
<i>KCNQ10T1</i>	24	38.34 ± 2.58	24	37.99 ± 3.87	0.714
<i>SNRPN</i>	24	34.85 ± 3.85	25	33.83 ± 5.46	0.457
<i>ZIM2</i>	25	37.20 ± 4.70	24	36.96 ± 5.07	0.863

Altered methylation levels of imprinted genes in CHD classifications

CHD was classified into four groups including atrial and ventricular septal defect (ASD and VSD), valvular malformation, vascular malformation and Tetralogy of Fallot (ToF). In ASD and VSD patients, 9 genes showed methylation changes, as compared with control group, whereby 2 imprinted genes (the *GRB10*

and *MEST*) were hypermethylated (Fig. 2A-B; Additional file 9, Fig. S9A) and 7 (*PEG10*, *NAP1L5*, *INPP5F*, *PLAGL1*, *NESP*, *MEG3* and *H19*) hypomethylation (Fig. 2C-I; Additional file 9, Fig. S9A). In valvular malformation group, the methylations of 5 imprinted genes were lower than that of control group, including *PEG10*, *NAP1L5*, *INPP5F*, *NESP* and *MEG3* (Fig. 3; Additional file 9, Fig. S9B). In ToF group, methylation of 5 imprinted genes changed, including *GRB10* with higher methylation than that in control group (Fig. 4A; Additional file 9, Fig. S9C) and *PEG10*, *INPP5F*, *MEG3* and *H19* with lower methylation (Fig. 4B-E; Additional file 9, Fig. S9C). There was differences in methylation of 6 imprinted genes between vascular malformation and control groups, including hypermethylation of the *GRB10* gene and hypomethylation of *PEG10*, *INPP5F*, *NESP*, *MEG3* and *KCNQ10T1* respectively (Fig. 5; Additional file 9, Fig. S9D). Taken together, 3 imprinted genes (*PEG10*, *INPP5F* and *MEG3*) were hypomethylated in all four CHD classifications. *NESP* hypomethylation occurred in all CHDs except ToF, while *GRB10* hypermethylation occurred in all CHDs except valvular malformation.

Risk Analysis between differential methylation regions (DMRs) methylation levels of imprinted genes and CHD

In order to develop a model for assessing the risk of CHD based on the amplitude of methylation levels of DMRs of imprinted genes, CHD samples were categorized as Q1-Q4 according to the quartiles of methylation levels in control group. As shown in Table 2, the DMRs of 8 differentially imprinted genes were further analyzed to examine the relationship between methylation changes of these genes and risk of CHD. Compared to control group, the DMR methylation level of *NESP* gene was lower in all CHDs. Methylation modification of two imprinted genes were elevated at 75th percentile level in CHD, with *GRB10* in 21 of 25 patients and *MEST* in 10 of 20 patients.. There were 4 imprinted genes whose methylation level of DMRs were lower at 25th percentile level in CHD including 14 of 25 cases of *PEG10*, 17 of 22 cases of *INPP5F*, 11 of 26 cases of *PLAGL1* and 16 of 20 cases of *MEG3* respectively. When logistic regression analysis was used with adjustment to sex and compared with control group, the risk of CHD was 21 times higher than that of control group when *GRB10* methylation was higher than 44% ($P < 0.05$). When the methylation levels of *PEG10*, *INPP5F*, *PLAGL1* and *MEG3* were lower than 45.2%, 70.39%, 40.76% and 41.83%, respectively, the risk of CHD increased by 16.8 [$P < 0.05$], 11.06 [$P < 0.01$], 11 [$P < 0.05$] and 12 [$P < 0.01$] times.

Table 2
Risk Analysis results of DMRs methylation levels of imprinted genes and CHD occurrence

Imprinted genes	Quantile percentages	Controls n (%)	CHD n (%)	Adjusted OR ^a (95%CI ^b)	Adjust P value
<i>GRB10</i>	Q ₁ ^c (< 39.67%)	6(26.1%)	1(4%)	1(reference)	
	Q ₂ -Q ₃ ^d (39.67%-44%)	11(47.8%)	3(12%)	1.64(0.14–19.39)	0.696
	Q ₄ ^e (> 44%)	6(26.1%)	21(84%)	21.00(2.10-210.14)	0.01
<i>MEST</i>	Q ₁ (< 49.36%)	5(22.7%)	1(5%)	1(reference)	
	Q ₂ -Q ₃ (49.36%-56.07%)	12(54.5%)	9(45%)	3.75(0.371–37.95)	0.263
	Q ₄ (> 56.07%)	5(22.7%)	10(50%)	10.00(0.907–110.28)	0.06
<i>PEG10</i>	Q ₁ (< 45.2%)	5(22.7%)	14(56%)	16.80(1.60-176.23)	0.019
	Q ₂ -Q ₃ (45.2%-55.7%)	11(50%)	10(40%)	5.46(0.56–53.52)	0.145
	Q ₄ (> 55.7%)	6(27.3%)	1(4%)	1(reference)	
<i>NAP1L5</i>	Q ₁ (< 67.14%)	6(26.1%)	9(45%)	3.50(0.64–19.20)	0.149
	Q ₂ -Q ₃ (67.14%-71.29%)	10(43.5%)	8(40%)	1.87(0.36–9.64)	0.456
	Q ₄ (> 71.29%)	7(30.4%)	3(15%)	1(reference)	
<i>INPP5F</i> ^f	Q ₁ (< 70.39%)	4(23.5%)	17(77.3%)	11.06(2.47–49.53)	0.002
	Q ₂ -Q ₄ (> = 70.39%)	13(76.5%)	5(22.7%)	1(reference)	
<i>PLAGL1</i>	Q ₁ (< 40.76%)	6(24%)	11(42.3%)	11.00(1.06-114.09)	0.045
	Q ₂ -Q ₃ (40.76%-45.82%)	13(52%)	14(53.8%)	6.46(0.68–61.16)	0.104

a, odds ratio; b, confidence interval; c, 25th percentile; d, 25th percentile to 75th percentile; e, 75th percentile. Cut-off value was defined as 25th and 75th percentiles of the control group methylation level. Adjusted for sex by logistic regression. f indicates that the maximum value of the imprinting gene cases was less than 75% of control group and could not be calculated, so there were divided into two groups.

Imprinted genes	Quantile percentages	Controls n (%)	CHD n (%)	Adjusted OR ^a (95%CI ^b)	Adjust P value
	Q ₄ (> 45.82%)	6(24%)	1(3.8%)	1(reference)	
<i>MEG3</i> ^f	Q ₁ (< 41.83%)	6(25%)	16(80%)	12.00(2.86–50.31)	0.001
	Q ₂ -Q ₄ (> = 41.83%)	18(75%)	4(20%)	1(reference)	

a, odds ratio; b, confidence interval; c, 25th percentile; d, 25th percentile to 75th percentile; e, 75th percentile. Cut-off value was defined as 25th and 75th percentiles of the control group methylation level. Adjusted for sex by logistic regression. f indicates that the maximum value of the imprinting gene cases was less than 75% of control group and could not be calculated, so there were divided into two groups.

Discussion

It is well known that the epigenetic regulation of imprinted genes is involved in many embryonic developmental processes, including cardiac development [17]. At present, DNA methylation disorder is considered as one of the important factors in the occurrence and development of CHD [6]. However, the specific role of the imprinted gene methylation regulation on cardiac development remains unclear. In this study, we found for the first time the abnormal methylation changes of 8 imprinted genes (*GRB10*, *MEST*, *PEG10*, *NAP1L5*, *INPP5F*, *PLAGL1*, *NESP* and *MEG3*) in CHD patients. The methylations of imprinted genes varied with the heterogeneity of CHDs. Six of them were associated with significantly higher risk of CHD except *MEST* and *NAP1L5*.

The etiology of CHD is complex, involving the interaction of environmental and genetic factors, leading to developmental phenotypes that regulate morbidity and severity. The potential relationship between alterations of genomic methylation and CHD is increasingly recognized. The dynamic changes of DNA methylation in cardiomyocyte-related gene sets during cardiac development suggest that DNA methylation modification is essential for the occurrence of cardiac diseases [5, 18]. One study has confirmed that there are significant differences in methylation of genes associated with muscle contraction and cardiomyopathy in CHD [6]. Genomic imprinting is a process of epigenetic modification, and loss of imprinting may lead to abnormal embryonic development [15, 19]. Our data provide the first evidence of methylation changes of imprinted gene in CHD. In CHD patients, altered methylation was found in 8 imprinted genes associated with embryonic development, namely, *GRB10*, *MEST*, *PEG10*, *NAP1L5*, *INPP5F*, *PLAGL1*, *NESP* and *MEG3*. The relationship between CHD classification and methylation modification of imprinted genes was further revealed by stratified analysis. Overall, our data clearly illustrated an association between altered methylation of imprinted genes and CHD.

Some imprinted genes have key functions influencing the proper embryonic growth and development by forming "imprinted gene network", including *GRB10* and *MEST* as the core molecules [15]. *GRB10*, as an

important growth-limiting factor, has a wide range of effects on embryonic development. Maternal *GRB10* knockout results in embryonic overgrowth, and the change of ICR of *GRB10* gene results in obvious dysplasia [20, 21]. Previous studies have found an increase methylation of *GRB10* in fetal samples of spontaneous abortion [22]. It has been found that the *Ddc* gene, involved in the development of trabecular cardiomyocytes of the embryonic and neonatal heart, displays tight conserved linkage with the methylated *GRB10* gene [23]. In the present study, we found that the DMR methylation of *GRB10* imprinted gene in CHD was increased except valvar defects. It is hinted the methylation disorder of *GRB10* may be involved in the development of heart through the interaction with *DDC*. Furthermore, hypermethylation of *GRB10* may inhibit its expression, and then regulated the growth through insulin pathway such as *IGF2*, leading to cardiac dysplasia. *MEST* is a maternal imprinted gene and widely expressed throughout the embryonic period [24]. *Mest*-knockout mice exhibited embryonic and placental growth retardation and postnatal growth inhibition, while loss of the imprinted gene resulted in postnatal weight gain and multiple organ hypertrophy [25, 26]. King et al. have found that *Mest* specifically expressed myocardial trabeculae in developing atria and ventricles of mice. *Mest*-knockout mice showed subtle changes of myocardial trabeculae, which displayed an increase in thickness and reduction in density of the compact myocardium, similar to that in human heart disease, which hinted *MEST* is closely related to heart disease [27]. The increased methylation of DMR of *MEST* imprinted gene in CHD in our data, especially in ASD and VSD. It is suggested that *MEST* expression disorder via altered methylation of the gene may be involved in the occurrence of CHD by affecting the development of myocardial trabecula.

PEG10, a paternally expressed imprinted gene, is expressed in embryonic tissues and placenta, and participates in cell proliferation, differentiation and apoptosis [28, 29]. The *Peg10*-knockout mice exhibited early embryo death and placental defects [30]. A study has found that the expression of *PEG10* in the heart was very low, indicating that trace *PEG10* could maintain the normal development of the heart [28]. On the contrary, we found abnormal hypomethylation of *PEG10* in CHD. Overexpression of *PEG10* may be involved in the invasion and metastasis of malignant tumors, such as hepatocellular carcinoma and endometrial cancer etc., through the epithelial mesenchymal transition (EMT) [31–33][1–3]. Therefore, the low methylation of *PEG10* may cause abnormal elevation of expression, and induce abnormal migration of related cells through EMT pathway, leading to abnormal cardiac development.

INPP5F encodes inositol 1,4,5-trisphosphate (InsP₃) 5-phosphatase, which is an important functional endogenous regulator. It has been found that *INPP5F* regulated the size of cardiac myocytes and cardiac stress response, increased the hypertrophy and activation of the fetal gene program in the stress response in *Inpp5f*-knockout mice, and decreased the hypertrophic response in *Inpp5f*-overexpressed mice [34, 35]. *INPP5F* can be used as a negative feedback regulator of insulin signal and downregulation of *INPP5F* in diabetes mellitus has cardio protective effect [36]. Consistent with previous studies, we found that *INPP5F* methylation was reduced in a variety of CHDs and increased CHD risk. This result further confirms that *INPP5F* may be a protective factor for cardiac development.

MEG3 is a maternal-expressed imprinted gene that is expressed in many normal tissues. It inhibits the proliferation of tumor cells in vitro. It also interacts with tumor suppressor p53 to regulate the expression of p53 target genes [37]. Consistent with this effect, *MEG3* has an impact on cardiac remodeling induced by pathological hypertrophy by regulating the binding of p53 to other gene promoters [38]. Other studies have shown that *MEG3* participates in the development of myocardial fibrosis and prevents myocardial remodeling by regulating the production of MMP-2 by CFs in vitro and in vivo [39]. In consistency to previous studies, our results also found that methylation of *MEG3* was reduced and associated with the increased risk of CHD.

We also detected hypomethylation of DMRs of other 3 imprinted genes including *NAP1L5*, *PLAGL1* and *NESP* in CHD. Stratified analysis revealed that different CHDs showed specific methylation patterns, such as hypomethylation of the 3 genes in ASD and VSD, hypomethylation of *NAP1L5* and *NESP* in valvular malformation, hypomethylation of *NESP* in vascular malformation, and no significant change in ToF.

As a pilot study, it is subject to a couple of limitations. First, methylation alterations of imprinted genes were found in CHD patients, but the causal relationship remains to be explored in further experimental studies; second, we studied a small heterogeneous group of CHD patients and found different methylation alterations of imprinted genes in different types of CHD. A large study sample is warranted to identify the methylation characteristics of imprinted genes in relation to the types of CHD.

Conclusion

Methylation of DMRs of 8 imprinted genes was altered in white blood cells in patients with CHD. Further experiments are warranted to identify the methylation characteristics of imprinted genes in different types of CHD and clarify the pathogenesis mechanism of imprinted genes in CHD.

Methods

Sample Collection

The study was approved by the Ethics Committee of the Capital Institute of Pediatrics. Written informed consent was obtained from parents of the children. These samples were obtained from children with CHD who visited the Capital Institute of Pediatrics during the biennium 2014–2015, and were diagnosed as CHD by echocardiography and other related tests, ranging in age from 1 to 120 months. According to the International Classification Standard of Diseases, the diagnosis of existing cases was strictly classified, and a complete database of CHD research is established. Moreover, the children with CHD were excluded from other congenital and acquired abnormal diseases, such as congenital mental retardation, genetic metabolic diseases, etc. As control group, 28 controls of blood samples were selected from the physical examination children. They were same age and race and had no developmental deformities or other diseases. Peripheral blood samples were collected from 27 children with CHD and 28 healthy children as controls in Capital Institute of Pediatrics (Table 3). In details, 2 ml venous blood was collected into the

evacuated tubes without anticoagulant (Becton Dickinson). Blood samples were immediately centrifuged at 2500 rpm for 10 min. The separated plasma was aliquot without reducing agent and stored at -20°C .

Table 3
Type of CHD and gender distribution in CHD and control groups

Type of CHD		Number of cases		P value
		Male	Female	
CHD	VSD ^a	10	7	0.71
	ASD ^b	6	6	0.40
	Valvular malformation	3	4	0.30
	ToF ^c	4	0	0.15
	PDA ^d	1	3	0.13
	CoA ^e	2	1	0.93
	Others	2	1	0.93
	Total CHD	17	10	0.92
Controls		18	10	

a: ventricular septal defect; b: atrial septal defect; c: Tetralogy of Fallot; d: patent ductus arteriosus; e: constriction of aortic arch.

DNA extraction

Genomic DNA was extracted from peripheral blood with the Blood and Tissue DNA Kit (65904, QIAGEN, Dusseldorf, Germany) according to manufacturer's instructions. The concentration and purity of the DNA were determined by nanodrop and agarose gel electrophoresis. DNA with an OD_{260/280} absorbance ratio of 1.8 was used for subsequent work.

Bisulfite treatment

A total of 500 ng genomic DNA from each sample underwent bisulfite treatment using the EZ DNA methylation kit (D5001, Zymo Research, Irvine, CA, USA) according to the manufacturer's instruction. The quality of the bisulfite conversion was controlled by using PCR products that had no methyl group. Sequencing results confirmed that 96.6% of cytosine residues were converted in previous research [40].

PCR amplification

Eighteen imprinted genes (Table 1) were selected according to WAMZDEX, Gene imprint, Catalogue of Imprinted Genes and other imprinted gene analysis resources. DMRs sequences of the selected imprinted genes were retrieved to confirm using the UCSC database (Additional file 10, Table S1). The primers were designed for the quantitative analysis of the methylation level with Methprimer (Additional file 11, Table S2, <http://epidesigner.com/>). For each reverse primer, an additional T7 promoter tag for in vivo transcription was added, as well as a 10-mer tag on the forward primer to adjust for melting temperature differences.

Sap reaction

The sap reaction was mainly to eliminate the residual dNTP in the PCR product to reduce its interference with the mass spectrometry detection. The specific steps were to add 2 ul sap mixture to the 5 ul PCR product previously, and then centrifuge after vortex mixing. The reaction conditions were 37°C for 20 minutes and then 85°C for 5 minutes.

Methylation analyses

The Sequenom MassARRAY platform (CapitalBio, Beijing, China) was used to perform the quantitative methylation analysis of the imprinted genes. This system uses matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry in combination with RNA base-specific cleavage (MassCLEAVE). A detectable pattern was then analyzed for its methylation status. The spectral methylation ratios were generated using EpiTyper software version 1.0 (Sequenom, San Diego, CA, USA)

Statistical analyses

Data are presented as mean and standard deviation. The methylation levels in the DMRs of imprint genes were compared between CHD and control groups by independent samples t-test. One-way ANOVA was performed to evaluate the differences among different CHD subtypes and control groups. Odds ratios (ORs) was performed to evaluate the incidence risk of CHD in relation to methylation levels. $P < 0.05$ was considered significant. GraphPad Prism 7 software (GraphPad Software, San Diego, California, USA) was used to visually display the results of analysis. Data were stored in the EPI 3.1 Database (EpiData Association, Odense, Denmark) and analyzed with the SPSS 18.0 software package (McGraw-Hill Inc., New York, NY, USA).

Supplementary Information

Additional file 1: Fig. S1: Analysis of methylation level of GRB10

Additional file 2: Fig. S2: Analysis of methylation level of PEG10

Additional file 3: Fig. S3: Analysis of methylation level of MEST

Additional file 4: Fig. S4: Analysis of methylation level of NAP1L5

Additional file 5: Fig. S5: Analysis of methylation level of INPP5F

Additional file 6: Fig. S6: Analysis of methylation level of PLAGL1

Additional file 7: Fig. S7: Analysis of methylation level of NESP

Additional file 8: Fig. S8: Analysis of methylation level of MEG3

Additional file 9: Fig. S9: Abnormal methylation level of the imprinted genes between multiple CHDs classification and control groups

Additional file 10: Table S1. DMR sequence information of 18 imprinted genes

Additional file 11: Table S2. Primer sequence information of 18 imprinted genes

Abbreviations

ASD, atrial septal defect; CHD, congenital heart disease; DMRs, differential methylation regions; ToF, Tetralogy of Fallot; VSD, ventricular septal defect.

Declarations

Ethics approval and consent to participate

The study was approved by the Ethics Committee of the Capital Institute of Pediatrics. Written informed consent was obtained from the parents of the participants.

Consent for publication

Not applicable.

Availability of data and materials

The data that supported the findings of this study are available from the corresponding authors upon reasonable request.

Competing interests

The authors declare that they have no conflict of interest.

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Authors' contributions

S. Chang wrote the manuscript; Y. Wang and Y. Xin conducted the experiment; Y. Wang and S. Chang analyzed data; S. Wang, Y. Luo and H. Zhang provided essential reagents and materials; L. Wang and J. Li designed the study and finalized the manuscript.

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Figures

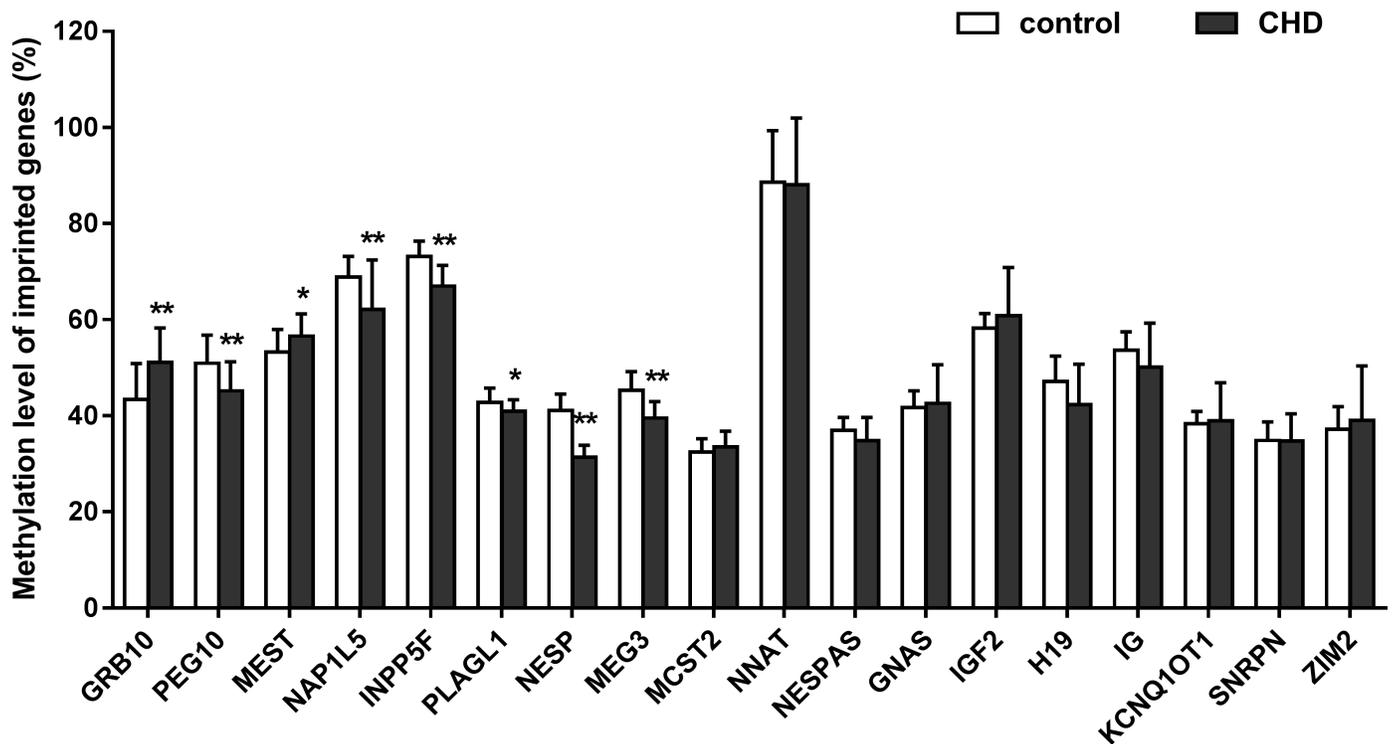


Figure 1

Altered methylation level of 18 imprinted genes between CHD and control groups. A) Methylation level of 18 imprinted genes between CHD and control groups. Compared with the normal group, 8 of the 18 imprinted genes in CHD showed abnormal methylation modification including GRB10, MEST, PEG10, NAP1L5, INPP5F, PLAGL1, NESP and MEG3. Among them, the two genes including GRB10 and MEST showed increased methylation, while the others showed decreased methylation. B) Methylation level of 8 different imprinted genes among single CHD, complex CHD and control groups. CHD was classified as simple CHD and complex CHD according to the degree of disease complexity. Among the 8 differential imprinted genes, methylation level of 6 genes showed statistical significance including GRB10, PEG10, NAP1L5, INPP5F, NESP and MEG3 compared with control group, and methylation modification of PEG10 and NESP were lower in complex CHD than single CHD. Independent sample t-test and One-way ANOVA were used to assess the difference among multiple CHD groups and control groups. All the values was shown as the mean \pm SD. *, $P < 0.05$; **, $P < 0.01$.

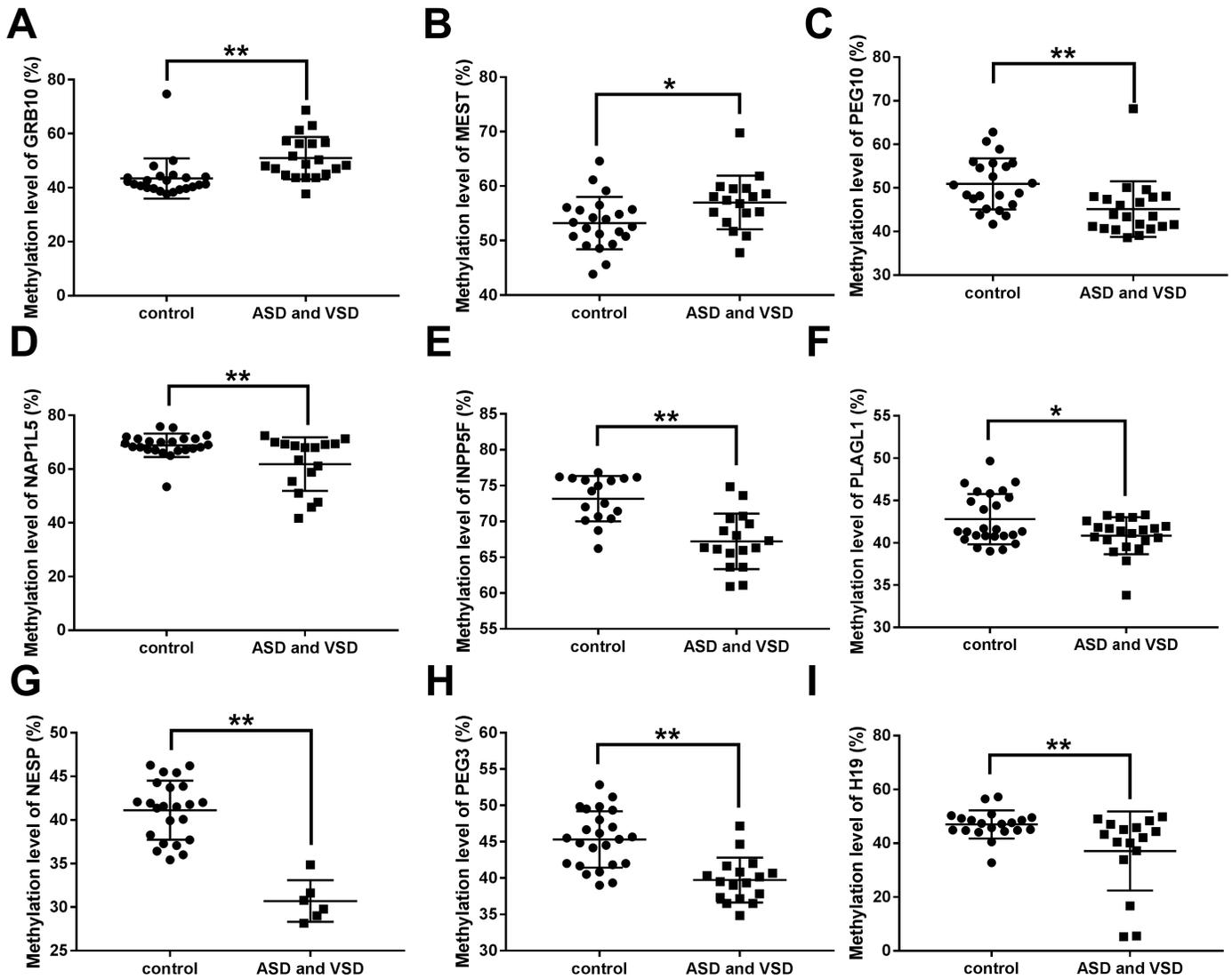


Figure 2

Altered methylation level of the imprinted genes between ASD and VSD and control groups. A-B) Imprinted genes up-regulated by methylation between ASD/VSD and control groups. There were two imprinted genes with increasing methylation including GRB10 and MEST in ASD and VSD groups, compared with the normal group. C-I) Imprinted genes up-regulated by methylation between ASD and VSD and control groups. Compared with the normal group, 7 of the 18 imprinted genes in ASD and VSD showed decreasing methylation modification including PEG10, NAP1L5, INPP5F, PLAGL1, NESP, MEG3 and H19. Independent sample t-test was used to assess the difference between ASD and VSD and control groups. All the values was shown as the mean \pm SD. ASD and VSD, atrial and ventricular septal defect. *, P<0.05; **, P<0.01.

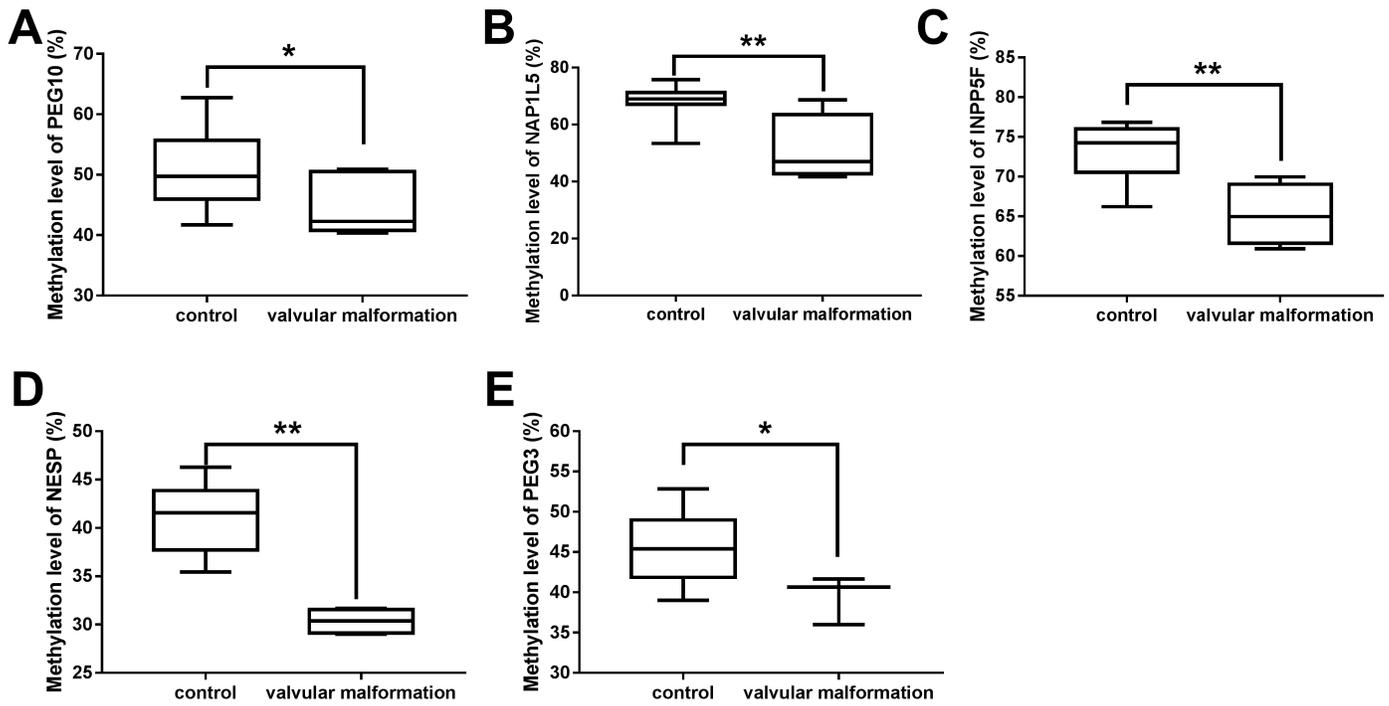


Figure 3

Altered methylation level of the imprinted genes between valvular malformation and control groups. A-E) Hypomethylation of imprinted genes between ASD and VSD and control groups. Compared with the normal group, 5 of the 18 imprinted genes in ASD and VSD showed decreasing methylation modification including PEG10, NAP1L5, INPP5F, NESP and MEG3. Independent sample t-test was used to assess the difference between valvular malformation and control groups. All the values was shown as the mean \pm SD. *, $P < 0.05$; **, $P < 0.01$.

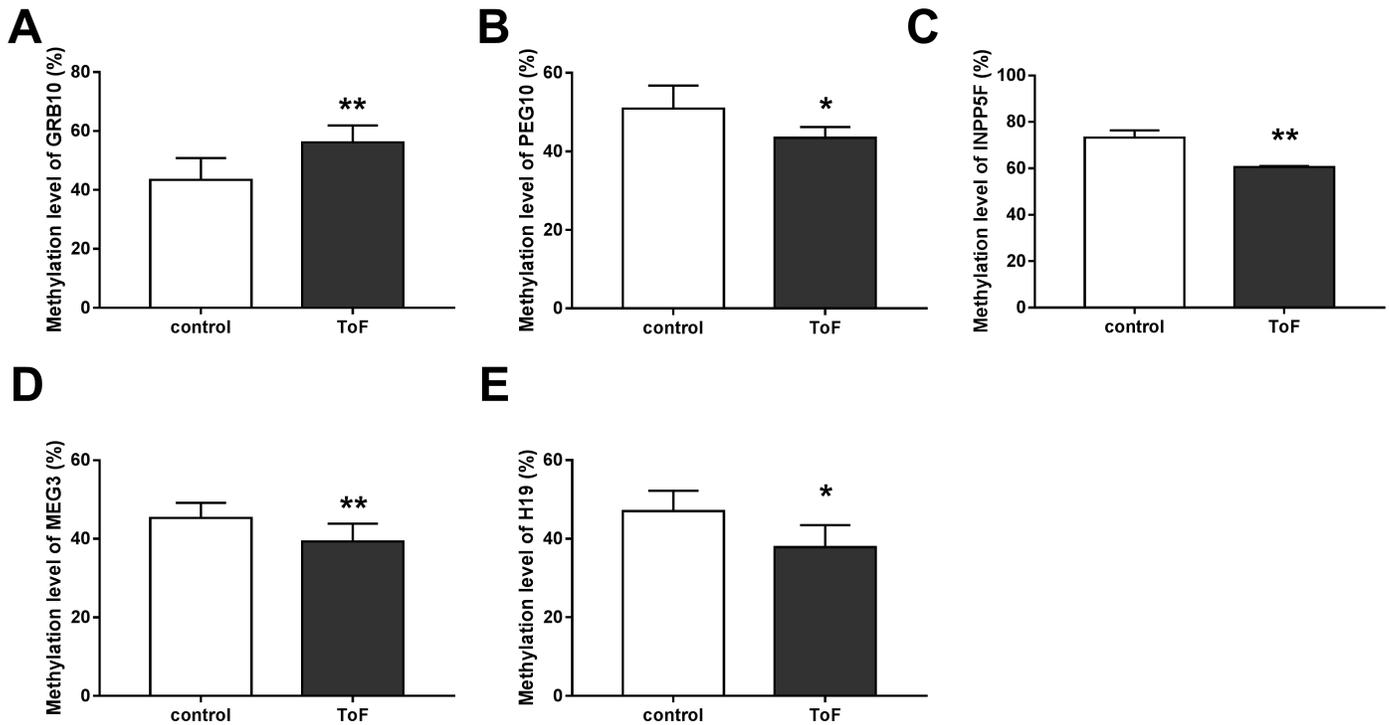


Figure 4

Altered methylation level of the imprinted genes between ToF and control groups. A) Hypermethylation level of GRB10 gene between ToF and control groups. B-D) Hypomethylation of imprinted gene between ToF and control groups. Compared with the normal group, 4 of the 18 imprinted genes in ToF showed decreasing methylation modification including PEG10, INPP5F, MEG3 and H19. Independent sample t-test was used to assess the difference between CHD and control groups. All the values was shown as the mean \pm SD. ToF, Tetralogy of Fallot. *, P<0.05; **, P<0.01.

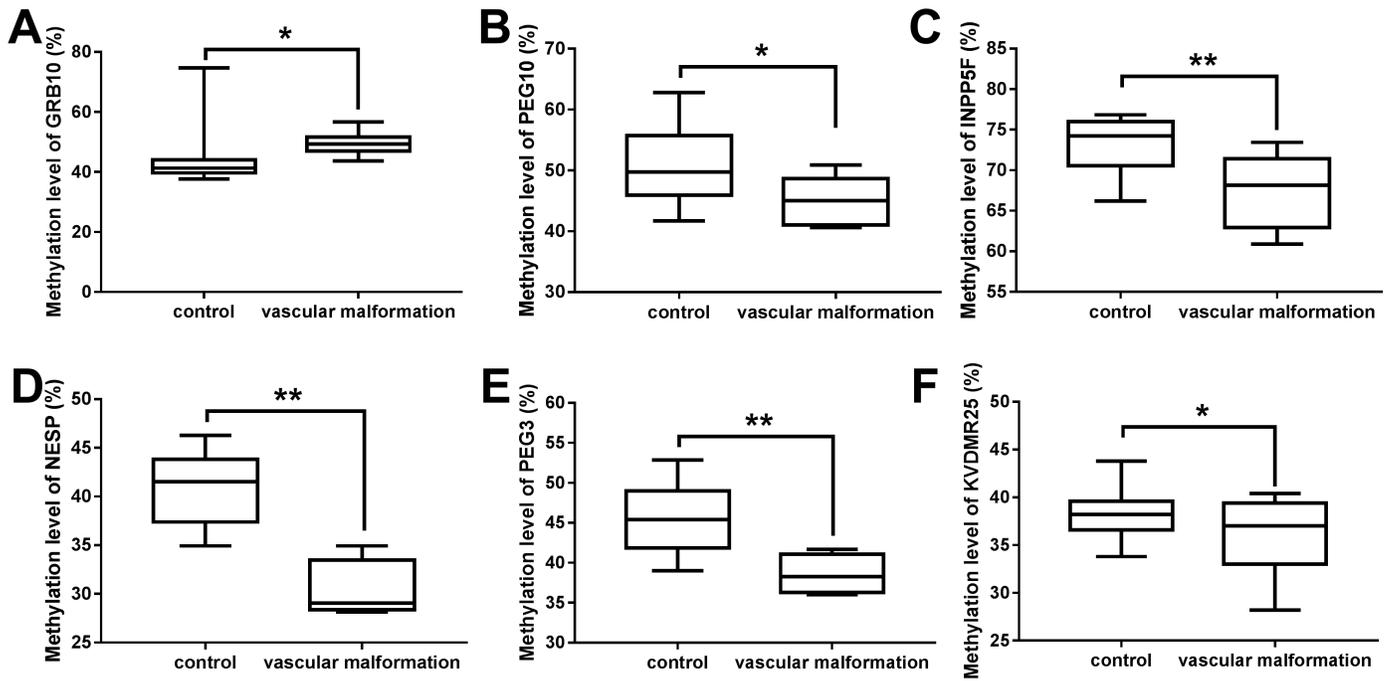


Figure 5

Altered methylation level of the imprinted genes between vascular malformation and control groups. A) Up-regulation of methylation level of GRB10 gene between vascular malformation and control groups. B-D) Down-regulation of imprinted gene methylation between vascular malformation and control groups. Compared with the normal group, 5 of the 18 imprinted genes in vascular malformation showed decreasing methylation modification including PEG10, INPP5F, NESP, MEG3 and KCNQ10T1. Independent sample t-test was used to assess the difference between vascular malformation and control groups. All the values was shown as the mean \pm SD. *, $P < 0.05$; **, $P < 0.01$.

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