

Endometrial Mesenchymal Stem/Stromal Cells: The Enigma to Code Messages for Generation of Functionally Active Regulatory T Cells

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Research

Keywords: MenSCs, Regulatory T cells, Proliferation, Endometrium, Pregnancy, Immunomodulation

Posted Date: June 1st, 2021

DOI: <https://doi.org/10.21203/rs.3.rs-559124/v1>

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Abstract

Background

Regulatory T cells (Tregs) play an important role in fine tuning immune responses and is pivotal for a successful pregnancy. Recently, the importance of mesenchymal stem cells in regulation of immune responses in general and Tregs in particular has been highlighted. Here, we hypothesized that menstrual stromal/stem cells (MenSCs) contribute to uterine immune system regulation through induction of functionally active Tregs.

Methods

MenSCs were collected from 18 apparently healthy women and characterized. Bone marrow mesenchymal stem cells (BMSCs) served as a control. The effect of MenSCs on proliferation of anti-CD3/CD28-stimulated T CD4 + cells and generation of Tregs with and without pre-treatment with mitomycin C, IFN- γ and IL-1 β was evaluated by flow cytometry. The potential role of IDO, PGE2, IL-6, IL-10, and TGF- β on proliferation of T CD4 + cells and generation of Tregs was assessed using blocking antibodies or agents. IDO activity was evaluated in MenSCs and BMSC culture supernatants by a colorimetric assay. IL-10 and IFN- γ production in MenSCs-primed T CD4 + was measured using intracellular staining. To investigate the functional properties of Tregs induced by MenSCs, Treg cells were isolated and their functional property to inhibit proliferation of anti-CD3/CD28-stimulated PBMCs was assessed by flow cytometry.

Results

According to the results, proliferation of T CD4 + lymphocytes were enhanced in the presence of MenSCs, while pre-treatment of MenSCs with pro-inflammatory reversed this effect. PGE2 and IDO were the major players in MenSCs-induced T cell proliferation. Non-treated MenSCs decreased frequency of Tregs, whereas after pre-treatment with IFN- γ and IL-1 β they induced Tregs, which was mediated through IL-6, IL-10, TGF- β and IDO. IFN- γ /IL-1 β -treated MenSCs induced IL-10 and IFN- γ production in CD4 + T cells.

Conclusion

Collectively, these finding indicates that immunomodulatory impact of menstrual blood stem cells (MenSCs) on generation of Tregs and inhibition of T cells proliferation is largely dependent of pre-treatment with IFN- γ and IL-1 β . This is the first report on immunomodulatory impact of MenSCs on Tregs and highlights the pivotal role of endometrial stem cells in regulation of local endometrial immune responses.

Background

Regulatory T cells (Tregs), as one of the main regulators of the immune system, play an important role in fine-tuning immune responses and preventing autoimmunity. The presence of Tregs in the uterus and their positive impact in successful pregnancy have been fully understood (1, 2). The frequency of circulatory Tregs in non-pregnant fertile women reaches to the highest level between days 9 to 13 of follicular phase and reduces during luteal phase of the menstrual cycle (3) which is believed to be essential to prepare the uterus for encountering the paternal antigens and possible implantation (4). Along with that, it is known that same alteration in Treg population can be observed in murine estrus cycle (5).

In very early stages of human pregnancy, systemic and local frequency of Tregs in decidua, lymph nodes and spleen are increased and reaches its peak on second semester of gestation (2, 6, 7). Treg frequency is then diminished throughout the weeks prior to the childbirth and a clear decrease is seen after the delivery, even though it is still slightly higher than non-pregnant women (2, 8). Frequency of murine endometrial Tregs peaks in the estrus phase and then declines during metestrus and remains low until the next estrus (5, 9). In pregnant mice, Tregs are quickly recruited to uterus draining lymph nodes and are activated during the first 2 days after the implantation (1). Tregs accumulate in thymus and draining lymph nodes in the early stages of mouse pregnancy and relocate to the periphery in later pregnancy stages (10). The importance of Tregs for successful pregnancy in mice is substantiated by exploiting an adoptive transfer model. In this model, transferring Treg-depleted T cell population to nu/nu BALB/c mice led to fetus rejection in allogeneic mating; whereas transferring total T cell population resulted in successful pregnancy (1).

Antigen-dependent and -independent mechanisms contribute to induction of Tregs during pregnancy resulting in immunological tolerance toward the fetus without endangering the integrity of the mother's immune system. The most important factors that triggers antigen-independent pathway of Tregs induction include estrogen (11), progesterone (12), PGE₂ (13) and TGF- β (14). In human, a sharp increase in the number of Tregs in follicular phase is strictly associated with high serum estradiol (E₂) level (3), which, not only can affect Treg Foxp3 mRNA expression but also can cause elevation of chemokines responsible for recruitment of Treg cells to the uterus. In parallel, progesterone can induce development of Tregs and suppresses differentiation of Th1 and Th17 cells (12, 15). In the antigen-dependent pathway, the role of seminal fluid in keeping antigenic and environmental signals is quite essential (16). Seminal fluid contains steroid hormones such as estrogen and testosterone, prostaglandin E (17) and TGF- β (18, 19) which induces uterine hypo responsiveness to sperm alloantigen's through induction of Tregs (16, 20).

Recently, the importance of mesenchymal stem cells in regulation of immune responses in general and Tregs in particular has been highlighted. Several studies have shown the ability of mesenchymal stem cells (MSCs) from different origins in induction of Tregs. BM-MSC are able to inhibit T cell proliferation and to shift T cell profile toward Tregs (21, 22). MSCs from other sources such as those derived from

adipose tissue also mediate generation of Tregs with immunosuppressive ability (23, 24). As one of the most regenerative tissues in the human body, the endometrium provides a unique source of adult stromal/stem cells. The immunomodulatory capacity of endometrial stromal/stem cells (eMSCs) has not been fully elucidated. Reportedly, this cell type produces different array of chemokines (25, 26) with capacity to recruit immature dendritic cells (26) and to inhibit proliferation of mouse spleen T lymphocyte stimulated with mitogen (27). eMSC population shed during menstruation into menstrual fluid. Menstrual blood stromal/stem cells (MenSCs) are newly-identified adult stem cells with multi-lineage differentiation capacity (28–33). In 2008, Murphy et al. proposed the immunomodulatory ability of MenSCs to suppress mixed lymphocyte reaction (MLR) and to attenuate pro-inflammatory responses (34), a finding which was supported by our investigations showing the suppressive effects of MenSCs on allogeneic MLR in a dose-dependent manner. (35). These cells share phenotypic and functional properties with other MSCs; yet have their own specificities (36, 37). Moreover, we also reported the potency of these cells to inhibit dendritic cell maturation (38). In a recent report, we showed that MenSCs are able to modulate functional features of natural killer (NK) cells and induce a pregnancy-friendly phenotype in these cells (39). Here, we hypothesized that MenSCs contribute to uterine immune system regulation through induction of functionally active regulatory T cells.

Materials And Methods

Sample collection

Menstrual blood was collected from 18 apparently healthy women aging between 25 and 35 years after getting signed inform consent. We were not able to do all experiments on each MenSCs sample because experiments had to be performed on MenSCs with low passage number. Menstrual blood samples did not yield a sufficient number of cells suffice to do all experiments simultaneously. Indeed, doing all experiments simultaneously was practically impossible. So, we did a part of the experiments on a set of samples and the rest of experiments with another sets. Nonetheless, not less than 6 samples were tested in any experiment. Participants had at least one live birth, no history of pregnancy complications including abortion and preeclampsia, no background diseases and no consumption of contraceptive, anti-inflammatory drugs, corticosteroids or vitamin D in the last three months before sampling. They also had negative test results for such blood transmittable viruses as HCV, HBV, and HIV and had no sign or symptoms of endometriosis and autoimmune diseases. Menstrual blood sample collection was performed as we described recently (40) in the second day of menstruation using Diva Cup (Lunette, Stockholm, Sweden) and immediately transported to the laboratory in a cold chain. Bone marrow-derived mesenchymal stem cells (BMSCs) from four healthy donors was obtained from Avicenna Research Institute (ARI) biobank and buffy coats were collected from Iranian blood transfusion organization (IBTO).

MenSCs isolation and culture

Isolation of MenSCs was carried out according to the protocol we described recently (41). In brief, menstrual blood samples were washed and cultured in DMEM/F12 supplemented with penicillin-streptomycin and 10% fetal bovine serum (FBS) (Gibco). After 48 hrs., non-adherent cells were removed and MenSCs were allowed to propagate. After confluency of about 80%, cells were detached using trypsin-EDTA (Gibco), and stored at liquid nitrogen. BMSCs were cultured in the same condition as with MenSCs. All experiments were performed on MenSCs and BMSCs at passages below five.

Multi-lineage Differentiation & Immunophenotyping of MenSCs and BMSCs

Differentiation of MenSCs toward osteocytes, chondrocyte, and adipocyte was carried out according to the protocols we published elsewhere (32, 33, 41). Immunophenotype of MSCs was examined by flow cytometry using a panel of antibodies against mesenchymal and embryonic stem cells and hematopoietic cell markers (Table I). All immunostainings were done by direct staining with fluorochrome-labelled antibodies except for Oct-4 which was done by two step indirect intracellular staining using rabbit anti-human Oct-4 antibody (Abcam, UK) and FITC-conjugated goat anti-rabbit Ig (Abcam, UK). Signals were analyzed using Attune NxT flow cytometer (Thermo Fisher Scientific, USA) in reference to appropriate isotype controls.

T CD4+ Isolation and MenSCs-T CD4+ co-culture

Peripheral blood mononuclear cells (PBMCs) were isolated from buffy coat using Ficoll-Hypaque density gradient medium (Biosera, USA). Upon harvesting and washing, CD4+ T cells was isolated from PBMCs using human T CD4+ cell Isolation kit, (Miltenyi Biotec, Germany) according to manufacturer's instruction. Isolated cells had viability and purity of >95% as assessed by trypan blue dye exclusion and CD4+ staining by flow cytometry. For MenSCs-T CD4+ co-culture, 2×10^5 freshly-isolated CD4+ T cells were co-cultured with titrating amounts of MenSCs in 24-well plates (Corning, USA) in a total volume of 1 mL/well. Culture medium consisted of RPMI-1640 (Gibco) supplemented with penicillin (100 µg/mL), streptomycin (100 IU/mL), L-glutamine (2 mM) (all from Sigma), non-essential amino acids, sodium pyruvate, and 10% FBS (all from Gibco). Several different settings were tested in MenSCs-T CD4+ co-culture. In one setting, MenSCs were remained untreated or mitotically inactivated with 25 µg/ml of Mitomycin C (Sigma) for 1 hr. before co-culturing. In parallel, to verify the effect of IFN-γ and/or IL-1β on MenSCs function, MenSCs were treated with either 25 ng/mL IFN-γ (BD, USA) and/or 10 ng/mL IL-1β (BD) 48 hrs. prior to co-culture with T CD4+ cells. At initiation of the co-culture, T cells were activated with activation beads containing 5 µg/mL anti-CD3 and 5 µg/mL anti-CD28 at 4:1 ratio (CD4+ T cell to bead) according to manufacturer's instruction (Miltenyi biotec, Germany). In the third day of co-culture, half of the culture media was gently replaced with fresh RPMI-1640, containing 100 U/ml recombinant IL-2 (R&D systems, USA). In most settings, BMSCs were co-cultures in parallel.

Measurement of IDO activity

A colorimetric method was used to determine the activity of IDO in MenSCs and BMSCs. MenSCs were treated with 100 ng/ml IFN-γ (BD) in the presence or absence of 10 ng/ml IL-1β (BD) or remained

untreated. Similarly, BMSCs were treated with IFN- γ or remained untreated as a control. Untreated cells or those treated with 100 $\mu\text{g}/\text{mL}$ L-Tryptophan (Trp) served as controls. The activity of IDO was measured as described earlier (42).

Proliferation Assay and Treg generation

To assess the effects of MenSCs on CD4⁺ T cells proliferation, CD4⁺ T cells were co-cultured in different settings as mentioned above. Before co-culturing, CD4⁺ T cells were washed and labeled with 5 μM 5,6-carboxyfluorescein diacetate succinimidyl ester (CFSE) (Molecular Probes, USA).

To investigate the effects of MenSCs secretome on CD4⁺ T cell proliferation, a transwell system (Corning, USA) was utilized. Briefly, IFN- γ /IL-1 β -treated MenSCs were seeded in the lower chamber of a 24-well transwell plate. Later, CD4⁺ T cells were added into the upper chamber and plates were incubated for five days in incubator. In some settings, 1 mM 1-Methyl-DL-tryptophan (Sigma, USA) as IDO blocker, 20 mM Indomethacin (Sigma, USA) as PGE2 inhibitor, 10 $\mu\text{g}/\text{ml}$ anti-IL-6 antibody (R&D, USA), 10 $\mu\text{g}/\text{ml}$ anti-IL10 antibody (R&D, USA) or 10 $\mu\text{g}/\text{ml}$ anti-TGF- β (R&D, USA) were added to the co-cultures in order to investigate the involvement of aforesaid molecules on modulatory effects of MenSCs on T cell proliferation or Treg generation. After 5 days, proliferation of CD4⁺ T cells and frequency of Tregs was assessed by flow cytometry.

Isolation of Tregs and functional assays

To investigate the percentage and functional properties of Tregs induced by co-culture of CD4⁺ T cells with MenSCs, Treg cells were isolated from 5-day co-cultures by Treg isolation kit (Miltenyi Biotec, Germany) according to manufacturer's instruction and their functional property to inhibit stimulated PBMCs was assessed. For this purpose, PBMCs autologous to the Tregs were first stained with CFSE and added to the wells of 96-well U bottom plates (Corning, USA) (5×10^4 cells/well/50 μl). Isolated Treg cells were then added to the wells at the ratios of 1:5 and 1:10 (Treg to PBMCs) in a volume of 50 μl /well. Finally, activation/expansion beads (Miltenyi biotec, Germany) containing 5 $\mu\text{g}/\text{ml}$ anti-CD3 and 5 $\mu\text{g}/\text{ml}$ anti-CD28 at 1:2 ratio (bead to PBMC) were added to the wells and total volume of each well was adjusted to 200 μl . In the second day of culture, 100 μl of culture medium was gently replaced with fresh culture medium containing 200 U/ml IL-2 (R&D systems, USA) and incubation was continued for 2 days at %CO₂ incubator. Proliferation of PBMCs was then measured by flow cytometry.

Flow Cytometry

Flow cytometry experiments were performed to assess T cells co-cultured with MenSCs. The fluorochrome-conjugated antibodies included Alexa Fluor 647 anti FoxP3 (BD Bioscience, USA), FITC anti-human IFN- γ , PerCP/Cy5.5 anti-CD4, APC anti-human IL-10, and PE anti-CD25 (All from Biolegend, USA). Antibodies were used at the concentrations recommended by the manufacturers. For assessment of cytokines, cells were treated with 50 $\mu\text{g}/\text{ml}$ PMA (Sigma, USA), 1 $\mu\text{g}/\text{ml}$ ionomycin (Sigma, USA) and 0.7 $\mu\text{g}/\text{ml}$ Monensin (BD Bioscience, USA) 6 hrs. before staining procedure. For intracellular staining of

Foxp3 and cytokines, transcription factor buffer set (BD Bioscience, USA) was used for cell permeabilization and fixation according to the manufacturer's instruction. Cells were also stained with Live/Dead fixable near red fluorescent dye (Molecular Probe, USA) to separate the alive and dead populations before antibody-specific gating. The stained cell was then read by flow cytometer Attune NxT flow cytometer and analyzed by FlowJo software (FlowJo, LLC, USA).

Statistics

All statistical analyses and group comparisons were performed using GraphPad Prism 8. Data were compared with Mann-Whitney tests. Descriptive data were reported as mean \pm standard error of the mean (SEM). P values less than 0.0332 were interpreted as statistically significant. P values less than 0.0332, 0.0021, 0.0002, and 0.0001 were shown as *, **, ***, and ****, respectively.

Results

Characterization of MenSCs

MenSCs isolated from menstrual blood were characterized by multi-lineage differentiation and immunophenotyping. A considerable fraction of MenSCs were able to differentiate toward osteocyte, chondrocyte and adipocyte, as judged by specific staining's (Fig.1A). Cells grown in the absence of induction media failed to differentiate. MenSCs expressed markers of mesenchymal origin, but failed to express hematopoietic-specific markers. Unlike BMSCs, MenSCs expressed Oct-4 but were negative for SSEA-4 (Fig.1B). The results obtained here were almost the same with those we reported earlier (40, 43).

Pro-inflammatory cytokines are needed to induce inhibitory capacity of MenSCs on T cell proliferation

The capacity of MenSCs to modulate CD4⁺ T cell proliferation was tested at different conditions by flow cytometry. First, the impact of Mitomycin C and IFN- γ treatment of MenSCs on T cell proliferation was assessed. Untreated MenSCs were shown to increase proliferation of CD4⁺ T cells compared to CD4⁺ T cells cultured alone ($p < 0.0002$). Mitomycin C inactivation of MenSCs had no effect on increased proliferation of CD4⁺ T cells co-cultured with MenSCs. IFN- γ treatment of MenSCs decreased their capacity to induce T cell proliferation compared to untreated MenSCs ($p < 0.0332$) (Fig.2B). Expectedly, IFN- γ -treated BMSCs strongly inhibited T cell proliferation ($p < 0.0021$). However, inactivation of BMSCs with Mitomycin C prior to IFN- γ -treatment reversed this effect and caused significant T cell proliferation ($p < 0.0021$). Based on these findings, all subsequent tests were done without Mitomycin treatment. Next, we tested the effect of MenSCs on CD4⁺ T cells proliferation at different ratios. As with our earlier report (44), we observed that MenSCs significantly promoted T cell proliferation compared to the biological control at 1:2-1:8 ratios ($p < 0.0002$ - 0.0001) (Fig.3A). Regarding the simultaneous presence of multiple pro-inflammatory cytokines in early pregnancy decidua, the effect of simultaneous treatment of MenSCs with IFN- γ and IL-1 β on T cell proliferation was tested at the next step. As depicted on Fig.3B, simultaneous treatment of MenSCs with IFN- γ and IL-1 β inhibited proliferation of CD4⁺ T lymphocytes compared to the biological control ($p < 0.0002$ - 0.0001) at 1:2-1:8 ratios. In order to evaluate the effect of IFN- γ - and IL-1 β -

treated MenSCs on the proliferation of CD4⁺ T lymphocytes without cell-cell contact, transwell plates were used. As shown in Fig.3C, although the proliferation of CD4⁺ T cells in the absence of cell-cell contacts was inhibited, the differences were not significant.

Inhibition of PGE2 & IDO increased the proliferation of CD4⁺ T cells

To explore the mediators responsible for inhibition of T cell proliferation by pre-treated MenSCs, different blockers including anti-IL-6, anti-IL-10, TGF- β , indomethacin, 1 methyl tryptophan (1MT) or their double combinations were added to the MenSCs-T cell co-cultures at 1:8 ratio. Untreated MenSCs and MenSCs treated IFN- γ and IL-1 β served as untreated and Treated controls, respectively. As illustrated in Fig.4A, anti-IL-6, anti-IL-10 and combination of anti-IL-10 and anti-TGF- β failed to restore T cell proliferation. In contrary, treatment of MenSCs with indomethacin or double combination of indomethacin and 1-MT significantly restored T cell proliferation to the level observed in Treated control ($p < 0.0002$).

IFN- γ /IL-1 β -treated MenSCs induced IL-10 and IFN- γ production in CD4⁺ T cells

To see how MenSCs affect the balance of TH1/TH2, the levels of intracellular IL-10 and IFN- γ were measured in CD4⁺ T cells after co-culture with MenSCs at 1:8 ratio (MenSCs:CD4⁺ T cells). As showed in Fig.4C MenSCs significantly ($p < 0.0021$) increased production of both cytokines in CD4⁺ T lymphocytes.

MenSCs treated with IFN- γ and IL-1 β induced Tregs

In the next step, the effect of untreated MenSCs on induction of Tregs was examined. T cells cultured alone served as biological control. As shown in Fig.5B, untreated MenSCs decreased the frequency of Tregs at 1:4 and 1:8 co-culture ratios ($p < 0.0002$). The mean percentage of Tregs decreased from (3.21 ± 0.05) to (2.16 ± 0.24) and (2.49 ± 0.15) when T cells were co-cultured with untreated MenSCs at 1:4 and 1:8 ratios, respectively. Based on the results presented above, effect of IFN- γ and IL-1 β treatment of MenSCs on induction of Tregs was tested in the next step. As shown in Fig.5C, MenSCs treated with IFN- γ and IL-1 β significantly augmented Treg frequency at all co-culture ratios tested ($p < 0.0001$). The percentage of Tregs increased two folds when T cells were co-cultured with MenSCs pre-treated with pro-inflammatory cytokines.

MenSCs-induced Tregs were functionally active to inhibit PBMC proliferation

Functional properties of Tregs generated in the presence of MenSCs was tested by inhibition of PBMC proliferation. Wells containing only Tregs (generated in the absence of MenSCs), PBMC or MenSCs-induced Tregs served as negative controls. Co-cultures containing Tregs (generated in the absence of MenSCs) and PBMC were considered as positive control. Tregs were added at 1:5 and 1:10 ratio to the CFSC-labeled PBMC cultures activated with activation beads. All control wells showed only a negligible level of cell proliferation. As shown in Fig.5D, MenSCs-induced Tregs were able to significantly ($p < 0.0021$) inhibit proliferation of PBMCs in both 1:5 and 1:10 ratios with a mean percentage of 19.50 ± 0.98 and 30.73 ± 0.89 compared to positive control groups 33.90 ± 0.40 and 37.1 ± 0.40 , respectively.

Blocking of IL-6, IL-10 and TGF- β decreased MenSCs ability to induce Tregs

To find the cytokines involved in the induction of Tregs after co-culture with IFN- γ /IL-1 β -treated MenSCs, co-cultures at 1:8 ratios (MenSCs:CD4 $^+$ T cells) were treated with blocking antibodies against IL-6, IL-10 or TGF- β or with indomethacin or 1-MT to inhibit PGE2 and IDO, respectively. Differentiation of CD4 $^+$ T cells toward Tregs in each group was compared to the control group containing the same co-culture without blocking agents. As illustrated in Fig.6A-B, treatment with blocking antibodies against IL-6, IL-10 and TGF- β or 1-MT significantly decreased the percentage of Tregs (3.06 ± 0.35 , 2.65 ± 0.38 , 2.91 ± 0.40 , and 3.40 ± 0.42 respectively vs. 4.34 ± 0.14 in biological control group) ($p < 0.0021$). Co-treatment with anti-IL-10 and anti-TGF- β further decreased the percentage of Tregs (2.07 ± 0.33 , $p < 0.0002$). However, inhibition of PGE2 did not affect Treg induction. Co-treatment with both blockers had no significant effect on the induction of Tregs.

Pre-treatment of MenSCs with inflammatory cytokines induced IDO activity

To study the activity of IDO secreted by MenSCs, these cells were pre-stimulated with IFN- γ as a potent inducer of IDO. We observed that treatment of MenSCs with IFN- γ and IL-1 β or IFN- γ alone could induce IDO activity ($p < 0.0001$) (Fig.6C). Treatment of MenSCs with IL-1 β alone did not induce IDO activity in these cells. As expected, IFN- γ -treated BMSCs produced a considerable amount of active IDO. Untreated cells or those treated with tryptophan did not exert a significant IDO activity.

Discussion

Despite tremendous effort to unravel the immunology of reproduction in species with hemochorial placentation, the mystery of uterus as being a privileged site during pregnancy has not been resolved yet. Several hypotheses have been put forth so far and a dozen of mediators with immunosuppressive properties have been introduced to explain the state of uterine immune system hypo responsiveness during pregnancy (45). Nonetheless, with a closer view to the pathways and networks proposed in this regard, it became clear that many of them are interconnected and failure or dysfunction in one pathway might affect the proper functioning of the other pathways. Up to now, it remained unclear which component in the uterus is the main trigger of the delicate uterine immune regulatory network during pregnancy.

In recent years the central role of mesenchymal stem cells in regulation of immune responses has been highlighted. BM-MSCs have been proved to exert an array of immune regulatory activities including inhibition of T cell proliferation and skewing T cell profile toward Tregs (21, 22). MSCs from other sources such as those derived from adipose tissue also mediate generation of Tregs with immunosuppressive ability (23, 24). Human endometrium is a dynamic tissue with ability for hundreds of cycles of growth, differentiation and shedding during reproductive age. It contains a population of mesenchymal stem/stromal cell (eMSC) which shed during menstruation. While eMSC and MenSCs have several biological features with BM-MSCs, they also show some differences in immunophenotype, functional aspects, and differentiation capacities (46). We recently showed that MenSCs are able to inhibit

functional features of NK cells and TH17 differentiation to exert a pregnancy-friendly phenotype in this cell population (40, 43). Here we were about to explore how biological features of Tregs, as the central cells in pregnancy tolerance, are affected by MenSCs.

We observed that MenSCs promoted proliferation of CD4⁺ T cells in a dose-dependent manner. This results was in line with proliferation promoting effect of MenSCs on NK and T cells (40, 44). In our previous study, using protein microarray, we showed that MenSCs condition media contained considerably lower level of IGFBP1-4 (Insulin-like growth factor-binding protein) compared to BMSCs leaving higher functional free from of IGFs to exert trophic activity (40). We showed that blocking IL-6 in the co-culture system resulted in reduced proliferation of T cells indicating supportive role of this cytokine for T cell proliferation. IL-6 has been shown to have a particular role in survival of resting and activated T cells (47, 48) and prevent apoptosis in T cells through a mechanism which is completely independent of IL-2 (48).

Several molecules have been reported to be involved in MSC-mediated immunoregulation. In this regard, pro-inflammatory cytokines exert a substantial role. IFN- γ , alone or in combination with other pro-inflammatory cytokines TNF- α , IL-1 α or IL-1 β have profound effects on stimulation of immunoregulatory phenotype in MSC (49, 50). Therefore, in the next step we examined the potential effect of pro-inflammatory cytokines on MenSCs-induced T cell proliferation. We observed that IFN- γ treatment alone did not prevented proliferation of T cells. However, combinational treatment of MenSCs with IFN- γ /IL-1 β significantly reduced T cell proliferation. It was demonstrated that human embryonic stem cell-derived mesenchymal stromal cells (hES-MSCs) has no significant effect on lymphocyte proliferation despite pretreatment with IFN- γ (51). Depending on IFN- γ dose, umbilical cord blood mesenchymal stem cells have been reported to induce or suppress allogenic lymphocytes after pretreatment with IFN- γ (52). Responsiveness to pro-inflammatory cytokine treatment is largely dependent on MSC source. We showed that pre-treatment of BMSCs with the same concentration of IFN- γ resulted in substantially reduced T cell proliferation. Recent studies show that compared to BMSCs, MenSCs are less responsive to IFN- γ due to fewer expression of IFN- γ R1 and IFN- γ R2 (50). Some studies show that Adipose-derived stem cells (ASCs) could gain promoted inhibitory properties in the presence of IFN- γ , IL-1 β , TNF- α and IFN- α (53). Our results showed that MenSCs have to be pre-treated with both IFN- γ and IL-1 β to gain the capacity to suppress T cell proliferation. We also showed that, in contrast to conventional co-culture system, MenSCs pre-treated with IFN- γ /IL-1 β did not significantly suppress T cell proliferation in a transwell system. Reportedly, IFN- γ along with TNF- α or IL1 β increased ICAM-1 and VCAM-1 expression and strengthen the attachment of MSCs and T cells in co-culture. Inhibition of these two molecules led to decreased suppressive effects of MSCs (54, 55). In this regard, it seems that cell-cell contact plays a significant role in the inhibitory function of IFN- γ /IL-1 β -treated MenSCs on T cell proliferation.

To investigate factors contributed in MenSCs-induced suppression of T cell proliferation following IFN- γ /IL-1 β treatment, IL-6, TGF β , IL-10, PGE₂, and IDO were blocked. We found that PGE₂ was involved in suppression of T cells proliferation in co-culture system. PGE₂ is a crucial player in chronic inflammation and could influence the functions of DC, NK, and lymphocytes. High concentrations of PGE₂ inhibits T

lymphocytes proliferation through reducing both IL-2 and IL-2 receptor synthesis (56). It is also shown that inhibition of PGE2 reduced the immunosuppressive effects of human cord blood-derived mesenchymal stem cells (CB-MSCs) on T lymphocytes proliferation (57). Simultaneous pre-treatment of MenSCs with IFN- γ and IL-1 β resulted in increased expression of COX-2 (50). Moreover, IFN- γ and IL-1 β are well-known inducers of PGE2 because both of them can stimulate COX-2 mRNA expression (58, 59). Inhibition of IDO had not a significant effect on T cell proliferation. This data is in line with Gieske et al. that reported anti-proliferative properties of Human multipotent mesenchymal stromal cells (MSCs) is independence of IDO (60). Similarly, inhibition of TGF- β alone showed negligible effect on the proliferation of co-cultured CD4 + T cells. This finding is in line with what reported earlier for umbilical cord MSCs (58). However, we showed simultaneous inhibition of IL-10 and TGF- β or inhibition of IL-10 alone could significantly reduce proliferation of CD4 + T cells. In this regard, Harizi and Gualde reported that IL-10, as an anti-inflammatory cytokine, can inhibit COX-2 induction by inhibiting protein expression and thus PGE2 production in BMDC (Bone marrow-derived DC) (61). As mentioned above, PGE2 plays a pivotal role in suppressive activity of MSCs, therefore by blocking IL-10, PGE2 production could be enhanced leading to further suppression of T cell proliferation. Nonetheless, based on the complex nature of *in vivo* microenvironment, it is evident that none of these molecules has an exclusive role and that MSC-mediated immunoregulation is mediated by a combination of several molecules which do not exert their effects in the same direction.

One potential way that MSCs cells could inhibit proliferation of immune cells is their ability to induce Treg cells. Therefore, we next aimed to investigate the effect of MenSCs on induction of Treg lymphocyte population. We found that in the absence of inflammatory cytokines, MenSCs did not induce Tregs, neither did they decrease their frequency. MenSCs can secrete a baseline levels of IL-6 and TGF β in proinflammatory environment (62) which could be increased in an allogeneic cell culture system. In a mouse GVHD model, inhibition of the IL-6 signaling pathway markedly reduced pathologic damage through a significant increase in the absolute number of Tregs (63). Indeed, IL-6 could induce STAT3-mediated T cell production of IL-10, a cytokine with Treg induction capacity (64). Similarly, our results showed that higher levels of IL-10 were produced in MenSCs-T cell co-culture supernatant and that inhibition of IL-10 or TGF- β significantly inhibited the induction of Tregs. The crucial impact of these cytokines on Treg generation has already been reported. Moreover, induction and propagation of Tregs by MSCs are closely related to IL10 and TGF- β cytokines (65). We showed that treatment of MenSCs with IFN- γ and IL-1 β led to increased level of IDO generation, a finding which is in line with our previous findings (42). IDO dominantly controls the generation and functional status of Tregs in response to inflammatory stimuli which could potentially lead to generation of Tregs.

Finally, we showed that Tregs generated in the presence of MenSCs are functionally active and could prevent proliferation of PBMCs activated with anti-CD3/CD28 antibodies and IL-2. Similar results have also been reported for other sources of MSCs either in mixed lymphocyte reaction (22, 66–68). It seems that higher immunosuppressive phenotype in Tregs generated in the presence of MSCs is due to the induction of these cells in an inflammatory milieu.

Conclusion

Collectively, we showed here that, in contrast to other sources of stem cells, MenSCs have a supportive effect on the proliferation of CD4 + T lymphocytes and reduced percentage of Tregs unless they were first primed with pro-inflammatory cytokines. Pro-inflammatory cytokines-primed MenSCs could enhance functionally active Tregs and suppress T cell proliferation, a finding which support their role as the main regulator of the endometrial immune microenvironment during pregnancy.

Abbreviations

Tregs: Regulatory T cells; MenSCs: Menstrual stromal/stem cells; BMSCs: Bone marrow mesenchymal stem cells; IDO: Indoleamine-pyrrole 2,3-dioxygenase; PGE2: Prostaglandin E2; IL-6: Interleukin 6; IL-10: Interleukin 10; TGF- β : Transforming growth factor beta; IFN- γ : Interferon gamma; IL-1 β : Interleukin 1 beta; eMSCs: Endometrial stromal/stem cells; MLR: Mixed lymphocyte reaction; NK: Natural killer cell; HCV: Hepatitis C virus; HBV: Hepatitis B virus; HIV: Human immunodeficiency virus; Oct-4: Octamer-binding transcription factor 4; PBMCs: Peripheral blood mononuclear cells; IL-2: Interleukin 2; CFSE: 5,6-carboxyfluorescein diacetate succinimidyl ester; SSEA-4: Stage-specific embryonic antigen 4; 1-MT: 1-Methyl tryptophan; TH1: Type 1 T helper cells; TH2: Type 2 T helper cells; IGFBP: Insulin-like growth factor-binding protein; IGFs: Insulin-like growth factors; ASCs: Adipose-derived stem cells; TNF- α : Tumor necrosis factor alpha; hES-MSCs: Human embryonic stem cell-derived mesenchymal stromal cells; AT-MSCs: Adipose tissue-derived multipotent stromal cells; ICAM-1: Intercellular adhesion molecule 1; VCAM-1: Vascular cell adhesion protein 1; DC: Dendritic cell; CB-MSCs: Cord blood-derived mesenchymal stem cells; COX-2: Cyclooxygenase-2; BMDC: Bone marrow-derived DC; GVHD: Graft versus host disease; STAT3: Signal transducer and activator of transcription 3; DSCs: Decidua-derived stem cells

Declarations

Acknowledgements

We would like to express our sincere thanks to department of immunology, Tehran university of medical sciences (TUMS) which provided all facilities needed to perform the experiments conducted in this research project.

Author Contributions

M.A., collection and/or assembly of data, data analysis, manuscript writing. M.B. and S.N., data analysis and interpretation, A.G., M.R.S., and S.M., collection and/or assembly of data, F.S., and A.H.Z., conception and design, data analysis and interpretation, manuscript writing, final approval of manuscript.

Funding

This research was supported by grants from Tehran University of Medical Sciences (No:95-04-27-33245) and Avicenna Research Institute (No: 940402-002).

Availability of data and materials

All the datasets used or analyzed in this study are available from corresponding author on reasonable request.

Ethics approval and consent to participate

Ethics Committee for medical research of the Avicenna Research Institute (ARI) approved the study (Ethical code: 940402-002). All participants gave informed consent before menstrual blood donation.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

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Tables

Table 1
Antibody Panel for Immunophenotyping of MenSCs

Antibody	Fluorochrome	Clone	Company
Anti-CD9	FITC	M-L13	BD Bioscience
Anti-CD10	PE	H110a	BD Bioscience
Anti-CD29	PE	MAR4	BD Bioscience
Anti-CD34	FITC	581	BD Bioscience
Anti-CD38	FITC	HIT2	BD Bioscience
Anti-CD44	PE	515	BD Bioscience
Anti-CD45	PE	HI30	BD Bioscience
Anti-CD73	PE	AD2	BD Bioscience
Anti-CD105	PE	166707	BD Bioscience
Anti-CD133	PE	W6B3C1	BD Bioscience
Anti-Oct-4	-	Polyclonal	Abcam
Anti-SSEA-4	-	MC813-70	BD Bioscience

Figures

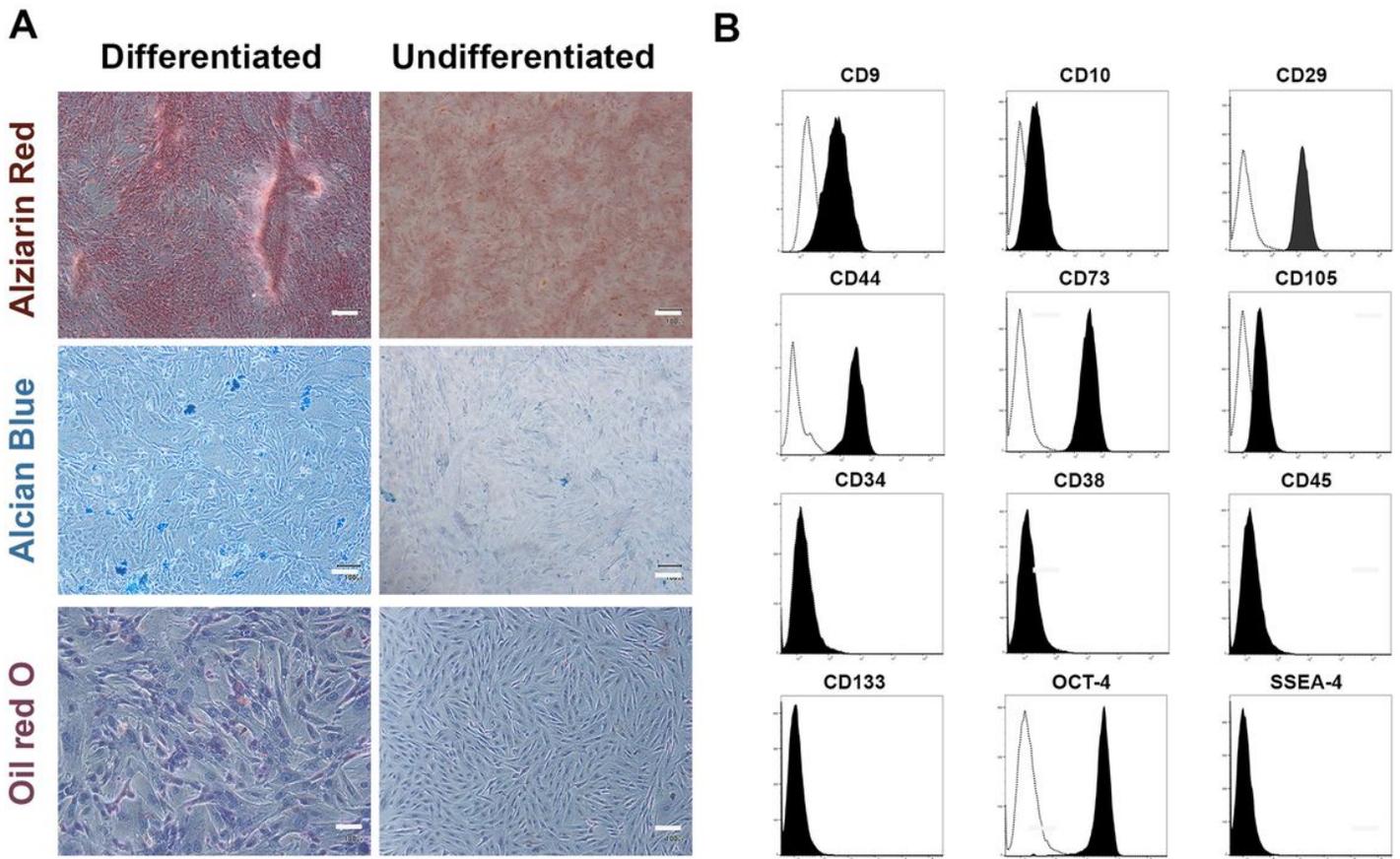


Figure 1

Differentiation of MenSCs into adipocytes, osteoblasts and chondrocytes. MenSCs were differentiated toward adipocytes, osteoblasts and chondrocytes and the extend of differentiation was evaluated using Alizarin red, Alcian blue and Oil Red O staining, respectively. Undifferentiated cells served as controls (A). Immunophenotyping of MenSCs was carried out using a panel of mesenchymal and hematopoietic markers (B).

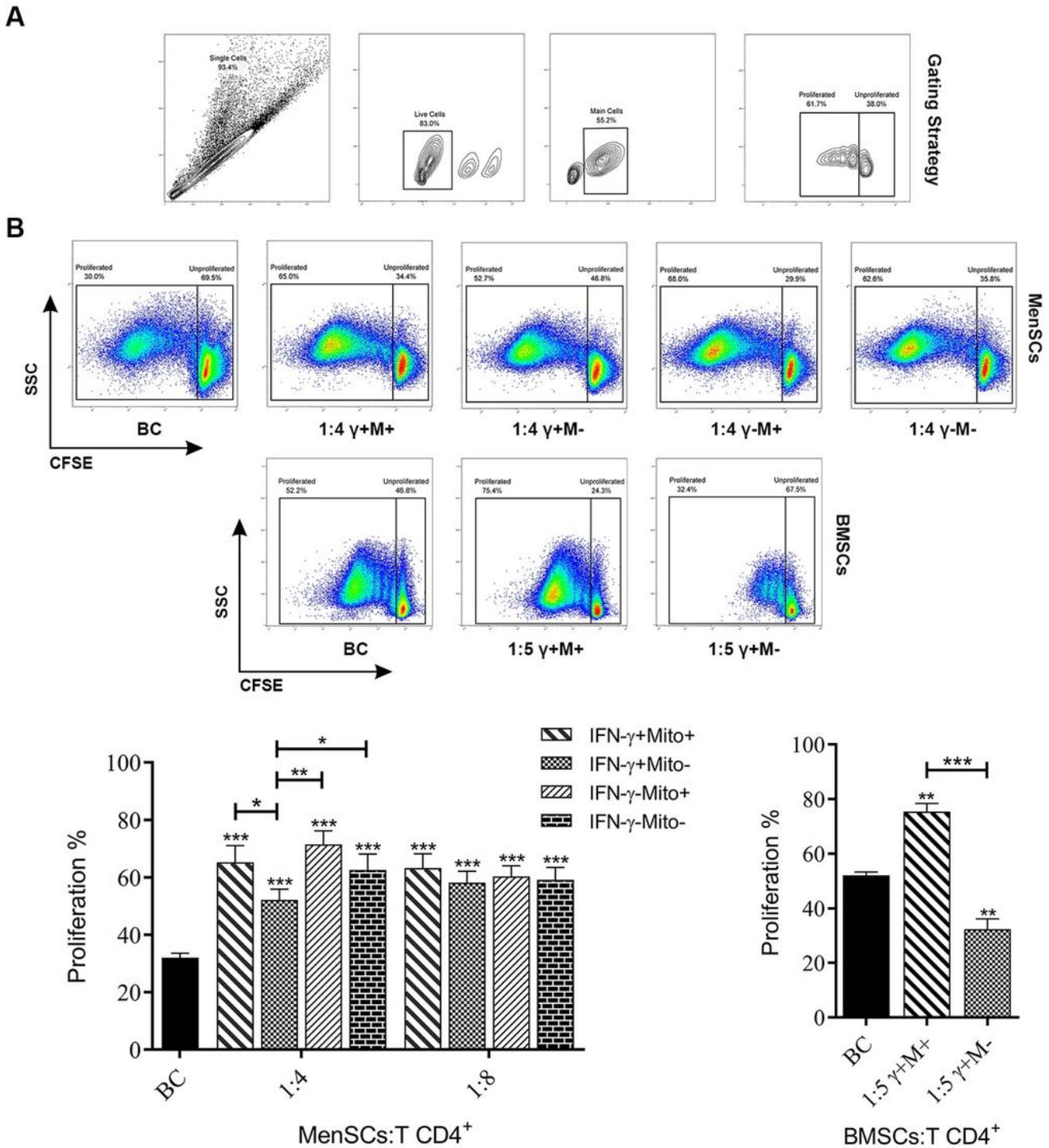


Figure 2

MenSCs induced T cell proliferation. MenSCs were either inactivated with Mitomycin C or remained intact. Some wells were also primed with IFN- γ prior to co-culture with T cells. CFSE-labeled purified CD4⁺ T cells were then co-cultured with MenSCs at different ratios and activated with T cell activation beads. The extent of T cells proliferation was then tested after five days by flow cytometry. The effect of BMSCs on T cell proliferation was also tested in the same conditions. Gating strategy is shown in panel (A).

Representative flow cytometry dot plots and bar charts for MenSCs and BMSCs are shown in (B). T cells cultured in the absence of MenSCs or BMSCs served as biological control (BC). Data was expressed as mean \pm SEM of 8 experiments. The signs (*), (**), (***) and (****) stand for $p < 0.0332$, $p < 0.0021$, $p < 0.002$, and $p < 0.0001$, respectively.

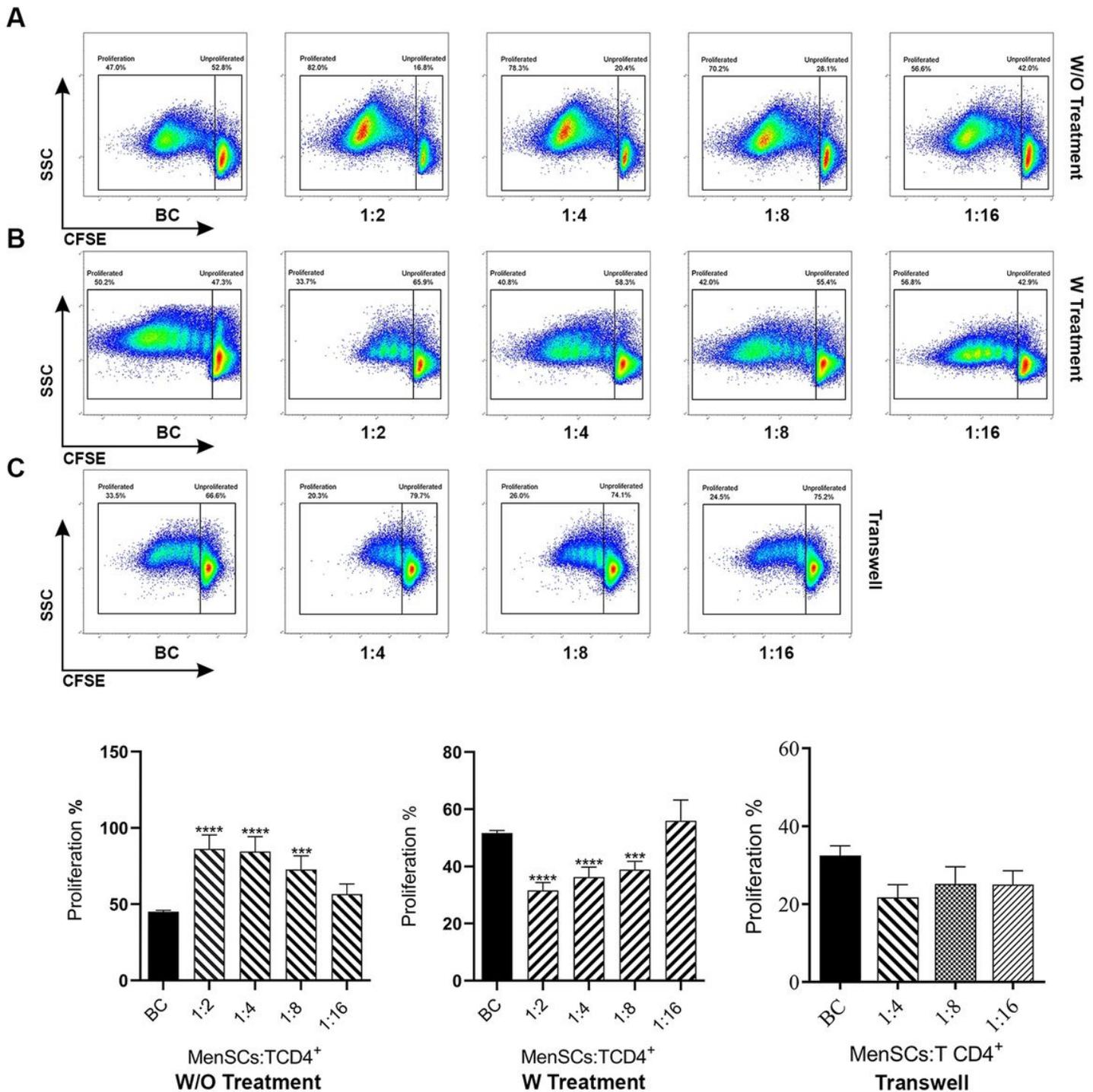
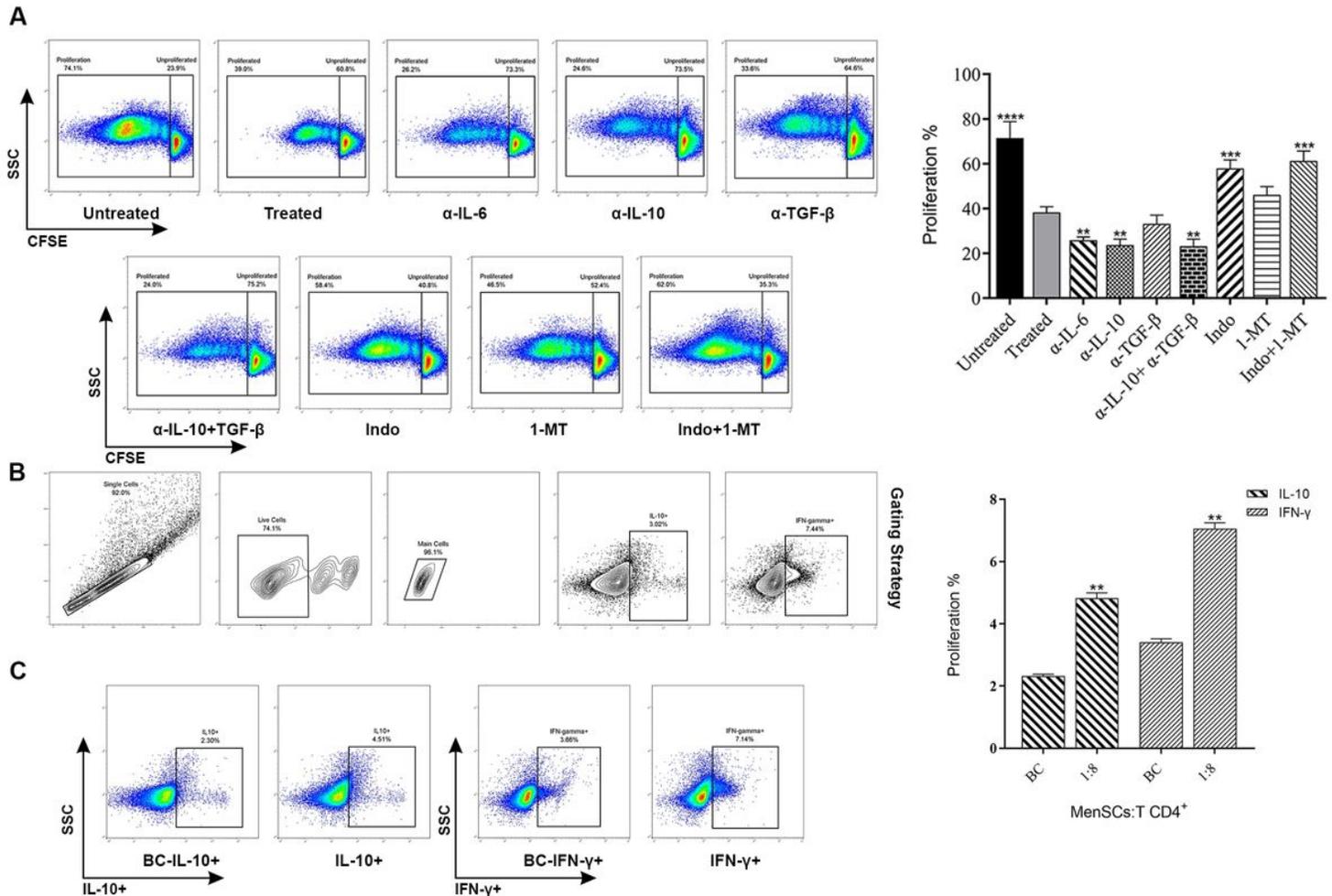


Figure 3

MenSCs pre-treated with IFN- γ and IL-1 β inhibited T cell proliferation. MenSCs were either remained untreated (A) or primed with IFN- γ and IL-1 β (B). CFSC-labeled purified CD4⁺ T cells were then co-cultured

with MenSCs at different ratios and activated with T cell activation beads. Proliferation of T cells was then tested after five days by flow cytometry. In order to evaluate the effect of secretome of IFN- γ /IL-1 β -treated MenSCs on CD4 $^+$ T cell proliferation, transwell plates were used with the aforesaid settings (C). T cells cultured in the absence of MenSCs served as biological control (BC). Data was expressed as mean \pm SEM of 10 experiments. The signs (*), (**), (***) and (****) stand for $p < 0.0332$, $p < 0.0021$, $p < 0.0002$, and $p < .0001$, respectively.



control. The results were displayed as mean \pm SEM of 8 different experiments. The signs (*), (**), (***), and (****) stand for $p < 0.0332$, $p < 0.0021$, $p < 0.0002$, and $p < .0001$, respectively.

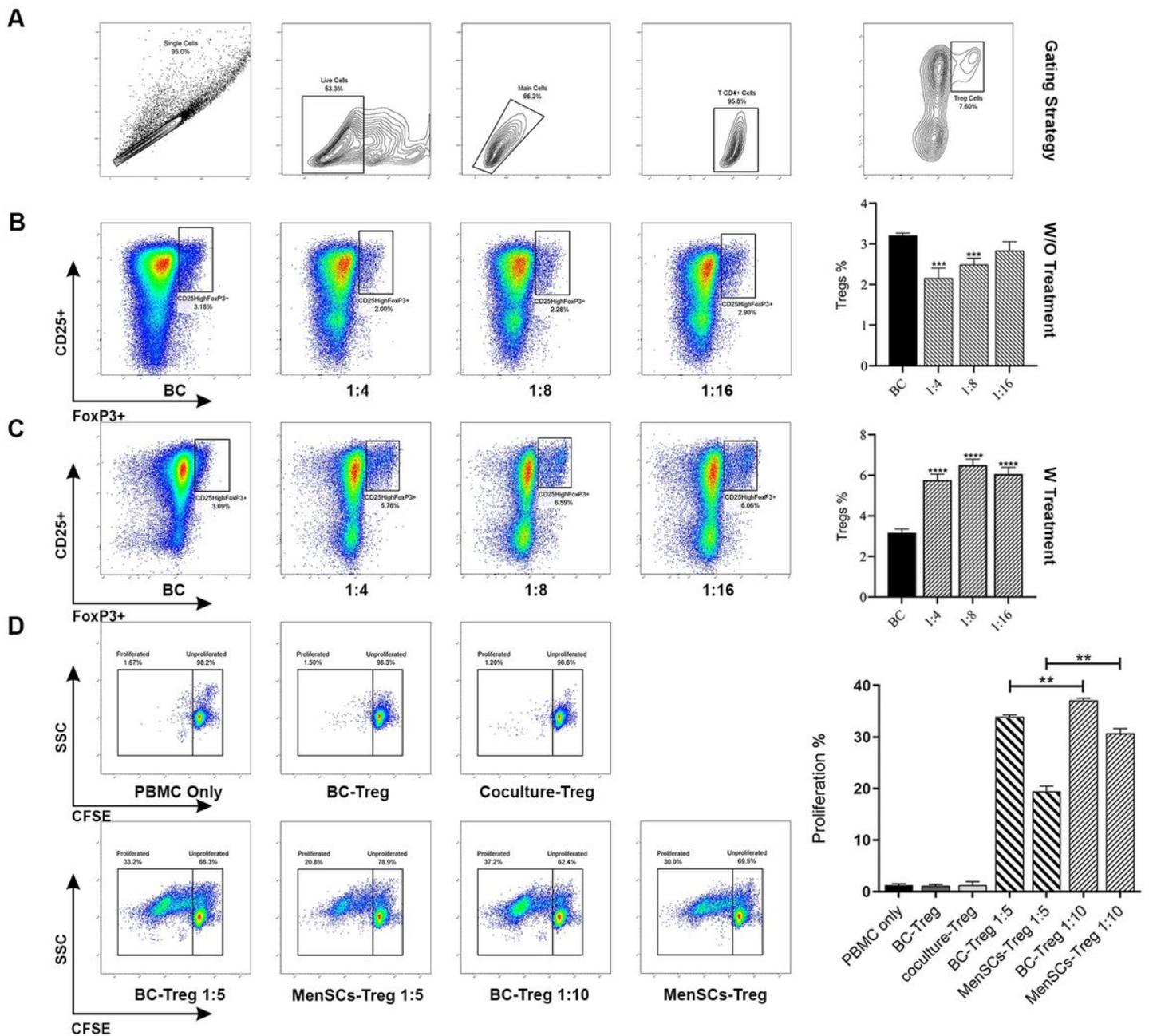


Figure 5

MenSCs pre-treated with IFN- γ and IL-1 β augmented functional Treg generation. Purified CD4⁺ T cells were co-cultured with MenSCs at different ratios and activated with T cell activation beads. After five days, percent of CD4⁺CD25^{high}FOXP3⁺ Tregs was quantified by flow cytometry. Gating strategy for Tregs is shown in panel (A). MenSCs were either remained untreated (B) or primed with IFN- γ and IL-1 β (C). T cells cultured in the absence of MenSCs served as biological control (BC). The data obtained from each group were compared with biological control, and the data was expressed as mean \pm SEM of 12 experiments. IL-1 β /IFN- γ -treated MenSCs were co-cultured with CD4⁺ T cells for five days and induced

Tregs were then isolated by MACS. Tregs were added at 1:5 and 1:10 ratio to the CFSC-labeled autologous PBMC cultures activated with activation beads and proliferation of PBMC were measured after 4 days by flow cytometry (D). Wells containing only Tregs (generated in the absence of MenSCs, BC-Treg), PBMC or MenSCs-induced Tregs served as negative controls. Co-cultures containing Tregs (generated in the absence of MenSCs) and PBMC were considered as positive control. Results were displayed as Mean \pm SEM of 6 different experiments. The signs (**) and (***) stand for $p < 0.0021$, and $p < 0.0002$, respectively.

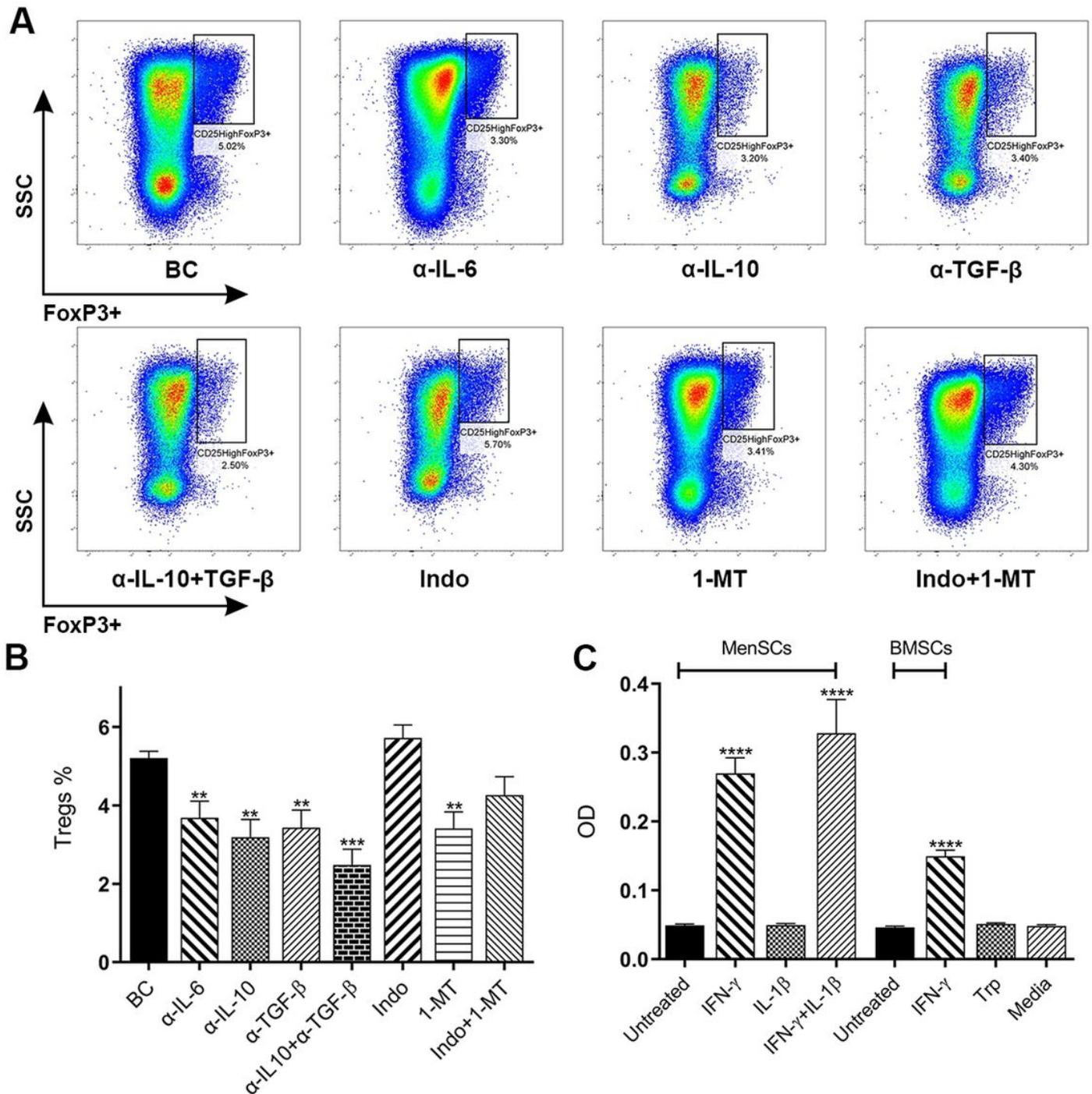


Figure 6

IL-6, IL-10 and TGF- β were involved in MenSCs-mediated Treg generation. MenSCs treated with IL-1 β and IFN- γ were co-cultured with CD4⁺ T lymphocytes at 1:8 ratio and activated with T cell activation beads. Different blockers were added to the co-cultures. After five days, percent of CD4⁺CD25^{high}FOXP3⁺ Tregs was quantified by flow cytometry (A-B). T cells cultured in the absence of MenSCs served as biological control (BC). The data obtained from each group were compared with biological control, and the results were displayed as mean \pm SEM of 8 experiments. Functional activity of IDO in MenSCs was tested using a colorimetric assay (C). Cells were treated with IFN- γ and/or IL-1 β for 48 hrs. IFN- γ -treated BMSCs served as positive control. Wells containing untreated cells or cells treated with Trp were considered as negative controls. Data was expressed as mean \pm SEM of 16 different samples of MenSCs and BMSCs. The signs (*), (**), and (***) stand for $p < 0.0332$, $p < 0.0021$, and $p < 0.0001$, respectively. Indo: Indomethacin, 1-MT: 1-methyl tryptophan, Trp: Tryptophan.