

Multi-HLA class II tetramer analyses of citrulline-reactive T cells and early treatment response in rheumatoid arthritis

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Abstract

Background: HLA class II tetramers can be used for *ex vivo* enumeration and phenotypic characterization of antigen-specific CD4+ T cells. They are increasingly applied in settings like allergy, vaccination and autoimmune diseases. Rheumatoid arthritis (RA) is a chronic autoimmune disorder for which many autoantigens have been described. Results: Using multi-parameter flow cytometry, we developed a multi-HLA class II tetramer approach to simultaneously study several antigen specificities in RA patient samples. We focused on previously described citrullinated HLA-DRB1*04:01-restricted T cell epitopes from α -enolase, fibrinogen-b, vimentin as well as cartilage intermediate layer protein (CILP). First, we examined inter-assay variability and the sensitivity of the assay in peripheral blood from healthy donors (n=7). Next, we confirmed the robustness and sensitivity in a cohort of RA patients with repeat blood draws (n=14). We then applied our method in two different settings. We assessed lymphoid tissue from seropositive arthralgia (n=5) and early RA patients (n=5) and could demonstrate autoreactive T cells in individuals at risk of developing RA. Lastly, we studied peripheral blood from early RA patients (n=10) and found that the group of patients achieving minimum disease activity (DAS28 <2.6) at 6 months follow-up displayed a decrease in the frequency of citrulline-specific memory T cells. Conclusions: Our study demonstrates the development of a sensitive tetramer panel allowing simultaneous characterization of antigen-specific T cells in *ex vivo* patient samples including RA 'at risk' subjects. This multi-tetramer approach can be useful for longitudinal immune-monitoring in any disease with known HLA-restriction element and several candidate antigens.

Background

HLA class II tetramers allow direct *ex vivo* detection and phenotypic characterization of antigen-specific T cells [1] and are increasingly used in a range of different settings, like allergy, vaccination and autoimmune diseases [2-6]. This technology requires knowledge of both the antigenic peptide as well as the HLA restriction element, and hereby is utilised in conditions with known antigenic targets and a strong HLA component.

Rheumatoid arthritis (RA) is a chronic inflammatory disease and in a majority characterized by the presence of so called anti-citrullinated protein antibodies (ACPA). It is highly associated with a certain set of HLA-DR alleles [7-9] and a number of citrullinated T cell targets have recently been identified [5 10-14]. To detect rare antigen-specific CD4+ T cells *ex vivo*, others and we made use of a protocol first published by Wucherpfennig and colleagues [15] that combines tetramer staining with magnetic bead enrichment. By this means, we previously quantified and phenotypically characterized T cells specific for epitopes from several RA-associated candidate antigens in a variety of different RA samples, including peripheral blood mononuclear cells (PBMC), synovial mononuclear cells (SFMC) and synovial tissue [5 6]. However, a major shortcoming of this method is the need for large samples, where the study of one epitope typically requires 20-30 million PBMC.

Based on the combinatorial HLA class II tetramer staining approach demonstrated for immunodominant viral epitopes by Uchtenhagen *et al* [16], we thus set out to develop a multi-tetramer assay for detection of autoreactive T cells that would render it possible to investigate numerous specificities simultaneously and in parallel retain high sensitivity allowing identification of rare autoreactive CD4+ T cells. We here focused on known HLA-DRB1*04:01-restricted citrullinated T cell epitopes from some of the most common RA-associated autoantigens, namely α -enolase, vimentin, fibrinogen beta chain (FGB) as well as cartilage intermediate layer protein (CILP) [5 14].

ACPAs have been shown to be present in individuals developing RA decades before onset of disease [17]. Moreover, studies on a large twin cohort examining genetic and environmental factors in the development of RA suggested that specific HLA class II alleles and thus probably CD4+ T cells are likely to be involved in the maturation of the ACPA response, *i.e.* epitope spreading shortly before disease onset, and the subsequent initiation of arthritis [18]. However, studies on autoreactive CD4+ T cells *ex vivo* prior to or at disease onset are currently not available in RA and the role of autoreactive CD4+ T cells in the pathogenesis of RA needs to be further investigated. We therefore studied the autoreactive T cell profile, specificity, phenotype and frequency, in individuals with seropositive arthralgia who are at risk of developing RA, and in early, untreated RA patients at disease onset. To this end we screened lymph node (LN) core needle biopsies from arthralgia patients and compared them with early, untreated RA patients. Next we investigated PBMC collected from early RA patients that were sampled prior to treatment initiation and at six months follow-up visit [19].

Altogether, this study demonstrates that a detailed analysis of citrulline-specific T cells using a multi-tetramer assay is feasible in both individuals at risk of developing RA and patients at disease onset. Even though at low frequencies, these autoreactive T cells mainly displayed a memory phenotype at time of RA onset. Moreover, certain autoreactive T cells were found to decrease in frequency in patients achieving minimum disease activity following six months treatment. Hereby, we could substantiate the utility of this approach for future longitudinal immune-monitoring.

Results

The multi-tetramer approach allows stable detection of autoreactive T cells at low frequencies in healthy controls

To study citrulline-reactive T cell specificities with our multi-tetramer assay, we created a panel of HLA-DRB1*04:01 tetramers loaded with eight citrullinated RA-associated self peptides derived from four candidate autoantigens, namely α -enolase, vimentin, fibrinogen-b as well as CILP. Additionally, we employed two commonly used viral control epitopes from influenza matrix proteins [20 21]. Table 2 lists the sequence and position of the peptides as well as the tetramer fluorophores. As autoreactive T cells have been shown to be present at low frequencies in the circulation of RA patients and healthy individuals [5 6] we first determined the sensitivity of our multi-tetramer assay by applying it to PBMC from seven HLA-DRB1*04:01-positive healthy control subjects. Here, we unambiguously detected T cells

specific for α -enolase, CILP and fibrinogen at frequencies ranging up to 10 per million CD4+ T cells (median frequency 4.7 and 1.7 per million CD4+ T cells, respectively; Figure 1a). As expected, the number of cells reactive to our positive influenza peptide controls was higher, typically in a range of 20 to 1000 per million CD4+ T cells (median frequency 61 per million CD4+ T cells). Repeat experiments performed on cells from the same time points and donors demonstrated on average comparable frequencies of tetramer-positive cells in the two experiments for each donor (Figure 1a).

Besides enumerating the tetramer-positive CD4+ T cells, we also determined their differentiation state by examining the surface expression of CD45RA and CCR7 (Figure 1b). As expected, T cells specific for influenza were predominantly of a memory phenotype and distributed between a T_{cm}, central memory (51%) and a T_{em}, effector memory phenotype (44%). Conversely, the majority of autoreactive T cells in these healthy subjects displayed a naïve phenotype, expressing CCR7 and CD45RA simultaneously (Figure 1b and c). Still, it should be noted that we also detected central memory type T cells in a subset of the samples, while effector memory T cells were consistently a minor phenotype.

Autoreactive T cells are found in most RA patients, even in the absence of concurrent disease activity

Next, in order to further validate our panel also in patient samples, we analysed a longitudinal cohort of 14 RA patients from which we obtained samples from repeat blood draws approximately 2-3 weeks apart and therefore could analyse intra-individual variance. The patients included in this cohort were recruited according to the following criteria: having ACPA-positive RA and at least one HLA-DRB1*04:01 allele. All patients had long disease duration (>5 years), overall no signs of active disease around the time of sampling and stable anti-rheumatic treatment according to standards (see Additional file 1: Table S1.1).

We detected frequencies between 1 and 35 tetramer-positive cells per million CD4+ cells of CILP/fibrinogen- and α -enolase-specific T cells in these RA patients (Figure 2a). These frequencies were slightly increased in patients compared to healthy controls (Figure 1a and 2a). Not all specificities were present in all patients, with α -enolase-specific T cells being detected in eight out of fourteen and CILP/fibrinogen-specific T cells in thirteen out of fourteen patients. Specificities within individual patients were reliably detected in the repeat blood draws in half of the individuals. Other patients showed citrulline-specific T cells only at one or two of the three time points, as indicated by single dots and dotted lines connecting the frequencies of the remaining time points in Figure 2a. In contrast, influenza-specific T cells were steadily found in all patients in each of the three repeats and always at 10-20 times higher frequency compared to autoreactive T cells (Figure 2a). Examining the overall distribution of the cells within the different memory and naïve states, we detected - similarly to healthy subjects - a high proportion of influenza-specific T cells in the central and effector memory compartment and very little amounts of naïve T cells (Figure 2b). Again, we found a broad distribution of the proportion of naïve citrulline-reactive T cells between different subjects. Within the memory subset, central memory type T cells were overrepresented among CILP/fibrinogen- compared to α -enolase-reactive T cells (Figure 2b). To

a lower extent (<20%) we also detected citrulline-specific T cells with a terminal effector memory phenotype, such Temra cells were virtually absent amongst the influenza-specific T cells (Figure 2b).

Analysing the expression of chemokine receptors on the tetramer-positive cells revealed a higher percentage of CXCR3+ cells among α -enolase- and CILP/FGB-specific compared to influenza-specific T cells (Figure 2c). CXCR5 expressing cells on the other hand were mainly detected among CILP/fibrinogen- and influenza-specific T cells and rather less among cells recognizing α -enolase. We found little expression of both CD25 and CCR6 on the influenza-specific as well as the general CD4 population, while such phenotypes were detected in α -enolase- and CILP/fibrinogen-specific T cells (Figure 2c). Of note, among influenza- and citrulline-specific cells as well as in the general CD4 population around half of the CXCR3+ cells also expressed CXCR5 and to a lesser extent CCR6 (data not shown).

Citrulline-specific T cells are present in lymph node biopsies of arthralgia and early RA patients

Next, we applied the multi-tetramer assay to lymph node biopsies from HLA-DRB1*04:01-positive individuals. Included in this part of the study were arthralgia patients with elevated RF and ACPA levels as well as early RA patients. Samples from one patient with undifferentiated arthritis and one healthy control were also included in this analysis (see Additional file 1: Table S1.2)

Due to the limited cell numbers that can be retrieved from a core needle biopsy we decided against the general *ex vivo* protocol but opted instead for analysing cells after *in vitro* expansion using PHA and IL-2. We could detect citrulline-reactive T cells in all of the tested individuals albeit with varying number of specificities (Figure 3). All five RA patients had vimentin- and α -enolase-specific T cells while CILP/fibrinogen-specific T cells were only found in four patients. Three of the five arthralgia patients displayed reactivity against all citrullinated peptides. The remaining two had no CILP/fibrinogen-specific T cells, but both displayed vimentin-reactive and one also α -enolase-specific T cells. Also for the healthy control we could demonstrate the presence of vimentin- and α -enolase-specific T cells, however no CILP/fibrinogen-reactive ones. The patient diagnosed with undifferentiated arthritis on the other hand had all T cell reactivities we tested for. It should be noted that since these cells required cell culture expansion, we could not calculate the original frequencies nor analyse the unmanipulated phenotypes.

Citrulline-specific T cells decline in early RA patients achieving drug-induced remission

Finally, we applied the multi-tetramer assay to peripheral blood samples from ten patients enrolled in the LURA study at the Karolinska University Hospital. This cohort comprises 134 newly diagnosed patients with duration of patient-reported symptoms ≤ 1 year and naïve to treatment with disease-modifying anti-rheumatic drugs (DMARDs) and oral glucocorticoids (GC). The selection of the patients for this substudy was based on HLA-DRB1*04:01-positivity and the availability of blood samples from both baseline and six months follow-up.

T cells specific for both influenza and citrullinated peptides were found to be part of the peripheral T cell repertoire at both baseline and following six months of standard medication with methotrexate in this early RA cohort (Figure 4a). Citrulline-specific T cells were detected in all but one patient, who interestingly turned out to be negative for both RF as well as ACPAs. The specificity that was detected least often was towards vimentin, with 3/10 and 4/10 positive patients for baseline and follow-up, respectively. Both α -enolase- and CILP/fibrinogen-specific cells were found in 50-60% of the patients at both time points (Figure 4b). The general frequencies determined for influenza-specific as well as autoreactive T cells were in line with previous results and ranged from 10 to 230 and 1 to 7 tetramer-positive cells per million CD4+ cells, respectively. Notably, three of the four patients for whom the frequency of citrulline-specific T cells declined from baseline to follow-up, were also patients that achieved a DAS28-value <2.6 which corresponds to minimal disease activity and clinical remission (Figure 4a, coloured in green and table 1). Another patient could, due to a missing DAS28-value at follow-up, not be added to the group of responders, although we could demonstrate a decrease in citrulline-reactive T cells also for this patient (Figure 4, coloured in grey and table 1).

Subsequent analysis of CD45RA and CCR7 expression revealed a significant decrease of central memory type T cells after six months amongst CILP/fibrinogen-specific cells. Along the same line we detected a general trend towards lower proportions of memory type T cells for both α -enolase- and CILP/fibrinogen-specific cells and concurrently higher proportions of naïve as well as terminal effector memory T cell phenotypes (Figure 4c and Additional file 1: Figures S2c, right panel, and S3a). Conversely, vimentin-specific T cells displayed a trend for increased central memory and decreased naïve T cells at six months follow-up (Figure 4c and Additional file 1: Figure S3a).

To investigate the activation status of these cells in more detail we analysed expression of CD25 and CD38, markers of recent T cell activation and detected no significant changes upon comparing baseline and follow-up for neither of the two markers. As expected for patients not subject to ongoing infections or recent vaccination, the level of expression on the influenza-specific control as well as the general CD4+ population was very low for both markers. However, trends are pointing towards lower proportion of CD25+ cells among both α -enolase- and CILP/fibrinogen-specific T cells after six months of treatment. For CD38 we found the opposite feature with an increased expression on both α -enolase- and vimentin-specific memory cells after six months (see Additional file 1: Figure S3b).

Similarly, we examined the surface expression of the chemokine receptors CCR6, CXCR3 and CXCR5 and found a general decrease in the frequency of CXCR5+ cells for all autoreactive as well as the influenza-specific T cells at follow-up. In general, CXCR5, compared to CXCR3 and CCR6, is the chemokine receptor that was most prominently expressed on tetramer-positive cells with mean levels of up to 40% for citrulline-specific and 54% for influenza-specific populations (see Additional file 1: Figure S3b) while for the general CD4+ population it was only around 10%. In the case of CXCR3 the average expression level did not exceed 22% and was lower at follow-up for CILP/fibrinogen- and vimentin-specific cells. Among α -enolase-reactive cells on the other hand we could not detect changes in the proportion of CXCR3+ cells. The frequency of CCR6-expressing cells was generally highest among vimentin-specific cells (mean

between 17 and 33%) but decreased from baseline to follow-up while it increased for both α -enolase- and CILP/fibrinogen-specific cells (see Additional file 1: Figure S3b).

Among all citrulline-specific cells we observed a number of cells co-expressing two or three of the chemokine receptors. Here it was most common to find CXCR5 in combination with either CXCR3 or CCR6 (data now shown). In contrast to this we found only a few influenza-specific cells co-expressing these chemokine receptors with a majority of CXCR3+ cells carrying CCR6 and/or CXCR5, while among CXCR5+ cells the proportion of cells carrying a second chemokine receptor was rather small (data not shown).

Discussion

With this study we demonstrate the utility of a multi-HLA class II tetramer approach to allow the unambiguous and simultaneous detection of rare self-reactive T cells in patients with seropositive arthralgia, i.e. individuals at risk of RA development, and those with early, untreated RA at the time of diagnosis. This technology could be useful in several settings where precise immune-monitoring would be beneficial, including longitudinal screening in the context of the stepwise development of RA, as well as follow-up studies in conjunction with therapy response or tolerance induction. This is demonstrated within the present study through the assessment of frequency and phenotype of citrulline-reactive T cells in RA patients at time of diagnosis and after six months of methotrexate therapy. Here, the numbers of citrulline-reactive T cells were diminished and the phenotype of certain specificities, but not others, was changed in patients responding to therapy and reaching minimal disease activity (DAS28 <2.6).

Self-reactive CD4+ T cells are known to be rare, and several autoimmune diseases can be initiated through recognition of multiple target antigens. This is true *e.g.* for type 1 diabetes (T1D) and multiple sclerosis [22-24], but even more so for rheumatoid arthritis where common posttranslational modifications, like citrullination and carbamylation, are the target of the aberrant immune response [25 26]. Clinical immune-monitoring of autoreactive CD4+ T cells in this setting will hence not only depend on sensitive methods to capture these rare events but also on the possibility to determine multiple specificities in the same assay. Multi-parameter flow cytometry is well-suited for these challenges and we here performed a first proof of principle study building on previous work in the setting of influenza vaccines [16]. We built a panel focusing on eight citrullinated T cell epitopes from four different proteins, that have previously been shown to be presented on HLA-DRB1*04:01 in the context of RA [5 11 14], and also included two viral control peptides [20 21] (Table 2). For the phenotypic markers we cover subdivisions of naïve versus central and effector memory cell states as well as the main signature T helper cell subsets including Th1 (CXCR3), Th17 (CCR6), Tfh (CXCR5), all of which have been postulated as interesting in the context of RA [27-32]. In fact, when examining the expression of these chemokine receptors on the citrulline-specific T cells we found the cells to be double- or even triple-positive for these markers in several instances, which also conformed with our observations for cells from the general CD4+ cell population.

For general validation of the assay we started out staining PBMC from healthy controls making use of the fact that autoreactive T cells, albeit in low numbers, form part of the normal T cell repertoire [5 33-38]. As expected, the frequencies of citrulline-specific T cells were low but stably detectable in repeat experiments performed on samples from the same donors. Thus, besides the sensitivity in detecting these rare autoreactive T cells, we also demonstrated its robustness by replicating the stainings. To further corroborate these results, we applied our approach to repeat blood samples – three per patient – taken from 14 RA patients over the time course of 4-6 weeks. As all patients had stable long-standing disease we expected the autoreactive CD4+ T cell responses to be stable over this period of time. This was the case for around half of the patients for which we detected citrulline-specific T cells at all three time points. When investigating the differentiation state of the tetramer-positive cells we decided to focus on CD45RA and CCR7 to discriminate defined memory phenotypes from naïve T cell subsets and also included an activation marker besides the above-mentioned chemokine receptors. In contrast to influenza-specific T cells that were mainly found in effector and central memory subsets, a large proportion of the citrulline-specific T cells in these quiescent RA patients belonged to the naïve compartment with only few of the cells displaying a memory phenotype. However, interestingly, there is increasing evidence that the naïve T cell compartment is not homogeneous and can also include memory T cells such as the so called stem cell memory T cells, Tscm [39]. In our study, we defined naïve T cells as CD45RA and CCR7 double-expressing cells and did not include further markers to subcategorise such Tscm cells. It is nonetheless possible that said cells exist also in the autoreactive T cell repertoire in RA and explain the large proportion of citrulline-reactive T cells we detect in the naïve compartment.

The above-mentioned *ex vivo* staining approach relies on the availability of viable cells in the magnitude of around 20 million per time point. Still, in a clinical setting some samples would never reach such numbers. Small needle biopsies represent such scarce, but clinically interesting material. We had the opportunity to utilise samples of the Amsterdam lymph node biopsy cohort for our study [40], and in this context used cells that had been *in vitro* propagated prior to cryopreservation and tetramer staining. Nevertheless, we managed to find citrulline-reactive T cells of different specificities in all the RA patients and in many of the arthralgia patients. We also included some LN-relevant phenotypic markers, focusing on PD-1, CXCR5 as well as ICOS, to query the presence of a classical follicular T helper phenotype [41]. The phenotypic data was, probably due to the extended *in vitro* culture, inconclusive; nonetheless for some individuals we still detected prominent PD-1 positive populations (data not shown). Importantly, the protocol allows easy adjustment of the panel to capture the most relevant phenotypic markers based on the compartment from which the cells originate. So when *e.g.* studying RA synovial fluid (SF), a different set of markers would be interesting to stain for compared to peripheral blood. An interesting marker would in that case be PD-1 that has recently been shown to characterize a specific subset of T helper cells in SF, the peripheral T helper cells, that in contrast to T follicular helper cells do not express CXCR5 and target inflamed tissues which is why they are close to absent in the periphery [42].

To further explore the capacity of our multi-tetramer panel in detecting differences over time, we utilized samples from early RA patients [19] which included peripheral blood obtained at time of RA diagnosis, prior to treatment initiations, and the subsequent 6 months follow-up visit. This gave us the possibility to

ex vivo study untreated early RA patients and their T cell reactivities as well as the phenotype of these cells. We found for instance that three out of the four patients achieving remission by anti-rheumatic treatment had lower frequencies of citrulline-specific T cells at the follow-up. Notably, patient 5, for whom we also detected a decline in the frequency of citrulline-reactive CD4+ T cells at follow-up, could due to a missing DAS28-value at 6 months not be classified as achieving remission. Nevertheless, we suggest based on low DAS28-values at 3 and 13 months (2.24 and 2.53, respectively) and on the fact that no change in therapy was done at the 6 months follow-up visit that this patient probably had low disease activity if not achieved remission. We also demonstrate the benefit of having different HLA class II tetramers in various channels in order to make specific sub-analyses here. Like this, we could detect a decrease in the frequency of memory T cells among α -enolase- and CILP/fibrinogen-specific T cells, but not in vimentin-reactive T cells. Generally, it has been difficult to predict clinical response in RA patients with both synthetic and biological DMARDs [43-51], and our small study implies that clinical response may parallel or result from the restriction of the autoimmune component of the disease.

Conclusions

In our present study, we have used the multi-tetramer approach to demonstrate citrulline-reactive T cells in RA patients. Still, it should be noted that we consider the assay applicable to any disease restricted by HLA where there are a number of antigens involved or even when screening for T cells specific for different peptides of the same antigen. Other disease settings could for example be T1D with its restriction to both HLA-DR and -DQ loci [52] and the numerous autoantigens involved, like glutamic acid decarboxylase (GAD) and 60kDa heat shock protein (HSP60) [53 54] as well as coeliac disease with the HLA-DQ2 and -DQ8-restricted gluten-derived peptides [55 56]. Importantly, we have addressed a number of candidate autoantigens in RA, but consider this only the beginning. As the disease starts developing many years prior to clinical onset and is likely to affect many organs, additional antigens are presumably involved in the disease. More studies are needed in order to get an overview of which specificities are important in each step of disease development and also whether it is possible to distinguish which T cells are public, *i.e.* present in many or the majority of patients, and which are private. Such information would be invaluable for further refinement of an immune-monitoring protocol and for designing future immunotherapies based on the exciting progress being made in RA in this regard [57 58].

Methods

Patients and cell samples

All subjects (healthy donors as well as RA patients) were included based on their genetic profile by carrying one or two HLA-DRB1*04:01 alleles known to be associated with increased risk for rheumatoid arthritis. Fourteen ACPA-positive RA patients (men, %: 29), recruited at the Rheumatology clinic at the Karolinska University Hospital, were asked to participate in a longitudinal validation cohort and donate peripheral blood (PB) at three time points with intervals of 2-3 weeks (see Additional file 1: Table S1.1). Moreover, we had the opportunity to use baseline and six months follow-up PBMC samples from ten HLA-

DRB1*04:01-positive patients (median age and range at inclusion, years: 43 [26-71]; men, %: 40) enrolled in the lung investigation in newly diagnosed RA (LURA) study at Karolinska University Hospital [19]. General characteristics for the selected newly diagnosed early RA patients, including antibody status, disease activity score (DAS28) at baseline and follow-up as well as medication after baseline sampling are displayed in Table 1. Seven HLA-DRB1*04:01-positive control subjects were recruited at the Uppsala Bioresource. PBMC, obtained from heparinized blood, were prepared by centrifugation over Ficoll-Hypaque gradients (GE Healthcare). The samples were cryopreserved in liquid nitrogen in 10% DMSO and 90% heat-inactivated fetal bovine serum.

Lymphoid tissue samples were derived from individuals undergoing ultrasound-guided inguinal LN core needle biopsy [40] recruited at the Academic Medical Center, Amsterdam, Netherlands (median age and range at inclusion, years: 50 [19-81]; men, %: 17). Table S2 (Additional file 1) shows the demographics of the 12 included subjects that were selected based on the presence of at least one HLA-DRB1*04:01 allele. The general study cohort consists of individuals with arthralgia (joint pain without inflammation) and elevated levels of IgM-rheumatoid factor (RF) and/or ACPA as well as early arthritis patients with a disease duration ≤ 6 months and naïve to treatment with biologics. Besides this, healthy controls without any joint problems and RA-specific antibodies were included. Of the five arthralgia patients included in this study none developed RA within a follow-up period of 20-28 months after initial biopsy sampling.

Production of HLA-DRB1*04:01 Tetramers

HLA-DRB1*04:01 protein used within this study was recombinantly produced in S2 insect cells as previously described [20]. All peptides were synthesized at >95% purity by GenScript (Piscataway, NJ, USA). The biotinylated monomer was loaded with the different peptides by incubation in the presence of n-octyl-b-D-glucopyranoside and Pefabloc SC (Sigma-Aldrich) and subsequently tetramerized using streptavidin conjugated to PE, PE-Cy5, PE-CF594 (BD Biosciences) or APC (Biolegend), respectively.

***Ex vivo* detection of T cells by HLA class II tetramers**

PBMC samples from RA patients were stained as previously described [16] using the tetramers loaded with the peptides displayed in Table 2 and subsequently labelled with anti-CD14 (MΦP9), anti-CD19 (4G7), Annexin V, anti-CD4 (SK3), anti-CD45RA (HI100), anti-CD25 (2A3), anti-CCR7 (3D12), anti-CCR6 (11A9), anti-CXCR3 (1C6), anti-CXCR5 (J252D4) antibodies. The inclusion of the more specific markers, like CD25 and the different chemokine receptors in the staining panel allowed us an in-depth characterization of the tetramer-positive cells from the samples. The early RA samples were furthermore labelled with anti-CD38 (HIT2) antibodies. Additional inclusion of an anti-CD3 antibody is recommendable, as it will facilitate gating of a pure CD4+ T lymphocyte population. Samples were then

acquired on a BD LSRFortessa flow cytometer and data was analysed using FlowJo, version 10.0.7 (Tree Star) and GraphPad Prism, version 6.0h. The gating strategies for the different cohorts are presented in Figure S1 (Additional file 1). For calculating the frequency of antigen-specific cells we divided the total number of tetramer-positive cells in the bound fraction by the total number of CD4+ T cells. A cut-off of $1/10^6$ CD4+ T cells was applied when analysing the RA

PBMC samples.

Detection and sorting of lymph node T cells using HLA class II tetramers following *in vitro* cultures

As the number of cells that can be retrieved during a core needle biopsy is limited, we cultured the cells *in vitro* with phytohemagglutinin (PHA) and interleukin-2 (IL-2) to specifically promote T cell activation and proliferation. After 5 days in culture the cells were counted and cryopreserved in liquid nitrogen. The expanded cells were stained directly after thawing using the citrulline-specific tetramers listed in Table 2 and incubated for 1h at 37°C. Thereafter, the cells were labelled with anti-CD14, anti-CD19, Annexin V, anti-CD4, anti-PD-1 (EH12), anti-CXCR5 and some (n=7) also with anti-ICOS (DX29) antibodies. Tetramer-positive cells were subsequently sorted on a BD influx cell sorter using the gating and sorting strategy shown in Figure S1b (Additional file 1).

Statistics

Statistical analysis was performed when applicable and if so, is indicated in the respective figure legends. Comparing the frequency of tetramer-positive cells at baseline with the follow-up time point in early RA patients we applied Mann-Whitney test. p-values less than 0.05 were considered significant and marked with an asterisk.

Abbreviations

RA	Rheumatoid arthritis
CILP	cartilage intermediate layer protein
ACPA	anti-citrullinated protein antibody
PBMC	peripheral blood mononuclear cells
SFMC	synovial fluid mononuclear cells
FGB	fibrinogen beta chain

LN	lymph node
PB	peripheral blood
LURA	lung investigation in early RA patients
DAS	disease activity score
RF	rheumatoid factor
HA	hemagglutinin
MP	matrix protein
PHA	phytohemagglutinin
IL-2	interleukin-2
Tcm	central memory T cell
Tem	effector memory T cell
Temra	terminal effector memory T cell
DMARDs	disease-modifying anti-rheumatic drugs
GC	glucocorticoids
T1D	type 1 diabetes
Tscm	stem cell memory T cell
SF	synovial fluid
GAD	glutamic acid decarboxylase (GAD)
HSP60	60kDa heat shock protein

Declarations

Ethics approval and consent to participate

This study has been approved by the local ethical review boards at Karolinska University Hospital, Stockholm, Uppsala University, Uppsala and Academic Medical Center, Amsterdam and was performed according to the guidelines of the Declaration of Helsinki. Written informed consent was obtained from all participants prior to inclusion in the studies.

Consent for publication

Not applicable.

Availability of data and material

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Competing interests

The authors declare that they have no competing interests.

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Authors' contributions

CG and VM designed the experiments and wrote the manuscript. CG, ST and KC acquired and analysed data. THR processed samples. AHH, AD, GK, LR, AIC and LGMvB provided reagents or samples, respectively. AHH, HU, WWK, AA, AIC and LGMvB contributed to discussion. AHH, ST, KC, HU, AD, WWK, AA, AIC and LGMvB reviewed and edited the manuscript. All authors read and approved the final manuscript.

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References

1. Altman JD, Moss PA, Goulder PJ, Barouch DH, McHeyzer-Williams MG, Bell JI, et al. Phenotypic analysis of antigen-specific T lymphocytes. *Science*. 1996;274(5284):94-6.
2. Wambre E, DeLong JH, James EA, LaFond RE, Robinson D, Kwok WW. Differentiation stage determines pathologic and protective allergen-specific CD4+ T-cell outcomes during specific immunotherapy. *The Journal of allergy and clinical immunology*. 2012;129(2):544-51, 51 e1-7.
3. Raki M, Fallang LE, Brottveit M, Bergseng E, Quarsten H, Lundin KE, et al. Tetramer visualization of gut-homing gluten-specific T cells in the peripheral blood of celiac disease patients. *Proceedings of the National Academy of Sciences of the United States of America*. 2007;104(8):2831-6.
4. Kwok WW, Roti M, DeLong JH, Tan V, Wambre E, James EA, et al. Direct ex vivo analysis of allergen-specific CD4+ T cells. *The Journal of allergy and clinical immunology*. 2010;125(6):1407-9 e1.
5. James EA, Rieck M, Pieper J, Gebe JA, Yue BB, Tatum M, et al. Citrulline-specific Th1 cells are increased in rheumatoid arthritis and their frequency is influenced by disease duration and therapy. *Arthritis & rheumatology*. 2014;66(7):1712-22.
6. Pieper J, Dubnovitsky A, Gerstner C, James EA, Rieck M, Kozhukh G, et al. Memory T cells specific to citrullinated alpha-enolase are enriched in the rheumatic joint. *Journal of autoimmunity*. 2018.
7. Gregersen PK, Silver J, Winchester RJ. The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. *Arthritis Rheum*. 1987;30(11):1205-13.
8. Huizinga TW, Amos CI, van der Helm-van Mil AH, Chen W, van Gaalen FA, Jawaheer D, et al. Refining the complex rheumatoid arthritis phenotype based on specificity of the HLA-DRB1 shared epitope for antibodies to citrullinated proteins. *Arthritis and rheumatism*. 2005;52(11):3433-8.
9. Klareskog L, Stolt P, Lundberg K, Kallberg H, Bengtsson C, Grunewald J, et al. A new model for an etiology of rheumatoid arthritis: smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. *Arthritis and rheumatism*. 2006;54(1):38-46.
10. von Delwig A, Locke J, Robinson JH, Ng WF. Response of Th17 cells to a citrullinated arthritogenic aggrecan peptide in patients with rheumatoid arthritis. *Arthritis and rheumatism*. 2010;62(1):143-9.
11. Snir O, Rieck M, Gebe JA, Yue BB, Rawlings CA, Nepom G, et al. Identification and functional characterization of T cells reactive to citrullinated vimentin in HLA-DRB1*0401-positive humanized mice and rheumatoid arthritis patients. *Arthritis and rheumatism*. 2011;63(10):2873-83.
12. Freed BM, Schuyler RP, Aubrey MT. Association of the HLA-DRB1 epitope LA(67, 74) with rheumatoid arthritis and citrullinated vimentin binding. *Arthritis and rheumatism*. 2011;63(12):3733-9.

13. Law SC, Street S, Yu CH, Capini C, Ramnoruth S, Nel HJ, et al. T-cell autoreactivity to citrullinated autoantigenic peptides in rheumatoid arthritis patients carrying HLA-DRB1 shared epitope alleles. *Arthritis research & therapy*. 2012;14(3):R118.
14. Gerstner C, Dubnovitsky A, Sandin C, Kozhukh G, Uchtenhagen H, James EA, et al. Functional and Structural Characterization of a Novel HLA-DRB1*04:01-Restricted alpha-Enolase T Cell Epitope in Rheumatoid Arthritis. *Frontiers in immunology*. 2016;7:494.
15. Day CL, Seth NP, Lucas M, Appel H, Gauthier L, Lauer GM, et al. Ex vivo analysis of human memory CD4 T cells specific for hepatitis C virus using MHC class II tetramers. *The Journal of clinical investigation*. 2003;112(6):831-42.
16. Uchtenhagen H, Rims C, Blahnik G, Chow IT, Kwok WW, Buckner JH, et al. Efficient ex vivo analysis of CD4+ T-cell responses using combinatorial HLA class II tetramer staining. *Nature communications*. 2016;7:12614.
17. Brink M, Hansson M, Mathsson L, Jakobsson PJ, Holmdahl R, Hallmans G, et al. Multiplex analyses of antibodies against citrullinated peptides in individuals prior to development of rheumatoid arthritis. *Arthritis Rheum*. 2013;65(4):899-910.
18. Hensvold AH, Magnusson PK, Joshua V, Hansson M, Israelsson L, Ferreira R, et al. Environmental and genetic factors in the development of anticitrullinated protein antibodies (ACPAs) and ACPA-positive rheumatoid arthritis: an epidemiological investigation in twins. *Annals of the rheumatic diseases*. 2015;74(2):375-80.
19. Reynisdottir G, Joshua V, Nyren S, Hensvold A, Karimi R, Harju A, et al. Lung changes detected by high resolution tomography are present in ACPA positive RA patients already at disease onset. *Annals of the rheumatic diseases*. 2012;71(Suppl 1):A23-A.
20. Novak EJ, Liu AW, Nepom GT, Kwok WW. MHC class II tetramers identify peptide-specific human CD4(+) T cells proliferating in response to influenza A antigen. *The Journal of clinical investigation*. 1999;104(12):R63-7.
21. Ge X, Tan V, Bollyky PL, Standifer NE, James EA, Kwok WW. Assessment of seasonal influenza A virus-specific CD4 T-cell responses to 2009 pandemic H1N1 swine-origin influenza A virus. *Journal of virology*. 2010;84(7):3312-9.
22. Spanier JA, Sahli NL, Wilson JC, Martinov T, Dileepan T, Burrack AL, et al. Increased Effector Memory Insulin-Specific CD4(+) T Cells Correlate With Insulin Autoantibodies in Patients With Recent-Onset Type 1 Diabetes. *Diabetes*. 2017;66(12):3051-60.
23. Blahnik G, Uchtenhagen H, Chow IT, Speake C, Greenbaum C, Kwok WW, et al. Analysis of pancreatic beta cell specific CD4+ T cells reveals a predominance of proinsulin specific cells. *Cellular immunology*. 2018.
24. Compston A, Coles A. Multiple sclerosis. *Lancet*. 2008;372(9648):1502-17.
25. Wegner N, Lundberg K, Kinloch A, Fisher B, Malmstrom V, Feldmann M, et al. Autoimmunity to specific citrullinated proteins gives the first clues to the etiology of rheumatoid arthritis. *Immunological reviews*. 2010;233(1):34-54.

26. Shi J, Knevel R, Suwannalai P, van der Linden MP, Janssen GM, van Veelen PA, et al. Autoantibodies recognizing carbamylated proteins are present in sera of patients with rheumatoid arthritis and predict joint damage. *Proceedings of the National Academy of Sciences of the United States of America*. 2011;108(42):17372-7.
27. Qin S, Rottman JB, Myers P, Kassam N, Weinblatt M, Loetscher M, et al. The chemokine receptors CXCR3 and CCR5 mark subsets of T cells associated with certain inflammatory reactions. *The Journal of clinical investigation*. 1998;101(4):746-54.
28. Chabaud M, Durand JM, Buchs N, Fossiez F, Page G, Frappart L, et al. Human interleukin-17: A T cell-derived proinflammatory cytokine produced by the rheumatoid synovium. *Arthritis and rheumatism*. 1999;42(5):963-70.
29. Pene J, Chevalier S, Preisser L, Venereau E, Guilleux MH, Ghannam S, et al. Chronically inflamed human tissues are infiltrated by highly differentiated Th17 lymphocytes. *Journal of immunology*. 2008;180(11):7423-30.
30. Ronnelid J, Berg L, Rogberg S, Nilsson A, Albertsson K, Klareskog L. Production of T-cell cytokines at the single-cell level in patients with inflammatory arthritides: enhanced activity in synovial fluid compared to blood. *British journal of rheumatology*. 1998;37(1):7-14.
31. Kanik KS, Hagiwara E, Yarboro CH, Schumacher HR, Wilder RL, Klinman DM. Distinct patterns of cytokine secretion characterize new onset synovitis versus chronic rheumatoid arthritis. *The Journal of rheumatology*. 1998;25(1):16-22.
32. Ma J, Zhu C, Ma B, Tian J, Baidoo SE, Mao C, et al. Increased frequency of circulating follicular helper T cells in patients with rheumatoid arthritis. *Clinical & developmental immunology*. 2012;2012:827480.
33. Burns J, Rosenzweig A, Zweiman B, Lisak RP. Isolation of myelin basic protein-reactive T-cell lines from normal human blood. *Cellular immunology*. 1983;81(2):435-40.
34. Tournier-Lasserre E, Hashim GA, Bach MA. Human T-cell response to myelin basic protein in multiple sclerosis patients and healthy subjects. *Journal of neuroscience research*. 1988;19(1):149-56.
35. Viglietta V, Kent SC, Orban T, Hafler DA. GAD65-reactive T cells are activated in patients with autoimmune type 1a diabetes. *The Journal of clinical investigation*. 2002;109(7):895-903.
36. Danke NA, Koelle DM, Yee C, Beheray S, Kwok WW. Autoreactive T cells in healthy individuals. *Journal of immunology*. 2004;172(10):5967-72.
37. Reijonen H, Kwok WW, Nepom GT. Detection of CD4+ autoreactive T cells in T1D using HLA class II tetramers. *Annals of the New York Academy of Sciences*. 2003;1005:82-7.
38. Roep BO, Atkinson MA, van Endert PM, Gottlieb PA, Wilson SB, Sachs JA. Autoreactive T cell responses in insulin-dependent (Type 1) diabetes mellitus. Report of the first international workshop for standardization of T cell assays. *Journal of autoimmunity*. 1999;13(2):267-82.
39. van den Broek T, Borghans JAM, van Wijk F. The full spectrum of human naive T cells. *Nature reviews Immunology*. 2018;18(6):363-73.

40. de Hair MJ, Zijlstra IA, Boumans MJ, van de Sande MG, Maas M, Gerlag DM, et al. Hunting for the pathogenesis of rheumatoid arthritis: core-needle biopsy of inguinal lymph nodes as a new research tool. *Annals of the rheumatic diseases*. 2012;71(11):1911-2.
41. Fazilleau N, Mark L, McHeyzer-Williams LJ, McHeyzer-Williams MG. Follicular helper T cells: lineage and location. *Immunity*. 2009;30(3):324-35.
42. Rao DA, Gurish MF, Marshall JL, Slowikowski K, Fonseka CY, Liu Y, et al. Pathologically expanded peripheral T helper cell subset drives B cells in rheumatoid arthritis. *Nature*. 2017;542(7639):110-4.
43. Maini R, St Clair EW, Breedveld F, Furst D, Kalden J, Weisman M, et al. Infliximab (chimeric anti-tumour necrosis factor alpha monoclonal antibody) versus placebo in rheumatoid arthritis patients receiving concomitant methotrexate: a randomised phase III trial. ATTRACT Study Group. *Lancet*. 1999;354(9194):1932-9.
44. Moreland LW, Schiff MH, Baumgartner SW, Tindall EA, Fleischmann RM, Bulpitt KJ, et al. Etanercept therapy in rheumatoid arthritis. A randomized, controlled trial. *Annals of internal medicine*. 1999;130(6):478-86.
45. van de Putte LB, Atkins C, Malaise M, Sany J, Russell AS, van Riel PL, et al. Efficacy and safety of adalimumab as monotherapy in patients with rheumatoid arthritis for whom previous disease modifying antirheumatic drug treatment has failed. *Annals of the rheumatic diseases*. 2004;63(5):508-16.
46. Keystone E, Heijde D, Mason D, Jr., Landewe R, Vollenhoven RV, Combe B, et al. Certolizumab pegol plus methotrexate is significantly more effective than placebo plus methotrexate in active rheumatoid arthritis: findings of a fifty-two-week, phase III, multicenter, randomized, double-blind, placebo-controlled, parallel-group study. *Arthritis and rheumatism*. 2008;58(11):3319-29.
47. Emery P, Fleischmann RM, Moreland LW, Hsia EC, Strusberg I, Durez P, et al. Golimumab, a human anti-tumor necrosis factor alpha monoclonal antibody, injected subcutaneously every four weeks in methotrexate-naive patients with active rheumatoid arthritis: twenty-four-week results of a phase III, multicenter, randomized, double-blind, placebo-controlled study of golimumab before methotrexate as first-line therapy for early-onset rheumatoid arthritis. *Arthritis and rheumatism*. 2009;60(8):2272-83.
48. Cohen SB, Emery P, Greenwald MW, Dougados M, Furie RA, Genovese MC, et al. Rituximab for rheumatoid arthritis refractory to anti-tumor necrosis factor therapy: Results of a multicenter, randomized, double-blind, placebo-controlled, phase III trial evaluating primary efficacy and safety at twenty-four weeks. *Arthritis and rheumatism*. 2006;54(9):2793-806.
49. Genovese MC, Becker JC, Schiff M, Luggen M, Sherrer Y, Kremer J, et al. Abatacept for rheumatoid arthritis refractory to tumor necrosis factor alpha inhibition. *The New England journal of medicine*. 2005;353(11):1114-23.
50. Smolen JS, Beaulieu A, Rubbert-Roth A, Ramos-Remus C, Rovensky J, Alecock E, et al. Effect of interleukin-6 receptor inhibition with tocilizumab in patients with rheumatoid arthritis (OPTION study): a double-blind, placebo-controlled, randomised trial. *Lancet*. 2008;371(9617):987-97.

51. Lee EB, Fleischmann R, Hall S, Wilkinson B, Bradley JD, Gruben D, et al. Tofacitinib versus methotrexate in rheumatoid arthritis. *The New England journal of medicine*. 2014;370(25):2377-86.
52. Noble JA, Valdes AM. Genetics of the HLA region in the prediction of type 1 diabetes. *Current diabetes reports*. 2011;11(6):533-42.
53. Baekkeskov S, Aanstoot HJ, Christgau S, Reetz A, Solimena M, Cascalho M, et al. Identification of the 64K autoantigen in insulin-dependent diabetes as the GABA-synthesizing enzyme glutamic acid decarboxylase. *Nature*. 1990;347(6289):151-6.
54. Abulafia-Lapid R, Elias D, Raz I, Keren-Zur Y, Atlan H, Cohen IR. T cell proliferative responses of type 1 diabetes patients and healthy individuals to human hsp60 and its peptides. *Journal of autoimmunity*. 1999;12(2):121-9.
55. Lundin KE, Scott H, Hansen T, Paulsen G, Halstensen TS, Fausa O, et al. Gliadin-specific, HLA-DQ(alpha 1*0501,beta 1*0201) restricted T cells isolated from the small intestinal mucosa of celiac disease patients. *The Journal of experimental medicine*. 1993;178(1):187-96.
56. Lundin KE, Scott H, Fausa O, Thorsby E, Sollid LM. T cells from the small intestinal mucosa of a DR4, DQ7/DR4, DQ8 celiac disease patient preferentially recognize gliadin when presented by DQ8. *Human immunology*. 1994;41(4):285-91.
57. Baker KF, Isaacs JD. Novel therapies for immune-mediated inflammatory diseases: What can we learn from their use in rheumatoid arthritis, spondyloarthritis, systemic lupus erythematosus, psoriasis, Crohn's disease and ulcerative colitis? *Annals of the rheumatic diseases*. 2018;77(2):175-87.
58. Jansen D, Ramnoruth N, Loh KL, Rossjohn J, Reid HH, Nel HJ, et al. Flow Cytometric Clinical Immunomonitoring Using Peptide-MHC Class II Tetramers: Optimization of Methods and Protocol Development. *Frontiers in immunology*. 2018;9:8.
59. Sallusto F, Lenig D, Forster R, Lipp M, Lanzavecchia A. Two subsets of memory T lymphocytes with distinct homing potentials and effector functions. *Nature*. 1999;401(6754):708-12.

Tables

Table 1. Characteristics of selected newly diagnosed early RA patients

Subject	Antibody status (RF/ACPA)	DAS28 0m	DAS28 6m	Medication after baseline sampling
1	+/+	3.89	3.88	Mtx + GC
2	+/+	4.68	^{B, D} 2.58	Mtx+ GC
3	+/+	5.04	^{B, D} 1.05	Mtx+ GC
4	+/+	5.70	^C 3.44	Mtx
5	+/+	7.39	2.24/2.53 ^{#**}	Mtx+ GC
6	+/+	5.47	3.46 [*]	Mtx
7	-/+	4.90	2.84 ^{†**}	Mtx
8	-/-	5.52	2.39 ^{††**}	Mtx + GC
9	+/+	5.49	1.63 ^{††**}	Mtx + GC
10	+/+	4.31	2.63 ^{†**}	Mtx

Abbreviations: DAS28, disease activity score 28; Mtx, methotrexate; GC, glucocorticoids;

^A low disease activity; ^B remission; ^C moderate improvement; ^D good improvement; ^E DAS28 at 6m not available, displayed are DAS28 at 3m/13m

Table 2. Epitopes used in this study

Name	Protein	Sequence	Fluoro-chrome	Reference	
cit11-25	a-enolase	11-IFDSXGNPTVEVDLF	PE	5	
cit26-40	a-enolase	26-TSKGLFXAAVPSGAS		14	
cit326-340	a-enolase	326-KXIAKAVNEKSCNCL		5	
cit-CILP297-311	CILP	297-ATIKAEFVXAETPYM	PE-CF594	5	
cit-CILP982-979	CILP	982-GKLYGIXDVXSTRDR		5	
cit-fibb 69-80	fibrinogen b	69-GYXAXPAKAAAT		5	
cit-vim59-78	vimentin	59-GVYATXSSAVXLXSSVPGVR	APC	11	included in LURA and LN experiments
cit-vim418-431	vimentin	418-FSSLNLXETNLDSL		5	
HA306-318	influenza HA	306-PKYVKQNTLKLAT	PE-Cy5	21	excluded in LN experiments
MP97-116	influenza MP	97-VKLYRKLKREITFHGAKEIS		22	

Abbreviations: X, citrulline; CILP, cartilage intermediate layer protein; HA, hemagglutinin; MP, matrix protein

Figures

Figure 1

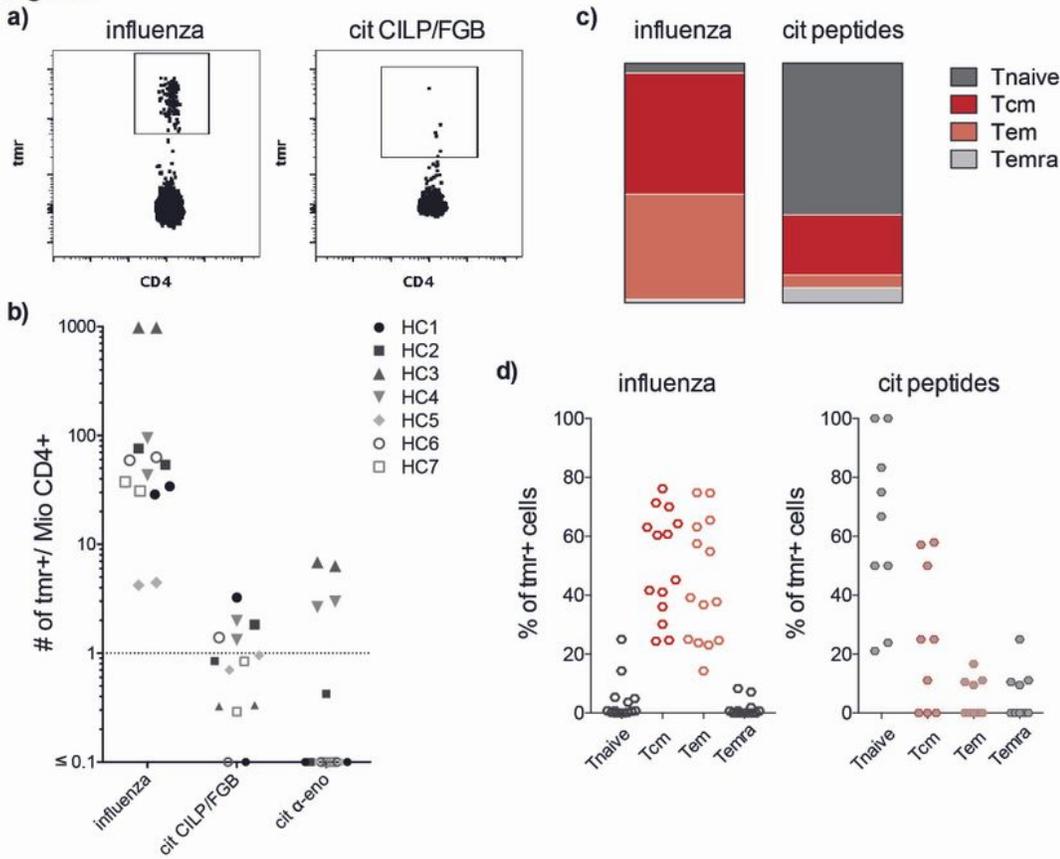


Figure 1

The multi-tetramer approach is sensitive enough to detect antigen-specific CD4+ T cells in healthy controls a) Representative flow plots depicting the gating strategy for CD4+ T cells reactive to influenza (left) and citrullinated CILP/FGB (right). b) Frequency of antigen-specific CD4+ T cells is shown for seven healthy controls (different symbols and shades of grey for each buffy coat). Plotted are tetramer-positive cells per million CD4+ T cells from all fourteen experiments (one technical replicate per healthy control).

Cut-off for positivity is one tetramer-positive cell per million CD4+ T cells, marked with a dotted line. c+d) Characterization of antigen-specific CD4+ T cells by differentiation status, determined by simultaneous or singular expression of CD45RA and CCR7 according to Sallusto et al (59) in naïve (Tnaïve), central memory (Tcm, coloured in red), effector memory (Tem, coloured in salmon) and CD45RA+ effector memory (Temra) T cells. We plotted the proportion of influenza- and citrulline-specific T cells among the four different phenotypes in (b) box plots showing the mean distribution and (c) scatter plots showing the detailed proportion and distribution of influenza- (left, open symbols) and citrulline-specific (right, closed symbols) T cells among the different phenotypes.

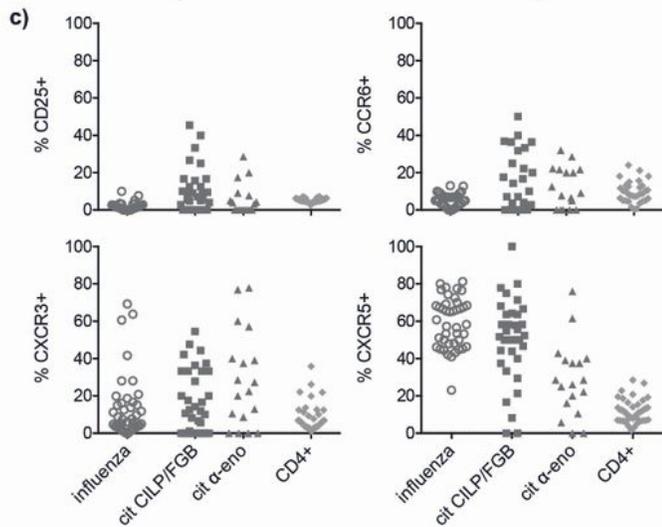
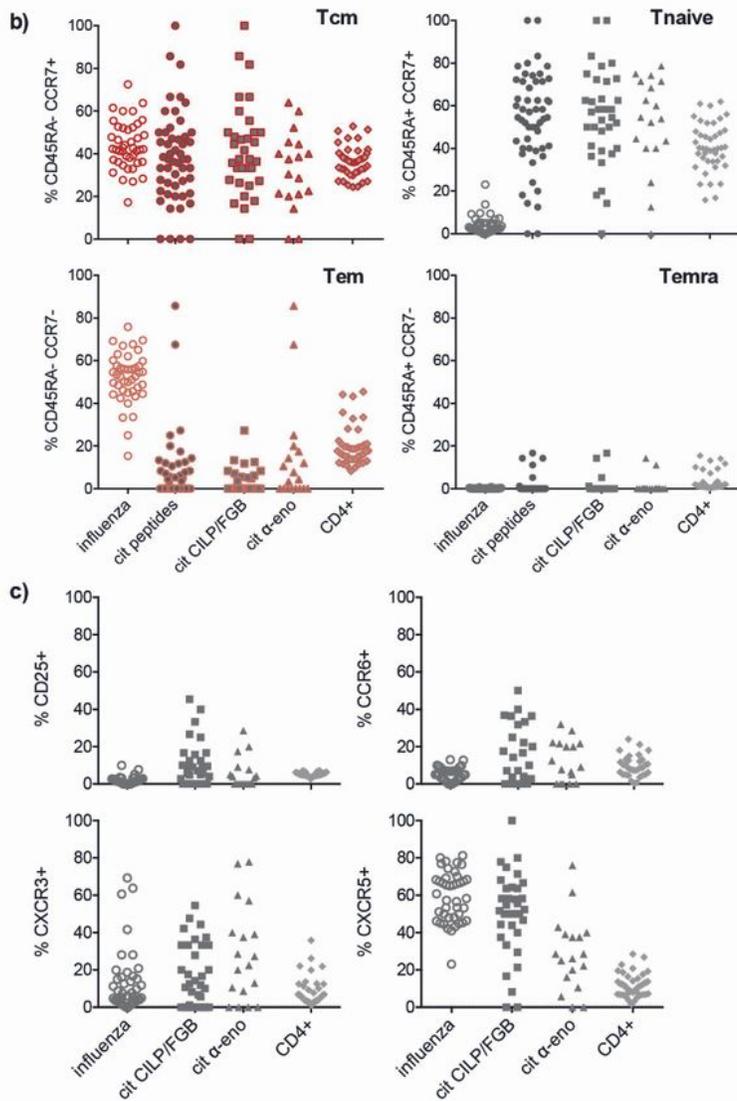
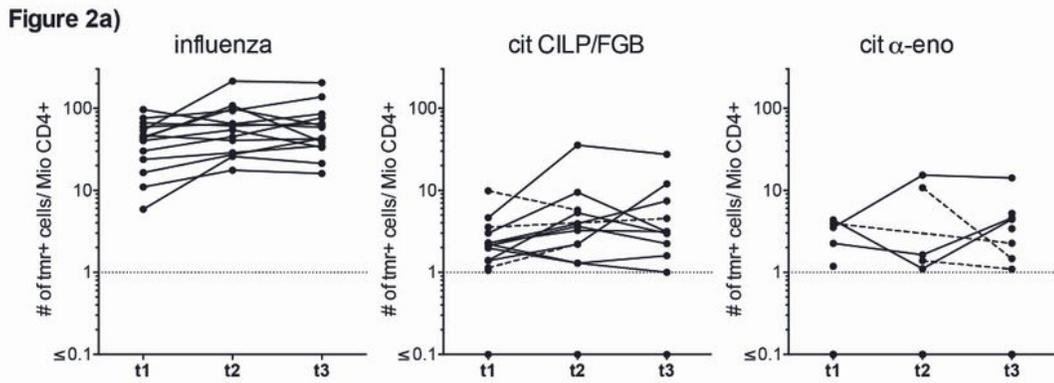


Figure 2

Autoreactive T cells in a cross-sectional cohort of RA patients a) Frequency of antigen-specific CD4+ T cells is shown for all 14 patients longitudinally. Plotted are tetramer-positive cells per million CD4+ T cells. Cut-off for positivity is one tetramer-positive cell per million CD4+ T cells and marked with a dotted line. The frequencies detected at the three time points (t1, t2, t3) are connected with a continuous line for each patient and with a dotted line in case there were no cells detected at one or two out of the three time

points. b+c) Phenotypic characterization of antigen-specific CD4+ T cells according to (b) simultaneous or singular expression of CD45RA and CCR7 and (c) expression of CD25, CXCR3, CXCR5 and CCR6. Plotted are frequencies from all 42 experiments (three per patient) for influenza- (open circles) and citrulline-specific (closed circles, not included in c) as well as for CILP/FGB- (squares) and α -enolase- (triangles) specific T cells and the general CD4+ population (light grey diamonds). Due to the setup of the experiment which resulted in there being up to three individual data points per patient plotted for each of the different antigen-specific T cell populations, we did not perform any specific statistical analysis.

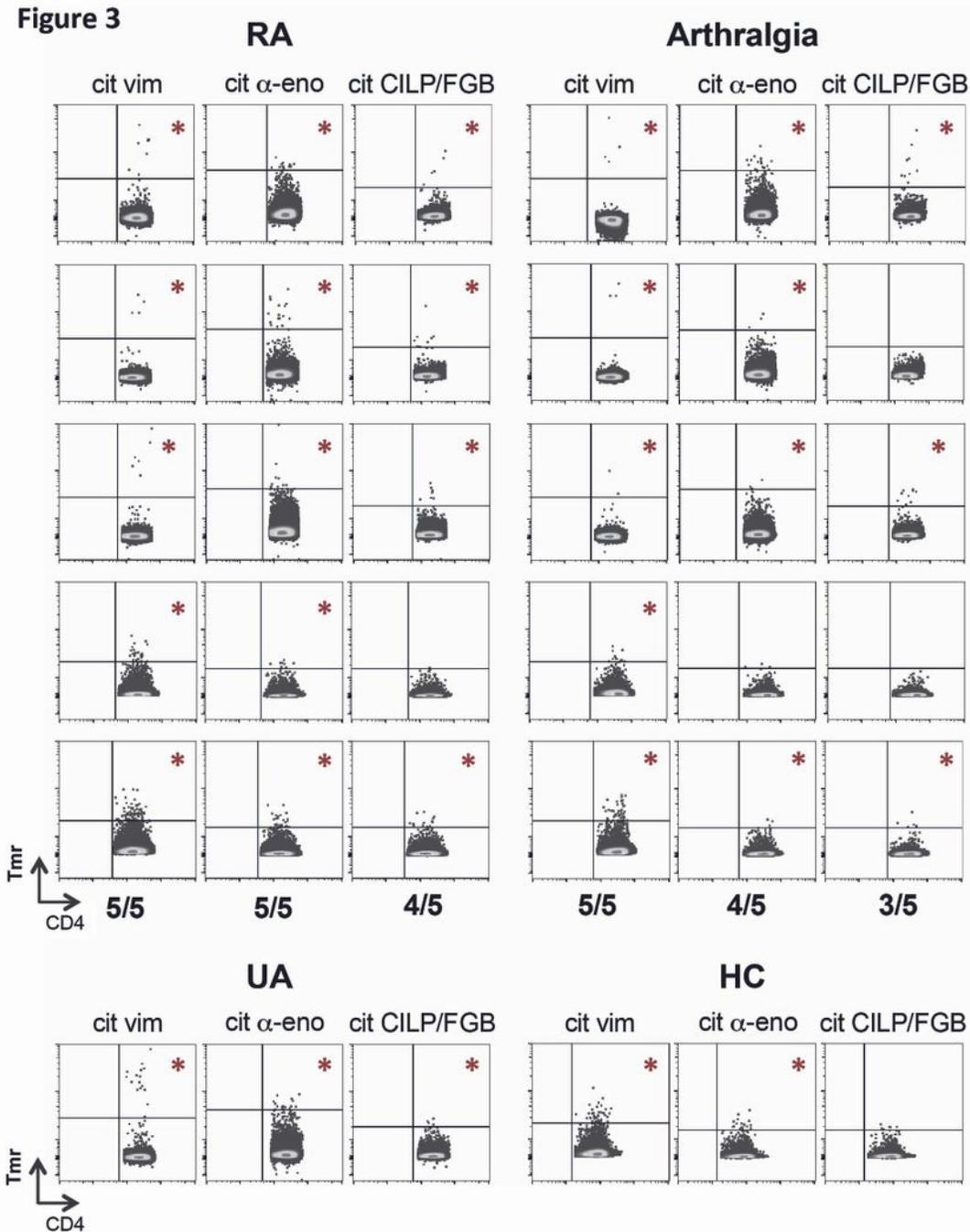


Figure 3

Citrulline-specific CD4⁺ T cells in LN biopsies from arthralgia and early RA patients Lymph node biopsies were stained for vimentin-, α -enolase- and CILP/FGB-reactive T cells after in vitro propagation. Displayed are plots from both RA and arthralgia patients (upper panel) and one patient with undifferentiated arthritis (UA) as well as one healthy control (HC, lower panel) showing tetramer-positive cells against CD4. Plots marked with a red asterisk show positive staining. The numbers of positive tested individuals/tested individuals in total is depicted below the plots for each specificity for both RA and arthralgia patients.

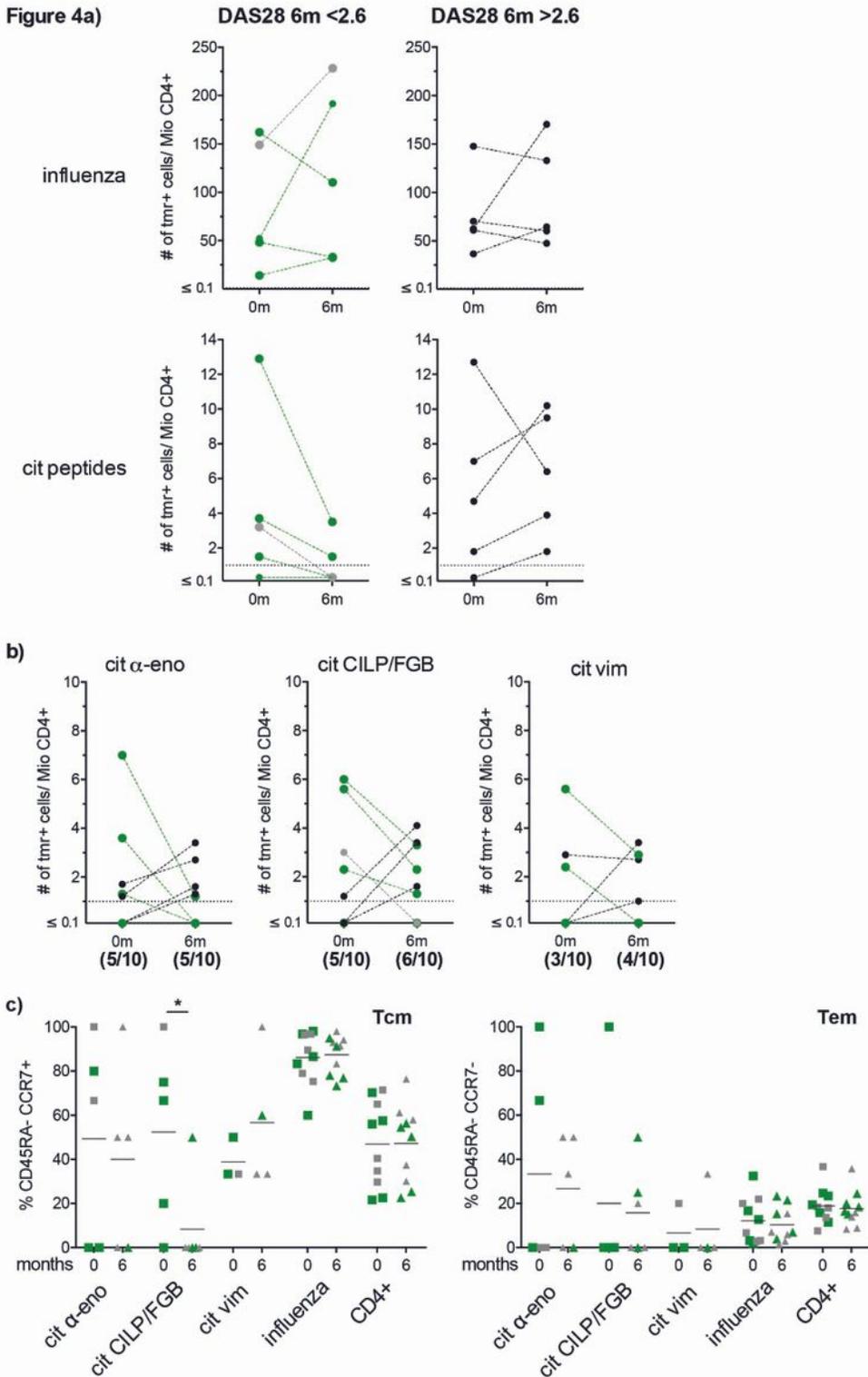


Figure 4

Early RA patients achieving drug-induced remission show a reduction in citrulline-specific T cells a+b) Frequency of (a) influenza- and citrulline-specific T cells in patients achieving remission (DAS28 at 6m <2.6) versus patients not achieving remission (DAS28 at 6m >2.6) and (b) of T cells reactive to α-enolase, CILP/FGB and vimentin is depicted at baseline (0m) and follow-up visit (6m) for 10 early RA patients. Plotted are tetramer-positive cells per million CD4+ T cells. Cut-off for positivity is one tetramer-positive

cell per million CD4+ T cells and depicted by a dotted line. Green symbols and lines depict the patients that achieved clinical remission at follow-up. The patient with missing DAS28-value at 6m is coloured in grey. The numbers of positive tested individuals/totally tested individuals for each time point is depicted below the graphs showing α -enolase-, CILP/FGB- and vimentin-specific T cells. c) Phenotypic characterization of antigen-specific CD4+ T cells according to simultaneous or singular expression of CD45RA and CCR7 in central memory (Tcm) and effector memory (Tem) T cells. Plotted are frequencies and the mean for α -enolase-, CILP/FGB-, vimentin- and influenza-specific T cells as well as the general CD4+ population at 0m (squares) and 6m follow-up (triangles). Green symbols depict the patients that achieved clinical remission at follow-up. Statistical analysis was performed using Mann-Whitney test when comparing the frequency of central memory T cells at baseline with the follow-up time point. p-values less than 0.05 were considered significant and marked with an asterisk.

Supplementary Files

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