

A neutralizing monoclonal antibody-based competitive ELISA for classical swine fever C-strain post-vaccination monitoring

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Abstract

Background: Virus neutralization test (VNT) is widely used for serological survey of classical swine fever (CSF) and efficacy evaluation of CSF vaccines. However, VNT is a time consuming procedure that requires cell culture and live virus manipulation. C-strain CSF vaccine is the most frequently used vaccine for CSF control and prevention. In this study, we presented a neutralizing monoclonal antibody (mAb) based competitive enzyme-linked immunosorbent assay (cELISA) with the emphasis on the replacement of VNT for C-strain post-vaccination monitoring. Results: One monoclonal antibody (6B211) which has potent neutralizing activity against C-strain was generated. A novel cELISA was established and optimized based on the strategy that 6B211 can compete with C-strain induced neutralizing antibodies in pig serum to bind capture antigen C-strain E2. By testing C-strain VNT negative pig sera (n=445) and C-strain VNT positive pig sera (n=70), the 6B211 based cELISA showed 100% sensitivity (95% confidence interval: 94.87 to 100%) and 100% specificity (95% confidence interval: 100 to 100%). The C-strain antibody can be detected in pigs as early as 7 days post vaccination with the cELISA. By testing pig sera (n=139) in parallel, the cELISA showed excellent agreement ($\text{Kappa}=0.957$) with VNT. The inhibition rate of serum samples in the cELISA is highly correlated with their titers in VNT ($r^2=0.903$, $p<0.001$). In addition, intra- and inter-assays of the cELISA exhibited acceptable repeatability with low coefficient of variations (CVs). Conclusions: This novel cELISA demonstrated excellent agreement and high level correlation with VNT. It is a reliable tool for sero-monitoring of C-strain vaccination campaign because it is a rapid, simple, safe and cost effective assay that can be used to monitor vaccination-induced immune response at the population level.

Background

Classical swine fever (CSF) is a highly contagious viral disease of swine, including wild (feral) and domestic pigs [1]. In many regions of the world, CSF is still endemic and is regarded as one of the major problems in pig industry [2]. CSF has the potential to cause devastating epidemics, particularly in countries free of the disease such as the United States [3, 4]. CSF control is primarily dependent on vaccination and the conventional Chinese vaccine (C-strain) is considered as one of the most effective vaccines because of its safety, quick protective immune response, cross-protection against challenge of CSF viruses from different genotypes, and ability to be used for oral immunization of wild boars [2, 5-9].

The etiologic agent of CSF, classical swine fever virus (CSFV), is one member of the genus *Pestivirus* of the *Flaviviridae* family [10]. CSFV genome is comprised of a single large open reading frame (ORF) coding for a polyprotein encompassing all the viral proteins: four structural (C, E^{ns}, E1, and E2) and eight nonstructural viral proteins (N^{pro}, p7, NS2, NS3, NS4A, NS4B, NS5A, and NS5B) [11, 12]. The envelope glycoprotein E2 is responsible for eliciting neutralizing antibodies which are protective against virulent CSF virus and is also the target antigen for development of CSF vaccines, molecular and serological tests [13-16]. High sequence variability has been found in E2 protein among CSFVs. Based on the full-length E2 gene sequences, CSFV isolates could be divided into three genotypes (1, 2, and 3) as well as 11 subgenotypes (1.1-1.4, 2.1a, 2.1b, 2.1c, 2.1d, 2.2, 2.3, and 3.4) [17].

Virus neutralization test (VNT) is considered as the gold standard for serological monitoring and efficacy evaluation of CSF vaccines. However, it has several limitations including time-consuming, requirement of cell culture, the need for live virus manipulation, and relatively expensive [2, 18-21]. Here, we described a competitive ELISA (cELISA) developed with a neutralizing anti-E2 monoclonal antibody. This novel cELISA is a rapid, simple, safe and cost effective approach for detection of C-strain CSF vaccine-induced immune response.

Results

Generation of suitable capture antigen and competitive monoclonal antibodies

The envelope glycoprotein E2 of C-strain CSFV was successfully expressed in insect cells by using Bacto-Bac® Baculovirus Expression System. The purified C-strain E2 protein mainly exists as homodimers (the native dimeric conformation) under non-reducing condition with a molecular weight of ~90 kDa (Fig.1A).

To generate suitable mAb for the cELISA, purified C-strain E2 protein was used as immunogen for mAb production using Balb/c mice. All mice maintained good physical health and no adverse event happened during the experiments. Spleen cells from one mouse with the highest anti-E2 antibody titer were collected for fusion. One panel of more than 5 mAbs against C-strain E2 protein was generated. After assessment by VNT, mAb 6B211 (IgG1 and kappa chain) showed the most potent neutralizing activity against C-strain CSFV. 6B211 only react with homodimer of E2 protein and cannot recognize the reduced proteins, which indicate that it recognize the conformational epitope of C-strain E2 protein (Fig. 1B). The neutralization titer (neutralization doses 50%, ND₅₀) of its purified supernatant (1mg/ml with 1,920 ND₅₀) is much higher than that of the commercial neutralizing E2 monoclonal antibody WH303 (1mg/ml with 480 ND₅₀) (Fig. 2A). In addition, 6B211 lacks cross-reaction with other viruses in genus *Pestivirus* such as Bovine viral diarrhea virus (BVDV) (Fig. 2B).

Establishment of competitive ELISA based on C-strain E2 protein and 6B211

Optimal concentrations of capture protein (C-strain E2) and competitive mAb (horseradish peroxidase conjugated 6B211, HRP-6B211) were determined by a systematic checkerboard procedure (Fig. 3A). A concentration of 0.625 µg/ml of C-strain E2 protein and a concentration of 1.25 µg/ml of HRP-6B211 were chosen as they consistently produced an OD₄₅₀ value around 1.7 and at a point in the linear range in the standard curve. 2% fetal bovine serum (FBS) in phosphate buffered saline (PBS) containing 0.05% Tween 20 (PBST) was chosen as the optimal blocking buffer as it consistently outperformed other blocking solutions (non-fat dry milk and BSA) in terms of high signal to noise ratio. To minimize the volume of serum required and background noise, serum dilution of 1:5 was chosen as the preferred dilution over lower dilutions that performed only slightly better (Fig. 3B). These conditions were used in all subsequent cELISA experiments.

Reproducibility of the cELISA

The reproducibility of the cELISA was determined by calculating the coefficient of variation (CV) of the percent of inhibition (PI) values by testing 20 C-strain VNT negative pig serum samples and 20 C-strain VNT positive pig serum samples. The intra-assay CVs of the C-strain VNT positive samples ranged from 0.02 to 1.68%. The inter-assay CVs of those same samples ranged from 1.02 to 8.66%. The intra-assay and inter-assay of the C-strain VNT negative samples also exhibited excellent repeatability, showing 0.01–0.35% and 0.66–4.29%, respectively (Table 1).

Table 1. Coefficient values of the samples tested by 6B211 based cELISA

Pig sera	No. of sera tested	CV range (%)		Comparison of 6B211 based cELISA with VNT
		Intra-assay	Inter-assay	
C-strain VNT positive	20	0.02-1.68	1.02-8.66	
C-strain VNT negative	20	0.01-0.35	0.66-4.29	

A total of 139 pig serum samples were tested using the established cELISA and VNT in parallel. Results (Table 2) showed that cELISA and VNT had an excellent agreement (Kappa=0.957) in identifying positive (from pigs vaccinated with C-strain vaccine) and negative samples. McNemar's test revealed that there was no significant difference in the results between the cELISA and VNT ($P>0.1$). The Pearson correlation coefficient between the inhibition rate in 6B211 based cELISA and titers in VNT was calculated (Fig.4) based on testing results of the 139 samples. The inhibition rate is highly correlated with VNT titers for these samples ($r^2=0.903$, $p<0.001$).

Table 2. Comparison of 6B211 based cELISA with VNT

	cELISA	VNT
Number positive	66	69
% Positive	47.5%	49.6%
Number negative	73	70
% Negative	52.5%	50.4%
Result agreement (Kappa)	0.957	
Significance	$P>0.1$	

Note. Significance: $P < 0.05$ = tests are significantly different at the 95% confidence level.

Standardization of the cut-off value of 6B211 based cELISA

A total of 515 pig serum samples were used to standardize the cut-off value of the established cELISA. Among them, 445 samples were C-strain VNT negative and 70 samples were C-strain VNT positive (21 to

56 days post vaccination, DPV). After testing these samples by the established cELISA, distributions of the cELISA PI values showing the frequency of positive and negative samples are calculated and shown in Fig. 5. The mean PI value (x-axis) of the negative sera detected by cELISA was -0.59%. When mean PI of negative sera plus two standard deviation (SD, 4.03%) was used as threshold, the sensitivity and specificity of the cELISA were 100% (95% confidence interval: 94.87 to 100%) and 98.43% (95% confidence interval: 96.79 to 99.37%), respectively. When mean PI of negative sera plus three SD (6.19%) was used as threshold, the sensitivity and specificity of the cELISA were 100% (95% confidence interval: 94.87 to 100%) and 100% (95% confidence interval: 100 to 100%), respectively.

Kinetics of antibody response of pigs at different time intervals post-vaccination

Serial serum samples (0 to 56 DPV at every 7 days) derived from two pigs vaccinated with C-strain were tested by the established cELISA. The C-strain antibody could be tested in both pigs as early as 7 DPV with inhibition values 24% and 56%, respectively. Significant increase of inhibition was observed between 7 DPV and 28 DPV. The levels of antibody titer kept relatively stable from 28 DPV to 56 DPV with inhibition values ranging from 68% to 86% (Fig.6).

Discussion

In this study, we developed a neutralizing mAb based cELISA with the emphasis on the replacement of VNT for C-strain post-vaccination monitoring. Several monoclonal antibody-based cELISAs have been developed for detection and titration of antibodies against different viruses. One study developed a cELISA based on a mAb to a neutralizing epitope of hemagglutinin protein of *peste des petits ruminants virus* (PPRV). Efficacy of the cELISA compared very well with VNT having high relative specificity (98.4%) and sensitivity (92.4%). Findings suggest that the cELISA test developed can easily replace VNT for sero-surveillance, sero-monitoring, diagnosis from paired sera samples and end-point titration of PPRV antibodies [22]. Another study developed a cELISA for detecting antibodies against canine distemper virus (CDV) and phocine distemper virus (PDV) in sera from dogs and various species of marine mammals. The authors suggested that as cELISA proved to be nearly as sensitive and specific as the VNT while being simpler and more rapid, it would be an adequate screening test for CDV or PDV cases [23]. The glycoprotein E2 exposed on the outer surface of CSFV is the major immuno-protective antigen of C-strain vaccine that is responsible for inducing neutralizing antibodies and eliciting protective immunity against CSFV [13-20]. Thus, it is not surprising that E2 protein has been successfully used to develop ELISAs to measure anti-CSFV antibody response in pigs after vaccination [13-16]. In this study, the purified insect cell expressed C-strain envelope glycoprotein E2 was used as the capture antigen. The insect cell expressed E2 provides proper refolding, post-translational modification, and oligomerization [24], which guaranteed the immunodominant epitopes of E2 protein exposed in their native state.

The mAb 6B211 showed potent neutralizing activity against C-strain CSFV and bound to a specific conformational epitope on C-strain E2 protein (Fig.1B & Fig.2A). In addition, it showed very high sensitivity when tested by IFA (mouse ascites of 6B211 produced a strong positive fluorescence signal at

dilution 1:16,000, data not shown). To test the hypothesis that neutralizing 6B211 can compete with C-strain induced neutralizing antibodies in pig serum to bind the capture antigen (C-strain E2), 445 C-strain VNT negative pig serum samples and 70 C-strain VNT positive pig serum samples were tested by the 6B211 based cELISA. The 6B211 based cELISA can effectively differentiate the C-strain VNT positive and C-strain VNT negative samples (Fig.5), which indicate that 6B211 can compete with C-strain induced neutralizing antibodies from pigs and can be used as the competitive antibody for cELISA.

By testing the pig sera in parallel and using mean PI of negative sera plus three SD as the threshold, the sensitivity and specificity of the cELISA were 100% (95% confidence interval: 94.87 to 100%) and 100% (95% confidence interval: 100 to 100%), respectively. The C-strain antibody can be tested in pigs as early as 7 DPV with the cELISA. The excellent agreement (Kappa=0.957, Table 2) between cELISA and VNT, and the high correlation VNT ($r^2=0.903$) between inhibition rate in cELISA and VNT titers of tested samples indicate that the 6B211 based cELISA can replace the VNT for C-strain post-vaccination monitoring. In addition, the intra-assay CVs and the inter-assay CVs of the 6B211 based cELISA (Table 2) are lower than 10% when tested with the negative and E2 antibody positive pig serum samples, which indicate that the established cELISA is repeatable with acceptable variations.

Conclusions

Through the aforementioned experiments and analysis, we concluded that the mAb 6B211 based cELISA showed excellent agreement and high correlation with the VNT. This cELISA is a reliable, rapid, simple, safe, and cost-effective tool for sero-monitoring of C-strain vaccination at a population level. We believe that the cELISA presented in this study could be used to assist in CSF control and eradication.

Methods

Animals

Five female Balb/c mice (six weeks old) weighing between 22 and 25g were purchased from Charles River Laboratories, Inc. Wilmington, MA, USA. The mice were fed with standard commercial diet and housed in a clean facility at the Kansas State University. Animal care and protocols were approved by Institutional Animal Care and Use Committee (IACUC#3517) at Kansas State University. All animal experiments were done under strict adherence to the IACUC protocols.

Generation of monoclonal antibody to C-strain E2 protein

Expression and purification of C-strain E2 protein using a baculovirus expression system were performed as previously described [18]. The purified C-strain E2 protein was concentrated using Amicon Ultra Centrifugal Filters 30,000 NMWL (Millipore, Billerica, USA) and measured using BCA assay kit (Pierce, USA) according to the manufacturer's recommendations.

For mAb production, 50 μ l (1 μ g/ μ l) purified E2 protein plus equal volume of 2% Alhydrogel (Invitrogen, CA, USA) was used as an immunogen to inject each of the Balb/c mice via intraperitoneal injection. Three booster immunizations with same dose were conducted at two week intervals. Three days after the final booster injection, one mouse with the highest anti-E2 antibody titer was humanely euthanized using carbon dioxide (CO₂) in euthanasia chambers and sprayed with 70% ethanol. Spleen cells were collected and fused with the mouse myeloma partner SP2/0-Ag14 (ATCC, MD, USA) by using polyethylene glycol 1500 (Boehringer Mannheim, IN, USA) at a ratio of 10:1. The hybridoma cells were maintained in RPMI1640 medium (Gibco, NY, USA) with 20% fetal bovine serum (FBS, Hyclone, UT, USA). Supernatants from growing hybridomas were screened by an ELISA for reactivity to E2 protein as previously described [18]. The positive hybridoma clones were subcloned three times by limiting dilution until monoclonal antibodies were obtained. Characterization of these monoclonal antibodies will be published elsewhere. One mAb, designated 6B211 was used in this study. Its isotype was classified with an antibody-isotyping kit (Roche Diagnostics Corporation, IN, USA). The reaction of 6B211 with native and β -mercaptoethanol treated C-strain E2 protein was analyzed by western blot as we described previously [18].

Indirect fluorescent antibody assay (IFA) test

The reactivity of mAb with different BVDVs was tested by IFA as described previously [25]. Briefly, MDBK cells (bovine kidney cells, ATCC, CCL22) grown in 96-well plate were infected with BVDVs (BVDV-32 strain, genotype 1; BVDV-0427 strain, genotype 1; BVDV-AV6 strain, genotype 1; BVDV-125 strain, genotype 2) at a multiplicity of infection (MOI) of 0.1 for 3 days. Cells were fixed in cold acetone and washed two times with PBST. Supernatant of hybridomas (1:50 diluted) was added and plates were incubated at 37°C for 1 hour (hr). Plates were washed three times with PBST and Alexa Fluor 488 goat anti-mouse IgG (H+L) (Life Technologies, MA, USA) was added at 1:200 dilution to each well and incubated at 37°C for 1 hr. Finally, the plate was washed three times with PBST and examined under a fluorescence microscope.

Neutralizing antibody test

Purified mAbs 6B211 (1mg/ml) and WH303 (1mg/ml, Animal and Plant Health Laboratories Agency, Weybridge, United Kingdom) were first diluted five-fold and then serially diluted two-fold. The diluted samples (in duplicate) were incubated with 100 TCID₅₀ (50% tissue culture infective dose) of CSFV C-strain in DMEM with 10 % FBS for 1 hr at 37 °C. Residual virus infectivity was determined by adding 1.0 \times 10⁴ ST cells to each well with serum-virus mixture in 96-well plate and incubated at 37 °C for 3 days. The cells were subjected to immunofluorescence staining with E2-specific mAb WH303 and Alexa Fluor 488 goat anti-mouse IgG (H + L) (Life Technologies, MA, USA). Neutralizing antibody titers (NAT) were expressed as the reciprocal of the highest dilution that caused 50 % neutralization.

Competitive enzyme-linked immunosorbent assay (cELISA)

The 6B211 was purified by HiTrap™ Protein G column (GE Healthcare Life Sciences, PA, USA) followed by conjugating with Horseradish Peroxidase (HRP) using EZ-Link™ Plus Activated Peroxidase (Thermo

Scientific, NJ, USA) according to the manufacturer's instruction. The HRP-6B211 was dialyzed with Slide-A-Lyzer Dialysis Cassettes (Thermo Scientific, NJ, USA) against PBS and stored in Pierce™ Peroxidase Conjugate Stabilizer (Thermo Scientific, NJ, USA).

The systematic checkerboard procedure was used to optimize the concentration of capture antigen and HRP-6B211. The optimal dilution of serum and blocking solution were determined experimentally. The established cELISA was performed in Corning® 96 Well Clear Flat Bottom Polystyrene High Bind Microplate (Corning, NY, USA). Briefly, plates were coated overnight with C-strain E2 (0.625µg/ml, 100µl/well) in PBS (without calcium and magnesium, pH7.4, Thermo Scientific, NJ, USA) at 4°C. After washing three times with PBST, the plates were blocked with blocking buffer by incubating at 37°C for 1 hr; after washing, 50 µl of diluted serum samples and 50 µl of diluted HRP-6B211 were added to each well and mixed well by pipetting. The plates were incubated at 37°C for 1 hr. After washing five times, 100 µl of room-temperature TMB Stabilized Chromogen (Invitrogen, CA, USA) were added and incubated at room temperature (RT) for 10 min; after adding 100 µl/well of 2N Sulfuric Acid (Ricca Chemical Company, TX, USA), the absorbance at 450 nm were obtained using SpectraMAX microplate reader (Molecular Devices, CA, USA). The OD₄₅₀ of the samples were converted to a percent inhibition (PI) value using the following formulation: $PI (\%) = (OD_{450} \text{ value of negative controls} - OD_{450} \text{ value of sample}) / OD_{450} \text{ value of negative controls} \times 100\%$.

The cut-off value that served as the threshold to separate VNT positive sera from VNT negative sera was determined by testing negative sera of unvaccinated pigs and VNT positive sera of C-strain or C-strain E2 subunit vaccinated pigs (21-56 DPV).

Serial derived serum samples after vaccination with C-strain from 0 to 56 DPV at every 7 days were used for testing the kinetics of antibody titers of pigs (n=2) at different time intervals post-vaccination.

Reproducibility and statistical analysis of the cELISA

Inter-assay and intra-assay reproducibility for the established cELISA was evaluated by testing CSFV antibody negative (n=20) serum samples and C-strain VNT positive pig serum samples (n=20). For the intra-assay reproducibility, each serum sample (in duplicate) was detected by the same batch of pre-coated ELISA plates. For the inter-assay reproducibility, each serum sample was detected by three batches of pre-coated ELISA plates. Sensitivity and specificity analysis were carried out by the web-based MedCalc statistical software (https://www.medcalc.org/calc/diagnostic_test.php). Statistical analysis of reproducibility was carried out by calculate the mean PI value and coefficient of variation (CV) of replications of each test. Statistical analysis of the degree of agreement (Kappa value) and correlation between the established cELISA and VNT were carried out by McNemar's test and Pearson correlation coefficient analysis in SPSS Statistics for Windows, version 25.0 (IBM Crop, NY, USA). Differences were considered statistically significant when $p < 0.05$. Scatter plots was generated using the same program.

Abbreviations

CSF, classical swine fever; CSFV, classical swine fever virus; cELISA, competitive enzyme-linked immunosorbent assay; mAb, monoclonal antibody; VNT, virus neutralization test; CV, coefficient of variation; ORF, open reading frame; ND50, neutralization doses 50%; BVDV, Bovine viral diarrhea virus; HRP, horseradish peroxidase; FBS, fetal bovine serum; PBS, phosphate-buffered saline; PBST, phosphate buffered saline containing 0.05% Tween 20; CO₂, carbon dioxide; PI, percent of inhibition; PPRV, peste des petits ruminants virus; CDV, canine distemper virus; PDV, phocine distemper virus; SD, standard deviation; IFA, indirect fluorescent antibody assay; MOI, multiplicity of infection; hr, hour; TCID₅₀: 50% tissue culture infective dose; RT, room temperature; DPI, days post infection; DPV, days post vaccination.

Declarations

Ethics approval and consent to participate

Animal care and use protocols were approved by Institutional Animal Care and Use Committee at Kansas State University (KSU).

Consent for publication

Not applicable.

Availability of data and materials

The datasets for the current study are available from the corresponding author on reasonable request.

Competing interests

The authors declare that they have no competing interests.

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Authors' contributions

LW and WG carried out the C-strain E2 expression and purification. LW, SM, RM, JR, AC, CC and WG participated in animal studies, optimizing and validating the cELISA. LW, SM, LG, MB, AC, HL, CT, WG, and JS conceived and coordinated the study, participated in data analysis and drafting the manuscript. All authors read and approved the final manuscript.

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References

1. Moennig V. Introduction to classical swine fever: virus, disease and control policy. *Vet Microbiol.* 2000; 73: 93-102.
2. Postel A, Austermann-Busch S, Petrov A, Moennig V, Becher P. Epidemiology, diagnosis and control of classical swine fever: Recent developments and future challenges. *Transbound Emerg Dis.* 2018; 65(Suppl.1): 248–61.
3. Edwards S, Fukusho A, Lefevre P, Lipowski A, Pejsak Z, Roehe P, [Westergaard J](#). Classical swine fever the global situation. *Vet Microbiol.* 2000; 73:103–19.
4. Penrith, ML, Vosloo W, Mather C. 2011. Classical swine fever (hog cholera): Review of aspects relevant to control. *Transbound Emerg Dis.* 2011; 58: 186 –97.
5. Blome S, Moß C, Reimann I, König P, Beer M. Classical swine fever vaccines: state-of-the-art. *Vet Microbiol.* 2017; 206: 10–20.
6. Graham SP, Everett HE, Haines FJ, Johns HL, Sosan OA, Salguero FJ, et al. Challenge of pigs with classical swine fever viruses after C-strain vaccination reveals remarkably rapid protection and insights into early immunity. *PLoS One.* 2012; 7:
7. van Oirschot JT. Vaccinology of classical swine fever: from lab to field. *Vet Microbiol.* 2003; 96: 367–84.
8. Vienna R. Brown, Sarah N. Bevins. A Review of Classical Swine Fever Virus and Routes of Introduction into the United States and the Potential for Virus Establishment. *Front Vet Sci.* 2018; 5: 31.
9. Zhao JJ, Cheng D, Li N, Sun Y, Shi Z, Zhu QH, Tu C, Tong GZ, Qiu HJ. Evaluation of a multiplex real-time RT-PCR for quantitative and differential detection of wild-type viruses and C-strain vaccine of classical swine fever virus. *Vet Microbiol.* 2008; 126: 1–1.
10. Rice L. Flaviviridae: the viruses and their replication. In Lippincott Williams and Wilkins, editors. *Fields Virology.* 2011.991-1041.
11. Thiel HJ, Stark R, Weiland E, Rumenapf T, Meyers G. Hog cholera virus: molecular composition of virions from a pestivirus. *J Virol.* 1991; 65: 4705–12.
12. Meyers G, Thiel HJ, Rumenapf T. Classical swine fever virus: recovery of infectious viruses from cDNA constructs and generation of recombinant cytopathogenic defective interfering particles. *J Virol.* 1996; 70(3): 1588-95.
13. Cheng TC, Pan CH, Chen CS, Chuang KH, Chuang CH, Huang CC, Chu YY, Yang YC, Chu PY, [Kao CH](#), [Hsieh YC](#), Cheng TL. Direct coating of culture medium from cells secreting classical swine fever virus E2 antigen on ELISA plates for detection of E2-specific antibodies. *Vet J.* 2015; 205: 107-9.

14. Clavijo A, Lin M, Riva J, Mallory M, Lin F, Zhou EM. Development of a competitive ELISA using a truncated E2 recombinant protein as antigen for detection of antibodies to classical swine fever virus. *Res Vet Sci.* 2001; 70: 1–7.
15. Moser C, Ruggli N, Tratschin JD, Hofmann MA. Detection of antibodies against classical swine fever virus in swine sera by indirect ELISA using recombinant envelope glycoprotein E2. *Vet Microbiol.* 1996; 51:
16. Kumar R, Barman NN, Khatoon E, Kumar S. Development of single dilution immunoassay to detect E2 protein specific classical swine fever virus antibody. *Vet Immunol Immunopathol.* 2016; 172: 50-54.
17. Hu D, Lv L, Gu J, Chen T, Xiao Y, Liu S. Genetic diversity and positive selection analysis of classical swine fever virus envelope protein gene E2 in east China under C-strain vaccination. *Front. Microbiol.* 2016; 7:85.
18. Madera R, Gong W, Wang L, Burakova Y, Llellish K, Galliher-Beckley A, Nietfeld J, Henningson J, Jia K, Li P, Bai J, Schlup J, McVey S, Tu C, Shi J. Pigs immunized with a novel E2 subunit vaccine are protected from heterologous classical swine fever virus challenge. *BMC Vet Res.* 2016; 12:
19. Madera R, Wang L, Gong W, Burakova Y, Buist S, Nietfeld J, Henningson J, Ozuna AGC, Tu C, Shi J. Towards the development of a one-dose classical swine fever subunit vaccine: antigen titration, onset and duration of immunity. *J Vet Sci.* 2018; 19: 393-405.
20. Laughlin RC, Madera R, Peres Y, Berquist BR, Wang L, Buist S, Burakova Y, Palle S, Chung CJ, Rasmussen MV, Martel E, Brake DA, Neilan JG, Lawhon SD, Adams LG, Shi J, Marcel S. Plant-made E2 glycoprotein single-dose vaccine protects pigs against classical swine fever. *Plant Biotechnol J.* 2018; 1–11.
21. Zhou B. Classical Swine Fever in China-An Update Minireview. *Front Vet Sci.* 2019; 6:187.
22. Singh PR, Sreenivasa BP, Dhar P, Shah LC, Bandyopadhyay SK. Development of a monoclonal antibody based competitive-ELISA for detection and titration of antibodies to peste des petits ruminants (PPR) virus. *Vet Microbiol.* 2004; 98: 3-15.
23. Saliki JT, Lehenbauer TW. Monoclonal antibody-based competitive enzyme-linked immunosorbent assay for detection of morbillivirus antibody in marine mammal sera. *J Clin Microbiol.* 2001; 39:1877-1881.
24. Jarvis DL. Baculovirus-insect cell expression Methods in Enzymology, 2009; 463: 191-222.
25. Wang L, Madera R, Burakova Y, Buist S, Sang Y, Nietfeld J, Henningson J, Galliher-Beckley A, Gong W, Tu C, Shi J. Construction of Recombinant Porcine Reproductive and Respiratory Syndrome Virus Expressing CSFV E2 Glycoprotein. *Int J Vaccine Res.* 2018; 3(1): 1-4.

Figures

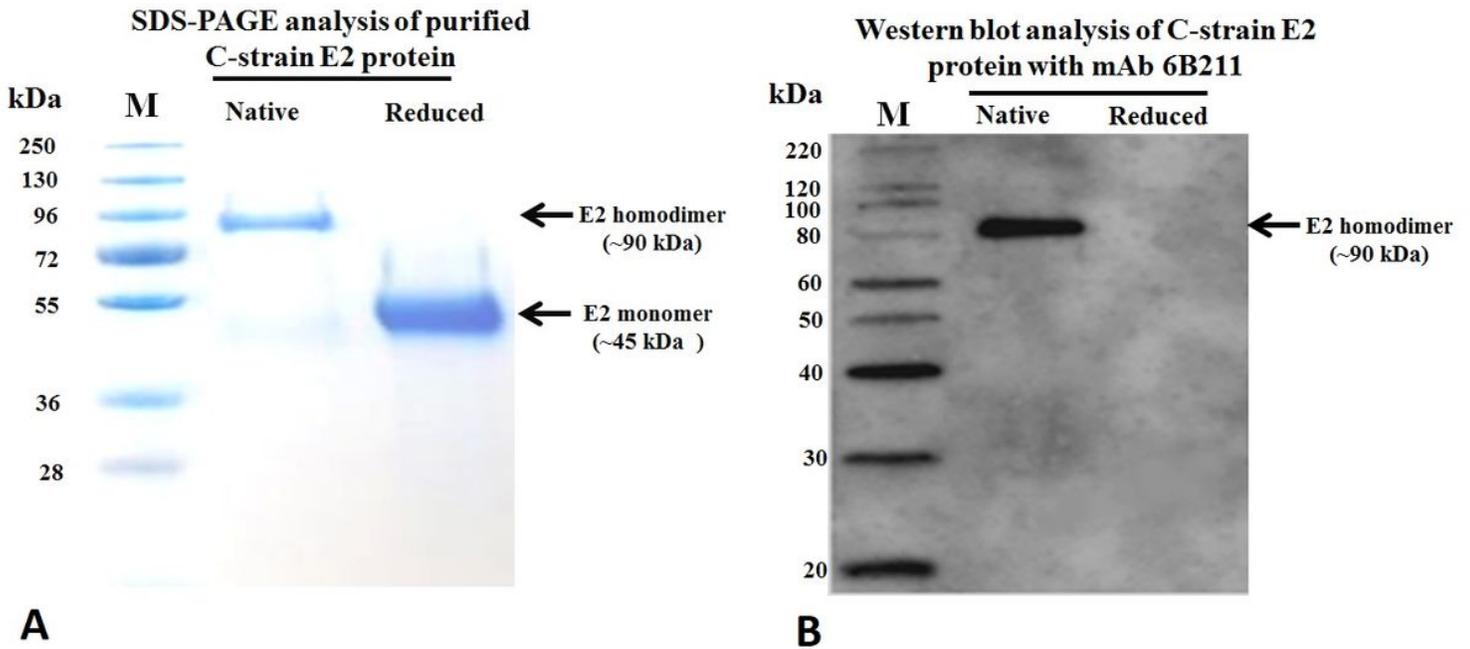


Figure 1

Analysis of purified C-strain E2 protein and monoclonal antibody 6B211. A) Purified C-strain E2 protein mainly exists in its native dimeric conformation. After purification steps, the purified insect cell expressed C-strain E2 protein were treated without (Native) or with β -mercaptoethanol (Reduced) and separated by SDS-PAGE in a Mini-Protean TGX Gel (Bio-Rad, CA, USA); B) 6B211 only reacted with the native C-strain E2 proteins. Purified E2 proteins (native or reduced) were loaded on Mini-Protean TGX Gel. The proteins were then transferred to PVDF membrane and the membrane were blocked and incubated with 6B211.

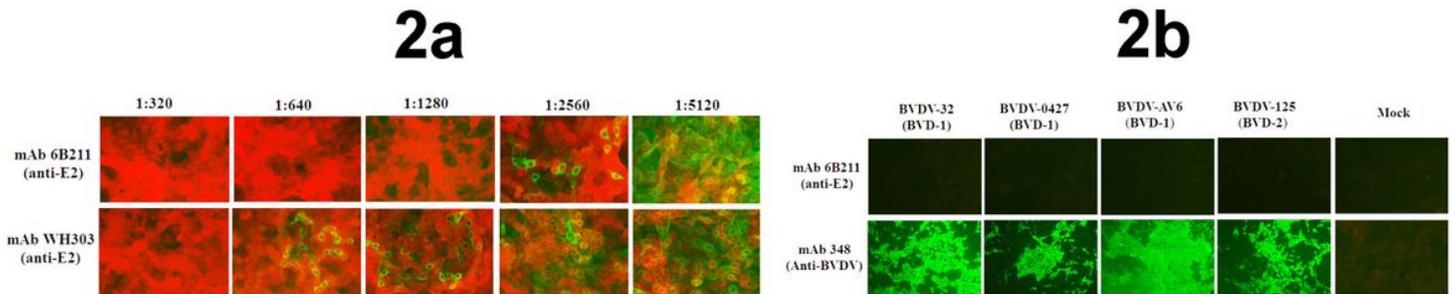


Figure 2

Neutralizing activity and cross-reaction testing of 6B211. A) 6B211 has potent neutralizing activity against C-strain; PK-15 cells were incubated with CSFV C-strain virus (100 TCID₅₀) and two-fold serial dilutions (1:320 to 1:5120) of mAb 6B211 (1mg/ml) or WH303 (1mg/ml); 3 days post infection (DPI); no green fluorescent signal means 100% inhibition of C-strain virus; B) 6B211 lacks cross-reactivity to BVDVs tested by IFA. Cells: MDBK; inoculated viruses: BVDV-32 (genotype 1, BVD-1), BVDV-0427 (BVD-1), BVDV-AV6 (BVD-1), and BVDV-125 (Genotype 2, BVD-2); 3 DPI; no green fluorescent signal means without reaction with BVDVs.

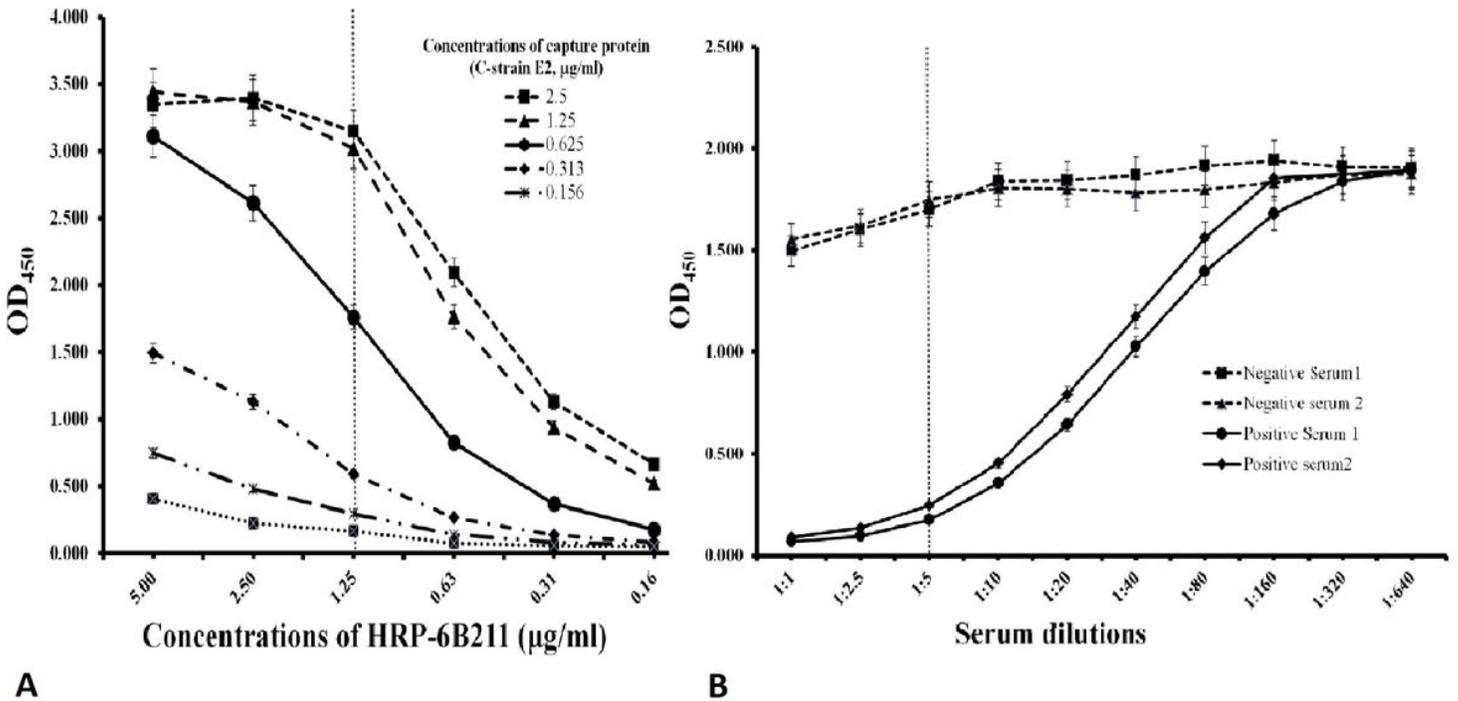


Figure 3

Determination of concentrations of capture antigen & competitive antibody, and dilution of serum. A) Determination of optimal concentrations of capture protein (C-strain E2) and competitive antibody (HRP-6B211); the optimal concentration of capture protein (0.625 µg/ml) and HRP-6B211 (1.25 µg/ml) was chosen as it consistently produced an OD₄₅₀ around 1.7 and at a point in the linear range that allowed optimal inhibition; B) Determination of an optimal dilution of serum; To minimize the volume of serum required and background noise, serum dilution of 1:5 was chosen as the preferred dilution over lower dilutions that performed only slightly better.

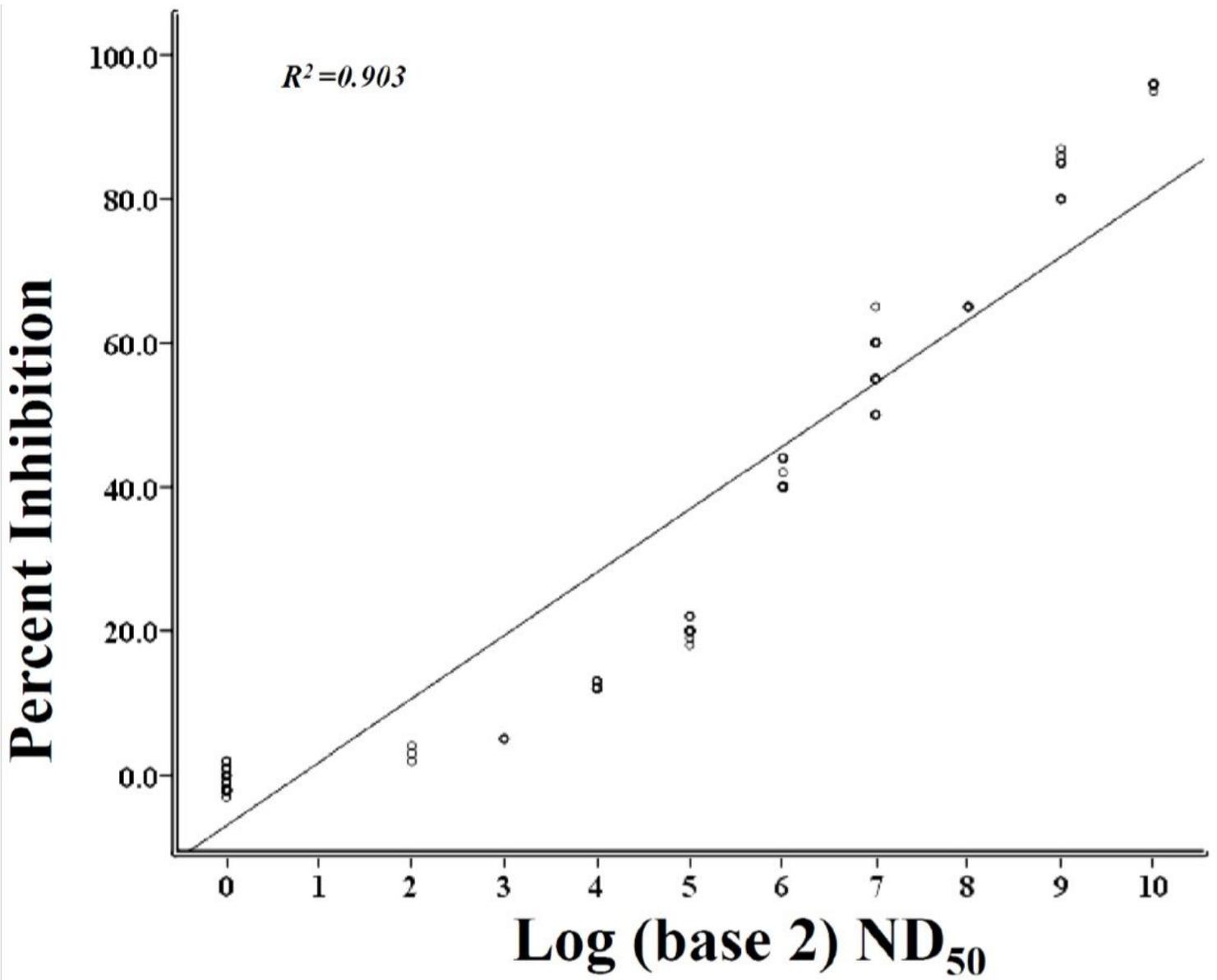


Figure 4

Correlation of inhibition rate of 6B211 based cELISA and titers of VNT against C-strain.

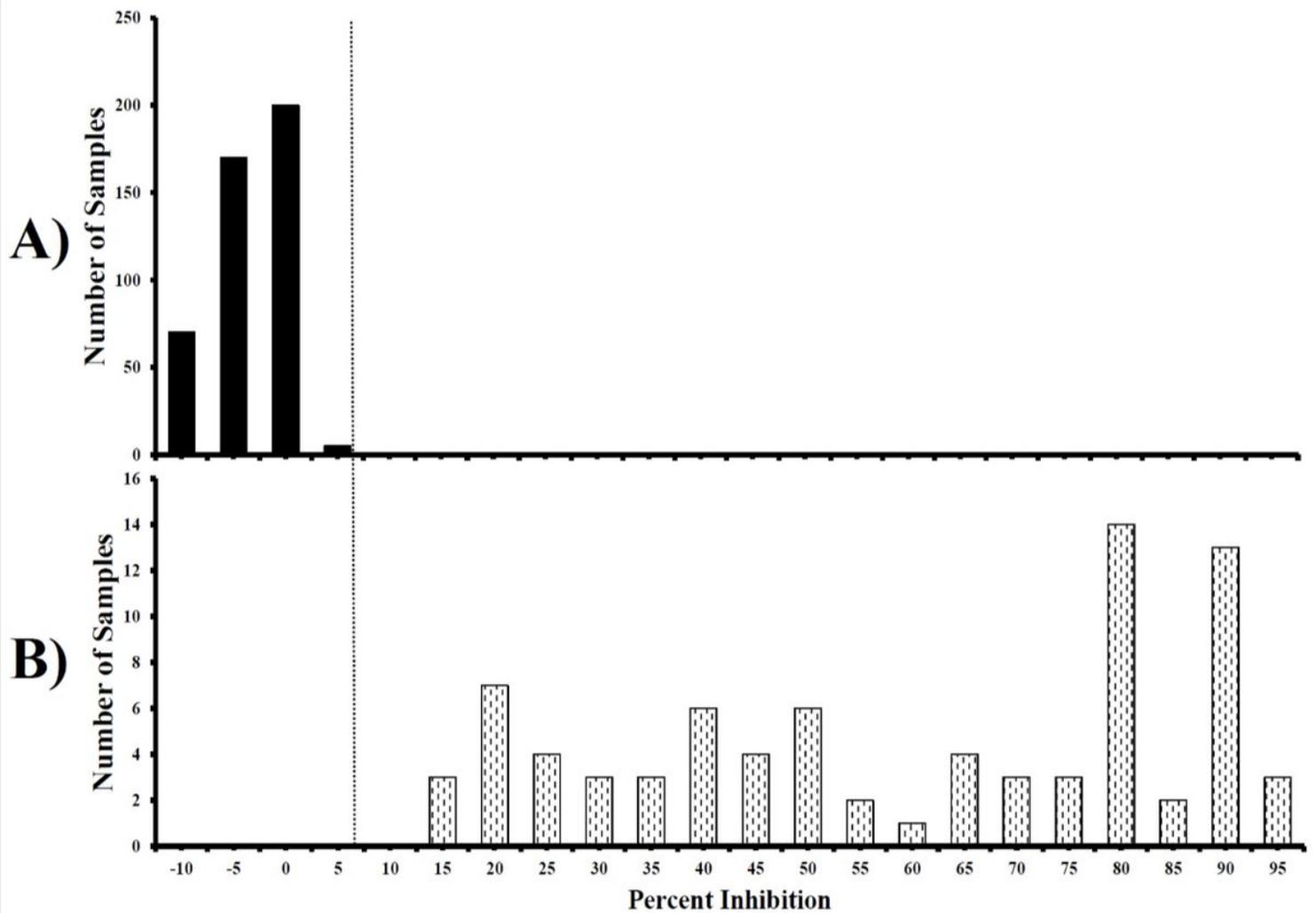


Figure 5

Standardization of the cut-off value of 6B211 based cELISA. A) C-strain VNT negative serum samples (n=445); B) C-strain VNT positive serum samples (n=70). The dotted line represents cut-off value of 6.19% inhibition when using the mean PI of negative sera plus three SD as the threshold.

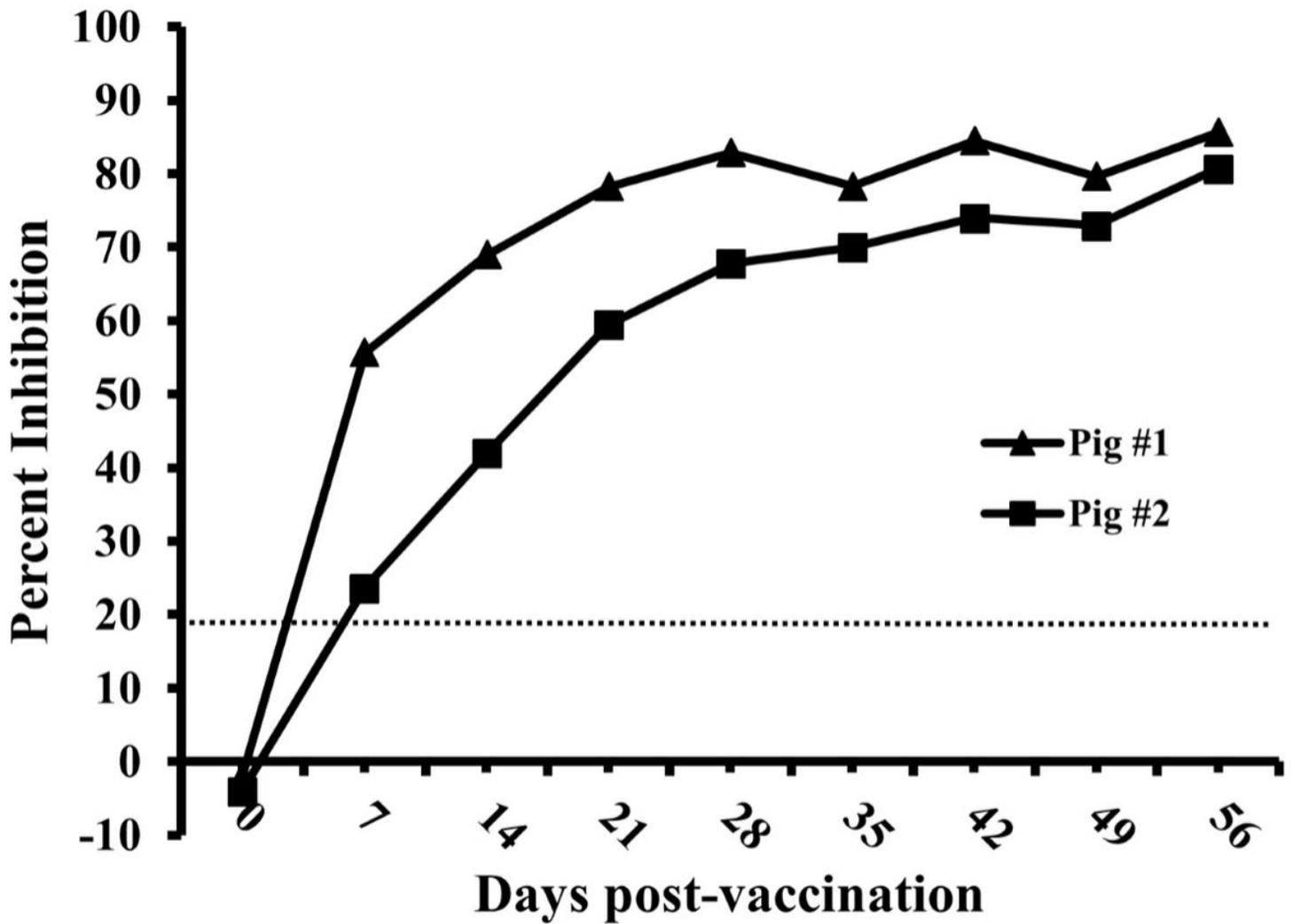


Figure 6

Kinetics of antibody responses of pigs determined by 6B211 based cELISA. Serum samples were derived from C-strain vaccinated pigs (n=2) at every 7 days. The dotted line represents cut-off value: 9.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [ARRIVEGuidelinesChecklist.pdf](#)