

High expression level of serpin peptidase inhibitor clade E member 2 is associated with poor prognosis in lung adenocarcinoma

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Research

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Abstract

Background: Recent studies have revealed that serpin peptidase inhibitor clade E member 2 (SERPINE2) is associated with tumorigenesis. However, SERPINE2 expression and its role in lung adenocarcinomas are still unknown.

Methods: The expression levels of SERPINE2 in 74 consecutively resected lung adenocarcinomas were analyzed by using immunostaining. Inhibition of *SERPINE2* expression by small interfering RNA (siRNA) was detected by quantitative PCR. Cell number assays and cell apoptosis assays were performed to clarify the cell-autonomous function of SERPINE2 in A549 and PC9 lung cancer cells.

Results: The overall survival of patients with high SERPINE2 expression was significantly worse than that of patients with low SERPINE2 expression ($P=0.0172$). Multivariate analysis revealed that SERPINE2 expression was an independent factor associated with poor prognosis ($P=0.03237$). The interference of *SERPINE2* decreased cell number and increased apoptosis in A549 and PC9 cells.

Conclusion: These results suggest that SERPINE2 can be used as a novel prognostic marker of lung adenocarcinoma.

Background

Serine proteinase inhibitor clade E member 2 (SERPINE2), also known as protease nexin-1 (PN-1), was first identified as a neurite-promoting factor released by cultured glioma cells [1]. In addition to glioma cells, various other cells secrete SERPINE2, including endothelial cells, fibroblasts, macrophages, platelets, smooth muscle cells, chondrocytes, astrocytes, and several types of tumor cells [2-6]. SERPINE2 was proven to be a member of the SERPINE family, which has serine protease activity [1, 7]. SERPINE2 is overexpressed in a variety of cancers, including breast cancer [3], pancreatic cancer [8], gastric cancer [9], osteosarcoma [10] and colorectal cancer [11], and its expression is correlated with the degree of cancer malignancy. A previous study demonstrated that SERPINE2 is upregulated by oncogenic activation of *RAS*, *BRAF* and *MEK1* and contributes to pro-neoplastic actions of ERK signaling in intestinal epithelial cells [11]. Therefore, SERPINE2 may be a potential therapeutic target for colorectal cancer treatment [11]. In lung adenocarcinomas in particular, high expression of SERPINE2 has been previously reported [12], but the relationship to prognosis or disease progression has never been reported. In this study, we examined the expression of SERPINE2 in 74 consecutive lung adenocarcinoma cases by immunohistochemistry using an anti-SERPINE2 antibody. In addition, we silenced *SERPINE2* in two kinds of non-small cell lung cancer (NSCLC) cell lines using siRNA and performed a proliferation assay and evaluation of apoptosis to clarify the cell-autonomous function of SERPINE2 in lung cancer cell lines.

Methods

Patients

During the period from January 2014 to December 2014, 78 consecutive patients were treated by complete surgical resection of lung adenocarcinoma at Kobe University Hospital, Kobe, Japan. Patients with a positive surgical margin were excluded from this study, and the remaining 74 patients were analyzed. The methods of data collection and analysis were approved by the institutional review board (permission number: 160117), and written informed consent was obtained from all patients.

Construction of the spiral array block and pathological studies

All surgical specimens were fixed with 10% formalin and embedded in paraffin. The paraffin-embedded block was sent to the Pathology Institute (Toyama, Japan) and processed into a spiral array block. The method of preparing the spiral array block is described in detail elsewhere [13]. Briefly, 50- to 100- μ m-thick slices of the sample block are cut and rolled up into cylindrical reels. These cylindrical reels are divided, and the reel containing the target site is embedded vertically in the recipient block. After that, the spiral array block is sliced and used for pathological examination. Serial 4- μ m sections were stained with hematoxylin and eosin (Fig. 1A, B). All histologic specimens that had been initially evaluated by pathologists were reviewed, and the expression levels of SERPINE2 were assessed independently by two pathologists (N.J. and T.N.) who were blinded to the clinical data. The histological diagnoses were based on the 2015 WHO classification [14]. Pathological stage was determined on the basis of the TNM classification of the International Union Against Cancer (UICC) [15].

Immunohistochemical staining

Anti-human SERPINE2 antibody (Lot no. 00014298, Proteintech, Rosemont, IL, USA) was used as the primary antibody. The spiral array block was cut into 4 μ m sections, which were mounted on silane-coated slides. The sections were deparaffinized in limonene (Nacalai, Kyoto, Japan) and dehydrated in a graded ethanol series. For antigen retrieval, the slides were heated for 20 min at 121 °C in 10 μ M citrate buffer (pH 6.0), and endogenous peroxidase was blocked with 3% hydrogen peroxide in absolute methyl alcohol. The slides were then blocked in 2.5% horse serum for 1 h and incubated with the primary antibodies (1:200). After overnight incubation, the slides were then washed with phosphate-buffered saline and incubated with ImmPRESS Reagent (Vector laboratories, Burlingame, CA, USA). The reaction products were stained with ImmPACT DAB reagent (Vector laboratories), and the sections were counterstained with hematoxylin. A high SERPINE2 case was defined as one in which there was at least a 50% increase in positive cells compared to that in normal tissue (Fig. 1C-F).

Cell culture

The lung adenocarcinoma cell lines A549 and PC9 were obtained from the American Type Culture Collection (Manassas, VA, USA). All cells were cultured in RPMI 1640 medium (Sigma, St. Louis, MO, USA) supplemented with 10% fetal bovine serum and 1% penicillin–streptomycin (Wako, Osaka, Japan) under 5% CO₂ at 37 °C.

SERPINE2 knockdown by small interfering RNAs (siRNAs)

The *SERPINE2* siRNAs (#1: s22188 and #2 s22189) and control siRNA (#14390843) were obtained from Thermo Fisher Scientific, MA, USA. Cells were plated in six-well plates at a density of 2×10^5 cells per well. The siRNAs or control siRNA duplexes were mixed with Lipofectamine RNAiMAX Transfection Reagent (Thermo Fisher Scientific) in Opti-MEM medium (Thermo Fisher Scientific) as described by the manufacturer's protocol and added to the plated cells. The cells were used for assessments 24 h after the addition of fresh medium.

Quantitative reverse transcription-polymerase chain reaction (qRT-PCR)

Preparation of total cellular RNAs and qRT-PCR of the RNAs by using them as a template were performed as described previously [16]. Relative mRNA levels were calculated with the $\Delta\Delta C_t$ method using *GAPDH* (glyceraldehyde-3-phosphate dehydrogenase) mRNA as an internal control. The primers used in this study were as follows: 5'-AATGAAACCAGGGATATGATTGAC-3' and 5'-TTGCAAGATATGAGAAACATGGAG-3' for *SERPINE2* and 5'-GCACCGTCAAGGCTGAGAAC-3' and 5'-ATGGTGGTGAAGACGCCAGT-3' for *GAPDH*.

Cell proliferation assay

To determine the knockdown effects of *SERPINE2* on cell growth, we used a cell proliferation assay (Cell Counting Kit-8 (CCK-8), Dojindo, Kumamoto, Japan) according to the manufacturer's instructions. A549 and PC9 cells were plated at a concentration of 5×10^3 cells/well in 96-well culture plates and analyzed 72 h later.

Fas ligand stimulation

To investigate the involvement of *SERPINE2* in the apoptosis pathway, lung cancer cell lines in which *SERPINE2* was knocked down were stimulated with 200 ng/ml soluble Fas ligand (Wako, Osaka, Japan), and the temporal changes in apoptosis-related proteins were evaluated by western blotting.

Western blotting

Primary antibodies against the following proteins were purchased from Cell Signaling Technology (Denver, MA, USA): β -actin (#4967), Bax (#2772), Bcl-2 (#4223), cleaved caspase 7 (#8438), and cleaved caspase 9 (#9505). To detect *SERPINE2*, we used the same antibody used for the immunohistochemical analysis at a different dilution of 1:1000. Cells were lysed in Cell Lysis Buffer (Cell Signaling Technology), and total cellular proteins (20 μ g) were separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE), which was followed by western blotting as described previously [17].

Evidence from TCGA database

To evaluate the relationship between *SERPINE2* mRNA expression and the overall survival of lung adenocarcinoma patients, we utilized The Cancer Genome Atlas (TCGA) dataset through the Gene Expression Profiling Interactive Analysis (GEPIA) web server[18]. We defined the patients in the 1st

quartile in terms of *SERPINE2* mRNA expression as the low *SERPINE2* group and those in the 4th quartile as the high *SERPINE2* group.

Statistical Analysis

All statistical analyses were performed with EZR version 1.37 (Saitama Medical Center, Jichi Medical University; <http://www.jichi.ac.jp/saitama-sct/SaitamaHP.files/statmed.html>; Kanda, 2018), which is a graphical user interface for R (The R Foundation for Statistical Computing, Vienna, Austria, version 3.4.1) [19]. Differences in patient characteristics between the two groups were tested for significance by the Pearson χ^2 test or the Fisher exact test. For the univariate analysis, the cumulative survival was estimated by the Kaplan-Meier method, and differences in variables were calculated by the log-rank test. A multivariate regression analysis was conducted according to the Cox proportional hazards model. All reported P values are 2-sided, and a P value less than 0.05 was considered significant.

Results

Relationship between *SERPINE2* expression and clinicopathologic factors

A consecutive series of 74 specimens of adenocarcinoma of the lung were examined for *SERPINE2* expression. *SERPINE2* expression in the cytoplasm of the cancer cells was observed in 19 of the 74 cases (26%). The relationship between *SERPINE2* expression and clinicopathologic factors is shown in Table 1. *SERPINE2* expression was significantly correlated with lymphatic invasion ($P = 0.0188$), suggesting that lung adenocarcinomas that express *SERPINE2* show aggressive features. There was no relationship between *SERPINE2* expression and EGFR mutation status ($P = 0.416$).

Relationship between *SERPINE2* expression and overall survival

The overall survival curve obtained by the Kaplan-Meier method is shown in Fig. 2A. *SERPINE2* expression was significantly correlated with a short survival time ($P = 0.0172$). Univariate analyses showed that high expression of *SERPINE2* ($P = 0.03228$) was correlated with a short survival time (Table 2). Table 3 shows the impact of potential prognostic factors on the survival of patients with adenocarcinoma with expression of *SERPINE2* based on the results of the multivariate analysis with the Cox proportional hazards model. All significant univariate parameters were included in the multivariate analysis simultaneously. Together, these results proved that *SERPINE2* was an independent predictor of a poor prognostic outcome. This is similar to the finding of the survival analysis of the TCGA dataset performed through the GEPIA webserver, which also showed that the high *SERPINE2* group had a worse prognosis ($P = 0.042$) than the low expression group (Fig. 2B).

Effect of *SERPINE2* siRNA on the autonomous behavior of cancer cells

Among lung cancer cell lines, H460, A549 and PC9 more highly express *SERPINE2* compared to BEAS-2B, which is a bronchial epithelial cell line (Fig. 3A). To test the effect of increased *SERPINE2* on cell

autonomous behavior in vitro, the proliferation ability of A549 and PC9 cells treated with *SERPINE2* siRNA or a negative control was examined by using a CCK-8 assay. After transfection, the interference efficiency of *SERPINE2* was first confirmed by qRT-PCR. When normalized to the expression of the negative control, *SERPINE2* expression was downregulated $6.3\pm 1.1\%$ and $4.0\pm 0.5\%$ in A549 and PC9 cells in the *SERPINE2* siRNA#1 group ($P<0.05$) and $3.9\pm 0.3\%$ and $8.5\pm 1.9\%$ in the siRNA#2 group ($p<0.05$), respectively (Fig. 3B). The protein levels of *SERPINE2* were also suppressed by gene knockdown (Fig. 3C). Then, the cell proliferation ability was monitored at 24, 48, and 72 h. The cell proliferation rate was significantly decreased in cancer cells with *SERPINE2* downregulation ($P < 0.05$) (Fig. 4). This result suggests that *SERPINE2* possibly plays a role in the proliferation of cancer cells.

The effect of *SERPINE2* knockdown on Fas ligand-mediated apoptosis

First, we performed cell migration and invasion assays, but we did not observe remarkable changes (Fig. S1 and S2). Furthermore, *BMP4*, which plays a crucial role in migration and invasion, was not activated by *SERPINE2* knockdown in A549 and PC9 lung cancer cells (Fig. S3). Then, to investigate the effect of *SERPINE2* on apoptosis, A549 and PC9 cells treated with *SERPINE2* siRNA or a negative control siRNA were stimulated with soluble Fas ligand (200 ng/ml). In both cell lines, Fas ligand-mediated apoptosis was more highly induced in cells with *SERPINE2* interference. Especially in A549 cells, the protein expression of bcl-2, an anti-apoptotic protein, was decreased (Fig. 5). Together, these results suggest that *SERPINE2* is not associated with cell migration and invasion but is associated with anti-apoptosis.

Discussion

The current study demonstrated the role of *SERPINE2* as an independent prognostic factor of NSCLC and its molecular mechanism for the first time. In terms of prognostic factors, evaluation of the pathological specimens showed that increased expression of *SERPINE2* was related to lymphatic invasion and poor overall survival. Using the GEPIA database, we found that high *SERPINE2* mRNA expression was also related to poor prognosis. The overall results for survival showed the same trend as that observed in previous reports on different cancers, including breast cancers [3], gastric cancers [9] and osteosarcomas [20]. Regarding the molecular mechanism, in vitro analysis revealed that *SERPINE2* plays a possible anti-apoptotic role, leading to the development of malignant features in cancer cells with high *SERPINE2* expression.

Our experiment showed that silencing of *SERPINE2* resulted in the inhibition of cell proliferation. Epidermal growth factor (EGF) induced *SERPINE2* expression through the EGF/MEK/ERK pathway, and *SERPINE2* knockdown reduced cell proliferation induced by EGF [21]. *SERPINE2* is thought to act as an effector of the EGF pathway to cause cell proliferation. Furthermore, *SERPINE2* is reported to inhibit plasminogen-induced apoptosis of Chinese hamster ovary fibroblasts (CHO-K1), which constitutively express tissue-type plasminogen activator (t-PA) [22]. In a previous study, transfection of the *SERPINE2* gene significantly inhibited the activity of plasmin and t-PA via the formation of inhibitory complexes and prevented cell detachment and apoptosis [22]. On the other hand, it was reported in a study using

prostate cancer cells that apoptosis was induced by SERPINE2 [5], and it is believed that the effect differs depending on the cancer type. The results of the current experiment suggest that SERPINE2 suppresses apoptosis in lung cancer cell lines and may be a target molecule for lung cancer treatment.

In esophageal squamous cell carcinomas, SERPINE2 inhibition resulted in a reduction in cell growth, migration and invasion [23]. In esophageal squamous cell carcinomas, SERPINE2 promotes tumor metastasis by activating bone morphogenetic protein 4 (BMP4) [23]. Hence, we revealed no remarkable changes in cell migration and invasion assays (Fig. S1 and S2) because the current study revealed that SERPINE2 was not associated with BMP4 in lung cancer cells (Fig. S3).

The limitation of the current study is that a relatively small sample size is included in the current study. Hence, other prognostic factors, such as T and N factors, were not significant factors associated with poor prognosis.

Conclusions

SERPINE2 can be a prognostic factor and might be a possible target of NSCLC by suppressing apoptosis.

Abbreviations

BMP4, bone morphogenetic protein 4; CCK-8, Cell Counting Kit-8; NSCLC, non-small cell lung cancer; qRT-PCR, quantitative reverse transcription-polymerase chain reaction; PN-1, protease nexin-1; SDS-PAGE, sodium dodecyl sulfate-polyacrylamide gel electrophoresis; SERPINE2, serine proteinase inhibitor clade E member 2; siRNA, small interfering RNA; UICC, International Union Against Cancer

Declarations

Ethics approval and consent to participate

The methods of data collection and analysis were approved by the institutional review board (permission number: 160117), and written informed consent was obtained from all the patients.

Consent for publication

Not applicable.

Availability of data and materials

All data generated or analyzed during this study are included in this article.

Competing interests

The authors declare that they have no competing interests.

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Authors' contributions

RD and TN wrote the manuscript. RD performed immunostaining. TK and HS made the database. TN and NJ performed pathological analysis. YM collected the samples. YY, MY and KK assisted with technical advice. MT and YN conducted the statistical analysis. All authors analyzed the data, conceived the study, read and approved the final manuscript.

Acknowledgments

The results shown here are in whole or in part based upon data generated by the TCGA Research Network: <https://www.cancer.gov/tcga>.

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Tables

Table 1. Patient characteristics

SERPINE2	high	low	P value
number of case	19	55	
age[median(range)]	69(54-83)	72(49-85)	
male	13	28	0.284
smoking history	14	30	0.181
lobectomy	16	40	0.372
partial resection	3	15	
pT1	10	26	0.847
pT2	9	28	
pT3	0	1	
pN0	16	43	0.121
pN1	0	8	
pN2	3	4	
pM0	19	55	
Pleural invasion	8	19	0.589
Lymphatic invasion	10	12	0.0188
Vascular invasion	9	20	0.425
pStagelA	11	25	0.163
pStagelB	5	13	
pStagelIA	0	11	
pStagelIB	0	2	
pStagelIIA	3	4	
R0	19	55	
EGFRmutation Ex21 L858R	5	7	0.416
Ex19 del	1	4	
Negative	9	23	
Unknown	12	13	
adjuvant chemotherapy	4	19	0.391

(Fisher's exact test)

Table 2. Univariate analysis

Variables	Cut off	Hazard ratio	95% CI for HR	P value
pT factor	≤2 vs. ≥2	3.08	0.596-15.91	0.1795
pN factor	0 vs. ≥1	2.787	0.6227-12.48	0.1801
Lymphatic invasion	0 vs. 1	2.375	0.5291-10.66	0.93750
Pleural invasion	0 vs. ≥1	3.383	0.7553-15.15	0.1111
Vascular invasion	0 vs. 1	2.623	0.5863-11.73	0.2072
SERPINE2	Low vs. High	5.273	1.151-24.16	0.03228

Table 3. Multivariate analysis

Variables	Cut off	Hazard ratio	95% CI for HR	P value
pT factor	≤2 vs. ≥2	2.2610	0.13140-38.900	0.57420
pN factor	0 vs. ≥1	3.5320	0.34880-35.770	0.28540
Lymphatic invasion	0 vs. 1	0.4981	0.06594-3.763	0.49940
Pleural invasion	0 vs. ≥1	0.6023	0.04930-7.358	0.69130
Vascular invasion	0 vs. 1	2.8550	0.31770-25.650	0.34910
SERPINE2	Low vs. High	9.0520	1.20300-68.090	0.03237

Figures

Figure 1

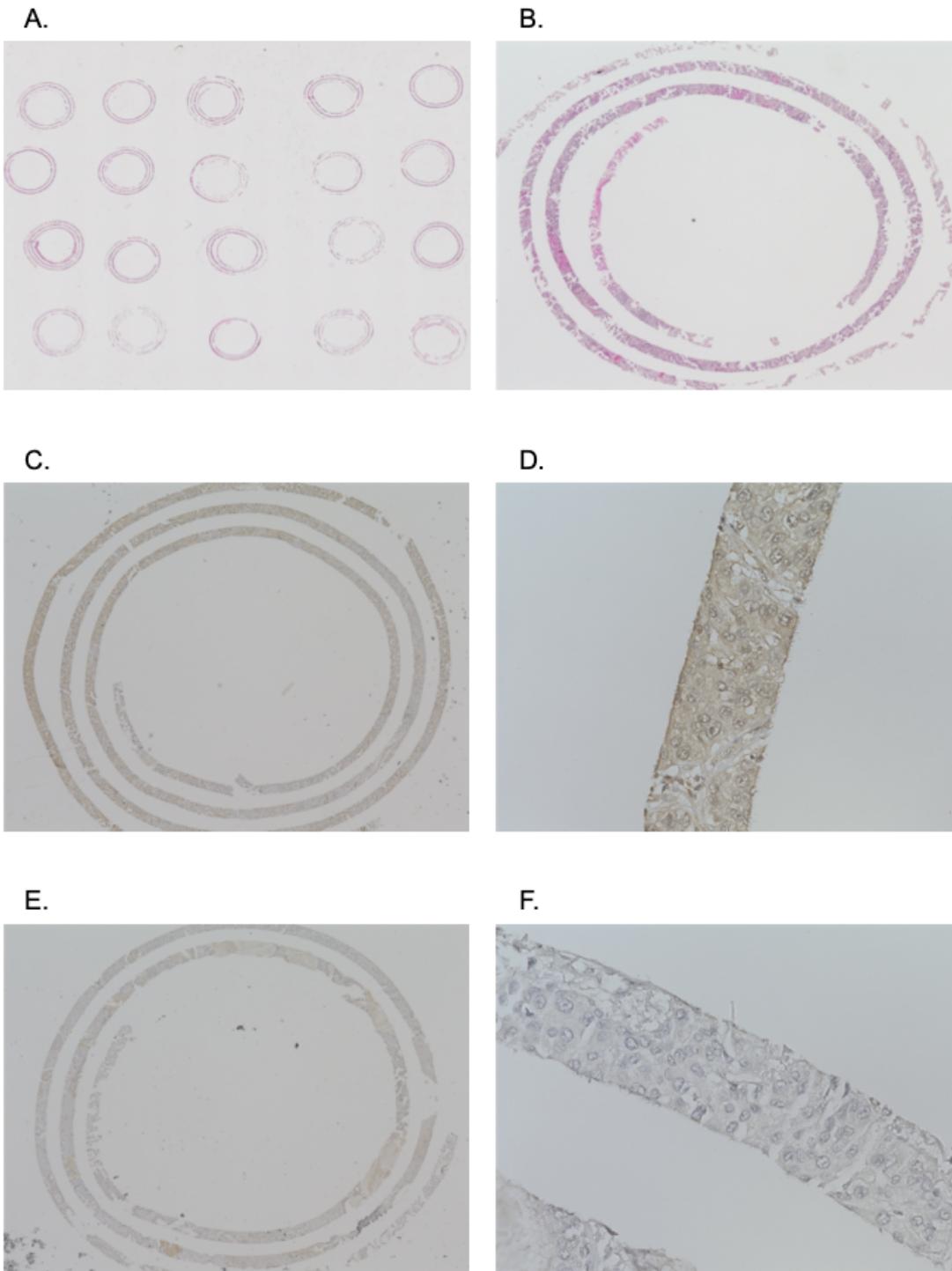


Figure 1

Microscopic images of spiral array specimens. Low power fields of (A) H&E staining, (C) SERPINE2 positive staining, and (E) SERPINE2 negative staining and high power fields of (B) H&E staining, (D) SERPINE2 positive staining, and (F) SERPINE2 negative staining are shown.

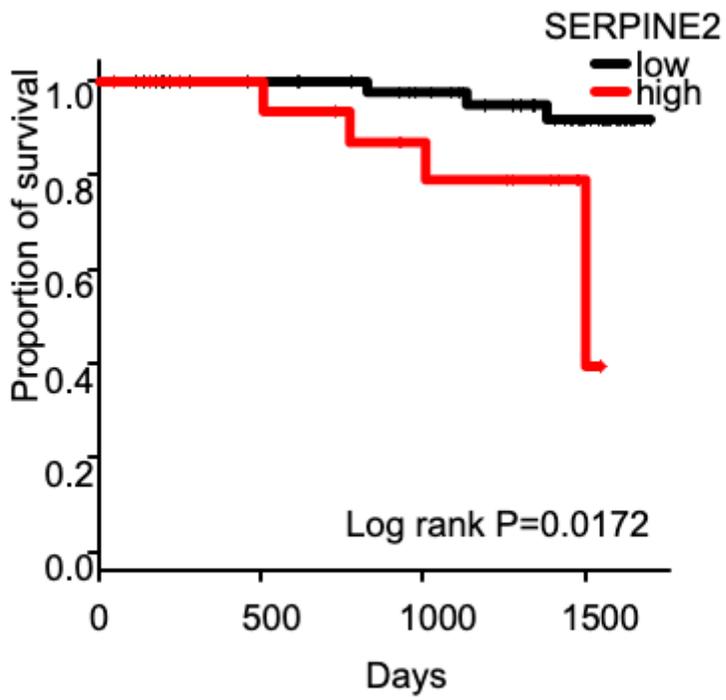


Figure 1

Kaplan-Meier survival curve. High SERPINE2 expression is correlated with poor prognosis.

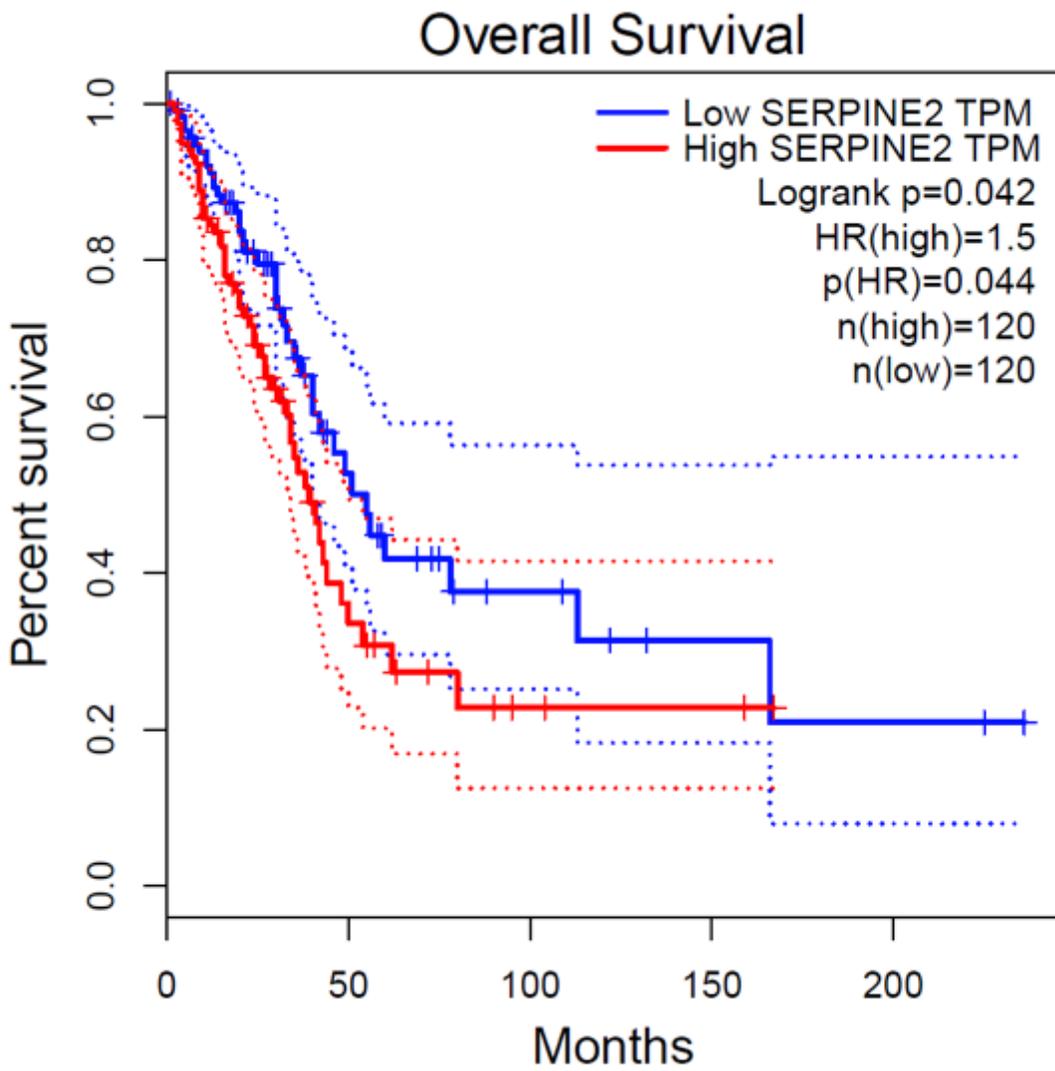
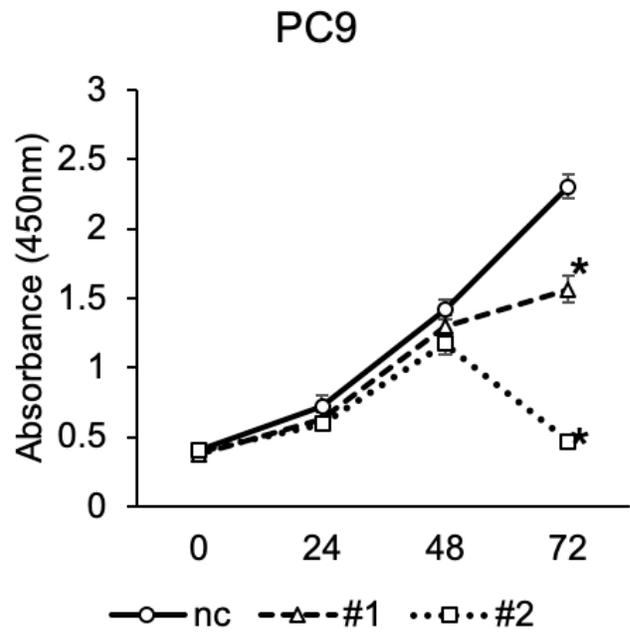
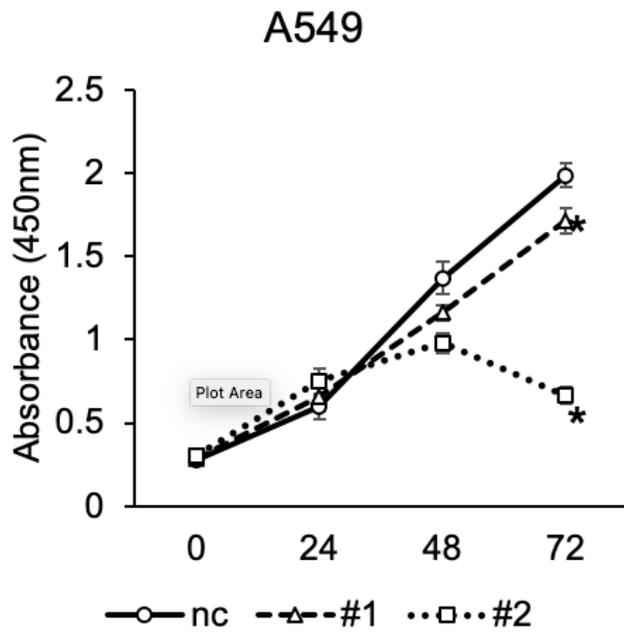


Figure 1

Survival analysis in TCGA dataset. The survival analysis based on TCGA dataset (LUAD) is shown.



*P<0.05 to negative control

Figure 1

Effect of SERPINE2 knockdown on cell number. The results of the cell number assay are shown. nc, negative control; #1, siRNA#1; #2, siRNA#2.

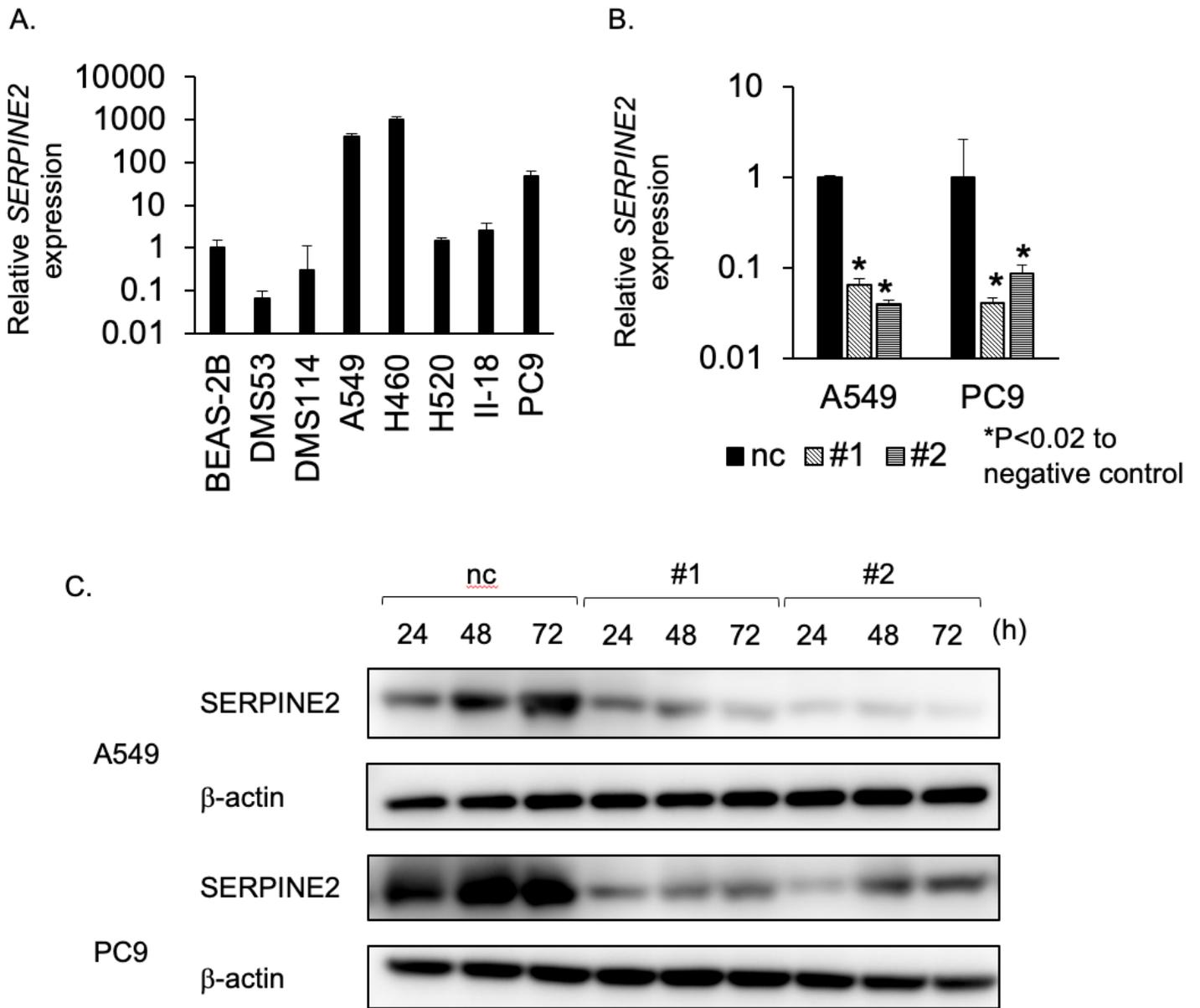


Figure 1

SERPINE2 expression and knockdown effect of SERPINE2. (A) SERPINE2 expression in lung cancer cell lines was normalized to BEAS-2B. (B, C) The efficacies of siRNA in A549 and PC9 cells were analyzed by qRT-PCR (B, relative to negative control) and western blotting (C). nc, negative control; #1, siRNA#1; #2, siRNA#2.

Figure 6

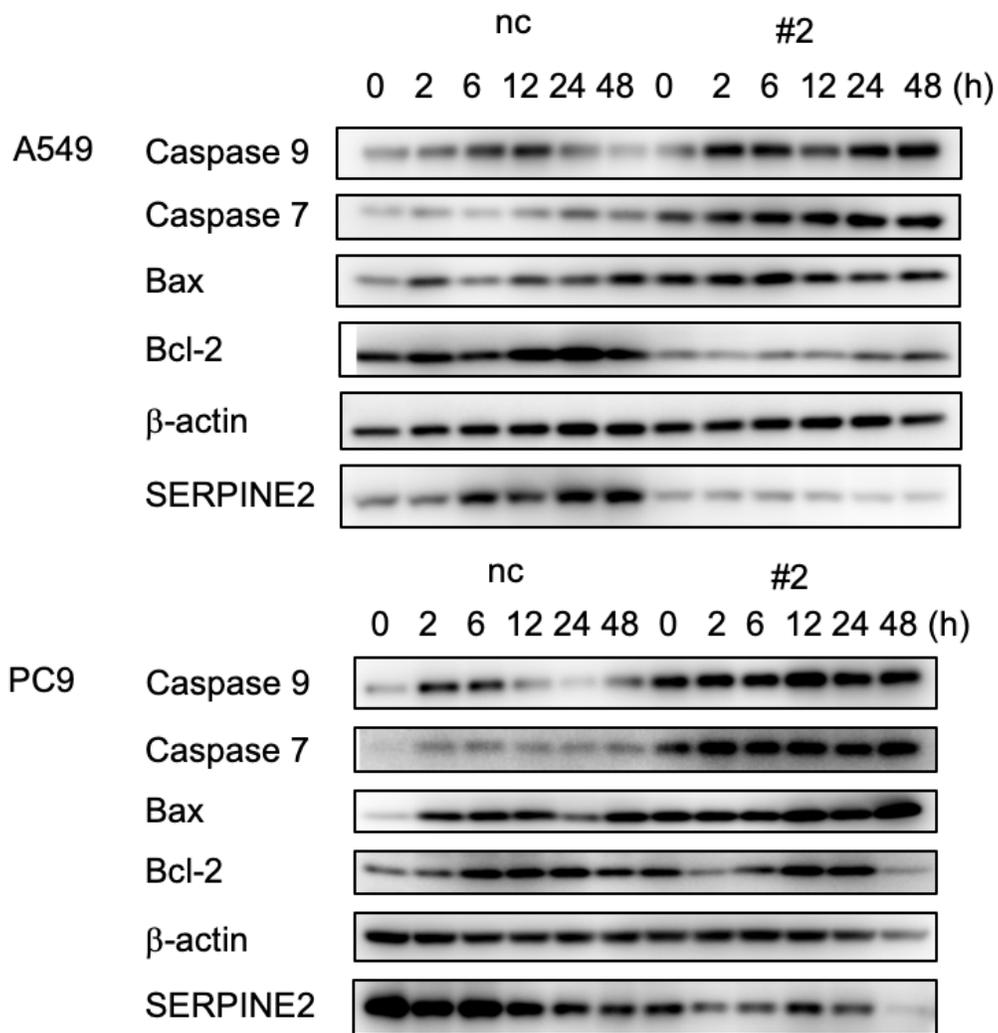


Figure 1

Effect of SERPINE2 knockdown on apoptosis. Western blotting analysis of apoptosis-related proteins at the indicated time points after Fas ligand stimulation is shown. A549 (upper) and PC9 (lower) cells pretreated with SERPINE2 siRNA were stimulated with Fas ligand (200 ng/ml). nc, negative control; #2, siRNA#2.

Supplementary Files

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