

Efficacy of Dual PI3K/mTOR Inhibitor Combined With Autophagy Suppression Against Laryngeal Squamous Cell Carcinoma

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Primary research

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Abstract

Objective: We explored the antitumor effect of dual PI3K/mTOR inhibitor combined with autophagy suppression on laryngeal squamous cell carcinoma (LSCC) and its underlying mechanism.

Methods: Hep-2 and AMC-HN-8 cell lines were treated with the Akt inhibitor LY294002, mTOR inhibitor rapamycin, and dual inhibitor NVP-BEZ235 separately. The biological characteristics of *in vitro* proliferation, cell cycle, apoptosis, migration, invasion, and autophagy were analyzed, and the expression levels of PI3K/Akt/mTOR pathway-related proteins were also measured. The *in vivo* effects of NVP-BEZ235 combined with inhibition of autophagy using pharmacological inhibitor was further assessed.

Results: Compared with Akt or mTOR inhibitor, NVP-BEZ235 had the most significant biological effects on LSCC cells. When combined with various autophagy inhibitors, along with siRNA against ATG7, NVP-BEZ235 showed a synergic antitumor effect in LSCC through increasing cell apoptosis and death both *in vitro* and *vivo*.

Conclusion: NVP-BEZ235 exerted stronger antitumor effects on LSCC, especially when combined with the autophagy inhibitor, providing convincing experimental data for new molecular targeted therapy for LSCC.

1. Introduction

Laryngeal squamous cell carcinoma (LSCC) is the most common head and neck squamous cell carcinoma (HNSCC) throughout the world, and its incidence has significantly increased in the past decade [1, 2]. Considering that the postoperative life quality and overall survival rate of patients with LSCC remain unsatisfactory, new chemotherapy strategies or targeted therapy are urgently required [3].

The mammalian target of rapamycin (mTOR) is overactivated in many malignancies and plays a crucial part in PI3K/Akt/mTOR pathway [4], one of the most frequently overactivated pathways in most human tumors, which can promote cell growth and proliferation [5, 6], and LSCC is no exception [7]. Moreover, PI3K/Akt/mTOR pathway seemed to be a prognostic factor and a predictor of response to radiotherapy and chemotherapy in clinical practice [8]. Since its critical influence on tumor activity, regulation of this signaling pathway has become a potential target for LSCC intervention.

Furthermore, recent studies have indicated that autophagy inhibition could enhance anticancer therapies in various malignancies [9–11], suggesting that autophagy is also emerging as a promising therapeutic target in oncotherapy. Considering that the PI3K/Akt/mTOR pathway is also involved in the regulation of autophagy, along with the possible limitation of monotherapy with PI3K or mTOR inhibitor [12], a combination of dual PI3K/mTOR inhibitor and autophagy inhibitor has become a novel research direction.

To identify a potential therapy strategy for LSCC, we compared the effects of three PI3K/mTOR signaling pathway inhibitors (LY294002, rapamycin, NVP-BEZ235) against LSCC cells *in vitro*, and the synergistic effect of the combined application of dual PI3K/mTOR inhibitor and autophagy inhibitor was evaluated *in vivo*.

2. Materials And Methods

2.1. Cell lines

Two LSCC cell lines, AMC-HN-8 and Hep-2, were acquired from the Stem Cell Bank of the Chinese Academy of Science. Both cells were cultured in RPMI-1640 medium (HyClone, Logan, UT) supplemented with 10% fetal bovine serum (FBS; Gibco, Grand Island, NY), 1% penicillin-streptomycin (Genom Biotechnology, China), and maintained in 5% CO₂ at 37°C.

2.2. Reagents

Antibodies against caspase 8, caspase 3, caspase 9, cyclin-D1, cyclin-E2, phospho-Akt (Ser473), Akt, phospho-P70S6K (Thr389), P70S6K, phospho-4E-BP1, 4E-BP1, phospho-S6RP, S6RP, LC3-II, LC3-I, and GAPDH (glyceraldehyde-3-phosphate dehydrogenase) were purchased from Sigma, USA. PI3K/mTOR inhibitors LY294002, rapamycin, NVP-BEZ235, and autophagy inhibitors 3-MA and CQ were purchased from Selleck, USA.

2.3. MTT detection for the calculation of drug IC₅₀

The cell lines were prepared and then treated with different concentrations of LY294002 (1, 5, 10, 20, 40, 80, 160 μM), rapamycin (1, 5, 10, 20, 40, 80, 160 μM), and NVP-BEZ235 (0.1, 0.5, 1, 2, 4, 8, 16 μM) for 72 h. DMSO served as the control group. Twenty microliters of MTT (5 mg/ml) was added to each well, and all plates were further incubated for 4 h. Later, 150 μl of DMSO was added, and the OD₄₉₀ values were measured. The inhibition rate was selected as the y-coordinate and the drug concentration (lnX) as the x-coordinate. The fitting curve formula $Y = A * \ln X + B$ was used to calculate the semi-inhibition rate of cells, $IC_{50} = \exp [(0.5-b)/a]$. All assays were performed in six replicates, and each experiment was repeated three times.

2.4. Western blotting

Hep-2 cells were treated with or without LY294002 (7.5 μM), rapamycin (15 nM), and NVP-BEZ235 (1.5 μM), while AMC-HN-8 cells were treated with or without LY294002 (6 μM), rapamycin (15 nM), and NVP-BEZ235 (0.75 μM). Whole cell protein lysates were obtained 72 h later using RIPA lysis buffer solution (Beyotime, China). Equivalent quantities of cell lysate protein (25 μg) were exposed to SDS-PAGE and transferred to a PVDF membrane (Millipore, USA), which was blocked (with 5% nonfat milk) and incubated with antibodies described above at 4°C overnight. The protein bands were visualized by using an enhanced chemiluminescent substrate (Millipore, USA). ImageJ software was used to analyze relative protein levels and normalize the internal reference protein GAPDH.

2.5. Apoptosis assay

An Annexin V-FITC/PI Detection Kit (Invitrogen, USA) was used to examine cell apoptosis according to the manufacturer's instructions. After treatment, the LSCC cells were collected and stained with Annexin V-FITC and PI. After 30 min of incubation in the dark, the cell apoptosis ratio was detected using FCM (Becton Dickinson, USA).

The TUNEL kit (Roche, Switzerland) was used to detect apoptotic cells in tumors according to the manufacturer's instructions. The proportion of TUNEL-positive cells showing a brown color on each section was counted to determine the occurrence of apoptosis.

2.6. Cell cycle assay

The cell cycle was examined using the propidium iodide (PI) technique. After treatment with LY294002, rapamycin, or NVP-BEZ235 72 h, the LSCC cells were starved for 24 h in RPMI-1640 without serum. Single cell suspensions were then fixed in 75% ethanol overnight at -20°C, washed twice with PBS, and incubated with PI/RNase staining buffer (BD Biosciences). Samples were tested on a flow cytometer (BD Biosciences) and analyzed with FlowJo software (FlowJo LLC).

2.7. Invasion and migration assay with a transwell chamber

The invasive and migratory capabilities of Hep-2 and AMC-HN-8 cells following treatment with the three inhibitors were evaluated by transwell assay. For invasion, 100 µl Matrigel (Becton Dickinson, USA) was added to the bottom of each upper chamber in advance and dried for 4 h at 37°C to a gelatinous form. Then, for both experiments, 5×10^5 /ml cells were placed in the upper chambers with LY294002, rapamycin, and NVP-BEZ235 incubated for 24 h at 37°C. The lower chambers were filled with a culture medium with 600 µl of 20% fetal bovine serum (FBS). Cells in the chambers were later fixed with 4% paraformaldehyde and stained with 0.1% crystal violet (Beyotime, China). Five random fields were selected to count, and the results were averaged and plotted.

2.8. ATG7 siRNA Interference

We transfected cells with siRNA oligos against ATG7 with the purpose of genetically inhibit autophagy. The sequences of ATG7 were as follows:

Atg7-1, 5'-GATCCGGTGCTGGTTTCCTTGCTTAACTCGAGTTAAGCAAGGAAACCAGCACCTTTTTTG-3' and antisense, 5'-AATTCAAAAAGGTGCTGGTTTCCTTGCTTAACTCGAGTTAAGCAAGGAAACCAGCACCG-3';
Atg7-2: 5'-GATCCGCAGCCTCTCTATGAGTTTGACTCGAGTCAAACCTCATAGAGAGGCTGCTTTTTTG-3' and antisense, 5'-AATTCAAAAAGCAGCCTCTCTATGAGTTTGACTCGAGTCAAACCTCATAGAGAGGCTGCG-3'.

A total of 1×10^5 AMC-HN-8 and Hep-2 cells were seeded in 6-well plates overnight before transfection. Cells were transfected with siRNA oligonucleotides at 4 µg/250 µl medium with 10 µl Lipofectamine 2000 (Invitrogen, USA) each well according to the manufacturer's instructions.

2.9. Xenograft nude mice of LSCC

All male 6- to 8-week-old BALB/c nude mice (SLAC Laboratory Animal Co., Ltd, Shanghai) were inoculated with 6×10^6 Hep-2 cells before being randomly divided into four groups (n = 6 each) as follows: (1) control group; (2) NVP-BEZ235 group, oral administration of NVP-BEZ235 (30 mg/kg/d); (3) CQ group, intraperitoneal (i.p.) administration of CQ (50 mg/kg/d) ; (4) NVP-BEZ235 + CQ group, rats were treated with NVP-BEZ235 (30 mg/kg/d, p.o.) and CQ (50 mg/kg/d, i.p.) simultaneously. The tumor sizes were measured twice a week with calipers. All surviving rats were sacrificed 28 days later to collect tumors for further detection. Tumor size and tumor cell apoptosis of each group were analyzed.

2.10. Statistical analysis

Data analysis was accomplished using SPSS 22.0 software. Differences between groups were assessed by the Student's t-test (two-tailed), and the results are shown as means \pm SEM. $P < 0.05$ was considered statistically significant.

3. Results

3.1. Dual PI3K/mTOR inhibitor NVP-BEZ235 leads to cell cycle arrest and apoptosis in LSCC cell lines

To clarify the role of PI3K/Akt/mTOR pathway in LSCC, Hep-2 and AMC-HN-8 cell lines were treated with PI3K inhibitor LY294002, mTOR inhibitor rapamycin, and dual PI3K/mTOR inhibitor NVP-BEZ235 separately. Cell apoptosis and the cell cycle were then assessed. NVP-BEZ235 elicited significant blockage at G2/M phase, and the protein levels of cyclin D1 and cyclin E2 also decreased in both cell lines than LY294002 and rapamycin (Fig. 1), which suggested its better effect on the cell cycle as a dual PI3K/mTOR inhibitor. Treatment with all three inhibitors resulted in significant apoptosis in Hep-2 and AMC-HN-8, especially with NVP-BEZ235 (Fig. 2A, 2B), whereas the protein level of caspases 3, 8, and 9 increased after inhibitor treatment (Fig. 2C).

3.2. NVP-BEZ235 shows more potent and dual blocking effects for mTORC1 and mTORC2

We next tested the activation of AKT, p70S6K, 4EBP1, and S6RP in the two cell lines after the treatment with the inhibitors. NVP-BEZ235 modulated not only mTORC1 targets p-70S6K, p-4EBP1 and p-S6RP, but also mTORC2 targets p-Akt (Fig. 3). To evaluate antitumor effects, we further manipulated the migration and invasion test by transwell chamber assay in Hep-2 and AMC-HN-8 treated with Ly294002, rapamycin, NVP-BEZ235, or DMSO (as control). All three inhibitors reduced cell migration and invasion *in vitro* (Fig. 4A, 4B, 4C, 4D); however, NVP-BEZ235 elicited the most significant effect.

Considering the above assessment, NVP-BEZ235 had a better block effect on PI3K/Akt/mTOR pathway as a dual PI3K/mTOR inhibitor on both LSCC cell lines than LY294002 and rapamycin.

3.3. Synergic antitumor effect of combination NVP-BEZ235 with autophagy inhibition *in vitro*

As NVP-BEZ235 can regulate cell autophagy in many tumors [13, 14], we investigated the regulatory effects of all three inhibitors on autophagy in LSCC cell lines. Autophagy control was observed using a confocal laser scanning microscope after acridine orange staining (Supplemental Fig. 1) and verified by LC3, a protein embedded in autophagosomes that became lipids during active autophagy. LC3II/LC3I ratio was obtained using western blotting analysis (Fig. 5D. E). We observed that all three inhibitors of PI3K/mTOR pathway induced autophagy, indicated by the rise of LC3II/LC3I ratio, with NVP-BEZ235 potentiating the ratio more effectively.

Furthermore, we tested the combination of NVP-BEZ235 with autophagy inhibitors, 3-methyladenine (3-MA), chloroquine (CQ) and ATG7-SiRNA. 3-MA inhibits the initiation of autophagy, CQ inhibits the fusion of autophagosomes and lysosomes; and ATG7-SiRNA, is a genetic silencing of Atg7, which inhibits the early phase of autophagy. The results showed that cell death was significantly increased when NVP-BEZ235 was combined with autophagy inhibitors (Fig. 5A. B. C), although LC3II/LC3I ratio and the expression level of caspase 3 were not significantly changed by the combination. This result strongly indicates that the combination of NVP-BEZ235 and autophagy inhibitors has a synergic antitumor effect through increasing LSCC cell apoptosis and death.

3.4. Synergic antitumor effect of combination NVP-BEZ235 with autophagy inhibition *in vivo*

We then tested the antitumor efficacy of the combination of NVP-BEZ235 with CQ *in vivo*. BALB/c mice bearing Hep-2 xenograft tumors were treated with NVP-BEZ235, CQ, NVP-BEZ235 + CQ, and physiological saline solution as a control for 4 weeks after tumor cell injection. First, we tested the drug toxicity of all groups by evaluating the changes in animal weight, behavior, and appearance. The tumor size was then measured weekly before the animals were sacrificed. The final tumor masses were dissected after the last measurement. Combination of NVP-BEZ235 and CQ demonstrated synergistic effect against autophagy, indicated by reduced tumor size and final tumor weight (Fig. 6). The interaction effect was elicited by the interaction index ($p < 0.0001$). Based on TUNEL assay of fragmented DNA of apoptotic cells in final tumor masses, the most apoptotic cells were found in the group treated with NVP-BEZ235 and CQ.

4. Discussion

In this study, the dual PI3K/mTOR inhibitor NVP-BEZ235 revealed a better effect on the cell cycle, cell apoptosis, migration, and invasion in LSCC. The expression changes in downstream molecules of PI3K/Akt/mTOR signaling pathway further explained its underlying mechanism. We demonstrated that the combination of NVP-BEZ235 and autophagy inhibitors has a synergic antitumor effect by promoting cell apoptosis and death both *in vitro* and *in vivo*.

Since PI3K/Akt/mTOR signaling pathway plays a crucial role in tumorigenesis by strikingly promoting cell growth and proliferation [15, 16], inhibition of this pathway is also postulated as a therapeutic option in LSCC[17]. The mTOR protein serves as an integral part in this signaling complex, which is involved in two major complexes: mTOR complex1 (mTORC1) and mTOR complex2 (mTORC2)[18]. Although discovered much later than mTORC1, mTORC2 is considered essential for transformation and viability of cancer cells while it appears more important in tumor cells, indicating mTORC2 is emerging as a promising candidate target in oncotherapy [19]. Studies have proved that mTORC2 can directly regulate Akt phosphorylation via activating a critical regulatory site required for maximal Akt kinase activity, which enables Akt to become a crucial downstream protein of the mTORC2 signal pathway [20, 21]. Due to multiple resistance mechanisms, such as the feedback activation of the PI3K/Akt signaling network and the insensitivity of the mTORC1 complex, inhibition of the mTORC1 pathway has met with limited success in many studies [22–25], which is also verified in our experiment. Compared with Akt inhibitor LY294002 and mTORC1 inhibitor rapamycin [26], dual PI3K/mTOR inhibitor NVP-BEZ235 showed better anticancer effects in arresting the cell cycle, promoting cell apoptosis, and inhibiting cell invasion and metastasis by suppressing mTORC1 and mTORC2 at the same time. Changes in downstream proteins, such as p-Akt, p-4E-BP1, p-S6RP, and p-p70S6K, also supported the remarkable anticancer effect of NVP-BEZ235 and its corresponding mechanism in two LSCC cell lines.

However, PI3K/mTOR inhibitors still have a moderate effect as single agents in clinical trials, which may be related to their induction of cell autophagy [27, 28]. Through degradation of damaged proteins and organelles, autophagy can protect tumor cells from stressful conditions, thus mediating resistance to anticancer therapies such as radiation, chemotherapy, and targeted treatments[29–32]. In this study, an increase of LC3II/I ratio was found in both LSCC cell lines treated with PI3K or mTOR inhibitors, among which the dual inhibitor NVP-BEZ235 is the most significant, supporting its induction of cell autophagy. Based on accumulating evidence that inhibition of autophagy can enhance the efficiency of antitumor drugs [33–35], we combined NVP-BEZ235 with autophagy inhibitors for further verification.

Surprisingly, the combination of NVP-BEZ235 with two autophagy inhibitors (3-MA, CQ) showed a synergetic antitumor effect in both cell lines. LSCC cells that inhibited autophagy in the early phase by silencing Atg7 also had increased cell apoptosis and death, further indicating that inhibition of autophagy can effectively improve the antitumor effect of PI3K/Akt inhibitors. We then combined NVP-BEZ235 with the less toxic autophagy inhibitor CQ to test their antitumor efficacy *in vivo*. The remarkably increased proportion of apoptotic cells in the tumor, along with the obviously decreased tumor size and tumor weight, were all consistent with those of our previous cell experiments.

In summary, we have proved the remarkable antitumor effect of dual PI3K/Akt inhibitor NVP-BEZ235 compared to other PI3K or Akt inhibitors in LSCC cells, which may be achieved by inhibiting mTORC1 and mTORC2 simultaneously. Combined treatment with CQ, which could inhibit autophagy induced by NVP-BEZ235, further promoted tumor cell apoptosis and death both *in vitro* and *vivo*, indicating their potential synergic antitumor effect in LSCC. Based on the current trend of combination therapy in cancer treatment [36], our experiments have solved the limitations of PI3K/Akt inhibitors to some extent, suggesting that

combined application of autophagy inhibitors could be an effective way to overcome resistance of LSCC to PI3K/Akt inhibitors, while providing a promising therapy with the combined application of targeted drugs in LSCC.

Declarations

Ethics approval and consent to participate:

This study did not involve the acquisition of HSCC tissue samples. The animal experiments were approved by the Animal Center, EENT Hospital, Fudan University

Consent for publication:

All the authors listed have approved the manuscript that is enclosed.

Availability of data and materials:

All the source data supporting the findings of this study are available from the corresponding author upon reasonable request.

Competing interests:

The authors declare no conflict of interest.

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Authors' contributions:

Hui-Ying Huang: Conceptualization, Formal analysis, Investigation, Methodology, Project administration, Validation, Writing - original draft, Writing - review & editing. Ke-Nan Li: Formal analysis, Investigation, Resources, Validation, Writing - original draft. Hui-Ching Lau: Formal analysis, Investigation, Resources, Validation. Chi-Yao Hsueh: Project administration, Resources. Ning Cong: Conceptualization, Project administration, Resources, Supervision, Writing - review & editing. Ming Zhang: Conceptualization, Funding acquisition, Resources, Supervision, Writing - review & editing

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The authors declare no conflict of interest.

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Figures

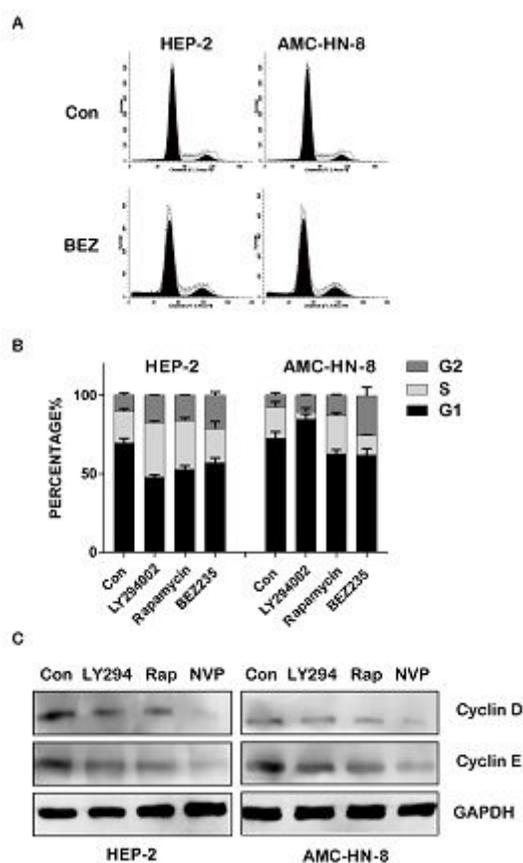


Figure 1

Cell cycle is affected by inhibitors of PI3K/ mTOR pathway. (A) Flow cytometric (FCM) analysis of cultured Hep-2 and AMC-HN-8 cells. (B) Cell cycle analysis of the Hep-2 and AMC-HN-8 cell lines. Cells in G2/M phase were remarkably increased by NVP-BE235 in both cell lines. Columns, means of duplicate measurements; bars, \pm SD. (C) Western blotting analysis of the expression levels of cyclin D1 and cyclin

E2 in two cell lines treated with three different PI3K/ mTOR inhibitors. Cyclin D1 and cyclin E2 decreased significantly after treatment with NVP-BEZ235.

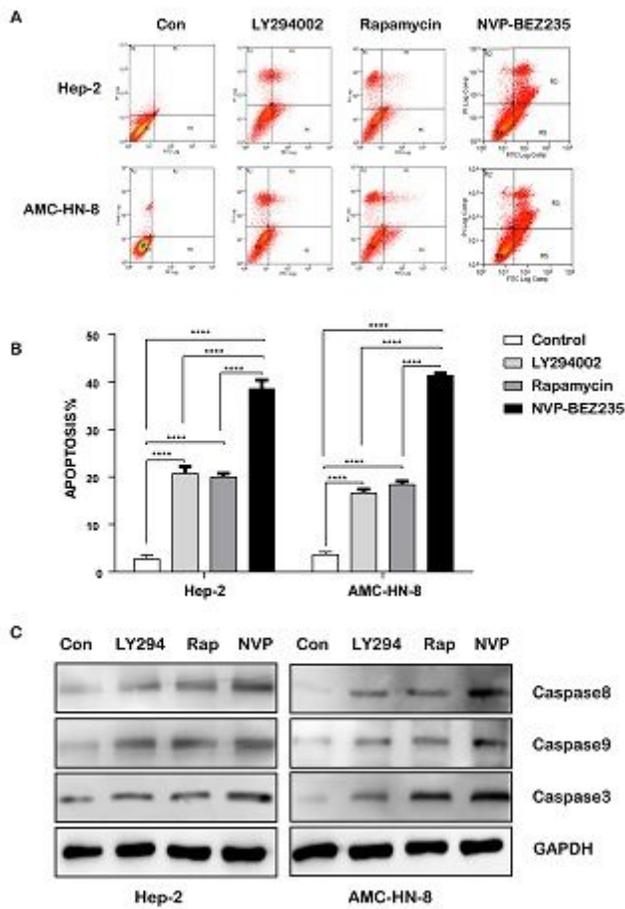


Figure 2

Cell apoptosis is increased by inhibitors of PI3K/mTOR pathway. (A) Flow cytometry (FCM) analysis of the cell apoptosis of cultured Hep-2 and AMC-HN-8 cell lines. (B) Cell apoptosis was increased by all three inhibitors in both cell lines ($p < 0.0001$). The apoptosis increase was similar with Ly294002 and rapamycin, but was more pronounced with NVP-BEZ235 ($p < 0.0001$). Columns, means of duplicate measurements; bars, \pm SD. (C) Caspases 8, 9, and 3 expression in the Hep-2 and AMC-HN-8 cell lines increased after NVP-BEZ235 treatment.

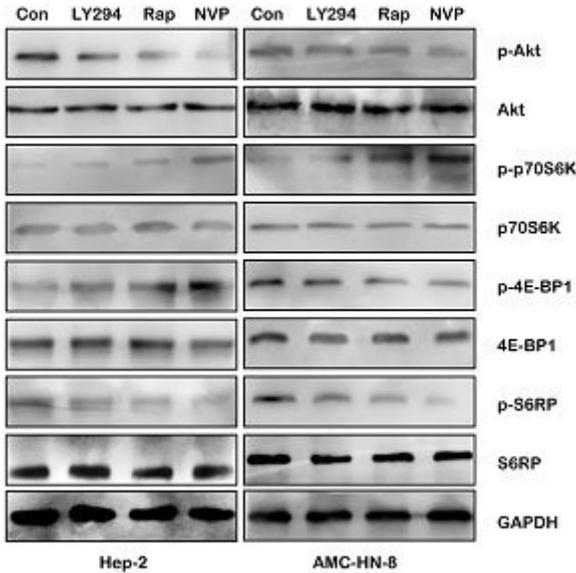


Figure 3

Protein expression and phosphorylation of members of the PI3K/Akt/mTOR pathway of Hep-2 and AMC-HN-8 cell lines, assessed by western blotting.

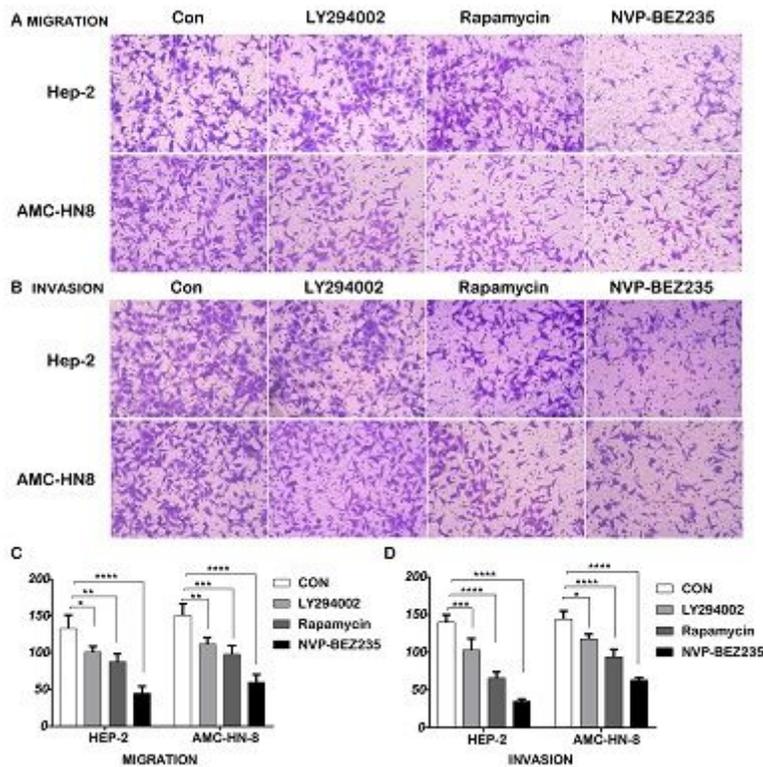


Figure 4

Cell migration and invasion is modulated by PI3K/ mTOR inhibitors. (A), (B) Migration and invasion affected by LY294002, rapamycin, and NVP-BE2235 in Hep-2 and AMC-HN-8 cell lines were separately tested by a trans-well chamber assay. (C), (D) Cell quantification showed a significant decrease in

migration and invasion by all three inhibitors in both cell lines, and NVP-BEZ235 showed a stronger effect.

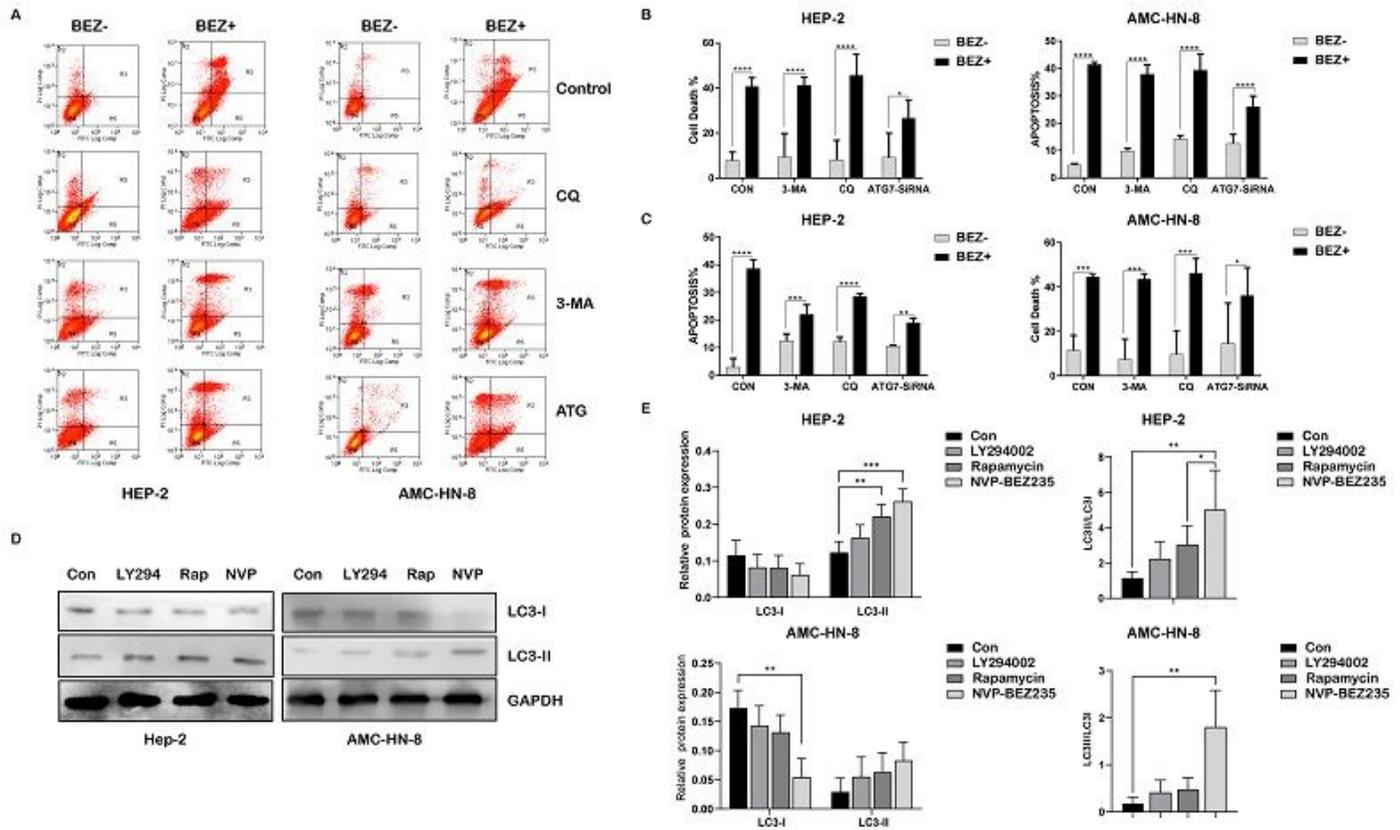


Figure 5

Cell autophagy and of PI3K/Akt/mTOR pathway inhibition. (A) Flow cytometric (FCM) analyses of the cell apoptosis of cultured Hep-2 cells or AMC-HN-8 cells. (B) Cell apoptosis increased in both cell lines treated with a combination of NVP-BEZ235 and autophagy inhibitors. Columns, means of duplicate measurements; bars \pm SD. (C) Cell death increased in both cell lines treated with a combination of NVP-BEZ235 and autophagy inhibitors. (D) Western blotting analysis of levels of LC3II and LC3I expression in two cell lines treated with three PI3K inhibitors. (E) Western blotting analysis showed LC3II/I ratio decreased significantly following treatment of both cell lines with Ly294002, rapamycin, and NVP-BEZ235.

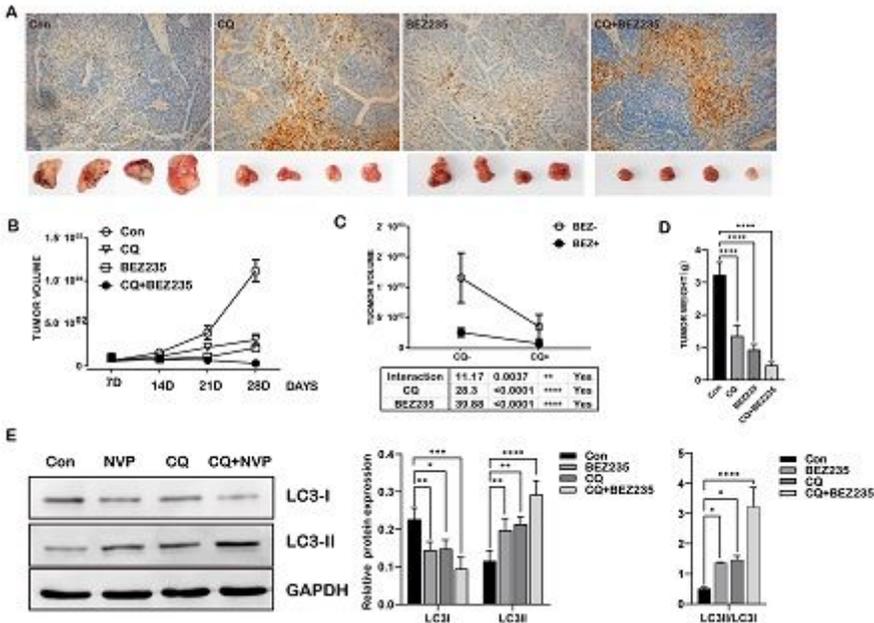


Figure 6

Anti-tumor efficacy of the combination of NVP-BE235 with autophagy inhibitor in vivo. (A) Tumors obtained from mice and their immunohistochemical results. (B) Tumor volumes increased with days, and their analysis results. Points, mean of four replicate determinations; bars \pm SD. (C), (D) Differences in tumor weights between the four groups. Points and columns, mean of four replicate determinations; bars \pm SD. (E) Western blotting testing for LC3II and LC3I expression of tumors from the four groups and the corresponding analysis.

Supplementary Files

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- [Fig.S1.tif](#)
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