

Are Hips Too Relaxed to Develop Healthy ?

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Abstract

Background: Developmental dysplasia of hip (DDH) etiology is still a controversial issue. While many factors involved in the etiology, one of which is the hormone relaxin. Relaxin concentrations in patients with DDH may lead to pathodynamic changes during hip development by altering the physiological nature of the ligament, as well as by long-term exposure to relaxin during pregnancy. Our objective in this study was to determine the quantity of relaxin receptors in the ligamentum teres and their part in causing DDH.

Methods: We identified infants between birth and 3 years of age who had undergone open reduction for DDH between 2010 and 2012. 12 hips of 12 miss abortus fetus between 20 to 35 weeks of gestation were used as control group. Specimens obtained from two groups were stained with Relaxin-2 antibody and the amount of staining for relaxin receptors was determined using an ordinal H score.

Results: The mean H scores of infants with DDH were significantly higher than those of controls ($p=0.00$).

Conclusion: As a result, increased number of relaxin receptors in the ligamentum teres could be a risk factor for DDH and the number of relaxin receptors is an important clue in explaining the female:male ratio and clinical prognosis of early dysplastic hips.

Level of Evidence: Level 2, Prospective comparative study

Introduction

The incidence of developmental dysplasia of the hip (DDH) is 2.7 to 17 per 1000 births worldwide [1].

Hormonal changes, as well as genetic diseases such as Ehler Danlos syndrome, may be involved in DDH associated with joint laxity [2]. The relaxin hormone, which is necessary for the preparation of the maternal pelvis for labor and pelvic relaxation, passes through the placenta to the infants and causes laxity. In addition, the fact that girls are more sensitive to this hormone explains the higher DDH rates in girls compared to boys and it is one of most common hormones associated with DDH [3]. Relaxin has collagenolytic effects on ligamentous tissue in both pregnant and non-pregnant women. It may also be associated with laxity of fetal ligaments [4]. Increased serum relaxin concentration may result in increased joint laxity, as shown by Steinetz et al. in 2008. In the study, they reported increased levels of relaxin in pregnant woman with pelvic joint instability or hip joint laxity compared with the control group. (5) 3 years later Dragoo et al. reported that increased level of serum relaxin resulted in reduced ligament integrity and increased risk of injury for anterior cruciate ligament (ACL). (6)

Two hypotheses address the importance of relaxin in joint laxity. One hypothesis is that it exerts a direct effect on fetal ligament laxity by mediating progesterone and estrogen concentrations in connective tissue [7]. Relaxin also stimulates collagen turnover by increasing the collagenase expression and down-

regulates collagen synthesis [8]. The second hypothesis is that it exerts an indirect effect that reduces relaxin concentrations in maternal blood. This reduction can impair relaxation of the pelvic ligaments and thus lead to malpositioning the fetus [9]. However, relaxin's effect on joints as mediated through weakened ligaments remains unclear. Although prolonged exposure to relaxin during pregnancy may increase joint laxity during late pregnancy, the acute effect of short-term exposure on ligaments has not been determined [10].

The ligament of head of femur is a cone-shaped ligament extending between the femoral head and the acetabulum. It is a collagenous structure more robust than the capsule in fetuses and may be important in stabilizing the fetal hip joint [11]. The ligament may prevent or slow the progression of an unstable hip to a dislocated hip, particularly at term, when the discrepancy between the size of the femoral head and socket depth is greatest [12]. In fact, Weinstein and Ponseti reported that the ligamentum teres in patients with DDH is longer and thicker than normal, as a result of laxity and the abnormal position of the femoral head [13].

We believe that higher relaxin concentrations in patients with DDH may lead to pathodynamic changes during hip development by altering the physiological nature of the ligament, as well as by long-term exposure to relaxin during pregnancy. We also believe that the number of relaxin receptors is an important clue in explaining the female:male ratio and clinical prognosis of early dysplastic hips. Accordingly, our objective in this study was to determine the quantity of relaxin receptors in the ligamentum teres and their part in causing DDH.

Patients And Methods

This prospective study was approved by the institutional review board of authors' affiliated institutions. Parents and guardians provided written informed consent to have their children enroll in the study and to have samples taken for analysis

We identified infants between birth and 3 years of age who had undergone open reduction for DDH between 2010 and 2012. 12 hips of 12 miss abortus fetus between 20 to 35 weeks of gestation were used as control group. Infants with teratologic hip dislocations, any accompanying syndrome, or a history of intra-articular intervention of the hip, such as hip surgery or arthrography, were excluded.

Immunohistochemical Analysis

The ligamentum teres ligaments were harvested from infants who had undergone open reduction through a medial or anterolateral incision, fixed in 10% formaldehyde for 24 hours, and embedded in paraffin blocks. Sections, 3- μ m thick, were stained with hematoxylin & eosin. Sections were mounted on lysine-coated slides and deparaffinized for immunohistochemical analysis (Fig. 1).

Antigen retrieval was performed in a citrate buffer solution, and relaxin-2 antibody (GPR106 antibody, Biorbyt, US) was used at 1:50 dilution. Placental tissue was used as a positive control (Fig. 2). We stained

the nucleus of cells with relaxin antibody. Preparations were analyzed under light microscopy at a magnification of 400 \times at the most intense staining area. The intensity of staining for relaxin was categorized as low, medium, and high, as indicated in Fig. 3. The proportion of cells in each staining category was determined from 100 cells in the densest area of staining by a single pathologist with experience in immunologic staining. Pathologist was blinded for cases and control groups.

The amount of staining for relaxin receptors was determined using an ordinal H score [14]

H score = 3 x (percent of cells stained at high intensity) + 2 x (percent of cells

stained at medium intensity) + 1 x (percentage of cells stained at mild intensity). The H scores ranges from zero (no relaxin receptors) to 300 (all cells are stained with relaxin at high intensity). They are significantly correlated with the PCR data and are also sensitive to small changes in receptor expression [15].

Statistical Methods

The H scores were compared between groups with two-tailed Student's *t*-test. Alpha was set at 0.05. Data were analyzed with the SPSS software package.

Results

We analyzed data from 26 infants (20 female) who underwent open reduction for DDH and 12 hips of 12 fetus (9 female) used as control group. The 26 infants ranged in age from 10 to 36 months (mean [SD] age, 22 [6] months).

The mean (SD) H scores of infants with DDH were significantly higher than those of controls: 215 (59) vs. 52 (48); *P* = 0.00; 95% CI, (Table 1). Statistically significant difference between the two groups in terms of gender was not found.

Table 1

Results of Staining the Ligamentum Teres for Relaxin Receptors in Infants with Developmental Dysplasia of the Hip Treated with Open Reduction and Control Infants with Hip Fractures Treated with Hip Replacement

| Characteristic | Infants with developmental dysplasia of the hip, n = 26 | Missed abortus fetuses as control n = 12 | <i>P</i> |
|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------------|------------------------------------------|----------|
| Age, mean (SD), months | 21,7 (5,74) | 27,8 (5,4) weeks | |
| Females, n (%) | 20 (77%) | 9 (75%) | > 0.05 |
| Right hip involved, n (%) | 10 (38%) | 5 (42%) | > 0.05 |
| H score ^a , mean (SD) | 215 (59) | 52 (48) | < 0.001 |
| ^a H score = 3x(percent of cells stained at high intensity) + 2x[(percent of cells stained at medium intensity) + 1x(percentage of cells stained at mild intensity)]. See text for details. | | | |

Discussion

The cause of developmental dysplasia of the hip is still debated, although ligamentous laxity is a contributing factor [1]. Few studies have found evidence for a hormonal effect on DDH. Only a few studies have found an association between DDH and serum or cord blood relaxin concentrations, and many others have found no association. However, our study appears to be the only one that measured relaxin receptors in the ligament of head of femur [16].

Relaxin acts on the extracellular matrix through metalloproteinases by enhancing collagen degradation [8]. Relaxin is associated with both local and generalized laxity [17]. In patients with laxity-associated arthrosis of the first carpometacarpal joint, the concentration of relaxin inhibitors was higher in the volar oblique and anterior oblique ligaments, indicating that relaxin was involved as a cause of laxity [18, 19]. Another study in healthy humans found that trapezium-metacarpal joint laxity was proportional to serum relaxin concentrations [17]. A study in rats found that recombinant human relaxin 30 ng/mL weakened the joint capsule and decreased collagen accumulation in a dose-dependent way [20].

In women athletes, the “monthly window of potential injury” has been associated with hormonal changes, as defined by Möller-Nielsen [21]. This window coincided with peak concentrations of relaxin during the menstrual cycle, and generalized joint laxity was increased during these peak times [21]. Relaxin also increased ligamentous laxity during pregnancy [22]. Relaxin receptors are up-regulated by estrogen therapy, and joint laxity occurring after the first trimester of pregnancy is synchronous with peak relaxin concentrations [22].

Steinetz et al. hypothesized that increased serum relaxin concentration may result in increased joint laxity. In their study, they reported that increased levels of relaxin in pregnant woman with pelvic joint instability or hip joint laxity compared with the control group. (Y) 3 years later Dragoo et al. reported that increased level of serum relaxin resulted in reduced ligament integrity and increased risk of injury for anterior cruciate ligament (ACL) (Y-1)

In one of two other separate studies by Dragoo, tibial translation in guinea pigs was increased by 12.8% (absolute increase, 1.09 mm) 21 days after relaxin administration. The maximum load of the anterior cruciate ligament was also weaker by 36%, dropping from 64.1 N to 40.4 N, and relaxin altered the structure of the ligament by collagenolytic effects [23]. In the second study, relaxin increased the synthesis of collagenase and many matrix metalloproteinases in anterior cruciate ligament rupture and decreased local collagen synthesis in women orthopedic patients [24]. Increases in estrogen and relaxin during the normal menstrual cycle last only a very short time and are insufficient to explain any collagenolytic effect. It appears that long-term fluctuations are required to remodel the collagen [24].

Prolidase activity is increased in patients with DDH [25]. High prolidase activity indicates increased collagen metabolism, which is consistent with relaxin's mechanism of action. In the present study, we found that the number of relaxin receptors was twice as high in infants with DDH than in the elderly control group. We believe that this higher number of receptors and long-term exposure to relaxin from the mother during pregnancy may alter the nature of the ligamentum teres, resulting in hyperlaxity, possibly predisposing the fetus to DDH. However, in this case, it appears that the higher number of fetal receptors, rather than the concentration of relaxin from the mother, might be the cause because Forst *et al.* found that relaxin concentrations in specimens from cord blood were lower in neonates with DDH than in those of a healthy control group [4]. In a similar study, Vogel *et al.* also found no relationship between relaxin concentrations and DDH in 15 infants with DDH [26].

The ligamentum teres is important in stabilizing the pathophysiology of DDH [27]. Wenger *et al.* suggested that the strength of the ligamentum teres was crucial in the biomechanics of the hip and recommended surgical reconstruction for patients with DDH [27]. Another study on rats reported that greater strength of the ligamentum teres helps proper remodeling of the acetabulum against the femoral head [28].

The risk of DDH is four times as high in females as in males [29]. Desteli *et al.* found a negative relationship between the number of estrogen receptors and collagen metabolism in the ligamentum teres harvested from infants with DDH [30]. Furthermore, relaxin receptor isoforms are down-regulated with progesterone and high-dose estrogen, increasing the laxity, whereas progesterone and estrogen are down-regulated with testosterone [31]. Testosterone concentrations are 8 times as high in men as they are in women, and testosterone reduces the expression of the relaxin receptor, as well as decreasing the passive range of motion in joints [32].

The following questions arise from these findings: because relaxin is a maternal hormone, is it likely that attenuated relaxin activity improves a hip from Graf type IIa to type I in neonates? In addition, in hips with

more relaxin receptors, although relaxin concentrations are reduced in time, does the long-term action of relaxin cause permanent dysplasia? Finally, can the down-regulation of relaxin receptors caused by testosterone in men be the reason for their lower rate of DDH? All these questions may guide future studies. In this preliminary study, we used a single evaluator to score the intensity of staining. Given that we found an association between relaxin receptors and DDH, however, future studies should reduce the chances of bias by using two or more evaluators.

Limitations

First, our study group and control group have a significant age difference which may influence serum relaxin levels and intraligamentous relaxin levels. Our study group consists of babies less than 3 months old, but our control group were miss abortus fetuses between 20 to 35 weeks of gestation. Second, as the hip development finishes before 20 weeks, we used to prefer fetuses over 20 weeks, but we cannot be sure that these fetuses have a genetic abnormality or would develop a DDH in future.

Conclusions

Our data support the conclusion that the number of relaxin receptors in the ligamentum teres is significantly higher in infants with DDH than in healthy control patients and increased number of relaxin receptors in the ligamentum teres could be a risk factor for DDH.

Abbreviations

DDH : Developmental Dysplasia of Hip, ACL : Anterior Cruciate Ligament

Declarations

Ethics Approval and Consent to Participate

This prospective study was approved by the institutional review board of authors' affiliated institutions. Parents and guardians provided written informed consent to have their children enroll in the study and to have samples taken for analysis.

Consent for Publication

Parents and guardians provided written informed consent to related informations to be published.

Availability of Data and Materials

Datasets are available in the form of an excel table.

Competing Interests

All authors declare that they have no conflict of interest.

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No funding has been received.

Author's Contributions

Semih Ayanoğlu M.D. is responsible for design of the work

Haluk Çabuk M.D. is responsible for design of the work and interpretation of data for the work

Fatmagül Kuşku Çabuk is responsible for examination of pathologic materials

Kubilay Beng is responsible for study desing and data analysis

Timur Yıldırım is responsible for medical writing and ethical considerations

Süheyla Uyar Bozkurt is responsible for examination of pathologic materials and data collection

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References

1. Tredwell SJ, Bell HM. Efficacy of neonatal hip examination. *J Pediatr Orthop.* 1981 Jun 1;(1):61–5.
2. Thieme WT, Wynne-Davies R. Clinical examination and urinary oestrogen assays in newborn children with congenital dislocation of the hip. *J Bone Joint Surg Br.* 1968 Aug;50(3):546–50.
3. Rhodes AML, Clarke NMP. A review of environmental factors implicated in human developmental dysplasia of the hip. *J Child Orthop.* 2014 Oct;8(5):375–9.
4. Forst J, Forst C, Forst R, Heller KD. Pathogenetic relevance of the pregnancy hormone relaxin to inborn hip instability. *Arch Orthop Trauma Surg.* 1997 Apr;116(4):209–12.
5. Steinetz BG, Williams AJ, Lust G, Schwabe C, Bullesbach EE, Goldsmith LT. Transmission of relaxin and estrogens to suckling canine pups via milk and possible association with hip joint laxity. *Am J Vet Res.* 2008;69(1):59–67.
6. Dragoo JL, Castillo TN, Braun HJ, Ridley BA, Kennedy AC, Golish SR. Prospective correlation between serum relaxin concentration and anterior cruciate ligament tears among elite collegiate female athletes. *Am J Sports Med* 2011a; 39 (10): 2175–2180.
7. Goldsmith L, Lust G, Steinetz B. Transmission of relaxin from lactating bitches to their offspring via suckling. *Biol Reprod.* 1994 Feb;50(2):258–65.
8. Unemori EN, Amento EP. Relaxin modulates synthesis and secretion of procollagenase and collagen by human dermal fibroblasts. *J Biol Chem.* 1990 Jun;25(18):10681–5. 265(.

9. Borthwick GM, Borthwick AC, Grant P. Relaxin levels in the human: an indicator of target, storage and production sites. In: MacLennan AH, Tregear GW, BryantGreenwood GD, editors. Progress in relaxin research. Singapore: World Scientific Publishing Co.; 1995. pp. 25–60.
10. Schauburger CW, Rooney BL, Goldsmith L, Shenton D, Silva PD, Schaper A. Peripheral joint laxity increases in pregnancy but does not correlate with serum relaxin levels. *Am J Obst Gynecol*. 1996 Feb;174(2):667–71.
11. Crelin ES. An experimental study of hip stability in human newborn cadavers. *Yale J Biol Med*. 1976 May;49(2):109–21.
12. Walker JM. Growth characteristics of the fetal ligament of the head of femur: significance in congenital hip disease. *Yale J Biol Med*. 1980 Jul-Aug;53(4):307–16.
13. Wolf JM, Williams AE, Delaronde S, Leger R, Clifton KB, King KB. Relationship of serum relaxin to generalized and trapezial-metacarpal joint laxity. *J Hand Surg Am*. 2013 Apr;38(4):721–8.
14. Hirsch FR, Varella-Garcia M, Bunn PA Jr, Di Maria MV, Veve R, Bremmes RM, Barón AE, Zeng C, Franklin WA. Epidermal growth factor receptor in non-small-cell lung carcinomas: Correlation between gene copy number and protein expression and impact on prognosis. *Clin Oncol*. 2003 Oct 15;21(20):3798 – 807.
15. Specht E, Kaemmerer D, Sängler J, Wirtz RM, Schulz S, Lupp A. Comparison of immunoreactive score, HER2/neu score and H score for the immunohistochemical evaluation of somatostatin receptors in bronchopulmonary neuroendocrine neoplasms. *Histopathology*. 2015 Sep;67(3):368–77.
16. Vogel I, Andersson J, Uldjberg N. Serum relaxin in the newborn is not a marker of neonatal hip instability. *J Pediatr Orthop*. 1998 Jul-Aug;18(4):535–7.
17. Wolf JM, Williams AE, Delaronde S, Leger R, Clifton KB, King KB. Relationship of serum relaxin to generalized and trapezial-metacarpal joint laxity. *J Hand Surg Am*. 2013 Apr;38(4):721–8.
18. Lubahn J, Ivance D, Konieczko E, Cooney T. Immunohistochemical detection of relaxin binding to the volar oblique ligament. *J Hand Surg*. 2006 Jan;31(1):80–4.
19. Wolf JM, Scher DL, Etchill EW, Scott F, Williams AE, Delaronde S, King KB. Relationship of relaxin hormone and thumb carpometacarpal joint arthritis. *Clin Orthop Relat Res*. 2014 Apr;472(4):1130–7.
20. Unemori EN, Beck LS, Lee WP, Xu Y, Siegel M, Keller G, Liggitt HD, Bauer EA, Amento EP. Human relaxin decreases collagen accumulation in vivo in two rodent models of fibrosis. *J Invest Dermatol*. 1993 Sep;101(3):280–5.
21. Möller-Nielsen J, Hammar M. Women's soccer injuries in relation to the menstrual cycle and oral contraceptive use. *Med Sci Sports Exerc*. 1989 Apr;21(2):126–9.
22. Petersen LK, Vogel I, Agger AO, Westergård J, Nils M, Uldbjerg N. Variations in serum relaxin (hRLX-2) concentrations during human pregnancy. *Acta Obstet Gynecol Scand*. 1995 Apr;74(4):251–6.
23. Dragoo JL, Padrez K, Workman R, Lindsey DP. The effect of relaxin on the female anterior cruciate ligament: analysis of mechanical properties in an animal model. *Knee*. 2009 Jan;16(1):69–72.

24. Dragoo JL, Lee RS, Benhaim P, Finerman GA, Hame SL. Relaxin receptors in the human female anterior cruciate ligament. *Am J Sports Med.* 2003 Jul-Aug;31(4):577–84.
25. Soran N, Altindag O, Aksoy N, Cakır H, Taşkın A, Soran M, Işıkan E. The association of serum prolidase activity with developmental dysplasia of the hip. *Rheumatol Int.* 2013 Aug;33(8):1939–42.
26. Vogel I, Andersson JE, Uldbjerg N. Serum relaxin in the newborn is not a marker of neonatal hip instability. *J Pediatr Orthop.* 1998 Jul-Aug;18(4):535–7.
27. Wenger DR, Miyanji F, Mahar A, Oka R. The mechanical properties of the ligamentum teres: a pilot study to assess its potential for improving stability in children's hip surgery. *J Pediatr Orthop.* 2007 Jun;27(4):408–10.
28. Canillas F, Delgado-Martos MJ, Martos-Rodriguez A, Quintana-Villamandos B, Delgado-Baeza E. Contribution to the initial pathodynamics of hip luxation in young rats. 2012 Sep;32(6):613–20.
29. de Hundt M, Vlemmix F, Bais JMJ, Hutton EK, de Groot CJ, Mol BW, Kok M. Risk factors for developmental dysplasia of the hip: a meta-analysis. *Eur J Obstet Gynecol Reprod Biol.* 2012 Nov;165(1):8–17.
30. Desteli EE, Pişkin A, Gülman AB, Kaymaz F, Köksal B, Erdoğan M. Estrogen receptors in hip joint capsule and ligamentum capitis femoris of babies with developmental dysplasia of the hip. *Acta Orthop Traumatol Turc.* 2013 May;47(3):158–61.
31. Dehghan F, Muniandy S, Yusof A, Salleh N. Sex-steroid regulation of relaxin receptor isoforms (RXFP1 & RXFP2) expression in the patella tendon and lateral collateral ligament of female WKY Rats. *Int J Med Sci.* 2014 Jan;9(2):180–91. 11(.
32. Dehghan F, Muniandy S, Yusof A, Salleh N. Testosterone reduces knee passive range of motion and expression of relaxin receptor isoforms via 5 α -dihydrotestosterone and androgen receptor binding. *Int J Mol Sci.* 2014 Mar;17(3):4619–34. 15(.

Figures

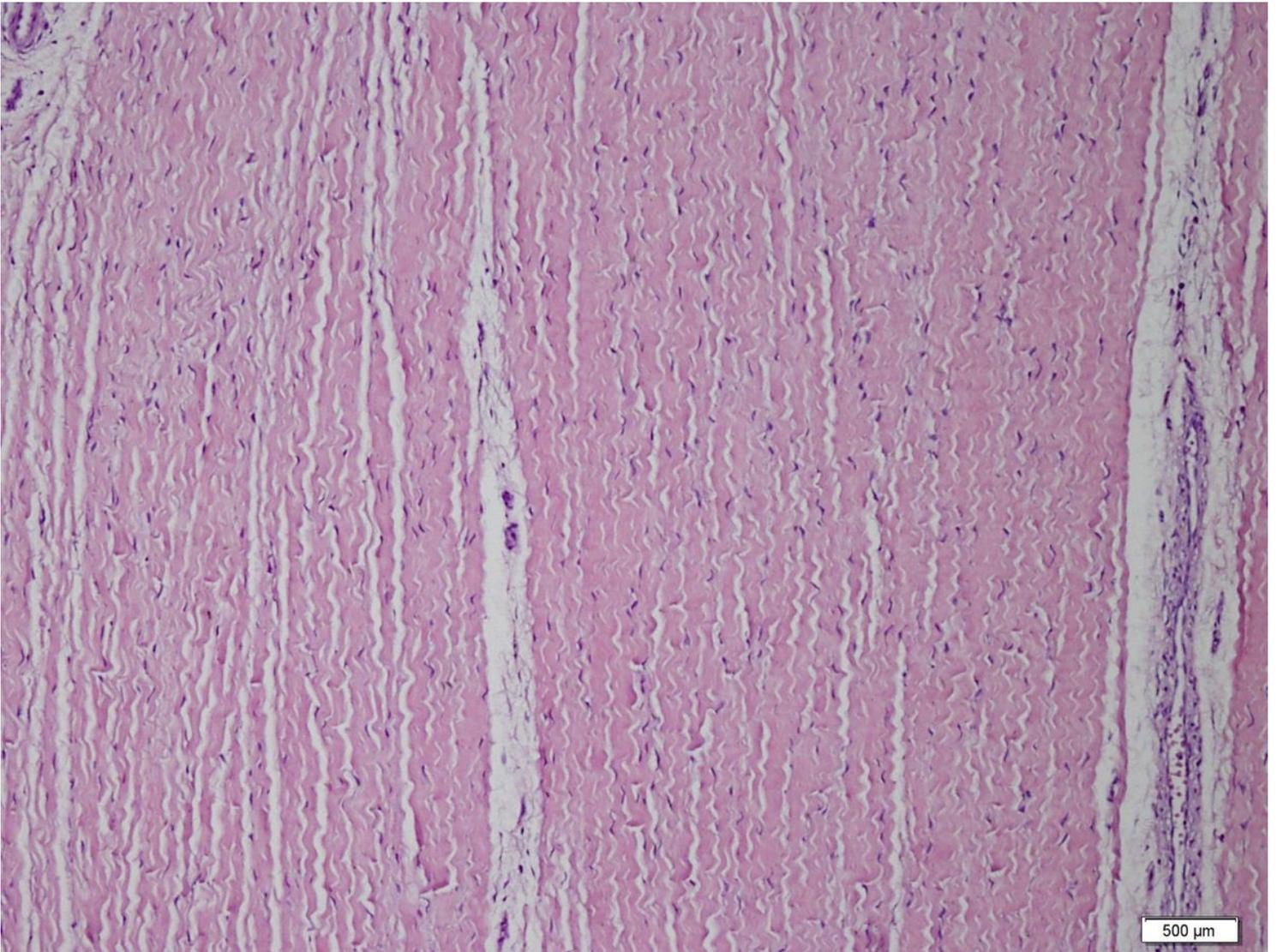


Figure 1

Morphology of the ligamentum teres. Note the collagen bundles extending parallel to each other (Hematoxylin & eosin;X100)

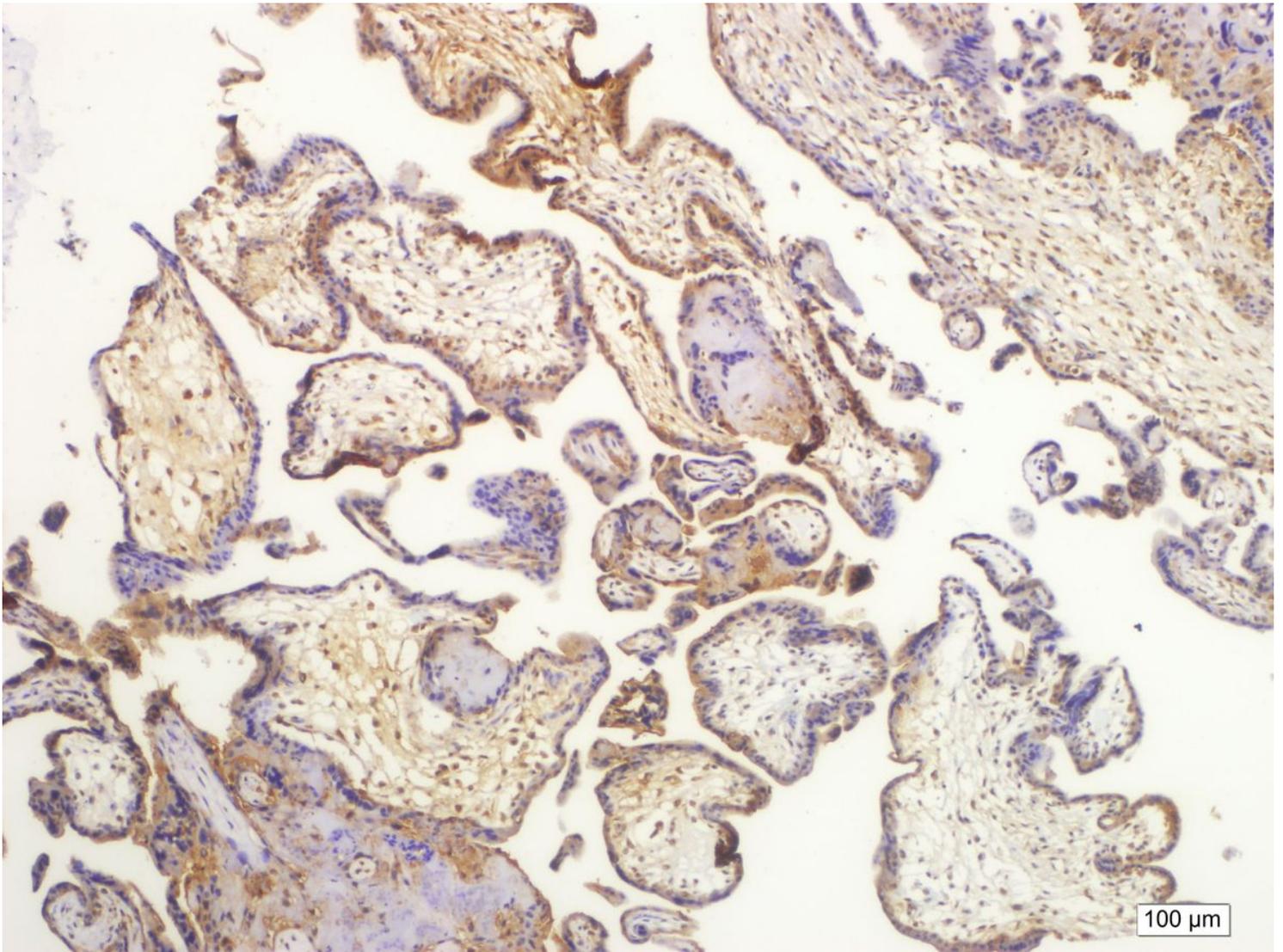


Figure 2

Placental tissue as a positive control for relaxin. Brown cytoplasmic staining was accepted as positive staining. (Hematoxylin & eosin; X200)

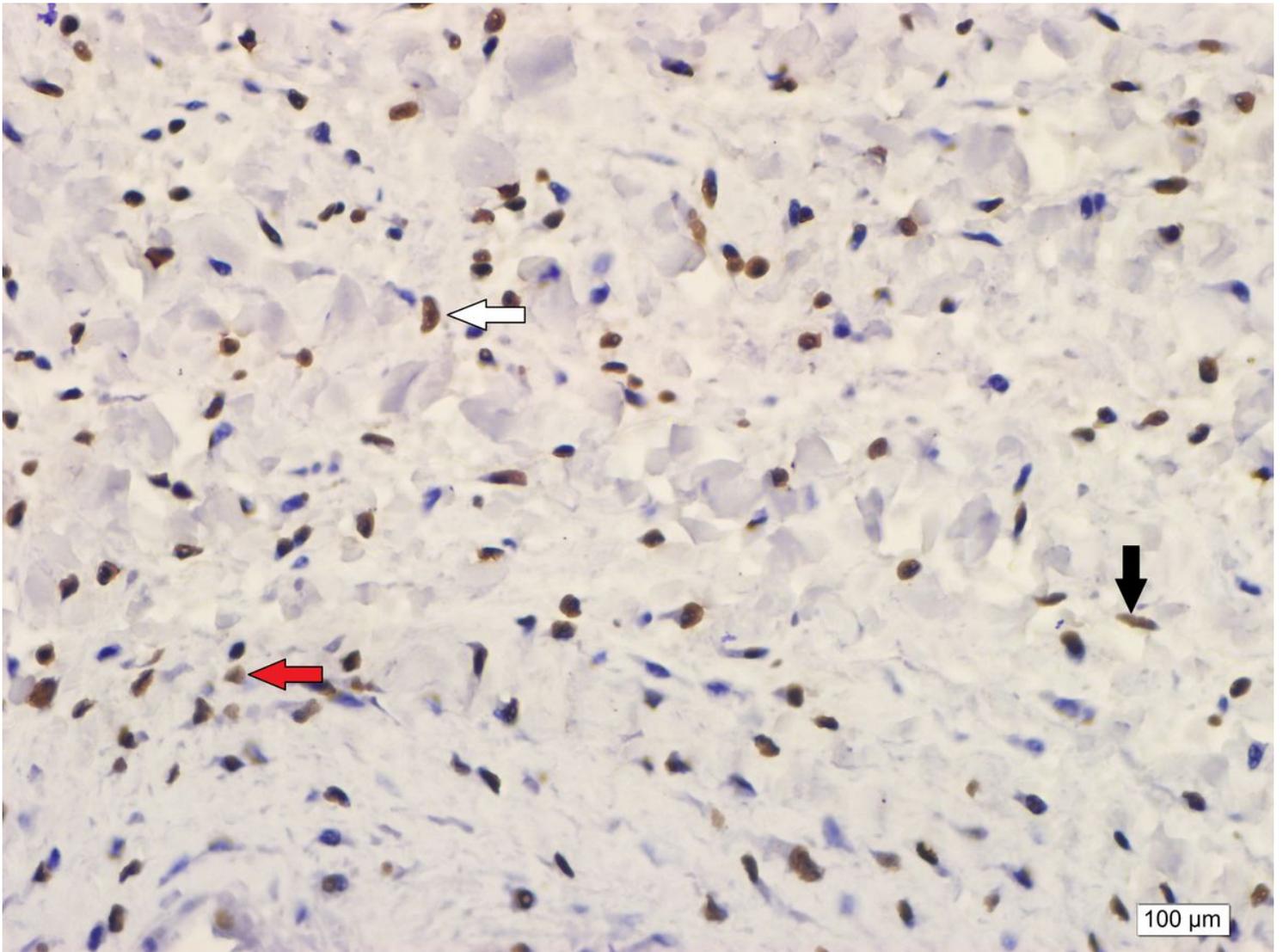


Figure 3

Examples of staining intensity of relaxin. White arrow, example of a 3+ result; black arrow, 2+; and red arrow, 1+ stained cells (Hematoxylin & eosin; X200)