

# Spectral Characterization, Antioxidant And Anti-Tumor Activity of Kaempferol- 3-O-Alpha-L-Rhamnoside; A Flavonol Glycoside Isolated From The Ethyl Acetate Extract of *Pithecellobium Dulce* Leaves

**Masuma Akter**

University of Rajshahi

**Mahadi Hasan**

University of Rajshahi

**Shahnaj Parvin**

University of Rajshahi

**Aziz Abdur Rahman**

University of Rajshahi

**Ekramul Islam** (✉ [ekram@ru.ac.bd](mailto:ekram@ru.ac.bd))

University of Rajshahi

---

## Research Article

**Keywords:** Antioxidant, Anti-tumor, NMR analysis, AAPH, EAC cell

**Posted Date:** June 2nd, 2021

**DOI:** <https://doi.org/10.21203/rs.3.rs-536879/v1>

**License:**   This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

---

# Abstract

**Background:** *Pithecellobium dulce* (Roxb.), an evergreen medium-sized, spiny tree which have vast nutritional values and widely used in ayurvedic medicines and home remedies. The plant has also been a rich source of biologically active compounds. The present study was designed to isolate pure compound and to know the efficacy as antioxidant as well as its anti-tumor activity on Ehrlich ascites carcinoma cell (EAC).

**Methods:** The isolation of the compound was carried out by column chromatography and structure was revealed by  $^1\text{H-NMR}$  and  $^{13}\text{C NMR}$ . The antioxidant activity was investigated by the scavenging of 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radicals as well as the inhibition of oxidative damage of pUC19 plasmid DNA, hemolysis and lipid peroxidation induced by 2, 2'-azobis (2-amidinopropane) dihydrochloride (AAPH) in human erythrocytes. *In vivo* anti-tumor activity of the compound was also evaluated by determining the viable tumor cell count, hematological profiles of experimental mice along with observing morphological changes of EAC cells by fluorescence microscope.

**Results:** The isolated compound showed strong antioxidant activity in DPPH radical scavenging with  $\text{IC}_{50}$  of  $14.6\mu\text{g/ml}$ . It also effectively inhibited AAPH induced oxidation in DNA and human erythrocyte model and lipid per oxidation. In anti-tumor assay,  $70.89\pm 6.62\%$  growth of inhibition of EAC was observed as compare to the control mice ( $p < 0.05$ ) at a dose  $50\text{mg/kg}$  body weight.

**Conclusion:** The compound may have a great importance as a therapeutic agent in preventing oxidative damage of biomolecules and therapeutic use in chemotherapy.

## Background

Wide variety of chemical compounds synthesized by plants may have important biological functions with defend against attack from predators such as insects, fungi and herbivorous mammals. Many of these phytochemicals have beneficial effects on long-term health when consumed by humans and can be used to effectively treat human diseases. It is well known that the chemical compounds of plants affect the human body by the same process as the chemical compounds of conventional medicine thus herbal medicines do not differ greatly from conventional drugs in terms of how they work. This enables herbal medicines as effective as conventional medicines, but they are less likely to cause harmful side effects [1]. Various studies have reported that medicinal plants are the sources of many nutrient and non-nutrient molecules that could have antioxidant, anti-inflammatory, and antimicrobial activities [2].

Oxidative stress caused by free radical and reactive oxygen species is associated with various diseases. A number of studies have been conducted worldwide to find natural antioxidants in plant sources. Plants containing phenolic and flavonoid compounds have been shown to have strong antioxidant activity. Natural plant-derived products such as flavonoids, terpenes, and alkaloids have received considerable attention in recent years due to their various pharmacological properties, including cytotoxic and cancer chemo protective effects [3].

About 50% of the drugs used for clinical trials were isolated from natural sources. DNA is present in every cells of body and human DNA remains continuously in exposure to free radicals attack that causes damage to DNA. Researchers all over the world are working on the aspect of relating the changes in DNA with evolutionary development [4–6]. Currently, there has been a growing interest in identifying free radical scavengers or antioxidants that prevent DNA from oxidative damage [7]. Another molecule, erythrocytes are considered the main target of free radical attack due to the high membrane concentration of polyunsaturated fatty acids [8]. Therefore, due to their sensitivity to oxidation, erythrocytes have been used as a cellular model to investigate oxidative damage in bio membranes.

The plant *Pithecellobium dulce* belongs to the fabaceae family and widely distributed in India, Huawei, and tropical Africa and especially along the coast. It has a long history of use in traditional system of medicine. The compound that was isolated from the leaves is a flavonoid glycosides inhibits lipid peroxidation and cyclooxygenase (COX)-1 and COX-2. Several recent studies have indicated that the compound Kaempferol-3-O-alpha-L-rhamnoside inhibits the growth of breast cancer cells by stimulating apoptosis and that it is relatively non-toxic to normal cells [9]. Ehrlich ascites carcinoma (EAC) is a rapidly growing experimental tumor with very aggressive behavior and resembles human tumors [10]. The potent inhibitory effects of many isolated compounds and crude extracts of different medicinal plants against EAC have also been reported in literature [11]. However, the effects of this compound on the AAPH induced oxidative damage of erythrocytes and pUC19 DNA and also anti-tumor activity against EAC cell has not been investigated.

In this study, we aimed to investigate the effect of kaempferol-3-O-alpha-L-rhamnoside on antioxidant by employing various test models like DPPH scavenging, DNA and erythrocytes damage protection and to check the anti-tumor activity of this compound against EAC cell.

## Methods

### Collection of plant materials

The plant material was collected by following the ethics standard for research activity on plants established at Department of Pharmacy, University of Rajshahi, Bangladesh. Fresh samples of *P. dulce* leaves were collected from the relevant area of Kumarkhali, Kushtia during the months of November and December and were authenticated by a taxonomist, Prof. AHM Mahbubur Rahman, Dept. of Botany, University of Rajshahi, Bangladesh and given Accession no. 48085. A sample specimen was preserved in National herbarium Dhaka, Bangladesh.

### Ethics of experimentation

The protocol for using human blood cells was approved by the Institutional Animal, Medical Ethics, Bio-Safety and Bio-Security Committee (IAMEBBC) for Experimentations on Animal, Human, Microbes, and Living Natural Sources at University of Rajshahi (Approval memo no-82/320/IAMEBBC/IBSc, 20 August, 2018). All procedures of this study adhere to the ARRIVE Guidelines for reporting animal research.

## Chemicals and reagents

2,2'-diphenyl-1-picrylhydrazyl (DPPH<sup>•</sup>), 2,2'-Azobis (2- amidinopropane) dihydrochloride(AAPH), catechin, 4, 6-diamidino-2-phenylindole; DAPI (Sigma-Aldrich), Dimethyl sulfoxide; DMSO (E-Merck, Germany), Trypan blue (Sigma, India), Phosphate buffer saline (PBS), pUC19 DNA (Thermo scientific) and Ethidium bromide (Etbr).

## Extraction and fractionation

The collected healthy leaves were washed with tap water and then with distilled water and were spread out and dried at room temperature for about 15 – 20 days and pulverized by mechanical grinder. 1.5kg of *P. dulce* leaves powder was first extracted with n-hexane. The residue after n-hexane extraction soaked in 3.5 liter of methanol and kept it for 12 days with occasional shaking and stirring. The whole mixture was then filtered through cotton and then through Whatmann No.1 filters paper and was concentrated with a rotary evaporator under reduced pressure at 45°C to collect the crude methanolic extract (CME). Then sequentially fractionated towards polarity using the solvent n-Hexane, chloroform and ethyl acetate. The eluents of the four different polarity solvents were collected separately and concentrated by rotary evaporation under vacuum.

## Isolation and characterization of the compound

*In vitro* biological assay, EAF had potent activity than other fraction and TLC assay also showed some distinct and prominent spot. Thus, later purification and isolation of active constituents were focused on EAF. This fraction was subjected a silica gel 60 column chromatography using n-hexane: ethyl acetate: methanol (1:2:1) and then ethyl acetate:n-hexane (95:5) as the eluent. Fractions 152 to 170 was combined due to similar spot on TLC plate. Then fraction 152 to 170 was further purified by preparative thin layer chromatography (PTLC) to isolate the target compound (1.2 g, R<sub>f</sub> = 0.65). The compound was dissolved either in methanol and ethyl acetate depending on its solubility for analysis. The structure of the isolated compound was elucidated from the data obtained from <sup>1</sup>H- and <sup>13</sup>C-NMR spectra and by comparing with those reported in the literature [12]. The spectra of the pure compound were recorded on the Jeol-Ex at 400 MHz and 100 MHz and on FT NMR spectrometers using methanol as solvent.

## Antioxidant activity

### DPPH radical scavenging activity

The antioxidant activity of kaempferol-3-O-alpha-L-rhamnoside at various concentrations (3.125µg/ml to 50µg/ml) was evaluated via the DPPH radical scavenging system using BHT as standard. Briefly, 3ml of 0.04% DPPH in methanol was mixed with the compound solution, incubated in dark for 30 min and finally absorbance was measured at 517nm [13]. The experiment was repeated in triplicate manner and mean absorption was taken to calculate percentage of DPPH radical scavenging activity as follows-

$$\% \text{ of Scavenging} = [(A_0 - A_1) / A_0] \times 100$$

Where,  $A_0$ -absorbance of blank,

$A_1$ - absorbance of extract

$IC_{50}$  (conc. in  $\mu\text{g/ml}$ ) value required for 50% scavenging of DPPH was calculated from the graph plotted for the % of scavenging against concentration of the sample.

### **Antioxidant assay in human erythrocytes**

#### **Inhibition assay of AAPH-induced hemolysis**

Protective effect of the compound against oxidative damage of erythrocytes induced by AAPH was determined by the method described by Ribeiro et al [14]. Here, hemolysis was performed with AAPH as free radical initiator. An aliquot of 200  $\mu\text{l}$  of 5% (v/v) erythrocytes suspension in PBS was mixed with 50  $\mu\text{l}$  of compound with different concentrations (10, 25 and 50 $\mu\text{g/ml}$  in PBS pH 7.4). Standard ascorbic acid was used at concentration of 50 $\mu\text{g/ml}$ . To this, 0.5 ml of 50mM AAPH (dissolved in PBS) was added. The reaction mixture was incubated at 37 $^{\circ}\text{C}$  for 6h in time dependent hemolysis. Each 1h interval sample was withdrawn, diluted with 2 ml PBS and centrifuged the mixture at 4000rpm for 10min. The supernatant portion was collected and absorbance was taken at 540nm. Reference values were determined using the same volume of erythrocytes in a hypotonic buffer (100% hemolysis) and kept under same condition as sample. The hemolysis percentage was calculated using the formula

$$\% \text{ of Hemolysis} = (\text{Absorbance of sample supernatant} / \text{Reference value}) \times 100$$

#### **Measurement of lipid peroxidation of erythrocytes**

The effect of the compound kaempferol-3-O-alpha-L-rhamnoside on the inhibition of the production of malondialdehyde (MDA), a by-product of lipid peroxidation, was evaluated by incubation of a human erythrocyte suspension (5%) in 50 mM of the oxidizing agent AAPH [15]. Briefly a solution of three different concentration of the compound (10, 25 and 50 $\mu\text{g/ml}$ ) was mixed separately with 0.5 ml of 50mM AAPH (dissolved in PBS). The reaction mixture was kept at 37 $^{\circ}\text{C}$  for 6h for time dependent hemolysis. Each 2h interval one set of sample was withdrawn and 300 $\mu\text{l}$   $\text{H}_3\text{PO}_4$  (0.44M), 750 $\mu\text{l}$  thiobarbituric acid (0.67%) were added to 1ml reaction mixture and incubated at 95 $^{\circ}\text{C}$  for 1h. This was then cooled in an ice bath for 10min and 450 $\mu\text{l}$  trichloroacetic acid (20%) was added. The mixtures were centrifuged at 4000 rpm for 10min. The supernatant portion was collected and absorbance was taken at 532nm. A control solution of buffer with same volume of erythrocytes was kept under same condition as sample. MDA level was determined using molar extinction coefficient is 156 $\text{mM}^{-1} \text{ cm}^{-1}$  at Beer Lambert Law and the value was expressed as pmol/g Hb.

$$\text{MDA level} = \left( \frac{\text{Absorbance of sample}}{\text{extinction coefficient}} \right) \times \text{Dilution factor}$$

#### **Antioxidant assay on plasmid DNA**

Protective effect on oxidative DNA damage Protective effect of the pure compound on DNA (pUC19) damage induced by AAPH was performed according to the Zhou *et al* [16]. The compound at the concentration (5, 10, 20, 40µg/ml) from a stock solution of 2mg/ml and 1µg DNA (2µl) were taken in eppendorff tubes after that 10µl AAPH was added to each mixture. The reaction mixture was adjusted to the total volume of 40µl by PBS and allowed to incubate for 30min at 37<sup>0</sup>C. After 30min incubation, 6X loading dye (1:1) were added. The reaction mixtures (10µl) were electrophoresed on 1 % agarose in 1X TAE buffer at 110 V for 40min. The gel was stained with EtBr solution for 30-45min, viewed under trans-illuminating UV light and photographed using gel documentation system.

## **Anti-tumor activity**

### **Animals**

Swiss albino mice of either sex, 3–4 weeks of age, weighting between 20 and 25 g were collected from the Jahangir Nagar University, Dhaka, Bangladesh. They were maintained under standard laboratory conditions (temperature 22-28 °C, humidity 55–3%) with 12h day-night cycles and also provided with standard dry pellet diet and water.

### **Experimental tumor model**

Transplantable tumor (Ehrlich's ascites carcinoma) used in this thesis were obtained from Department of Biochemistry, University of Rajshahi and were maintained in our laboratory in *Swiss albino* mice by intra-peritoneal transplantation.

### **Determination of median lethal dose (LD<sub>50</sub>)**

The median lethal dose (LD<sub>50</sub>) value was determined by injecting the solution of the compound intraperitoneally in mice at various doses (5, 10, 25, 50, 100, 200mg/kg) and mortality at the end of the 24 h experiment was recorded. Doses were selected for this anti-tumor study by fixed dose methods and were 25 and 50mg/kg [17].

### **Studies on *in vivo* EAC cell growth**

The pure compound kaempferol-3-O-alpha-L-rhamnoside at doses of 25 and 50mg/kg per day were given to every mouse of group-II and III, respectively. Group –IV was treated with anticancer drug vincristine. Treatment was continued for 6 days and on seventh day after EAC cell inoculation, animals were sacrificed. EAC cells were collected by repeated washing with 0.9% saline and viable EAC cells per mouse of the treated groups were compared with untreated control [17]. Here, vincristine was used as standard at a dose of 6.25mg/kg/day. Vincristine is a chemotherapeutic drug that is poisonous to other cells. When it is introduced into cells it binds to the cancer cell DNA which stops the cell division.

### **Morphological appearance of EAC cell**

Morphological changes of EAC cells were examined by DAPI (4, 6-diamidino-2-phenylindole) staining after collecting the cells from non-treated EAC-bearing mice and mice treated with kaempferol-3-O-alpha-L-rhamnoside (25 and 50mg/kg/day) for 7 days. Then visual images were taken using fluorescent microscope. Both fluorescent and optical views were observed.

### Studies on hematological parameters

To assess the haematological parameters, Swiss Albinomice were divided into four groups (n = 5). All the animals were injected with EAC cells (0.1 ml of  $1.6 \times 10^6$  cells/mouse) intraperitoneally except the normal group at the day zero. Group 1 served as the normal control, group II served as the untreated EAC control and group III served as the standard vincristine. Group IV and V were treated with compound kaempferol-3-O-alpha-L-rhamnoside at 25 and 50 mg/kg doses, respectively. On the 12th day, after tumor EAC cell inoculation, hematological parameters (Hemoglobin, RBC and WBC) were measured from freely flowing tail vein blood of each mice of each group [18-19].

### Statistical analysis

The data were analyzed by one-way ANOVA (analysis of variance) followed by multiple comparisons using Dunnett's post hoc and LSD test using SPSS software of 20version. All results were represented as mean  $\pm$  standard deviation (SD). Differences at  $p < 0.05$  level were considered to be statistically significant.

## Results

### Spectral characteristic

$^1\text{H-NMR}$  and  $^{13}\text{C-NMR}$  spectrum of the compound (Sz-02) are shown in **Fig. 1** and **Fig. 2**. The  $^1\text{H NMR}$  spectrum (MeOD, 400 MHz) of the compound revealed two signals at  $\delta$  6.19 (1H, d,  $J = 1.6$  Hz) and  $\delta$  6.36 (1H, d,  $J = 1.6$  Hz). Those are well known H-6, H-8 characteristic signals of a flavonoid glycoside [20]. Another two signals at  $\delta$  7.76 (2H, d,  $J = 8.4$  Hz) 6.93 (2H, d,  $J = 8.4$  Hz) are characteristic signals of H-3' & 5', H-2' & 6' in kaempferol-3-O-alpha-L-rhamnoside [20]. The signals at  $\delta$  5.36 (d, Overlapped),  $\delta$  0.91 (overlapped), 4.21, 3.69 are characteristic signals of 3-O-rhamnose. Presence of rhamnose was further confirmed by the anomeric carbon at  $\delta$  105.9. From the above mentioned data and by comparison with the literature, the compound (Sz-02) was identified as kaempferol-3-O-alpha-L-rhamnoside and the chemical structure of the compound are shown in **Fig. 3**.

On the basis of the  $^1\text{H NMR}$  spectral features (**Fig. 1**) as well as  $^{13}\text{C NMR}$  (**Fig. 2**) data, and comparison with the known kaempferol-3-O-alpha-L-rhamnoside [20], the compound was identified as kaempferol-3-O-alpha-L-rhamnoside.

### Antioxidant activity

#### DPPH radical scavenging activity

The antioxidant activity of the compound evaluated by DPPH radical scavenging is shown **Fig. 4**. The dose response curves of DPPH radical scavenging activities indicated that at lower concentration (3.125µg/ml) the scavenging activity of the compound was 19.40±0.55 % compare to standard BHT with 31.96±1.04% scavenging at the same concentration. At higher concentration (50µg/ml) it was 88.42±0.49% which was very near to BHT (93.20±0.36%). The compound exhibits IC<sub>50</sub> value of 14.6µg/ml whereas for BHT it was 1.45µg/ml.

### **Antioxidant assay on human erythrocytes**

#### **Inhibition assay of AAPH-induced hemolysis**

Oxidation of erythrocyte membrane by peroxy radicals generated by AAPH may result in the release of hemoglobin pigment into the medium. The color measurement mediated by hemoglobin at 410 nm serve the level of oxidative damage in cells. **Fig. 5** showed the effect of kaempferol-3-O-alpha-L-rhamnoside against peroxy radical-induced hemolysis in erythrocytes. When erythrocytes were incubated with AAPH in air at 37<sup>0</sup>C there was progressive hemolysis after 2 h of incubation and it was 96.73±6.56% after 6h of incubation. But this hemolytic process was inhibited in the presence of kaempferol-3-O-alpha-L-rhamnoside in a concentration and time dependent manner. After 6h the percentage of hemolysis was (46.31±2.16%) at maximum tested concentration (50µg/ml) of the compound whereas 65.44±1.37% haemolysis was observed with ascorbic acid at concentration of 50µg/ml.

#### **Inhibition of malondialdehyde production**

AAPH can also cause an increase in lipid peroxidation of erythrocytes. This was evaluated by malondialdehyde (MDA) formation and the results shown in **Fig. 6** indicated that AAPH could cause a significant increase in MDA from a range of 8.71± 0.34 to 19.90± 0.57nmol/g Hb. But preincubation of erythrocytes with the compound at different concentrations had no significant effect on MDA production. In this case the amount of MDA formation was measured as incubation for 2 to 6 hours, ranging from 2.62 ± 0.18 to 10.15 ± 0.22 nml/gHb at maximum concentration of the compound. In case of standard ascorbic acid MDA formation was from 2.08±0.22 to 7.29±0.26 nml/gHb.

#### **Antioxidant assay on plasmid DNA**

The ability of kaempferol-3-O-alpha-L-rhamnoside to protect oxidative damage of DNA was evaluated by analyzing the band pattern of pUC19 DNA on agarose gel as shown in **Fig. 7**. Lane-1 shows DNA in the native supercoil form, whereas lane-2 treated with AAPH, the super coil form has been converted in open circular DNA. Addition of the compound in lane 3, 4, 5, 6 and 7 at concentration of 5, 10, 20, 40 and 50µg/ml, respectively prevent the formation of circular form of plasmid DNA in a dose dependent manner. The compound showed the same effect at 40µg / ml as the standard gallic acid at 50µg / ml

#### **Anti-tumor activity**

#### **Studies on EAC cell growth inhibition**

*In-vivo* anti-tumor activity of kaempferol-3-O-alpha-L-rhamnoside against EAC cell bearing mice was assessed by the parameters such as viable EAC cell (% inhibition in cell growth). The average number of viable tumor cells per mouse of untreated EAC control group was found  $(7.51 \pm 0.83) \times 10^7$  cells/ml. Effects of the compound on EAC cells growth after tumor inoculation are shown in **Table 1**. Treatment with kaempferol-3-O-alpha-L-rhamnoside resulted in significant reduction of cell growth *in-vivo*. The percentage of cell growth inhibition of this compound was  $37.17 \pm 7.94\%$  and  $70.89 \pm 6.62\%$  at doses of 25 and 50mg/kg, respectively, whereas standard drug vincristine showed  $77.84 \pm 6.69\%$ .

Table 1: Effect of kaempferol-3-O-alpha-L-rhamnoside on viable Ehrlich ascites carcinoma (EAC) cell growth

Group	Treatment	Viable EAC cells on day 7 after inoculation ( $\times 10^7$ cells/ml)	Percentage (%) cell growth inhibition
1	EAC Cell	$7.51 \pm 0.83$	
2	EAC + Sz-02 (50 mg/kg)	$2.16 \pm 0.38^*$	$70.89 \pm 6.62^*$
3	EAC + Sz-02 (25mg/kg)	$4.674 \pm 0.28^*$	$37.17 \pm 7.94^*$
4	Vincristine (6.25 mg/kg)	$1.64 \pm 0.51^*$	$77.84 \pm 6.89^*$

Data are expressed as mean  $\pm$  SD (n = 5); Analysis of variance followed by LSD and Dunnett't post hoc test (IBM-SPSS/20) \*P < 0.05: Significance difference with respect to EAC control

### Morphological changes of EAC cell

Morphological changes of EAC cells were examined by DAPI staining after collecting the cells from non-treated EAC-bearing mice and mice treated with kaempferol-3-O-alpha-L-rhamnoside (25 and 50mg/kg/day) after 7 days. EAC cells nuclei were round, regular, and homogeneously stained with DAPI in control group (solvent treated) as shown in **Fig. 8**. Apoptotic morphologic alterations such as membrane and nuclear condensation were noted in Kaempferol-3-O-alpha-L-rhamnoside treated EAC cells. These results indicated treatment with this compound could induce apoptosis in EAC cells.

### Studies on hematological parameters

Hematological parameters of untreated EAC cell bearing mice on the day 12 were showed significant ( $P < 0.05$ ) changes when compared to normal mice (**Table 2**). The total WBC count was found to increase with a reduction in the hemoglobin content and total RBC count. At the same time interval, treatment of compound (25 and 50 mg/kg) could brought back these altered parameters to normal values. The overall results of this study clearly demonstrated the anti-tumor activity of the compound against EAC.

Table 2: Effect of kaempferol-3-O-alpha-L-rhamnoside on blood parameters of tumor bearing and normal mice

Parameters	Normal	Control	EAC+ Kaempferol (50mg/kg)	EAC+ Kaempferol (25mg/kg)	EAC+ Vincristine (6.25mg/kg)
Hgb (g/dl)	13.89±0.08	9.20±0.26*	12.99±0.33 <sup>#</sup>	10.24±0.48 <sup>#</sup>	13.52±0.28 <sup>#</sup>
RBC (×10 <sup>9</sup> cells/ml)	6.34±0.32	1.85±0.15*	3.35±0.165 <sup>#</sup>	3.2±0.118 <sup>#</sup>	6.05±0.52 <sup>#</sup>
WBC (×10 <sup>6</sup> cells/ml)	8.23±0.135	121.60±1.81*	28.80±0.72 <sup>#</sup>	51.47±0.59 <sup>#</sup>	17.87±0.48 <sup>#</sup>

Data are expressed as mean ± SD for five animals in each group. Analysis of variance followed by LSD and Dunnett't post hoc test (IBM-SPSS/20)

\*P<0.05: against normal group and <sup>#</sup>P<0.05: against EAC control group

## Discussion

*Pithecellobium dulce* is used as astringent in dysentery, abortifacient, antidiabetic, anticonvulsant, antiulcer, larvicide dermatitis, eye inflammation, indigestion, intestinal disorder, ear ache, leprosy and tooth ache. Phytochemical investigation of bark and leaves had revealed the presence of  $\beta$ -sitosterol, saponin glycosides, oleanolic and echinocystic acids as sapogenins, echinocystic acid, bisdesmoside, dulcin, triterpenoids, acylated triterpenoid saponin, flavanoids, saccharides, long chain aliphatic hydrocarbons, and tannins [21–22]. In our study, kaempferol-3-O-alpha-L-rhamnoside; a flavonoid that was isolated from leaves extract of *P. dulce*. To know its biological activity, we determined *in vitro* free radical scavenging and protection of oxidative damage on DNA and human erythrocytes as well as anti-tumor activity on EAC cells. The results of these experiments showed promising results. Antioxidants fight against free radicals and protect us from various ailments. Isolation and identification of many natural antioxidants from different plant materials have been studied. DPPH radical is one of the most common and stable chromogens used to estimate antioxidant activity of biological materials in a relatively short time. In our study, kaempferol-3-O-alpha-L-rhamnoside had potent DPPH radical scavenging activity and was higher than previously reported leaves extract of this plant [23]. For example, 28.9 and 83.2% DPPH inhibition for methanol and acetone (70%) of *P. dulce* leaves were reported at 100  $\mu$ g/ml concentrations. But our compound showed 88.42 ± 0.49% DPPH scavenging activity at concentration of 50 $\mu$ g/ml and is more potent and efficacious than those crude extract of leaves.

For a better understanding of the antioxidant activity of the compound, we used a human erythrocyte to measure hemoglobin oxidation and lipid peroxidation inhibition. Free radicals attacking of erythrocyte

membrane components (proteins and lipids) cause the alteration of membrane structure and function leading to hemolysis. The peroxy radicals generated from AAPH are capable of inducing lipid peroxidation and protein damage [23]. In the present study, the incubation of erythrocyte together with AAPH led to remarkable hemolysis that was consistent with previous findings [24]. The present results clearly show that the pure compound has high capacity to prevent the oxidative damage induced in the erythrocyte membrane and to minimize the lipid peroxidation. Lipid peroxidation from AAPH induced radical results in the production of MDA which is responsible for cellular damage and associated with many of pathological events [25]. Since the compound showed ability to protect oxidation of erythrocyte membrane so the inhibition of lipid peroxidation was also expected and we found that the compound could inhibit the malondialdehyde formation in a dose dependent manner. Therefore, the protective effects of erythrocyte membrane and inhibition of MDA production of this pure compound may result from diminishing peroxy radicals generated from AAPH during the incubation period as it was also effective in scavenging of DPPH free radicals.

Cellular DNA damage may cause alteration of replication and transcription causing cell death or mutations. Such modification of DNA is responsible for aging and various diseases, including Alzheimer, Parkinson and cancer etc. So, study on DNA damage protection is important. Oxidative DNA protective activities of flavonol type glycoside have been previously reported by other researches [26]. Here we reported the effectiveness of the isolated pure compound from leaves extract to prevent AAPH induced oxidative damage of DNA. AAPH converted the super coiled DNA strand into open circular. The compound could significantly inhibit the formation of the open circular form in a dose dependent manner as compared to standard gallic acid (Fig. 7). This result demonstrated the DNA damage inhibition potential of this compound, therefore, can be used in cancer prevention.

So, next we investigated the role of this compound in cell growth inhibition on EAC bearing Swiss albino mice. The prolongation of the life span & reduction of tumor weight of cancer bearing mice is a very important and reliable criterion for judging the potency of any drug as anticancer agent. The effectiveness of the compound against EAC cell bearing mice has further been verified by monitoring the change in hematological and biological parameters. This study indicates that the number of cell growth decreased and number of apoptotic cells increased significantly at different doses. Previously reported the effect of kaempferol-3-O-alpha-L-rhamnoside on the viability of MCF-7 and HC-04 cells was evaluated. The treatment of cancer (MCF-7) and non-cancer (HC-04) cell lines with kaempferol-3-O-alpha-L-rhamnoside resulted in a dose-dependent inhibition of cell growth, [27] which also support our findings. Anemia may occur during cancer chemotherapy. In our study, reduction in RBC or % in hemoglobin in tumor bearing mice may occur which is mainly due to iron deficiency or hemolytic or myelopathic conditions. Kaempferol-3-O-alpha-L-rhamnoside could significantly recover the hemoglobin content, RBC and WBC cell count that indicates the protective action of HM on the haemopoietic system. All these are measured are very important aspects in justifying the effectiveness of a compound in cancer chemotherapy.

## Conclusion

This study is the first to evaluate the antioxidant activity of kaempferol-3-O-alpha-L-rhamnoside present in the leaves of *P. dulce* in a comprehensive manner employing a range of assays. The results of the present study show that the pure compound possesses antioxidant activity evidenced by various methods as well as cell growth inhibition on EAC cells. The study showed the DNA damage inhibition potential of the compound, which could be used in cancer prevention. Further work should be carried out to establish the mechanism of the above activity and to determine its safety.

## Declarations

### Ethics approval and consent to participate

The protocol for using human blood cells and experimental mice was approved by the Institutional Animal, Medical Ethics, Bio-Safety and Bio-Security Committee (IAMEBBC) for Experimentations on Animal, Human, Microbes, and Living Natural Sources at University of Rajshahi (Approval memo no-82/320/IAMEBBC/IBSc, 20 August, 2018) in accordance with the declaration of Helsinki. Blood samples were collected from healthy adults with the informed consent of all participants in the study. All procedures of this study adhere to the ARRIVE Guidelines for reporting animal research.

The plant material was collected by following the ethics standard for research activity on plants established at Department of Pharmacy, University of Rajshahi, Bangladesh and were authenticated by a taxonomist, Prof. AHM Mahbubur Rahman, Dept. of Botany, University of Rajshahi and given Accession no. 48085.

### Consent for publication

The authors of the current work gave consent for publication to Dr. Ekramul Islam.

### Availability of data and materials

All data generated or analysed during this study are included in this published article (and its supplementary information files).

### Competing interests

The authors declare that they have no competing interests.

### Funding

None

### Authors' contributions

El designed and supervised the study the study. MA performed all the experiments. AR and SP analyzed the spectral characteristics of the compound and MH helped in conducting the experiments. All authors read and approved the final draft.

## Acknowledgement

The authors are thankful to department of Biochemistry and Molecular Biology, University of Rajshahi for providing us the EAC cells.

## Authors' information

Department of Pharmacy, Faculty of Science, University of Rajshahi, Rajshahi-6205, Bangladesh.

## References

1. Tapsell, L. C., Hemphill, I., Cobiac, L., Sullivan, D. R., Fenech, M., Patch, C. S., ... Inge, K. E. (2006). Health benefits of herbs and spices: the past, the present, the future.
2. Das, T., Sa, G., Saha, B., & Das, K. (2010). Multifocal signal modulation therapy of cancer: ancient weapon, modern targets. *Molecular and cellular biochemistry*, 336(1), 85–95.
3. Hefferon, K. L. (2018). Crops with improved nutritional content through agricultural biotechnology. In *Plant Micronutrient Use Efficiency* (pp. 279–294). Academic Press.
4. Ma, Q., Zhang, W., & Xiang, Q. Y. (2017). Evolution and developmental genetics of floral display—a review of progress. *Journal of Systematics and Evolution*, 55(6), 487–515.
5. Hall, B. K. (2012). Evolutionary developmental biology (Evo-Devo): Past, present, and future. *Evolution: Education and outreach*, 5(2), 184–193.
6. Sadrzadeh, S. M., Graf, E., Panter, S. S., Hallaway, P. E., & Eaton, J. W. (1984). Hemoglobin. A biologic fenton reagent. *Journal of Biological Chemistry*, 259(23), 14354–14356.
7. Lobo, V., Patil, A., Phatak, A., & Chandra, N. (2010). Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacognosy reviews*, 4(8), 118.
8. Yang, H. L., Chen, S. C., Chang, N. W., Chang, J. M., Lee, M. L., Tsai, P. C., ... Hseu, Y. C. (2006). Protection from oxidative damage using *Bidens pilosa* extracts in normal human erythrocytes. *Food and Chemical Toxicology*, 44(9), 1513–1521.
9. Imran, M., Salehi, B., Sharifi-Rad, J., Aslam Gondal, T., Saeed, F., Imran, A., ... Estevinho, L. M. (2019). Kaempferol: A key emphasis to its anticancer potential. *Molecules*, 24(12), 2277.
10. Kwiecinski, M. R., Felipe, K. B., Schoenfelder, T., de Lemos Wiese, L. P., Rossi, M. H., Gonçalez, E., ... Pedrosa, R. C. (2008). Study of the antitumor potential of *Bidens pilosa* (Asteraceae) used in Brazilian folk medicine. *Journal of ethnopharmacology*, 117(1), 69–75.
11. Nair, S. C., Panikkar, B., Akamanchi, K. G., & Panikkar, K. R. (1991). Inhibitory effects of *Ixora javanica* extract on skin chemical carcinogenesis in mice and its antitumour activity. *Cancer letters*, 60(3), 253–258.

12. Zhang, Y., Wang, D., Yang, L., Zhou, D., & Zhang, J. (2014). Purification and characterization of flavonoids from the leaves of *Zanthoxylum bungeanum* and correlation between their structure and antioxidant activity. *PloS one*, *9*(8), e105725.
13. Morelli, I. (2001). Antioxidant principles from *Bauhinia terapotensis*. *J. Nat. Pro*, *64*, 892895.
14. Ribeiro, A. B., Berto, A., Ribeiro, D., Freitas, M., Chisté, R. C., Visentainer, J. V., & Fernandes, E. (2015). Stem bark and flower extracts of *Vismia cauliflora* are highly effective antioxidants to human blood cells by preventing oxidative burst in neutrophils and oxidative damage in erythrocytes. *Pharmaceutical biology*, *53*(11), 1691–1698.
15. Suboh, S. M., Bילו, Y. Y., & Aburjai, T. A. (2004). Protective effects of selected medicinal plants against protein degradation, lipid peroxidation and deformability loss of oxidatively stressed human erythrocytes. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, *18*(4), 280–284.
16. Zhou, J., Li, P., Cheng, N., Gao, H., Wang, B., Wei, Y., & Cao, W. (2012). Protective effects of buckwheat honey on DNA damage induced by hydroxyl radicals. *Food and Chemical Toxicology*, *50*(8), 2766–2773.
17. Litchfield, J. J., & Wilcoxon, F. (1949). A simplified method of evaluating dose-effect experiments. *Journal of pharmacology and experimental therapeutics*, *96*(2), 99–113.
18. Habib, M. R., Aziz, M. A., & Karim, M. R. (2010). Inhibition of Ehrlich's ascites carcinoma by ethyl acetate extract from the flower of *Calotropis gigantea* L. in mice. *Journal of Applied Biomedicine*, *8*(1), 47–54.
19. Radha, R., Kavimani, S., & Ravichandran, V. (2008). Antitumour activity of methanolic extract of *Plumeria alba* L. leaves against Dalton lymphoma ascites in mice. *International Journal of Health Research*, *1*(2), 79–85.
20. Kim, Y. K., Kim, Y. S., Choi, S. U., & Ryu, S. Y. (2004). Isolation of flavonol rhamnosides from *Loranthus tanakae* and cytotoxic effect of them on human tumor cell lines. *Archives of pharmacal research*, *27*(1), 44–47.
21. Saxena, V. K., & Singhal, M. (1999). Novel prenylated flavonoid from stem of *Pithecellobium dulce*. *Fitoterapia (Milano)*, *70*(1), 98–100.
22. Sukantha, T. A., & Shubashini, K. (2015). Isolation and characterization of secondary metabolites from *Pithecellobium dulce* Benth fruit peel. *Int J Pharm Pharm Sci*, *7*, 199–203.
23. Giri, L., Belwal, T., Bahukhandi, A., Suyal, R., Bhatt, I. D., Rawal, R. S., & Nandi, S. K. (2017). Oxidative DNA damage protective activity and antioxidant potential of *Ashtvarga* species growing in the Indian Himalayan Region. *Industrial Crops and Products*, *102*, 173–179.
24. Wei, Q. Y., Chen, W. F., Zhou, B., Yang, L., & Liu, Z. L. (2006). Inhibition of lipid peroxidation and protein oxidation in rat liver mitochondria by curcumin and its analogues. *Biochimica et Biophysica Acta (BBA)-General Subjects*, *1760*(1), 70–77.
25. Niedernhofer, L. J., Daniels, J. S., Rouzer, C. A., Greene, R. E., & Marnett, L. J. (2003). Malondialdehyde, a product of lipid peroxidation, is mutagenic in human cells. *Journal of Biological Chemistry*,

278(33), 31426–31433.

26. Shim, J. S., Kim, S. D., Kim, T. S., & Kim, K. N. (2005). Biological activities of flavonoid glycosides isolated from *Angelica keiskei*. *Korean Journal of Food Science and Technology*, 37(1), 78–83.
27. Diantini, A., Subarnas, A., Lestari, K., Halimah, E. L. I., Susilawati, Y., Supriyatna, S. & Abdulah, R. (2012). Kaempferol-3-O-rhamnoside isolated from the leaves of *Schima wallichii* Korth. inhibits MCF-7 breast cancer cell proliferation through activation of the caspase cascade pathway. *Oncology Letters*, 3(5), 1069–1072.

## Figures

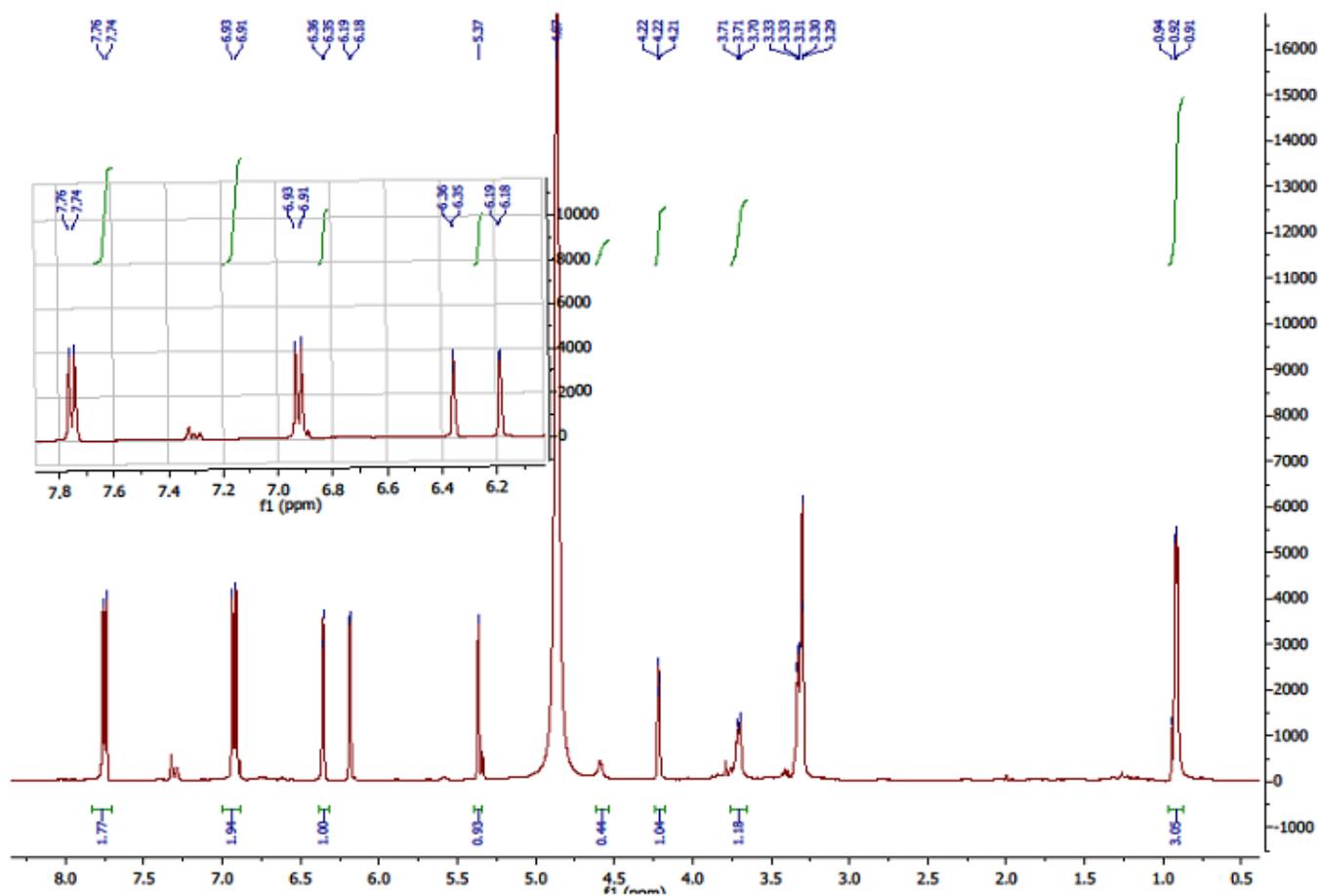


Figure 1

<sup>1</sup>H-NMR data of the isolated compound (Sz-02)

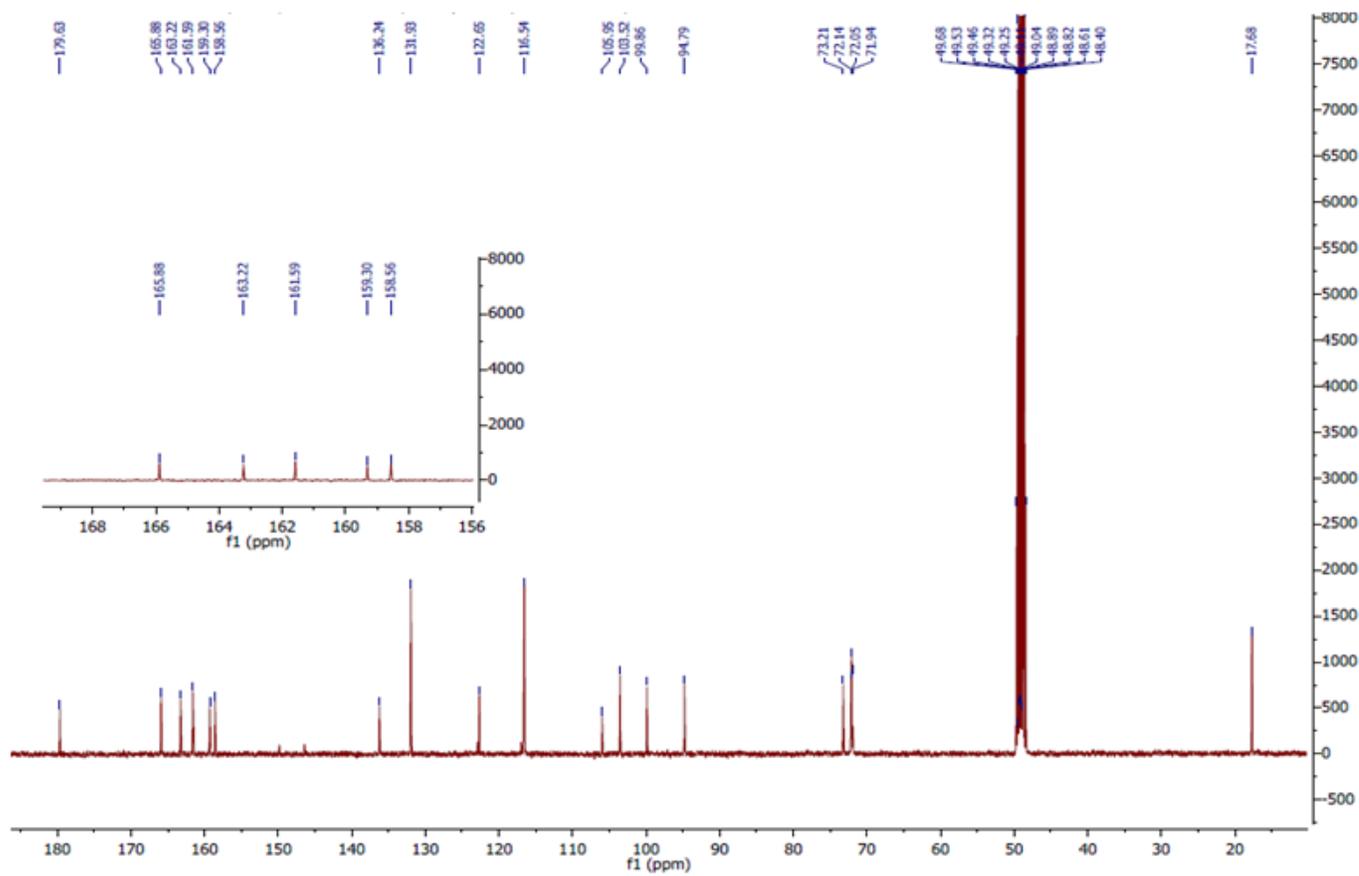


Figure 2

<sup>13</sup>C-NMR data of the isolated compound (Sz-02)

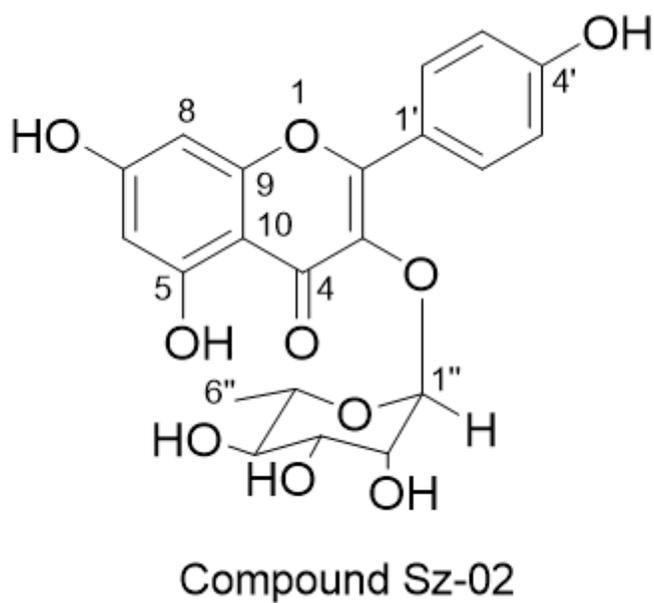
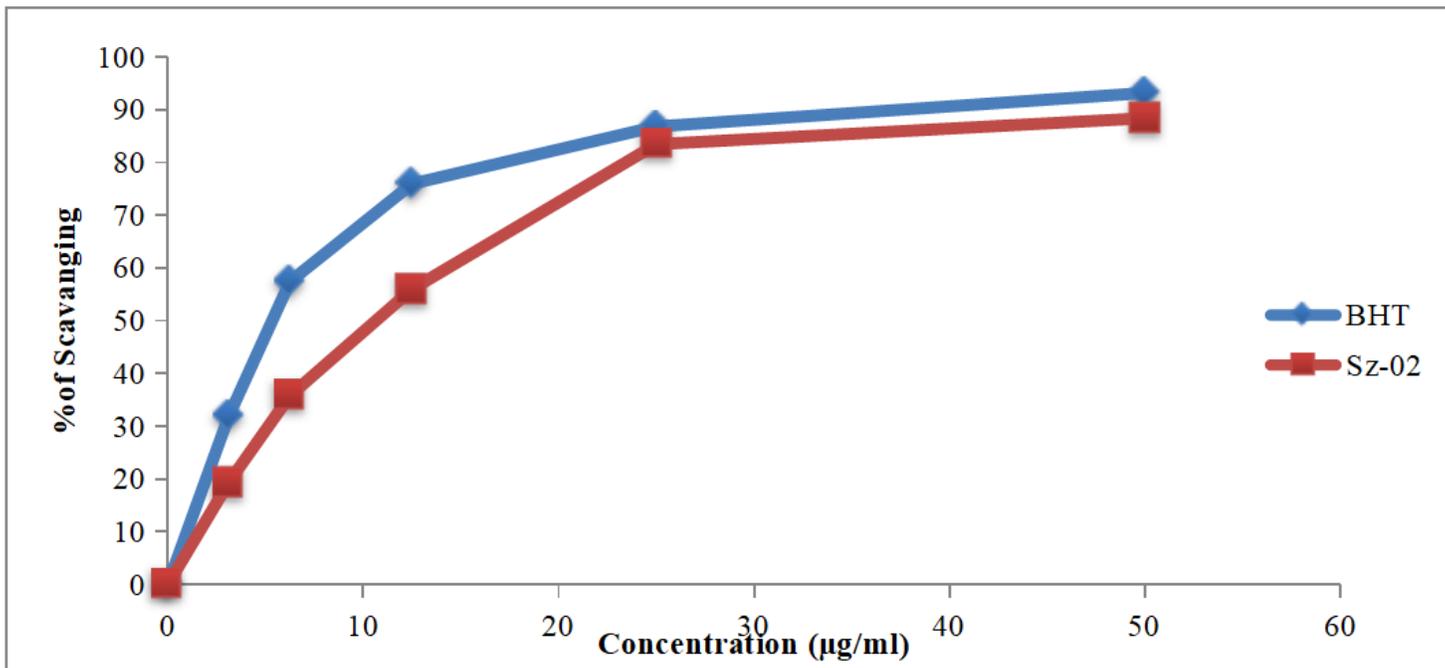


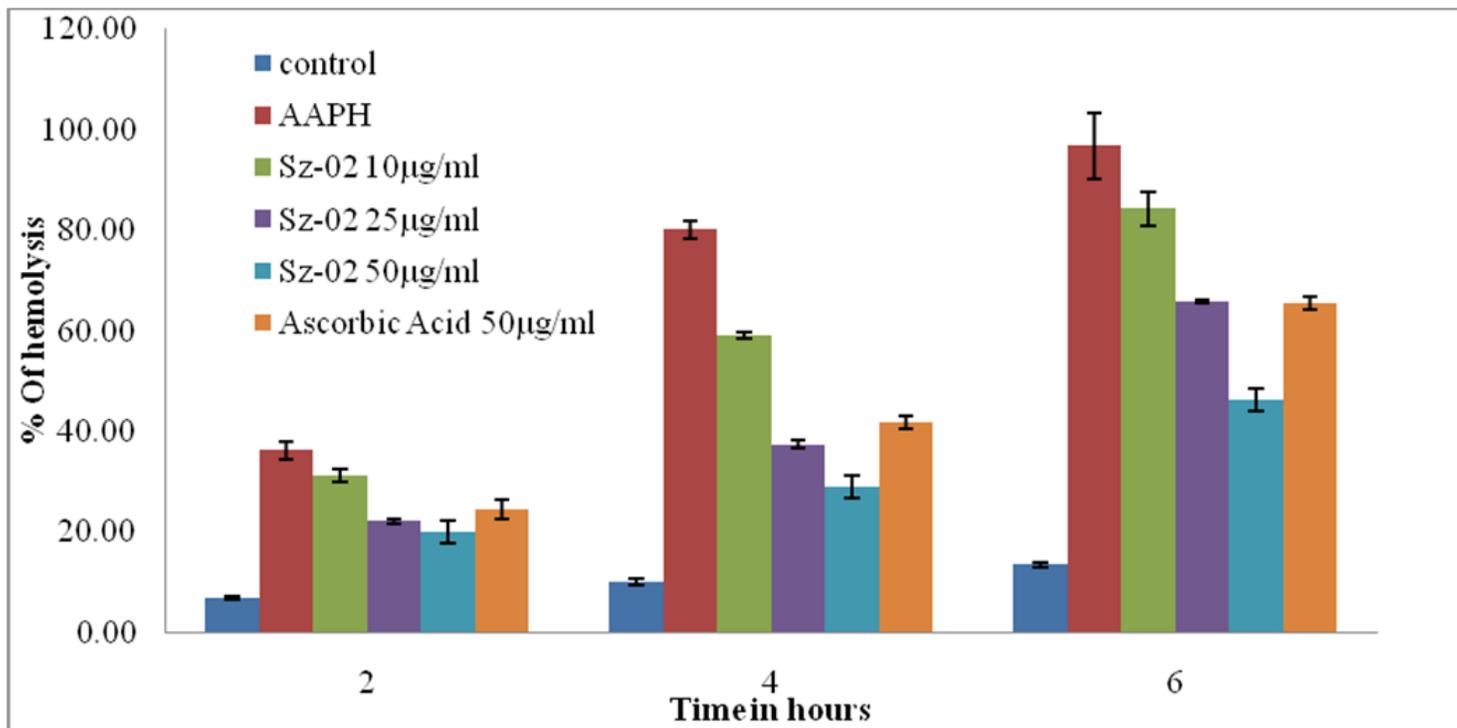
Figure 3

Chemical structure of the compound Sz-02



**Figure 4**

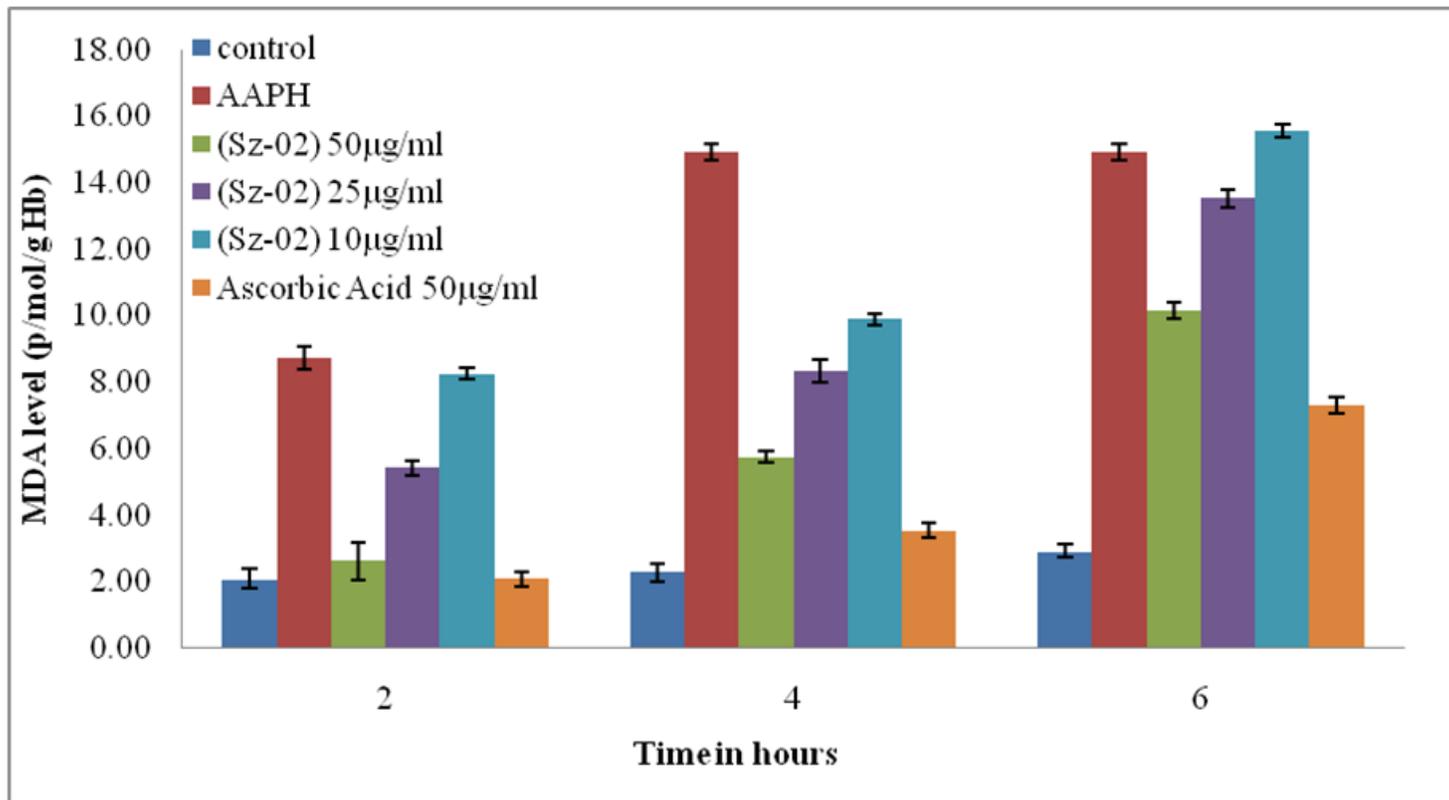
DPPH radical scavenging activity of the pure compound and standard BHT



**Figure 5**

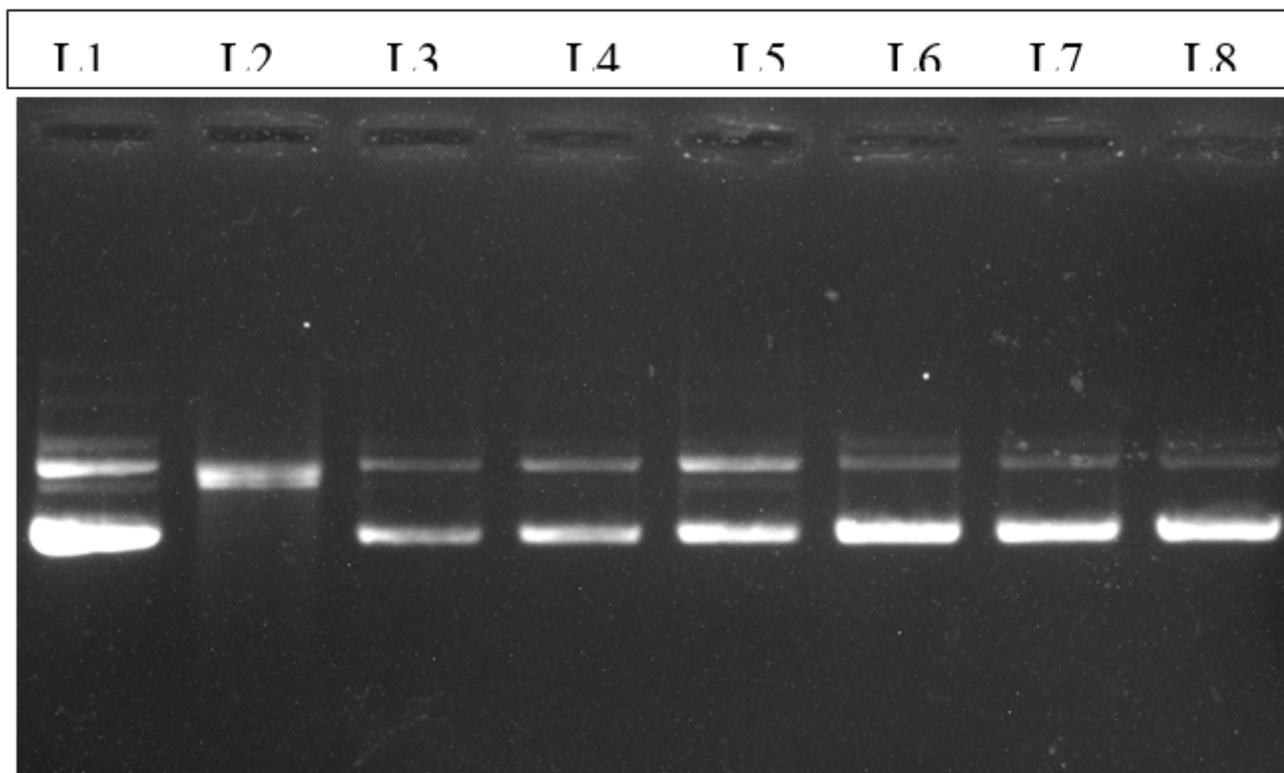
Time course effects of kaempferol-3-O-alpha-L-rhamnoside (Sz-02) and ascorbic acid on AAPH-induced hemolysis on erythrocytes. Erythrocyte suspension at 5% hematocrit was incubated with 50mmol/l AAPH

at 370C in the absence or presence of compound or ascorbic acid at the indicated concentrations. Values are expressed as mean±standard deviation.



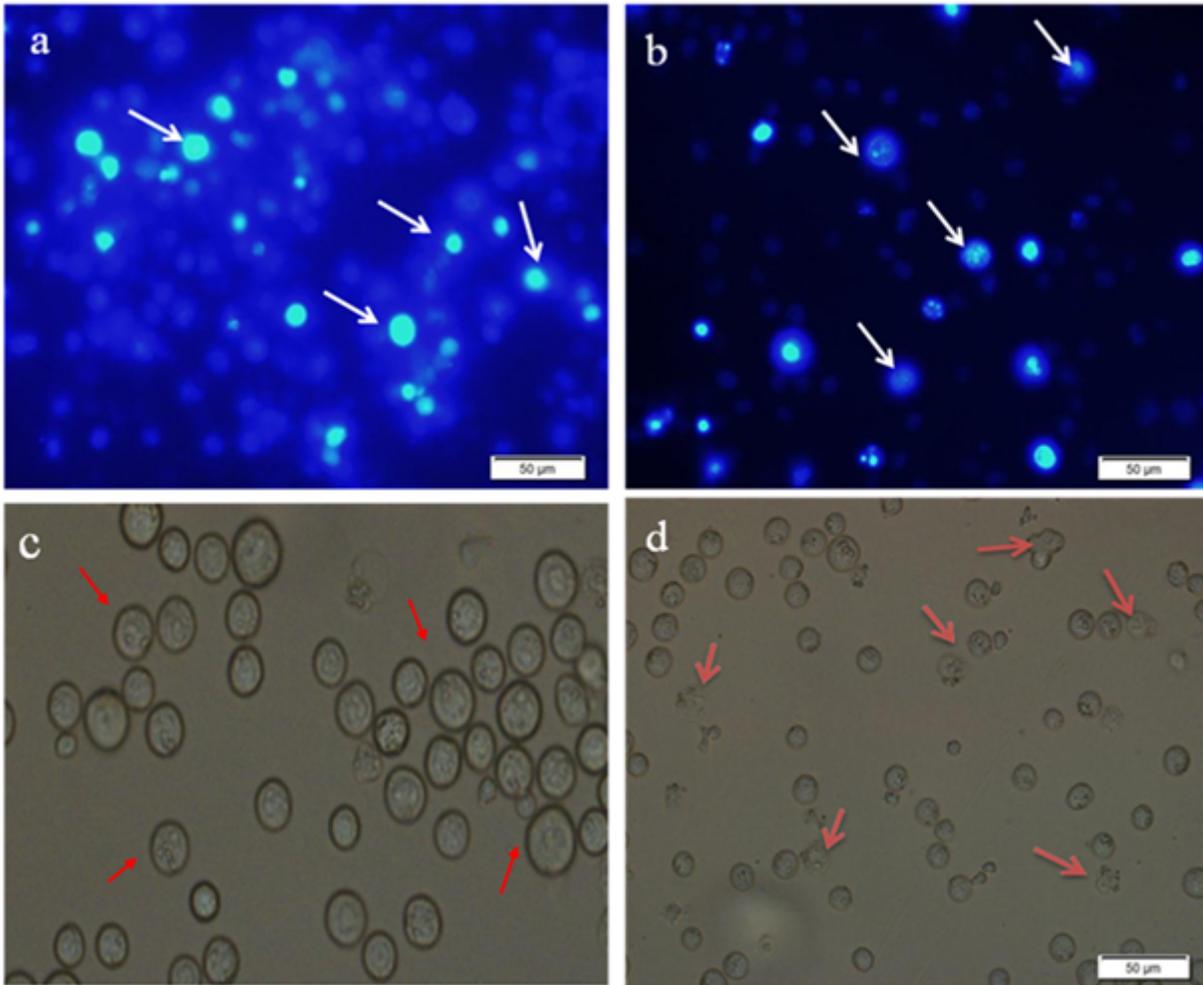
**Figure 6**

Effect of kaempferol-3-O-alpha-L-rhamnoside (Sz-02) and ascorbic acid on MDA formation in erythrocytes. Erythrocyte suspension at 5% hematocrit was incubated with 50 mmol/l AAPH at 370C in the absence or presence of compound or ascorbic acid at the indicated concentrations. Values are expressed as mean ± standard deviation.



**Figure 7**

Agarose electrophoretic pattern of pUC19 plasmid DNA with and without the treatment of pure compound and standard gallic acid. Lane 1: untreated DNA; Lane 2: DNA treated with AAPH; Lane 3: DNA+ AAPH + 5µg compound; Lane 4: DNA + AAPH + 10µg compound; Lane 5: DNA+AAPH+20µg compound; Lane 6: DNA+ AAPH+40µg compound; Lane 7: DNA+ AAPH+50µg compound; Lane 8: DNA+ AAPH+50µg gallic acid.



**Figure 8**

(a-b) Fluorescence and (c-d) optical microscopic observation of EAC cells for control mice and treated mice. (a) and (b) represent as fluorescence microscopic view of control and pure compound treated mice cell where (c) and (d) express the optical microscopic view of control and pure compound treated cells, respectively. Normal cell with round shape nucleuses appeared in control group indicated by white and red arrow in (a) and (c) whereas the mice treated with pure compound, condensed nucleus and fragmentation of cells (apoptotic characteristics) were found (b) and (d) indicated by white and red arrow, respectively.

## Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [S1.docx](#)