

Substitution Mapping of Two Closely Linked QTLs Controlling Grain Chalkiness and Grain Shape on Rice Chromosome 8

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1 **Substitution mapping of two closely linked QTLs controlling grain chalkiness and grain shape on**
2 **rice chromosome 8**

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12
13
14 **Abstract**

15 **Background:** Rice varieties are required to have high yield and good grain quality. Grain chalkiness
16 and grain shape are two important traits of rice grain quality. Low chalkiness slender grains are
17 preferred by most rice consumers. Here, we dissected two closely linked quantitative trait loci (QTLs)
18 controlling grain chalkiness and grain shape on rice chromosome 8 by substitution mapping.

19 **Results:** Two closely linked QTLs controlling grain chalkiness and grain shape were identified using
20 single-segment substitution lines (SSSLs). The two QTLs were then dissected on rice chromosome 8
21 by secondary substitution mapping. *qPGC8.1* was located in an interval of 1382.6 kb and *qPGC8.2* was
22 mapped in a 2057.1 kb region. The maximum distance of the two QTLs was 4.37 Mb and the space
23 distance of two QTL intervals was 0.72 Mb. *qPGC8.1* controlled grain chalkiness and grain width.
24 *qPGC8.2* was responsible for grain chalkiness and for grain length and grain width. The additive
25 effects of *qPGC8.1* and *qPGC8.2* on grain chalkiness were not affected by heat stress.

26 **Conclusions:** Two closely linked QTLs *qPGC8.1* and *qPGC8.2* were dissected on rice chromosome 8.
27 They controlled the phenotypes of grain chalkiness and grain shape. The two QTLs were insensitive to
28 high temperature.

29 **Key words:** grain chalkiness, grain shape, quantitative trait locus, heat stress, substitution mapping,
30 rice

31

32

33 **Background**

34 Rice is an important food crop in the world. Rice varieties with higher head rice yield, higher
35 transparency and less chalkiness are more popular in the market (Sreenivasulu et al. 2015; Misra et al.
36 2019). With the increase of living standard, rice varieties are required to have both a higher grain yield
37 and a better grain quality. Grain chalkiness not only affects grain appearance, but also has adverse
38 effects on milling and cooking performance. Chalkiness is a complex quantitative trait, which is easily
39 affected by environments. In the early and middle stages of seed development, the occurrence of
40 temperature stress will cause uneven seed filling and storage biosynthesis obstacles, leading to the
41 formation of chalkiness (Masutomi et al. 2015; Sreenivasulu et al. 2015; Morita et al. 2016). The
42 chalkiness of rice varieties varied greatly. It was found that the proportion of chalky grains of newly
43 developed varieties was higher than that of the old modern varieties, and the proportion of chalky
44 grains of the hybrid varieties was higher than that of other modern varieties (Laborte et al. 2015).
45 Therefore, high-yielding varieties usually have higher chalkiness levels (Misra et al. 2019).

46 Percentage of grain chalkiness (PGC) is a quantitative index of grain chalkiness controlled by
47 quantitative trait loci (QTLs) (Sreenivasulu et al. 2015; Misra et al. 2019). More than one hundred of
48 QTLs controlling chalkiness were reported in rice genome (Sreenivasulu et al. 2015; Yang et al. 2021).
49 Many QTLs for chalkiness were detected on chromosomes 5, 6 and 8 in different populations and
50 environments (He et al. 1999; Tan et al. 2000; Wan et al. 2005; Hao et al. 2009; Liu et al. 2011; Chen
51 et al. 2011; Guo et al. 2011; Liu et al. 2012; Li et al. 2014; Peng et al. 2014; Zhao et al. 2015; Chen et
52 al. 2016; Gao et al. 2016; Yun et al. 2016; Zhao et al. 2016; Wang et al. 2017; Zhu et al. 2018a; Misra
53 et al. 2019; Misra et al. 2020). On chromosome 8, the hotspot region of the QTLs for chalkiness was
54 located on the long arm (Li et al. 2003; Wan et al. 2005; Hao et al. 2009; Guo et al. 2011; Liu et al.
55 2012; Gao et al. 2016; Zhao et al. 2016; Wang et al. 2017). Although a larger number of the chalk
56 QTLs were detected on the hotspot chromosome region, only one of the QTLs was usually detected in
57 a population. Therefore, it is not clear how many chalk QTLs there are in the hotspot chromosome
58 region.

59 Grain chalkiness is a complex trait and easily affected by other genetic factors. Several
60 fine-mapped target genes and GWAS loci have been found to influence chalkiness, but many have

61 either low or moderate effect (Gong et al. 2017; Wang et al. 2017; Quero et al. 2018 Misra et al. 2019).
62 Although 11 GWAS loci for chalky grain rate were identified, the GWAS loci could only explain a
63 small part of the phenotypic variation (Gong et al. 2017). Grain chalkiness and grain shape are two
64 important traits for rice grain quality. Grain width was positively correlated with chalkiness (Zhao et al.
65 2015). It was found that some QTLs controlling grain width were overlapped with chalk QTLs (Wang
66 et al. 2017). Misra et al. (2020) reported 78 chalk QTLs, but only three QTL regions overlapped with
67 known grain width genes. The phenotypic variation for chalkiness showed a weak positive correlation
68 with grain width. They believed that the key genes of grain width are not causal factors of chalkiness.
69 Therefore, the genetic architecture of grain chalkiness and grain shape of rice remains unclear.

70 Substitution mapping is a powerful tool to detect QTLs in complex traits (Tan et al. 2021; Yang et
71 al. 2021). Like near-isogenic lines (NILs), single-segment substitution lines (SSSLs) carry only one
72 substitution segment from donors in the recipient genetic background (Zhang et al. 2004; Keurentjes et
73 al. 2007). We have developed a library of 2360 SSSLs, which were derived from 43 donors of 7
74 species of rice AA genome in the genetic background of Huajingxian 74 (HJX74), an *indica* elite
75 variety in southern China (Zhang et al. 2004; Xi et al. 2006; Zhang 2019). These SSSLs were widely
76 used to detect QTLs for complex traits, to clone QTLs of agronomic importance and to mine alleles of
77 different functions (Zeng et al. 2006; Teng et al. 2012; Wang et al. 2012; Zhang et al. 2012; Zhu et al.
78 2014; Yang et al. 2016; Zhou et al. 2017; Zhu et al. 2018b; Fang et al. 2019; Sui et al. 2019; Tan et al.
79 2020; Tan et al. 2021; Yang et al. 2021). Recently, the SSSLs were used to detect QTLs controlling
80 stigma exertion rate (SER) of rice. Two pairs of tightly linked QTLs for SER, *qSER-2a* and *qSER-2b*
81 on chromosome 2 and *qSER-3a* and *qSER-3b* on chromosome 3, were dissected by substitution
82 mapping (Tan et al. 2021). In previous study, we fine-mapped two QTLs *qPGC9* and *qPGC11* for grain
83 chalkiness on rice chromosomes 9 and 11, which were sensitive to high temperature (Yang et al. 2021).
84 In the present study, two closely linked QTLs for grain chalkiness on chromosome 8, *qPGC8.1* and
85 *qPGC8.2*, were detected. The two QTLs were showed effect on grain chalkiness and grain shape, and
86 insensitive to high temperature. Dissection of the two closely linked QTLs laid a foundation for
87 revealing the genetic architecture of grain chalkiness and the relationship between grain chalkiness and
88 grain shape in rice.

89

90 **Results**

91 **Grain chalkiness in SSSLs**

92 Two SSSLs 15-08 and 03-08 with lower grain chalkiness were selected from the HJX74-SSSL library
93 (Fig. 1a). The SSSLs were used to investigate grain chalkiness in consecutive 6 cropping seasons from
94 the first cropping season (FCS) of 2017 to the second cropping season (SCS) of 2019. On average, the
95 PGC of 15-08 and 03-08 was 11.9% and 10.3% respectively, which were significantly lower than 21.2%
96 of recipient HJX74 (Fig. 1b and Additional file 1: Table S1). It meant that the two SSSLs each carried a
97 QTL for PGC on their substitution segments.

98 The substitution segments of 15-08 and 03-08 were surveyed by densifying molecular markers
99 (Additional file 1: Table S2). The estimated length of substitution segments was 2726.7 kb in 15-08 and
100 4974.3 kb in 03-08. The two substitution segments overlapped in a 553.7 kb interval (Fig. 1c and
101 Additional file 1: Table S3).

102 Eight agronomic traits of 15-08 and 03-08 were investigated. Most traits of the SSSLs had no
103 significant difference with HJX74 (Fig. 1a and Additional file 1: Table S4). However, the grain shapes
104 of 15-08 and 03-08 were significantly different with HJX74. For 15-08, the grain width was
105 significantly narrower than that of HJX74, the former was 2.62 mm and 2.45 mm and the later was
106 2.69 mm and 2.64 mm in SCS of 2018 and in FCS of 2019, respectively. Compared with HJX74, 03-08
107 showed significantly longer in grain length and significantly narrower in grain width. In SCS of 2018
108 and in FCS of 2019, 03-08 had 9.14 mm and 8.88 mm in grain length and 2.55 mm and 2.40 mm in
109 grain width, respectively (Additional file 1: Table S4). It is noted that *GW8* is outside the substitution
110 segment of 03-08 (Fig. 1c). Therefore, the difference of grain shape between 03-08 and HJX74 was not
111 controlled by *GW8* gene.

112

113 **Substitution mapping of *qPGC8.1***

114 To map the QTL for grain chalkiness on the substitution segment of 15-08, the SSSL was used to
115 develop secondary SSSLs or NILs. Four NILs were developed from an $F_{2,3}$ population derived from the
116 cross of HJX74/15-08. The four NILs were then investigated for grain chalkiness. PGC levels of two
117 NILs, NIL15-08-26 and NIL15-08-43, were as low as 15-08, while those of other two NILs,
118 NIL15-08-4 and NIL15-08-14, were as high as HJX74. Substitution segments of the two NILs with low
119 PGC overlapped in the region between markers RM4815 and RM23137, while substitution segments of
120 other two NILs with high PGC located outside the region. These results indicated that the QTL for

121 grain chalkiness, *qPGC8.1*, was located in the region between markers RM4815 and RM23137 with
122 the estimated interval length of 1382.6 kb (Fig. 2a-b).

123 Using RM8264 marker in *qPGC8.1* interval, Chi-square test was performed in 86 individuals of
124 F₂ population. The results showed that the segregation ratio of the three marker genotypes was 1:2:1 (χ^2
125 = 0.51 < $\chi^2_{0.01, 2} = 9.21$). The effect of heterozygous genotype (*qPGC8.1/qpgc8.1*) was significantly
126 higher than that of dominant homozygous genotype (*qPGC8.1/qPGC8.1*) and significantly lower than
127 that of recessive homozygous genotype (*qpgc8.1/qpgc8.1*). The result showed that *qPGC8.1* was
128 incomplete dominance (Fig. 2c).

129 Like 15-08, the four NILs showed no-significant difference with HJX74 in grain length (Fig. 2d).
130 However, the grain width segregated in the NILs. Two NILs, NIL15-08-26 and NIL15-08-43, with low
131 chalkiness showed narrower in grain width as 15-08, while other two NILs, NIL15-08-4 and
132 NIL15-08-14, with high chalkiness were wider in grain width as HJX74, the former was 2.56 mm and
133 2.58 mm and the later was 2.66 mm and 2.67 mm, respectively (Fig. 2e). The results indicated that
134 *qPGC8.1* controlled grain width besides grain chalkiness.

135

136 **Substitution mapping of *qPGC8.2***

137 To map the QTL for grain chalkiness on the substitution segment of 03-08, the SSSL was used to
138 develop secondary SSSLs or NILs. Five NILs were developed from an F_{2:3} population derived from the
139 cross of HJX74/03-08. The five NILs were then investigated for grain chalkiness. PGC levels of three
140 NILs, NIL03-08-9, NIL03-08-15 and NIL03-08-19, were as low as 03-08, while those of other two
141 NILs, NIL03-08-14 and NIL03-08-55, were as high as HJX74. Substitution segments of the three NILs
142 with low PGC overlapped in the region between markers SNP8M54 and SNP8M28, while substitution
143 segments of other two NILs with high PGC located outside the region. These results indicated that the
144 QTL for grain chalkiness, *qPGC8.2*, was located in the region between markers SNP8M54 and
145 SNP8M28 with the estimated interval length of 2057.1 kb (Fig. 3a-b).

146 In the F₂ population of 100 individuals, segregation ratio of three marker genotypes of RM210 in
147 *qPGC8.2* region was 1:2:1 ($\chi^2 = 1.34 < \chi^2_{0.01, 2} = 9.21$). The effect of heterozygous genotype
148 (*qPGC8.2/qpgc8.2*) was significantly different from that of the homozygous genotypes
149 (*qPGC8.2/qPGC8.2* and *qpgc8.2/qpgc8.2*). The result showed that *qPGC8.2* was incomplete
150 dominance (Fig. 3c).

151 In substitution mapping, NILs segregated in grain length and grain width. Three NILs,
152 NIL03-08-9, NIL03-08-15 and NIL03-08-19, with low chalkiness showed longer in grain length and
153 narrower in grain width as 03-08, while other two NILs, NIL03-08-14 and NIL03-08-55, with high
154 chalkiness had shorter in grain length and wider in grain width as HJX74. The three NILs with low
155 chalkiness were 8.75 mm, 8.91 mm and 9.02 mm in grain length and 2.58 mm, 2.54 mm and 2.62 mm
156 in grain width, while the two NILs with high chalkiness were 8.46 mm and 8.48 mm in grain length
157 and 2.66 mm and 2.65 mm in grain width, respectively (Fig. 3d-e). The results indicated that *qPGC8.2*
158 controlled grain shape besides grain chalkiness.

159 It was noted that *qPGC8.2* was closely linked with *qPGC8.1* on chromosome 8. The maximum
160 distance of the two QTL location was 4.37 Mb, from markers RM4815 to SNP8M28. The space
161 distance between the two QTL regions was 0.72 Mb, from markers RM23137 to SNP8M54 (Fig. 1c,
162 Fig. 2b and Fig. 3b).

163

164 **Effects of different cropping seasons on grain chalkiness**

165 The grain chalkiness was tested in two cropping seasons per year. During flowering to harvest of rice,
166 the day and night temperatures of FCS and SCS were very different. In 2017-2019, the average values
167 of maximum, minimum and mean temperatures were 32.1°C, 25.9°C and 29.0°C in FCS, and 28.6°C,
168 21.0°C and 24.9°C in SCS, respectively. The mean temperature in SCS was 4.1°C lower than that in
169 FCS (Additional file 1: Table S5).

170 The grain chalkiness was lower in SCS than that in FCS in all lines. Percentage of chalky grains
171 (PCG) and PGC were significantly different in all lines. Percentage of chalk area (PCA) was
172 significantly different in 15-08, and no significantly different in 03-08 and HJX74. The PGC of HJX74,
173 15-08 and 03-08 was 25.8%, 15.9% and 13.6% in FCS, and 16.6%, 7.8% and 7.1% in SCS,
174 respectively (Fig. 4). It is obvious that high temperature had great influence on PGC of HJX74 and
175 SSSLs during seed filling period, mainly through the influence on PCG.

176

177 **Additive effects of *qPGC8.1* and *qPGC8.2* on grain chalkiness**

178 According to the estimation of chalkiness phenotype in 2017-2019, the additive effects of *qPGC8.1* and
179 *qPGC8.2* had no significant difference between SCS and FCS. For *qPGC8.1*, the additive effects on
180 PGC were -9.9% in FCS and -8.8% in SCS. For *qPGC8.2*, the additive effects on PGC in FCS and SCS

181 were -12.3% and -9.5%, respectively (Fig. 5). The results showed that *qPGC8.1* and *qPGC8.2* were
182 insensitive to high temperature of FCS.

183

184 **Discussion**

185 **Dissection of two closely linkage QTLs *qPGC8.1* and *qPGC8.2* on chromosome 8**

186 Grain chalkiness is a complex polygenic quantitative trait (Sreenivasulu et al. 2015). More than one
187 hundred of QTLs for the chalkiness traits were reported across all 12 chromosomes of rice genome
188 (Sreenivasulu et al. 2015). On chromosome 8, many QTLs for chalkiness were located on the long arm
189 in different populations and environments. In the hotspot region, however, only one chalk QTL was
190 detected in each population (Li et al. 2003; Wan et al. 2005; Hao et al. 2009; Guo et al. 2011; Liu et al.
191 2012; Gao et al. 2016; Zhao et al. 2016; Wang et al. 2017). In the present study, we detected two
192 closely linked QTLs, *qPGC8.1* and *qPGC8.2*, on the region of 19.01-23.38 Mb of chromosome 8. The
193 maximum distance of the two QTL location was 4.37 Mb and the space distance between the two QTL
194 regions was 0.72 Mb (Fig. 1c, Fig. 2b and Fig. 3b). Dissection of the two closely linked QTLs showed
195 that the hotspot region of chromosome 8 contained multiple QTLs for grain chalkiness. The results
196 revealed the genetic architecture of grain chalkiness on the hotspot region of chromosome 8.

197 Recently, two pairs of tightly linked QTLs controlling SER, *qSER-2a* and *qSER-2b* on
198 chromosome 2 and *qSER-3a* and *qSER-3b* on chromosome 3, were dissected by substitution mapping.
199 On chromosome 2, two linkage QTLs, *qSER-2a* and *qSER-2b*, were located in the region of 1288.0 kb,
200 and were respectively delimited to the intervals of 234.9 kb and 214.3 kb. On chromosome 3, two
201 QTLs, *qSER-3a* and *qSER-3b*, were detected in the region of 3575.5 kb and were narrowed down to
202 319.1 kb and 637.3 kb, respectively (Tan et al. 2021). Together, those results indicated that substitution
203 mapping using SSSLs is a powerful tool for dissection of closely linked QTLs for complex traits.

204

205 **Relationship of grain chalkiness and grain shape**

206 Grain chalkiness and grain shape are two important traits for rice grain quality, which relationship is
207 complicated. It was found that grain width had a negative pleiotropic effect on grain chalkiness (Zhao
208 et al. 2015). However, grain chalkiness was weakly positively correlated with phenotypic variation of
209 grain width, and the key genes affecting grain width were not the cause of chalkiness (Misra et al.
210 2020). It was also found that although several fine-mapped target genes and GWAS loci have been

211 found to influence chalkiness, many have either low or moderate effect (Gong et al. 2017; Wang et al.
212 2017; Quero et al. 2018; Misra et al. 2019). Recently, we detected two QTLs for grain chalkiness,
213 *qPGC9* and *qPGC11*, by substitution mapping, and found that they had no effect on grain shape (Yang
214 et al. 2021). Therefore, the relationship of grain chalkiness and grain shape in rice remains unclear. In
215 the present study, two closely linked QTLs, *qPGC8.1* and *qPGC8.2*, on chromosome 8 showed effects
216 on grain chalkiness and grain shape (Fig. 2 and Fig. 3). In addition, it is noted that the *fgr* gene for
217 fragrance (Bradbury et al. 2005) is located not only on the substitution segment of 15-08 but also in the
218 *qPGC8.1* mapped region, and the *GW8* gene controlling grain width (Wang et al. 2012) is located near
219 the *qPGC8.2* mapped region although it is outside the substitution segment of 03-08 (Fig. 1c). These
220 results indicated that the long arm of chromosome 8 may be a gene cluster area for grain development.
221 It is unclear that each of the regions of *qPGC8.1* and *qPGC8.2* carries two different genes, which
222 controlled grain chalkiness and grain shape respectively, or carries one gene controlling grain
223 chalkiness with pleiotropic effects on grain shape. Therefore, the mapping of *qPGC8.1* and *qPGC8.2*
224 laid a foundation for revealing the relationship between chalkiness and grain shape and its molecular
225 mechanism.

226

227 **Influence of high temperature on the effect of chalk QTLs**

228 It is found that the high temperature during the seed filling stage caused uneven grain filling and
229 resulted in chalk formation (Sreenivasulu et al. 2015; Masutomi et al. 2015; Morita et al. 2016;
230 Ishimaru et al. 2019). In Guandong province of China, rice is planted in two cropping seasons per year.
231 During the seed filling period, the air temperature of FCS is usually higher than that of SCS. In
232 2017-2019, the mean temperature of FCS was 4.1 °C higher than that of SCS (Additional file 1: Table
233 S5). It led to grain chalkiness of all lines in FCS was significantly higher than that in SCS (Fig. 4).
234 Obviously, the grain chalkiness of HJX74 and SSSLs was greatly affected by higher temperature
235 during the seed filling period.

236 Some QTLs for chalkiness were detected under high temperature stress (Nevame et al. 2018).
237 Kobayashi et al. (2007) detected three QTLs for chalkiness in *japonica* varieties under heat stress.
238 Tabata et al. (2007) identified four QTLs for chalkiness in a RIL population derived from a cross
239 between a heat stress-tolerant variety and a heat stress-sensitive variety. Wada et al. (2015) and
240 Miyahara et al. (2017) identified a set of chalk QTLs under heat stress condition using a RIL

241 population derived from a cross between a heat-tolerant variety and a heat stress-sensitive variety.
242 Recently, we mapped two QTLs for grain chalkiness, *qPGC9* and *qPGC11*, and found that the additive
243 effects of *qPGC9* and *qPGC11* on chalkiness in SCS were almost twice of those in FCS. The additive
244 effects of *qPGC9* and *qPGC11* on chalkiness were decreased by heat stress in FCS. Therefore, the
245 *qPGC9* and *qPGC11* were sensitive to high temperature (Yang et al. 2021). In the present study, we
246 found that although grain chalkiness of HJX74 and SSSLs was greatly affected by higher temperature
247 during the seed filling period, the additive effects of *qPGC8.1* and *qPGC8.2* on PCG, PCA and PGC
248 were no significant difference in different cropping seasons (Fig. 5). The results showed that the
249 additive effects of *qPGC8.1* and *qPGC8.2* on grain chalkiness were not affected by the high
250 temperature in FCS. It indicates that *qPGC8.1* and *qPGC8.2* were insensitive to high temperature of
251 FCS, which reactions are different from *qPGC9* and *qPGC11*. Therefore, *qPGC8.1* and *qPGC8.2* will
252 be useful for breeding of rice varieties with lower grain chalkiness under heat stress.

253

254 **Conclusion**

255 Two closely linked QTLs *qPGC8.1* and *qPGC8.2* controlling grain chalkiness and grain shape were
256 located on chromosome 8. The effect of *qPGC8.1* and *qPGC8.2* was incomplete dominance. The
257 additive effects of the two QTLs on grain chalkiness were no significant difference between FCS and
258 SCS. The *qPGC8.1* and *qPGC8.2* were insensitive to the high temperature in FCS. The mapping of
259 *qPGC8.1* and *qPGC8.2* laid a foundation for revealing the relationship between grain chalkiness and
260 grain shape and its molecular mechanism. The two QTLs will be useful for breeding of rice varieties
261 with lower grain chalkiness under heat stress.

262

263 **Materials and methods**

264 **Rice materials and cropping seasons**

265 Two SSSLs 03-08 and 15-08 with lower grain chalkiness were selected from the HJX74-SSSL library.
266 The substitution segment of 03-08 was from the donor Zhong4188 and that of 15-08 was from the
267 donor American Jasmine. The donors of both SSSLs are *indica* varieties. All rice materials were
268 planted at the farm of South China Agricultural University, Guangzhou (23°07'N, 113°15'E) from 2017
269 to 2019. The materials were planted in two cropping seasons per year, the FCS from late February to
270 middle July and the SCS from late July to middle November. Rice cultivation and controlling of

271 diseases and insect pests were common practices in southern China.

272

273 **Measurement of grain chalkiness**

274 The seeds of each line were harvested after full maturity. The dried seeds of 10 plants of each line were
275 processed into milled rice, and 200 head rice of each plant were randomly selected for measurement of
276 chalkiness (Yang et al. 2021). Images of the head rice were captured and the chalkiness parameters
277 were measured by Microtek ScanWizard EZ scanner and rice quality analyzer SC-E software
278 (Hangzhou Wanshen Detection Technology Co., Ltd., Hangzhou, China, www.wseen.com). PGC is the
279 product of PCG in total grains multiplied by PCA per chalky grain.

280

281 **Phenotyping of traits and statistical analysis**

282 Heading date, plant height and panicle number per plant was investigated in the field. Grain traits were
283 measured by the yield traits scorer (YTS), a rice phenotypic facility (Yang et al. 2014). The
284 percentages were converted to the arcsine square root for statistical analysis. The student's *t*-test was
285 used for comparison between two groups. Dunnett *t*-test was used to compare multiple groups with
286 control group. Least significance range (LSR) was used for multiple range test among multiple groups
287 (Duncan 1955). The data analysis and figure making were done by SPSS statistics 23.0 and OriginPro
288 9.0 (<https://www.originlab.com>).

289

290 **Genotyping of molecular markers and substitution mapping**

291 Molecular markers labeled "RM" were selected from online resources
292 (<https://archive.gramene.org/markers/>). New markers used in this study were designed using the
293 software of Primer Premier 5.0 (Lalitha 2000). The DNA samples were amplified by PCR method. The
294 PCR products were separated by gel electrophoresis on 6% denatured PAGE and detected by the silver
295 staining method (Tan et al. 2020). To develop secondary SSSLs or NILs, 03-08 and 15-08 were crossed
296 with the recipient HJX74. The NILs were developed from F_{2,3} populations derived from the crosses.
297 Minimum length, maximum length and estimated length of a substitution segment were estimated as
298 described by Tan (2020). When PGC showed significant difference between SSSL genotype and
299 HJX74 genotype, a QTL for PGC was detected on the substitution segment of SSSL. When multiple
300 substitution segments overlapped in NILs, the QTL was located in the overlapping region (Eshed and

301 Zamir, 1995; Tan et al. 2020). Additive effect of a QTL was defined as the phenotypic difference
302 between SSSL and HJX74 (Zhou et al. 2020). QTLs were named followed the method of McCouch et
303 al. (1997).

304

305 **Abbreviations**

306 FCS: First cropping season; HJX74: Huajingxian 74; NIL: Near-isogenic line; PCA: Percentage of
307 chalky area; PCG: Percentage of chalky grain; PGC: Percentage of grain chalkiness; QTL: Quantitative
308 trait locus; SCS: Second cropping season; SER: Stigma exertion rate; SSSL: Single-segment
309 substitution line.

310

311 **Declarations**

312 **Acknowledgments**

313 Not applicable.

314

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318

319 **Availability of data and materials**

320 All data generated or analyzed in this study are included in this published article and its additional
321 information files.

322

323 **Authors' contributions**

324 GZ and SW designed and supervised the works. WY performed most of the experiments and analyzed
325 the experimental data and prepared the draft of manuscript. LX, JL, QH, XL, QT, SL, ZL, SB, HZ, and
326 GL conducted a part of experiments. GZ analyzed the data and wrote the manuscript. All authors read
327 and approved the final manuscript.

328

329 **Ethics approval and consent to participate**

330 Not applicable.

331

332 **Consent for publication**

333 Not applicable.

334

335 **Competing interests**

336 The authors declare that they have no competing interests.

337

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495

496 **Figure legends**

497 **Fig. 1** Grain chalkiness in HJX74 and SSSLs. **a**, Plant types of HJX74 and SSSLs 15-08 and 03-08.
498 Scale bar: 15 cm. **b**, Percentage of grain chalkiness (PGC) (%) in HJX74 and SSSLs. Data were
499 presented as mean \pm S.E. in six cropping seasons, one-way ANOVA, two-tailed, Dunnett *t*-test was
500 used to test the differences, with HJX74 as control, ** represent $P \leq 0.01$. **c**, Chromosome locations of
501 the two SSSLs. Physical distance (Mb) is shown as rulers on the right of chromosome. Black bars on
502 the left of the chromosome 8 represent the estimated length of substitution segments in the SSSLs with
503 their code on the left. The functional markers *fgr* and *GW8* represent the loci of *fgr* gene for fragrance
504 and *GW8* gene controlling grain width. *Chr.* chromosome, *Mb* megabase.

505 **Fig. 2** Substitution mapping of *qPGC8.1* for grain chalkiness. **a**, The milled rice of the HJX74 and
506 SSSL 15-08. Scale bar: 1 cm. **b**, Substitution mapping of *qPGC8.1*. The positions of substitution
507 segments and the percentage of grain chalkiness (PGC) of 15-08 and its NILs are shown, with HJX74
508 as the control. The numbers under the chromosome indicate physical distance (Mb). White and black
509 blocks represent the homozygous genotypes of HJX74 and 15-08, respectively. PGC (%) was a mean \pm
510 S.E. in two cropping seasons. **c**, PGC of three genotypes of *qPGC8.1* in an F₂ population.

511 *qpgc8.1/qpgc8.1* represents homozygous genotype of HJX74 (n = 24); *qPGC8.1/qpgc8.1* represents
512 heterozygous genotype of HJX74/15-08 (n = 40); *qPGC8.1/qPGC8.1* represents homozygous genotype
513 of 15-08 (n = 22). **d**, Grain length of 15-08 and its NILs. **e**, Grain width of 15-08 and its NILs.
514 Significant difference analysis was by one-way ANOVA, Duncan, two-tailed. Values in the lines among
515 different letters are different at 1% level of significance in **b** and at 5% level of significance in **c**, **d** and
516 **e**.

517 **Fig. 3** Substitution mapping of *qPGC8.2* for grain chalkiness. **a**, The milled rice of the HJX74 and
518 SSSL 03-08. Scale bar: 1 cm. **b**, Substitution mapping of *qPGC8.2*. The positions of substitution
519 segments and the percentage of grain chalkiness (PGC) of 03-08 and its NILs are shown, with HJX74
520 as the control. The numbers under the chromosome indicate physical distance (Mb). White and black
521 blocks represent the homozygous genotypes of HJX74 and 03-08, respectively. PGC (%) was a mean \pm
522 S.E. in two cropping seasons. **c**, PGC of three genotypes of *qPGC8.2* in an F₂ population.
523 *qpgc8.2/qpgc8.2* represents homozygous genotype of HJX74 (n = 23); *qPGC8.2/qpgc8.2* represents
524 heterozygous genotype of HJX74/03-08 (n = 47); *qPGC8.2/qPGC8.2* represents homozygous genotype
525 of 03-08 (n = 30). **d**, Grain length of 03-08 and its NILs. **e**, Grain width of 03-08 and its NILs.
526 Significant difference analysis was by one-way ANOVA, Duncan, two-tailed. Values in the lines among
527 different letters are different at 1% level of significance in **b** and at 5% level of significance in **c**, **d** and
528 **e**.

529 **Fig. 4** The difference of chalky traits between first cropping seasons (FCS) and second cropping
530 seasons (SCS) in HJX74, 15-08 and 03-08. **a**, Percentage of chalky grain (PCG). **b**, Percentage of
531 chalky area (PCA). **c**, Percentage of grain chalkiness (PGC). *, ** and *** indicate the difference at
532 0.05, 0.01 and 0.001 levels of significance, respectively. *NS* no significance.

533 **Fig. 5** The additive effects of *qPGC8.1* and *qPGC8.2* on grain chalkiness in first cropping seasons
534 (FCS) and second cropping seasons (SCS). **a**, Percentage of chalky grain (PCG). **b**, Percentage of
535 chalky area (PCA). **c**, Percentage of grain chalkiness (PGC). *NS* no significance.

536

537 **Supplementary Information**

538 **Additional file 1: Table S1.** Phenotypes of chalky traits in SSSLs. **Table S2.** Markers developed to
539 detect the substitution segments of SSSLs. **Table S3.** Substitution segments of SSSLs. **Table S4.**
540 Phenotypes of agronomic traits in SSSLs. **Table S5.** Average temperatures for 30 days after flowering

541 of rice in two cropping seasons (XLSX 21 kb)

542

Figures

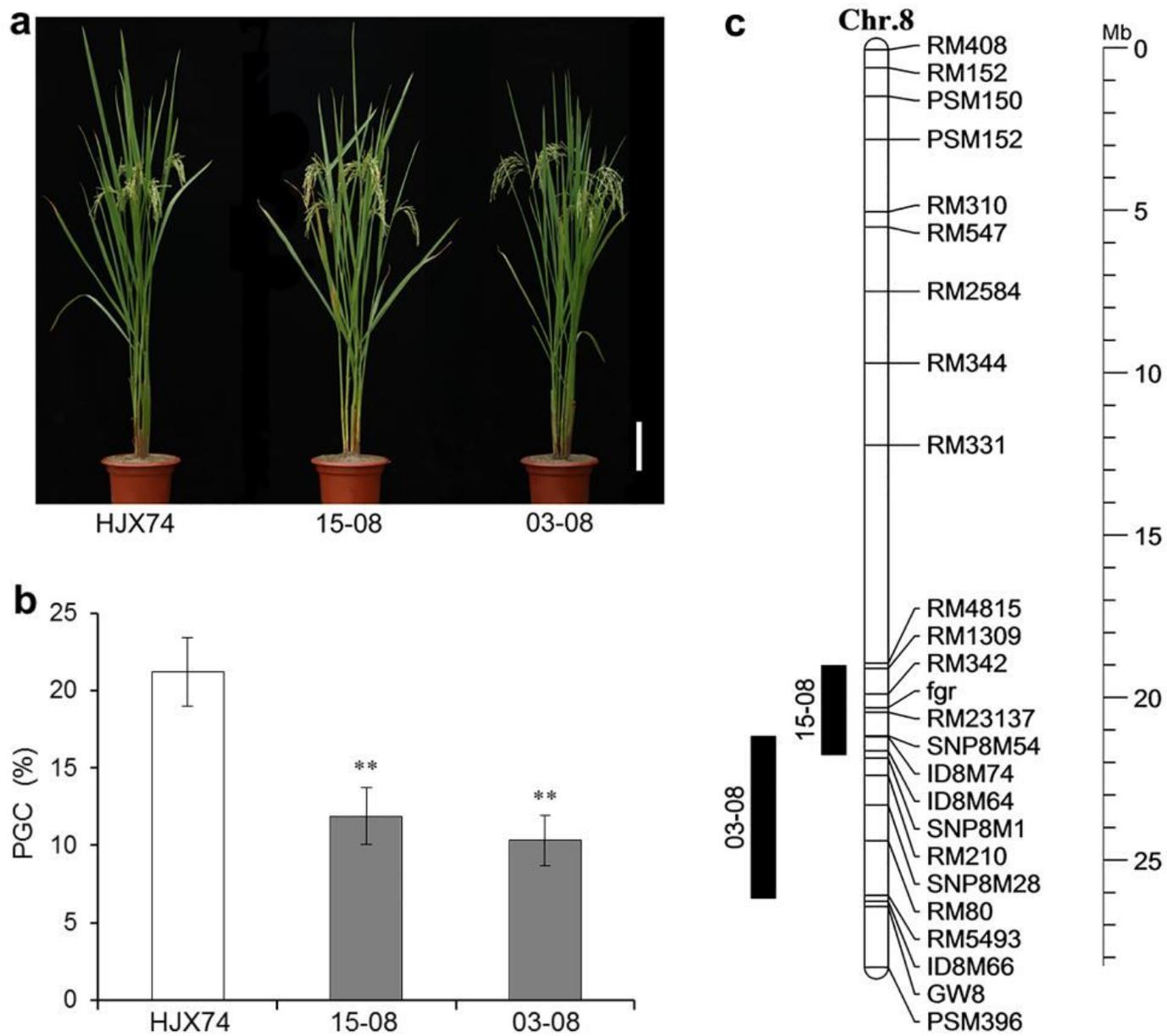


Figure 1

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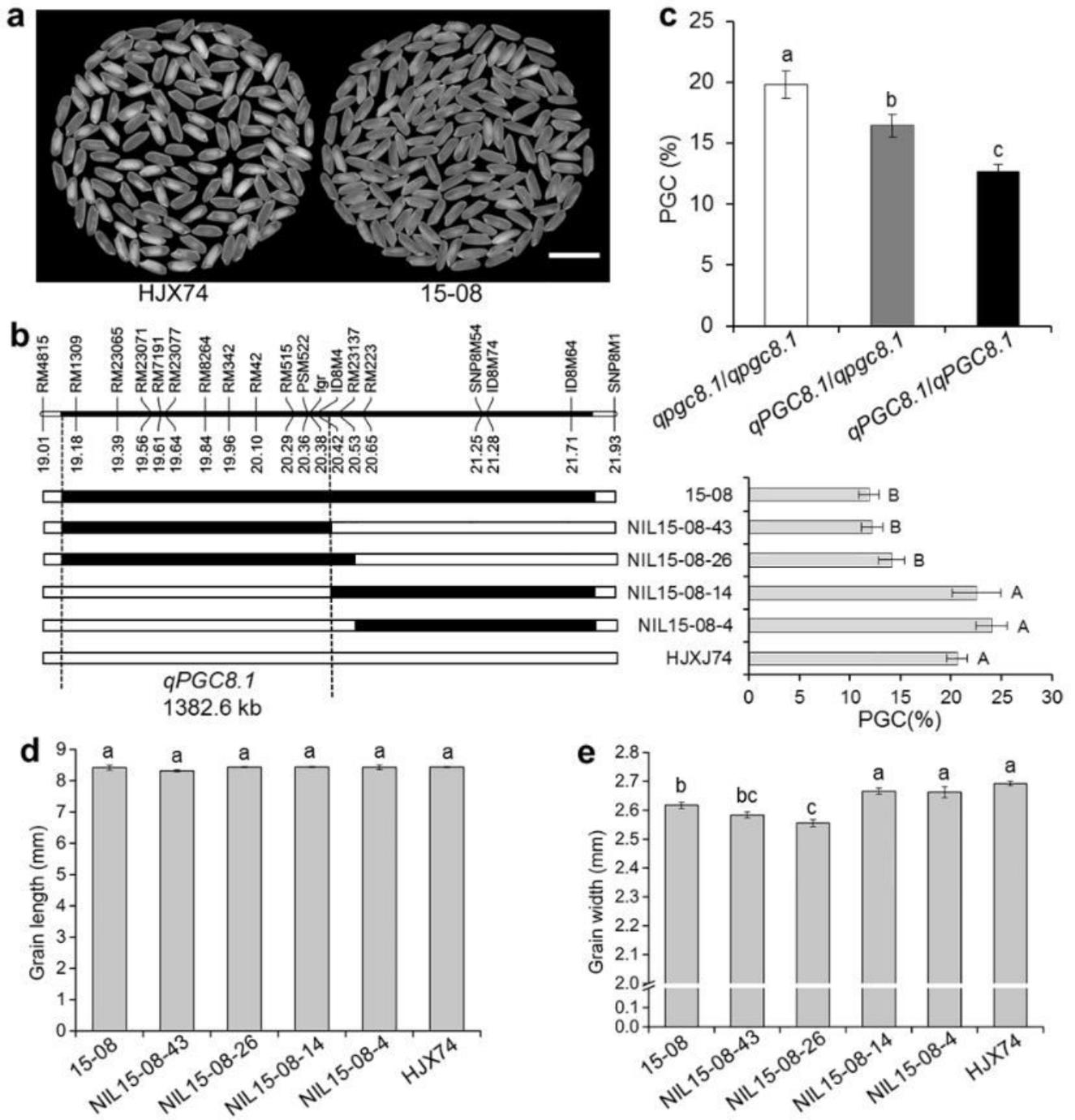


Figure 2

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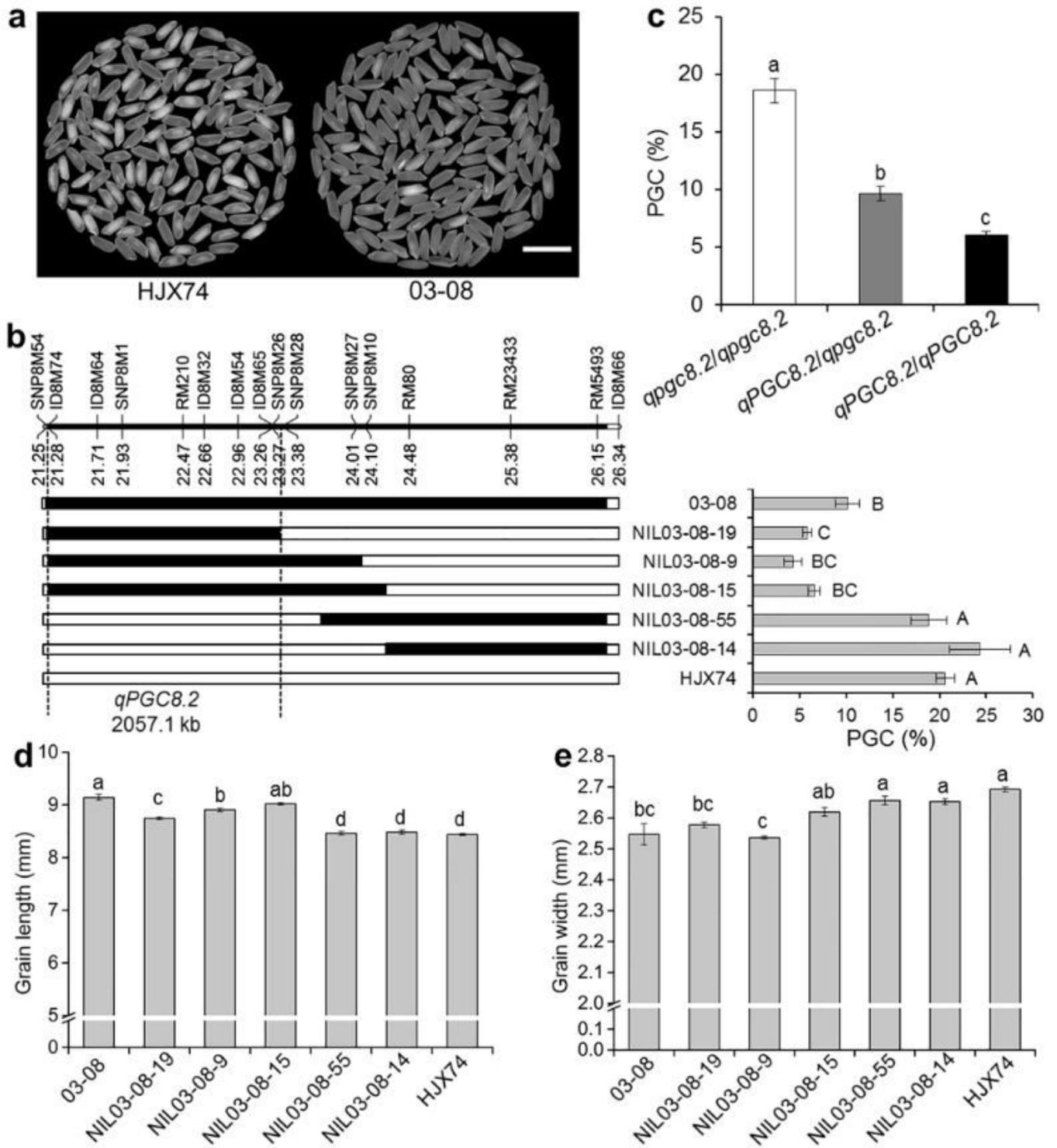


Figure 3

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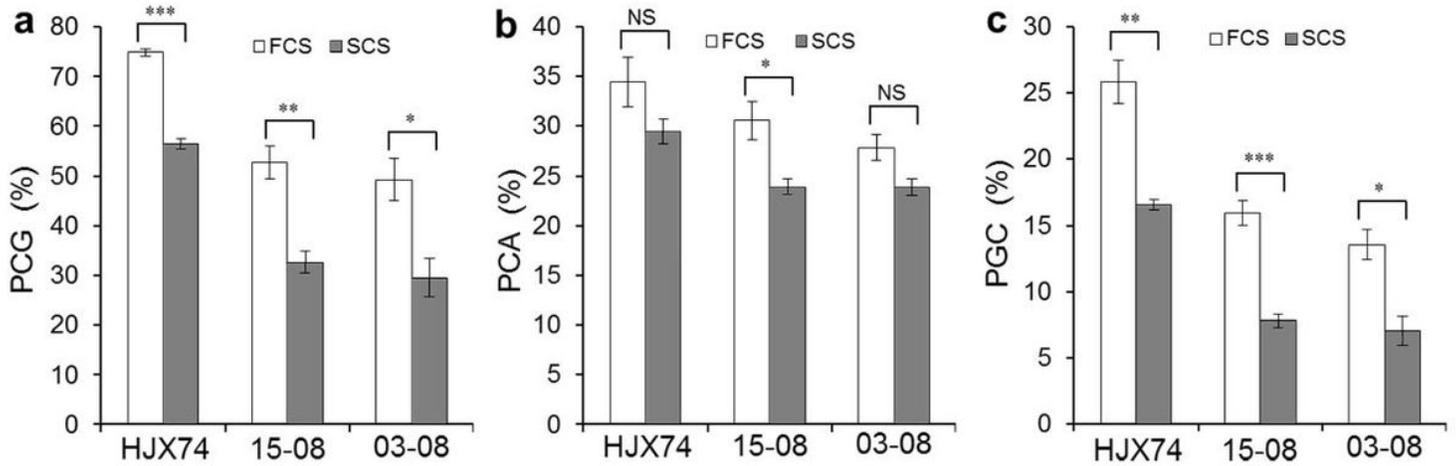


Figure 4

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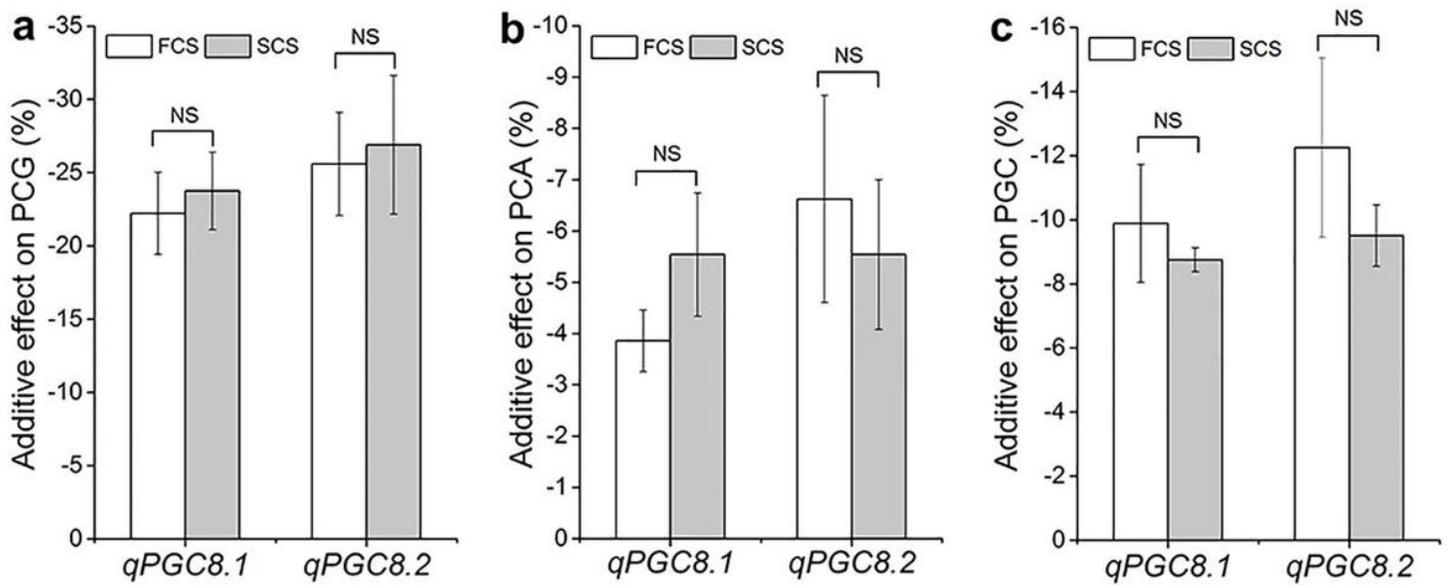


Figure 5

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