

# Pollution Resistance Assessment of Plants Around Chromite Mine Based on Anticipated Performance Index, Dust Capturing Capacity and Metal Accumulation Index

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## Research Article

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## Abstract

Plant species sustaining under a polluted environment for long time are considered as potentially resistant species. Those plant species can be considered as an eco-sustainable tool used to bio-monitor and mitigate pollution. This study was carried out on total ten commonly available plant species to assess their anticipated performance index (API), Dust capturing capacity (DCC) and Metal accumulation index (MAI) in chromite mine and control area. According to Anticipated performance index (API) *Macranga peltata*, *Holarrhena pubescens* and *Ficus hispida* are highly tolerant species while *Terminalia arjuna* and *Trema orientalis* are intermediate tolerant species. *Ficus hispida* was also showed the highest dust capturing capacity ( $5.94 \pm 0.43 \text{ mg/cm}^2$ ) and whereas *Woodfordia fruticosa* ( $1.03 \pm 0.11 \text{ mg/cm}^2$ ) was found to be lowest. Metal accumulation index ranged from 17.29 to 4.5 and 6.38 to 1.94 at mine and control area respectively. Two-way ANOVA analysis revealed area wise significant differences between biochemical and physiological parameters. Also, result showed that the pollution level and heavy metal affected different biochemical and physiological parameter of plant species at mining area. The plant species with highest API, DCC and MAI value could be recommended for greenbelt development in different polluted area.

## 1. Introduction

The economic growth and development of a nation is dependent on its mining sector to a great extent which is associated with the exploitation of the mineral resources (Lebre et al., 2017). Whereas mining plays a significant role in economic growth at the same time it's adversely impact on environment. Worldwide mining activity is one of the serious contributors to the environmental pollution and considered one of the significant sources of air, soil and water pollution (Oluwoye et al., 2017; Golui et al., 2019; Sahu and Basti, 2020). From the open cast mining, a massive amount of gaseous pollutant, toxic substance, fine ore particles and dust releases to the environment (Das et al. 2020). These releases pollutants mix with atmosphere and transported long distance through air from mine area and leads to pollution in the surrounding undisturbed area (Ni et al., 2018; Khazini et al., 2021)

Green vegetation around the mining area can act as an eco-friendly and cost-effective tool to reduce pollution. In polluted environment, green plant play a crucial function in improving environmental quality by accumulating toxic pollutant (Gheorghe and Ion 2011; Kaur and Nagpal, 2017). They have a great potential to absorption, adsorption and accumulation of pollutants from both soil and air (Manara, 2012; Remon et al., 2013). They transport toxic substances from soil and air to biotic environment. Green plants perform as pollution sink as they provides broad surface area for absorb and accumulation of pollutants hence, plantation of highly tolerant plant species can help in effective green belt development to clean up polluted areas, further improving the environmental quality (Remon et al., 2013; Selmi et al., 2016; Karmakar & Padhy, 2019;). Selection of suitable plant for greenbelt development differ from region to region.

Air pollution tolerance index (APTI) and anticipated performance index (API) are considered best and reliable indices to select appropriate tolerant and sensitive plant species. The highly tolerant plant species are chosen to perform against the environmental pollution and the sensitive species might be use for bioindicator (Rai, 2016; Javanmard et al. 2020; Roy et al. 2020; Molnár, 2020). Four biochemical parameters such as Total chlorophyll content (TCC), Leaf extract pH (pH), Ascorbic acid (AA) and Relative water content (RWC) were used to develop the APTI (Singh and Rao, 1991). API is an upgradation of APTI, which is more appropriate indices for selection of tolerant and sensitive species in a particular region. API is developed with the combination of APTI value of each individual species along with some morphological (Plant habit, canopy structure, types of plant, lamina structure) and socioeconomic characters (Mondal et al., 2011). Dust, particulate matter and heavy metal are the most prevalent and rapidly effecting environment causing critical problems to living organism (Xiu et al., 2020; Kong et al., 2021). Plants are the best efficient at accumulate and capture to heavy metal and dust particle. In order to combat such pollutant, Dust capturing capacity (DCC) and Metal accumulation index (MAI) are also considered to select the suitable plant species.

Sukinda chromite mine area is one of the most populated area in the world, having numbers of open cast chromite mine. This mining area holds 183 million tons of raw chromite reserves, which is total 97% of total reserves of the country and approximately 3.8 million tons of chromite produces per year (IBM 2010). Due to various mining activity a massive amount of ore particles and dust produced and get blown away through air to surroundings areas, subsequently causes chromium (Cr) contamination by the atmospheric deposition (Das et al., 2020). According to report of Blacksmith Institute (2007), extensive pollution of the area due to excessive chromite ore mining has made it fourth most polluted place in the world.

Through this study, we provide an integrated approach to the selection of both pollution tolerant and sensitive plant species. To know physiological and biochemical tolerance level of plants we studied air pollution tolerance index (APTI) and anticipated performance index (API). Heavy metal accumulation and dust capturing capacities of total ten plants were also examined. This study is confined only on commonly found native plant species in this area. So, our study will contribute convenient knowledge/ information for the establishment of greenbelt in this mining area as well as different other polluted areas to improve the soil and air pollution.

## 2. Materials And Methods

### 2.1. Study area

Present study work was done at two areas namely Sukinda chromite mine valley and Tomka forest, located in Jajpur district, Odisha, India. Sukinda Chromite mine valley is the largest chromite producer in India, almost 99% of country's chromium are produce from this valley. Stretching between 21 ° 00 ' N to 21 ° 05 ' N latitude and 85 ° 44 ' E to 85 ° 53 ' E, occupies an area of two hundred square kilometre holds an estimated 183 million tons of raw chromium deposits (Nayak and Kale 2020) and consists of an extensive plateau in the interior with a foreground of wide coastal plain and underlain by Precambrian rock. Currently, 20 open cast and 2 underground mines are functional in the valley producing 160 million tons of overburden. It also releases 11.73 tons of hexavalent chromium (Cr (VI)) to the environment every year, making it the fourth-worst polluted places in the world. (Dhal et al. 2011; Das et al. 2020). Tomka forest considered as control area due to pollution free atmosphere, located about 40 km away from the mining area.

### 2.2. Plant species sampling

In total, leaf samples of ten plant species namely: *Ailanthus excels*, *Bixa orellana*, *Ficus hispida*, *Macranga peltata*, *Woodfordia fruticose*, *Trema orientalis*, *Terminalia arjuna*, *Holarrhena pubescens*, *Callicarpa tomentosa* and *Combretum roxburghii* plant species were collected during winter session 2020. These plant species were considered on the basis of abundance and their socioeconomic importance. Three replicate of healthy, mature and fully developed plant leaves sample were collected for each species. The replicate plant species were chosen as those individual species having similar height and breast diameter. After being sampled leaves were placed into zipper locked bags and kept in a portable ice-box, then transferred to the laboratory. Leaves were collected and transferred very carefully form plant to zipper locked bag, for ensuring least disturbance to their surface dust. For further biochemical analysis leaves were washed with tap water followed by distil water, and stored in -20 °C.

### 2.3. Analysis of the biochemical parameters of the plants

#### 2.3.1. Relative water content (R)

After taking the fresh weight (FW), leaf were submerged in deionized water for 24 hours. Then turgid weight (TW) recorded by soak up the surface water of leaf. Overnight dried leaf in oven at 80° C used for taking dry weight (DW) (Liu and Ding, 2008).

$$RWC (\%) = [(FW - DW) / (TW - DW)] \times 100$$

#### 2.3.2. Total chlorophyll content (T)

Total chlorophyll content of leaves were calculated following the procedure mention by Arnon (1949). 1 g of fresh leaves were crushed by using 10 ml of 80% acetone and centrifuged at 5000 rpm for 5 min. After collection of supernatant volume were made up to 30 ml by 80% acetone and absorbance was taken at 645 and 663nm by using spectrophotometer (Analytical UV-Vis 3090V). Calculation of total chlorophyll content was estimate by following formula

$$T \left( \frac{mg}{g} \right) = \frac{(20.2 * A_{645} + 8.02 * A_{663}) * V}{1000 * W(g)}$$

Where, A<sub>645</sub> = Absorbance at 645 nm, A<sub>663</sub> = Absorbance at 663 nm, V = Total volume of extract (ml), W = Weight of leaf in gram.

#### 2.3.3. Ascorbic acid (A)

Ascorbic acid estimation was done by using modified colorimetric 2,6-dichlorophenol indophenol technique. 0.5g of fresh leaves was extracted in 4% oxalic acid and made up to 30 ml volume and centrifuged at 6000 rpm for 10 min. 10 ml of 4% oxalic acid added with 5 ml of the supernatant and titrate against the dye. Titration was done till pink colour appeared which presence for a few second.

$$A \left( \frac{mg}{g} \right) = \frac{0.5(mg) * V2(ml) * 30(ml)}{V1(ml) * 5(ml) * weight\ of\ sample(g)}$$

Where,

V1 = Volume of dye titrated against the working standard

V2 = Volume of dye titrated against the sample

#### 2.3.4. Leaves Extract pH (P)

The pH of the leaves were determined with the help of pH meter by crushed 1 g of fresh leaves and homogenizing in 40 mL of deionized water.

### 2.3.5. Air Pollution Tolerance Index (APTI)

APTI was calculated following the equation described by Sing and Rao (1991).

$$APTI = [A (T + P) + R] / 10$$

Where A = Ascorbic acid (mg/g), T = Total chlorophyll content (mg/g), P = pH of leaf extract, R = Relative water content (%).

### 2.3.6. Anticipated Performance Index (API)

Combining some morphological and socio-economic characteristics (plant habit, type of plant, laminar characters, canopy type, and economic value) with APTI value, API is estimated. The API score is calculated according to the following formula

$$API (\%) = (\text{Total positives}) \times 100 / 10$$

The maximum positive is allotted for a plant species is 16. According to API score plant species were categorise into eight different group (e.g., not recommended, very poor, poor, moderate, good, very good, excellent and best).

## 2.4. Dust Capturing Capacity (DCC)

Dust capturing capacity was quantified with the help of Petri dish. A petri dish was oven dried and weighed the initial weight (W1). Upper and lower surface dust of a leaf washed with deionized water and transferred to the petri dish. Then petri dish was completely dried and weighed to record final weight (W2). Surface area (A) of the leaf measured with the help of graph paper. Dust capturing capacity was calculated by the following formula

$$W = W2 - W1 / A$$

## 2.5. Analysis of metal concentration in leaves

The washed leaves were completely dried at 60°C in hot air oven. 1 g leaves material were digested using 2 acid i.e. HClO<sub>4</sub> and HNO<sub>3</sub> in the ratio of 1:2 (Samecka-Cymerman and Kempers 1999). Digested solutions were diluted with double distil water and final volume make up to 50 ml. Then digested solutions were analysed for Al, Cr, Fe, Ni, Pb and Zn by PerkinElmer ICP-OES (Model: Optima 2100 DV).

## 2.6. Metal Accumulation Index (MAI)

Metal accumulation capability of plant species are calculated by the using metal accumulation index (MAI) formula (Hu et al., 2014).

$$MAI = \left(\frac{1}{N}\right) \sum_{j=1}^n IJ$$

Where, N = Number of metals, IJ = is sub-index of J gained by dividing the metal concentration by its standard deviation. It is depend on collected sample number.

## 3. Results And Discussions

### 3.1. Ascorbic acid (A)

Ascorbic acid or vitamin C is an important low molecular weight non-enzymatic antioxidant present in plant chloroplasts. This antioxidant plays a crucial function in light reaction of photosynthesis, regulate various metabolic biosynthesis pathway and it also protect plant from biotic and abiotic stress (Singh and Verma, 2007). Throughout photosynthetic electron transfer, ascorbic acid react and scavenge reactive oxygen species (ROS). Therefore biosynthesis and concentration of this antioxidant directly proportionate to tolerance level of plant (Smrinoff and Wheeler, 2000; Wang et al., 2018). In mining area highest ascorbic acid content (mg/g in fresh leaves) was found in *H. pubescence* (26.83 ± 1.01) followed by *M. peltata* (24.66 ± 0.99), *T. orientalis* (22.81 ± 0.8), *F. hispida* (19.63 ± 2.77), and lowest content was noticed in *C. roxburghii* (8.76 ± 0.95), followed by *C. tomentosa* (9.46 ± 1.93) and *W. fruticosa* (9.86 ± 1.22). In comparison to the control area considerable difference was found in ascorbic acid contents in all plant species (Figure: 1). In control area highest amount found in *M. peltata* (14.28 ± 0.93) followed by *T. arjuna* (12.85 ± 0.57) and *H. pubescence* (11.36 ± 2.22) and lowest amount found in *C. tomentosa* (4.11 ± 0.7) followed by *C. roxburghii* (4.72 ± 1.03) and *A. excels* (5.56 ± 0.91).

Ascorbic acid react with various ROS (i.e. superoxide radical, singlet oxygen and hydroxyl radical) and protects pigments and nucleic acid. Elevated amount of ascorbic acid increases the tolerance level of plant under pollution environment as well as others stress condition (like, drought, salt, temperature etc.) (Wang et al., 2018). Hence, plants synthesized higher level ascorbic acid in polluted environment are evaluated as tolerant species. In our study, ascorbic acid content is higher at mining area and lower at control area for all examined plant species. The enhancement of ascorbic acid in all species from mining area might be due to elevated production of ROS, through accumulation of toxic metal in plant body.

### 3.2. Relative Water Content (R)

Relative water content signify the balance between plant water uptake and release. Water content maintain turgor pressure and cell wall permeability in plant body. During pollution condition different pollutant enhance cell permeability, which leads to deficiency of water and nutrients, derive to prior senescence of leaves and bark (Achakzai et al., 2017; Safari et al, 2019). So, relative water content of plants is a suitable physiological parameter to measure water condition in plant body. All the species had more relative water content in control area than mining area (Figure: 1). Highest relative water content in mining area was found in *M. peltata* ( $93.71 \pm 2.69$ ) and lowest in *A. excelsa* ( $64.81 \pm 7.76$ ). In control area, the relative water content of leaf of the plant species differed from maximum of  $98.22 \pm 0.33$  in *F. hispida* to a minimum of  $69.31 \pm 1.37$  in *A. excelsa* (Table).

The relative water content of plant leaves in mining area is decreased might be due to high pollutants and heavy metal contamination, it also specify unbalanced physiological condition of the plant. Heavy metal contamination in plant body reduce the water transport from root to leaves (Bercelo and Poschenrieder, 1990). Under any stress condition high water content help to sustain physiological and biochemical balance in a plant body. Higher level of relative water content in plants increases the tolerance strength of plant towards pollution stress (Gupta et al., 2016; Karmakar and Padhy, 2019).

### 3.3. Total chlorophyll content (T)

The photosynthetic pigment chlorophyll, found in the chloroplasts of green plants, is an index of productivity and is called a photoreceptor. Chlorophyll plays an important role in plant photosynthesis, so measurement of total chlorophyll content is a significant measure to assess the effect of pollution on the plant. The decreased level of total chlorophyll content of all species were noticed in the mining area as compared to the control area (Figure: 1). At mining area highest chlorophyll content (mg/g of fresh weight) was found in *H. pubescence* ( $1.81 \pm 0.01$ ) followed by *T. orientalis* ( $1.3 \pm 0.11$ ) and *M. peltata* ( $1.12 \pm 0.05$ ). Lowest found in *B. Orellana* ( $0.44 \pm 0.01$ ) followed by *W. fruticosa* ( $0.58 \pm 0.007$ ) and *C. roxburghii* ( $0.61 \pm 0.009$ ). At control area highest found in *H. pubescence* ( $2.33 \pm 0.22$ ) followed by *M. peltata* ( $1.43 \pm 0.24$ ) and *T. orientalis* ( $1.39 \pm 0.19$ ). Lowest found in *B. Orellana* ( $0.75 \pm 0.09$ ) followed by *F. hispida* ( $0.9 \pm 0.07$ ) and *A. excelsa* ( $1.06 \pm 0.13$ ). Chlorophyll have a highly organized state that may go through various photochemical reaction like oxidation, reduction and reversible bleaching in different pollutant condition (Karmakar et al., 2021). Decreased level of chlorophyll at mining area might be due to dust deposition on leaves surface and heavy metal accumulation. Dust deposition on leaves surface inhibit the sunlight absorption and blocks the stomatal pore, thus decreased the rate of photosynthesis which is important for chlorophyll synthesis (Roy et al., 2020). After accumulation of heavy metal,  $Mg^{++}$  ion of chlorophyll replace by the another metal ion and breakdown into phaeophytin (Karakoti et al, 2014). High pollution stress reduces the total chlorophyll content at the polluted site. But those species are able to maintain a high level of chlorophyll known as tolerant species.

### 3.4. Leaves Extract pH (P)

Leaves extract pH is a biochemical parameter of plant that signify as an indication of stress. The pH of the leaf extracts ranges from  $6.8 \pm 0.13$  to  $5.1 \pm 0.06$  and  $7.17 \pm 0.14$  to  $5.53 \pm 0.12$ , at polluted and control area respectively, with *F. hispida* and *B. orellana* having highest and lowest value. The pH of plants has shown a good relationship with the susceptibility to pollution. Low pH reduced the photosynthetic activity by altering the stomatal activity while a higher level of pH in plants can increase tolerance toward pollution by enhancing the synthesis rate of ascorbic acid from hexose sugar (Escobedo et al., 2008). The presence of acidic pollutant in the environment increases the acidic nature of the leaf by decreasing pH. Hence, those plant species have higher leaf extract pH are considered as pollution tolerant species. It is evidenced that all the plant species collected from mining area showed an acidic pH. Leaves pH decline in mining areas may be due to the existence of heavy metal in the environment. Karmakar & Padhy (2019) and Nadgorska-Sochaet al. (2017) has been reported that leaves pH influences in the presence of heavy metal like Cd, Cr, Ni, and Zn.

### 3.5. Air pollution tolerance index (APTI)

The APTI is an established method for evaluation of plants with regarding to their sensitivity to pollutants. Depending on APTI value plants species are categories into four different group i.e.  $\leq 15$  = sensitive,  $15-19$  = Intermediate,  $20-24$  = moderately tolerant and  $> 24$  = tolerant (Singh et al., 1991). APTI values of each plant species were calculated using the mean values of T, P, R and A values, so the standard deviation of APTI values are not mentioned (Table: 1). At mining area shown that APTI value range from 30.24 to 12.48. Highest value was

recorded in *H. pubescence* (30.24) followed by *M. peltata* (25.96), *T. orientalis* (25.62) and *F. hispida* (22.52). According to APTI range *H. pubescence*, *M. peltata* and *T. orientalis* are tolerant species as their APTI value  $\geq 24$ . *F. hispida* (22.52) is only moderately tolerant species. *C. roxburghii* (18.71), and *T. arjuna* (16.51) are the intermediate species. Whereas, *A. excelsa*, *B. Orellana*, and *C. tomentosa* are sensitive species (APTI  $\leq 15$ ). At control area the APTI showed a decline (Figure: 1) value ranging from 21.51 to 11.41, with *M. peltata* and *C. roxburghii* having highest and lowest value respectively. All species are grouped into three category i.e. sensitive, intermediate, moderately tolerant, no species could qualify as tolerant at control areas. A strong correlation between ascorbic acid and APTI at both mining ( $R^2 = 0.86$ ), and control area ( $R^2 = 0.83$ ) was noticed. Whereas remaining three parameters (pH, total chlorophyll, and relative water content) have insignificant low correlation with APTI. This indicates that, as the pollution level increases ascorbic acid in plants also increases to combat the pollution stress. Some previous study also have found similar kind of correlation between ascorbic acid and APTI value (Rai & Panda 2014, Bharti et al. 2018; Roy et al. 2020). Depending on the pollution condition, same species can act differently such as, tolerant, intermediate or sensitive. Plantation of tolerant plant species, to mitigate environmental pollution is a sustainable prospective to meet industrial and commercial growth.

### 3.6. Anticipated Performance Index (API)

Different species act differently to different pollution stress. Therefore grouping of species on the basis APTI value is only useful to evaluate tolerance status of plant. Hence, only APTI value is not enough for the selection of appropriate tolerant species to develop greenbelt in polluted area. By incorporate APTI value with economic value and morphological characters likes, plant habit, canopy structure, laminar character and plant types the API grade of different species are calculated. In the mining area *M. peltata* scored highest API value followed by *H. pubescens* and *F. hispida* qualified excellent, very good and good species respectively for greenbelt development (Table: 2). Among the rest of species two were (*T. arjuna* and *T. orientalis*) qualified as a moderate. *C. tomentosa*, *C. roxburghii*, *A. excels*, *Bixa orellana* and *W. fruticosa* are poor, very poor and not recommended for greenbelt development, because of their low API value (50 – 25). In control areas also similar result was found, *M. peltata* scored highest API value followed by *H. pubescens* and *F. hispida*. But all three species qualified as a good species for greenbelt development. Among the rest of the plant species, *T. arjuna* is moderate, *C. tomentosa* is poor, three species (*A. excelsa*, *B. orellana*, and *C. roxburghii*) are very poor and *W. fruticosa* is not recommended for greenbelt development. Screening all the species by using only APTI value *H. pubescence*, *M. peltata* and *T. orientalis* has been assessed as tolerant species and *F. hispida* was moderately tolerant species. But after combine APTI value with morphological and economic value, then *F. hispida* qualified as a tolerant species along with *H. pubescence* and *M. peltata* and *T. orientalis* qualified as a moderate along with *T. arjuna*.

### 3.7. Dust capturing capacity (DCC)

Variation of dust capturing capacity of collected species at mining and control areas were depicted in (Table: 3). *F. hispida* was found the highest dust capturing capacity (5.94 mg/cm<sup>2</sup>) in mining area followed by *C. tomentosa* (4.26 mg/cm<sup>2</sup>), *H. pubescence* (3.6 mg/cm<sup>2</sup>), *M. peltata* (3.51 mg/cm<sup>2</sup>) and *T. orientalis* (3.5 mg/cm<sup>2</sup>). Remaining species (*A. excels*, *B. orellana*, *C. roxburghii*, *T. arjuna*, *W. fruticosa*) showed the lower dust capturing capacities in mining area. In control areas *M. peltata* (0.39 mg/cm<sup>2</sup>) showed highest dust capturing capacities followed by *C. tomentosa* (0.37 mg/cm<sup>2</sup>) and *F. hispida* (0.36 mg/cm<sup>2</sup>) and lowest dust capturing capacities showed by *W. fruticosa* (0.14 mg/cm<sup>2</sup>) followed by *C. roxburghii* (0.16 mg/cm<sup>2</sup>) and *B. orellana* (0.17 mg/cm<sup>2</sup>). Dust capturing capacity of plant significantly depend on species to species. Several micro-characters, macro-characters and surrounding environment affect the plant's dust capturing capacity. Micro-characters such as stomatal size and density, presence of trichome, pubescence, wax layer, and macro-characters like height of plant, canopy structure, leaves arrangement on stem, petiole area significantly influence the dust capturing capacity of plant species (Mo et al., 2015; Leonard et al., 2016) *F. hispida*, *C. tomentosa*, *H. pubescence*, *M. peltata* and *T. orientalis* contribute high level of dust capturing capacities due to presence of trichome in both side of leaves surface. Similar findings were supported some recent studies by Roy et al. (2020) and Chaudhary et al. (2019) found higher dust capturing capacities in genus *Ficus*. Roy et al. (2020) also found that *Ailanthus excelsa* had lowest value due to its small and glabrous leaf petiole. According to single factor ANOVA showed significant differences in dust capturing capacities in both areas ( $F = 24.11$ ,  $F_{critical} = 4.41$ ), at 0.05 significance level. Area wise differences may be due to high atmospheric pollution and anthropogenic activity.

### 3.8. Assessment of heavy metal within the leaf

Heavy metals accumulation in plants can be from both, adsorption through stomata from areal dust deposition and root uptake from soil. So, it impossible to distinguish whether the accumulated heavy metal came from the soil or from the air (Serbula et al. 2012; Norouzi et al. 2015). In mining areas analysed plant leaves revealed that they were surpass the permissible limit of WHO (1996) standards. Al, Cr and Fe content were extremely higher within the all investigated plants leaves. Average metals accumulation ranges of all investigated plants at both the area are represented in (Table: 4). Fe concentration found higher in all plant leaves of mining area ranging from 1028.87  $\pm$  55.69 to 6793.93  $\pm$  797.15 mg/kg and in the control area ranged between 245.89  $\pm$  36.76 to 57.04  $\pm$  15.18. Highest Fe accumulation found in *F. hispida* leaves in mining area (6793.93  $\pm$  797.15 mg/kg) and in *C. tomentosa* leaves in control area (245.89  $\pm$  36.76 mg/kg). Al accumulation ranged between

159.43 ± to 1358.70 ± mg/kg at the mining area. The highest Al accumulation was detected in *F. hispida* followed by *M. peltata* (833.07 ± mg/kg) and lowest was detected in *W. fruticosa* followed by *T. orientalis* (183.20 ± mg/kg). Compared with control area highest accumulation found in *F. hispida* (5.23 ± mg/kg) and lowest was found in *T. orientalis* (0.62 ± mg/kg). The highest content of Cr accumulation were found in *F. hispida* (432.27 ± mg/kg) followed by *M. peltata* (328.03 ± mg/kg) and lowest accumulation were found in *C. tomentosa* (48.97 ± mg/kg) followed by *B. orellana* (52.83 ± mg/kg). Maximum accumulation of Ni were found in *C. roxburghii* (64.57 ± mg/kg) followed by *M. peltata* (62.57 ± mg/kg).

### 3.9. Metal Accumulation Index (MAI)

MAI is used to evaluate the heavy metals accumulation capacities in plants using standard formula (Hu et al., 2014; Karmakar & Padhy, 2019). The MAI value of analysed plant species are given in the (Table: 4). At mining area MAI ranged from 4.5 (*W. fruticosa*) to 17.29 (*C. tomentosa*). The trend followed was *C. tomentosa* (17.29) > *F. hispida* (10.31) > *M. peltata* (10.16) > *H. pubescens* (9.74) > *A. excelsa* (7.46) > *T. arjuna* (7.05) > *T. orientalis* (6.89) > *B. orellana* (5.17) > *W. fruticosa* (4.5). At control area the trend followed was *F. hispida* (6.38) > *C. roxburghii* (3.4) > *T. arjuna* (3.17) > *M. peltata* (3.15) > *C. tomentosa* (2.88) > *A. excelsa* (2.82) > *W. fruticosa* (2.57) > *H. pubescens* (2.34) > *T. orientalis* (2.26) > *B. orellana* (1.94). In both area *F. hispida* had higher MAI value (mining area = 10.31, control area = 6.38), indicating that it has higher metal accumulation capacity. However, MAI of *A. excelsa* (7.46) in Sukinda chromite mine area is higher than previously reported by Roy et al. (2020) in industrial area (2.13) and commercial area (2.03). In another study at China similar kind of result was found by Hu et al. (2014). MAI value of *Sophora japonica* in Beijing (9.0) was higher than reported in Yan'an city (2.56). This fluctuation of MAI value may mainly depending on local atmospheric chemistry and geology. It also depend on the other factors, such as amount of metal deposited on leaf surface, sampling height, time and plant characteristics as well as the capacity of removal air pollutant and accumulation of metal (Castanheiro et al., 2020; Jia et al., 2021). Plant species with higher MAI values have a better metal accumulation capacity and considered metal tolerant species, those species can use impediment between contaminated and uncontaminated area. Those plant species that have lower MAI value are considered sensitive to metal and can be used as bioindicators of metal pollution.

## 4. Conclusion

Based on dust capturing capacities and metal accumulation index as well as APTI and API indices *F. hispida*, *M. peltata* and *H. pubescens* can be use as tolerant species and could be recommended for greenbelt development to mitigate pollution. Those plant species (*A. excelsa*, *B. orellana*, *C. roxburghii*, *W. fruticosa*, *T. arjuna*, *T. orientalis*) have lower APTI and API value could be used as sensitive species for biomonitoring of environmental pollution. Our study also finds that *C. tomentosa* is the highest metal accumulation capacities, but according to anticipated performance index it is a poor performer toward pollution. So, these species could be used as hyper-accumulator to mitigate heavy metal contamination from air and soil. The physiological and biochemical response of plant varies on species to species and also area to area. But, more biochemical and physiological research is needed to confirm the effect and response of plant species in a particular pollution stress condition.

## Declarations

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## References

1. Achakzai K, Khalid S, Adrees M, Bibi A, Ali S, Nawaz R, Rizwan M (2017) Air pollution tolerance index of plants around brick kilns in Rawalpindi, Pakistan. *Journal of environmental management* 190:252–258
2. Arnon DI (1949) Copper enzymes in isolated chloroplasts. Polyphenoloxidase in *Beta vulgaris*. *Plant physiology* 24(1):1
3. Barceló JUAN, Poschenrieder C (1990) Plant water relations as affected by heavy metal stress: a review. *Journal of plant nutrition* 13(1):1–37
4. Bhaduri AM, Fulekar MH (2012) Antioxidant enzyme responses of plants to heavy metal stress. *Reviews in Environmental Science Bio/Technology* 11(1):55–69
5. Bharti SK, Trivedi A, Kumar N (2018) Air pollution tolerance index of plants growing near an industrial site. *Urban climate* 24:820–829
6. Black Smith Institute Report (2007) The world's worst polluted places. A project of Blacksmith Institute, pp 16–17
7. Castanheiro A, Hofman J, Nuyts G, Joosen S, Spassov S, Blust R, Lenaerts S, De Wael K, Samson R (2020) Leaf accumulation of atmospheric dust: Biomagnetic, morphological and elemental evaluation using SEM, ED-XRF and HR-ICP-MS. *Atmos Environ* 221:117082
8. Chaudhary IJ, Rathore D (2019) Dust pollution: Its removal and effect on foliage physiology of urban trees. *Sustainable Cities Society* 51:101696
9. Chen YM, Lucas PW, Wellburn AR (1991) Relationship between foliar injury and changes in antioxidant levels in red and Norway spruce exposed to acidic mists. *Environ Pollut* 69(1):1–15
10. Christou A, Georgiadou EC, Zissimos AM, Christoforou IC, Christofi C, Neocleous D, Dalias P, Torrado SO, Argyraki A, Fotopoulos V (2020) Hexavalent chromium leads to differential hormetic or damaging effects in alfalfa (*Medicago sativa* L.) plants in a concentration-dependent manner by regulating nitro-oxidative and proline metabolism. *Environ Pollut* 267:115379
11. Das PK, Das BP, Dash P (2020) Chromite mining pollution, environmental impact, toxicity and phytoremediation: a review. *Environmental Chemistry Letters*, 1–13
12. Das PK, Das BP, Dash P (2020) Chromite mining pollution, environmental impact, toxicity and phytoremediation: a review. *Environmental Chemistry Letters*, 1–13
13. DesMarias TL, Costa M (2019) Mechanisms of chromium-induced toxicity. *Current opinion in toxicology* 14:1–7
14. Dhal B, Das NN, Pandey BD, Thatoi HN (2011) Environmental quality of the Boula-Nuasahi chromite mine area in India. *Mine Water Environ* 30(3):191–196
15. Escobedo FJ, Wagner JE, Nowak DJ, De la Maza CL, Rodriguez M, Crane DE (2008) Analyzing the cost effectiveness of Santiago, Chile's policy of using urban forests to improve air quality. *Journal of environmental management* 86(1):148–157
16. Gheorghe IF, Ion B (2011) The effects of air pollutants on vegetation and the role of vegetation in reducing atmospheric pollution. *The impact of air pollution on health, economy, environment and agricultural sources*, 241–280
17. Golui D, Datta SP, Dwivedi BS, Meena MC, Trivedi VK, Jaggi S, Bandyopadhyay KK (2021) Assessing Geoavailability of Zinc, copper, nickel, lead and cadmium in polluted soils using short sequential extraction scheme. *Soil Sediment Contamination: An International Journal* 30(1):74–91
18. Gupta GP, Kumar B, Kulshrestha UC (2016) Impact and pollution indices of urban dust on selected plant species for green belt development: mitigation of the air pollution in NCR Delhi, India. *Arab J Geosci* 9(2):136
19. Hu Y, Wang D, Wei L, Zhang X, Song B (2014) Bioaccumulation of heavy metals in plant leaves from Yan'an city of the Loess Plateau, China. *Ecotoxicol Environ Saf* 110:82–88
20. Indian bureau of mines (2013) Monograph on chromite. 1–153
21. Javanmard Z, Kouchaksaraei MT, Hosseini SM, Pandey AK (2020) Assessment of anticipated performance index of some deciduous plant species under dust air pollution. *Environ Sci Pollut Res* 27(31):38987–38994
22. Jia M, Zhou D, Lu S, Yu J (2021) Assessment of foliar dust particle retention and toxic metal accumulation ability of fifteen roadside tree species: Relationship and mechanism. *Atmospheric Pollution Research* 12(1):36–45
23. Jones MB (1985) Plant microclimate. In *Techniques in bioproductivity and photosynthesis* 26–40
24. Karakoti N, Bajpai R, Upreti DK, Mishra GK, Srivastava A, Nayaka S (2014) Effect of metal content on chlorophyll fluorescence and chlorophyll degradation in lichen *Pyxine coccinea* (Sw.) Nyl.: a case study from Uttar Pradesh, India. *Environmental earth sciences* 71(5):2177–2183

25. Karmakar D, Padhy PK (2019) Air pollution tolerance, anticipated performance, and metal accumulation indices of plant species for greenbelt development in urban industrial area. *Chemosphere* 237:124522
26. Karmakar D, Deb K, Padhy PK (2021) Ecophysiological responses of tree species due to air pollution for biomonitoring of environmental health in urban area. *Urban Climate* 35:100741
27. Kaur M, Nagpal AK (2017) Evaluation of air pollution tolerance index and anticipated performance index of plants and their application in development of green space along the urban areas. *Environ Sci Pollut Res* 24(23):18881–18895
28. Keller T (1986) The electrical conductivity of Norway spruce needle diffusate as affected by certain air pollutants. *Tree physiology* 1(1):85–94
29. Khazini L, Dehkharghanian ME, Vaezihir A (2021) Dispersion and modeling discussion of aerosol air pollution caused during mining and processing of open-cast mines. *International Journal of Environmental Science and Technology*, 1–12
30. Kong SSK, Fu JS, Dong X, Chuang MT, Ooi MCG, Huang WS, Griffith SM, Pani SK, Lin NH (2021) Sensitivity analysis of the dust emission treatment in CMAQv5. 2.1 and its application to long-range transport over East Asia. *Atmospheric Environment*, p 118441
31. Lèbre É, Corder G, Golev A (2017) The role of the mining industry in a circular economy: a framework for resource management at the mine site level. *J Ind Ecol* 21(3):662–672
32. Leonard RJ, McArthur C, Hochuli DF (2016) Particulate matter deposition on roadside plants and the importance of leaf trait combinations, 20. *Urban Forestry & Urban Greening*, pp 249–253
33. Liu YJ, Ding HUI (2008) Variation in air pollution tolerance index of plants near a steel factory: Implication for landscape-plant species selection for industrial areas. *WSEAS Transactions on Environment development* 4(1):24–32
34. Manara A (2012) Plant responses to heavy metal toxicity. In: *Plants and heavy metals*. Springer, Dordrecht, pp 27–53
35. Mo L, Ma Z, Xu Y, Sun F, Lun X, Liu X, Chen J, Yu X (2015) Assessing the capacity of plant species to accumulate particulate matter in Beijing, China. *Plos one* 10(10):140664
36. Molnár V, Simon E, Tóthmérész B, Ninsawat S, Szabó S (2020) Air pollution induced vegetation stress—the air pollution tolerance index as a quick tool for city health evaluation. *Ecol Ind* 113:106234
37. Mondal D, Gupta S, Datta JK (2011) Anticipated performance index of some tree species considered for green belt development in an urban area. *International Research Journal of Plant Science* 2(4):99–106
38. Nadgórska–Socha A, Kandziora-Ciupa M, Trzęsicki M, Barczyk G (2017) Air pollution tolerance index and heavy metal bioaccumulation in selected plant species from urban biotopes. *Chemosphere* 183:471–482
39. Nayak S, Kale P (2020) A review of chromite mining in Sukinda Valley of India: impact and potential remediation measures. *Int J Phytoremediation* 22(8):804–818
40. Ni ZZ, Luo K, Zhang JX, Feng R, Zheng HX, Zhu HR, Wang JF, Fan JR, Gao X, Cen KF (2018) Assessment of winter air pollution episodes using long-range transport modeling in Hangzhou, China, during World Internet Conference, 2015. *Environmental pollution*, 236, 550–561
41. Norouzi S, Khademi H, Cano AF, Acosta JA (2015) Using plane tree leaves for biomonitoring of dust borne heavy metals: a case study from Isfahan, Central Iran. *Ecological indicators* 57:64–73
42. Oluwoye I, Dlugogorski BZ, Gore J, Oskierski HC, Altarawneh M (2017) Atmospheric emission of NO<sub>x</sub> from mining explosives: A critical review. *Atmos Environ* 167:81–96
43. Perl-Treves R, Perl A (2002) Oxidative stress: an introduction. *Oxidative stress in plants*, 1–32
44. Puckett KJ, Nieboer E, Flora WP, Richardson DHS (1973) Sulphur dioxide: its effect on photo-synthetic <sup>14</sup>C fixation in lichens and suggested mechanisms of phytotoxicity. *New Phytol* 72(1):141–154
45. Rai PK, Panda LL (2014) Dust capturing potential and air pollution tolerance index (APTI) of some road side tree vegetation in Aizawl, Mizoram, India: an Indo-Burma hot spot region. *Air Quality Atmosphere Health* 7(1):93–101
46. Rai PK (2016) Impacts of particulate matter pollution on plants: Implications for environmental biomonitoring. *Ecotoxicol Environ Saf* 129:120–136
47. Remon E, Bouchardon JL, Le Guédard M, Bessoule JJ, Conord C, Faure O (2013) Are plants useful as accumulation indicators of metal bioavailability? *Environ Pollut* 175:1–7
48. Roy A, Bhattacharya T, Kumari M (2020) Air pollution tolerance, metal accumulation and dust capturing capacity of common tropical trees in commercial and industrial sites. *Science of The Total Environment* 722:137622
49. Safari F, Akramian M, Salehi-Arjmand H, Khadivi A (2019) Physiological and molecular mechanisms underlying salicylic acid-mitigated mercury toxicity in lemon balm (*Melissa officinalis* L.). *Ecotoxicol Environ Saf* 183:109542

50. Sahu C, Basti S (2020) Trace metal pollution in the environment: a review. *International Journal of Environmental Science and Technology*, 1–14
51. Samecka-Cymerman A, Kempers AJ (1999) Bioindication of heavy metals in the town Wrocław (Poland) with evergreen plants. *Atmos Environ* 33(3):419–430
52. Selmi W, Weber C, Rivière E, Blond N, Mehdi L, Nowak D (2016) Air pollution removal by trees in public green spaces in Strasbourg city, France, 17. *Urban Forestry & Urban Greening*, pp 192–201
53. Serbula SM, Kalinovic TS, Ilic AA, Kalinovic JV, Steharnik MM (2012) Assessment of airborne heavy metal pollution using *Pinus* spp. and *Tilia* spp. *Aerosol Air Quality Research* 13(2):563–573
54. Shahid M, Shamshad S, Rafiq M, Khalid S, Bibi I, Niazi NK, Dumat C, Rashid MI (2017) Chromium speciation, bioavailability, uptake, toxicity and detoxification in soil-plant system: a review. *Chemosphere* 178:513–533
55. Singh SK, Rao DN, Agrawal M, Pandey J, Naryan D (1991) Air pollution tolerance index of plants. *J Environ Manage* 32(1):45–55
56. Singh SN, Verma A (2007) Phytoremediation of air pollutants: a review. *Environmental bioremediation technologies*, pp 293–314
57. Smirnoff N, Wheeler GL (2000) Ascorbic acid in plants: biosynthesis and function. *CRC Crit Rev Plant Sci* 19(4):267–290
58. Wang Y, Zhao H, Qin H, Li Z, Liu H, Wang J, Zhang H, Quan R, Huang R, Zhang Z (2018) The synthesis of ascorbic acid in rice roots plays an important role in the salt tolerance of rice by scavenging ROS. *Int J Mol Sci* 19(11):3347
59. WHO (1996) Denneman and Robberse 1990; Ministry of Housing, Netherlands 1994
60. Xiu Z, Nie W, Yan J, Chen D, Cai P, Liu Q, Du T, Yang B (2020) Numerical simulation study on dust pollution characteristics and optimal dust control air flow rates during coal mine production. *J Clean Prod* 482:119197
61. Zhang PQ, Liu YJ, Chen X, Yang Z, Zhu MH, Li YP (2016) Pollution resistance assessment of existing landscape plants on Beijing streets based on air pollution tolerance index method. *Ecotoxicol Environ Saf* 132:212–223

## Tables

Table 1  
Assessment of APTI of selected plant species from mining and control area

Species	Area	Total chlorophyll (mg/g)	pH	Relative water content (%)	Ascorbic acid (mg/g)	APTI
<i>Ailanthus excelsa</i>	Mining	0.73 ± 0.01	5.16 ± 0.04	64.81 ± 7.76	10.2 ± 0.81	12.48
	Control	1.06 ± 0.13	6 ± 0.29	69.31 ± 1.32	5.56 ± 0.91	10.85
<i>Bixa orellana</i>	Mining	0.44 ± 0.01	5.1 ± 0.06	70.90 ± 1.44	12.33 ± 2.86	13.92
	Control	0.75 ± 0.09	5.53 ± 0.12	79.62 ± 1.37	9.32 ± 0.92	11.82
<i>Callicarpa tomentosa</i>	Mining	0.63 ± 0.006	6.29 ± 0.11	80.37 ± 1.64	9.46 ± 1.93	14.58
	Control	1.36 ± 0.19	6.62 ± 0.16	95.87 ± 0.75	4.11 ± 0.7	12.86
<i>Combretum roxburghii</i>	Mining	0.61 ± 0.009	5.42 ± 0.05	71.3 ± 0.95	8.76 ± 0.95	18.71
	Control	1.03 ± 0.12	5.85 ± 0.24	74.71 ± 1.3	4.72 ± 1.03	10.71
<i>Ficus hispida</i>	Mining	0.82 ± 0.002	6.8 ± 0.13	75.63 ± 1.15	19.53 ± 2.69	22.52
	Control	0.9 ± 0.07	7.17 ± 0.14	98.22 ± 0.33	8.89 ± 0.78	16.99
<i>Holarrhena pubescens</i>	Mining	1.81 ± 0.01	6.13 ± 0.02	89.38 ± 2.63	26.83 ± 1.01	30.24
	Control	2.33 ± 0.22	6.97 ± 0.14	95.63 ± 1.6	11.36 ± 2.22	20.12
<i>Macaranga peltata</i>	Mining	1.12 ± 0.05	5.61 ± 0.03	93.71 ± 2.69	24.66 ± 0.1	25.96
	Control	1.43 ± 0.24	7 ± 0.09	94.72 ± 0.88	14.28 ± 0.93	21.51
<i>Terminalia arjuna</i>	Mining	0.87 ± 0.006	5.16 ± 0.09	74.1 ± 2.13	15.1 ± 1.61	16.51
	Control	1.31 ± 0.2	6.05 ± 0.17	77.87 ± 3.03	12.85 ± 0.57	17.24
<i>Trema orientalis</i>	Mining	1.31 ± 0.11	6.27 ± 0.03	83.56 ± 5.06	22.81 ± 0.8	25.64
	Control	1.39 ± 0.19	6.41 ± 0.04	81.47 ± 1.36	8.78 ± 1.07	15
<i>Woodfordia fruticosa</i>	Mining	0.59 ± 0.007	5.41 ± 0.02	86.16 ± 2.88	9.86 ± 1.22	14.52
	Control	1.16 ± 0.19	6.32 ± 0.12	86.12 ± 4.74	7.01 ± 0.33	13.85

Table 2  
Assessment of API of selected plant species

Species	APTI	Plant habit	Canopy structure	Type of plant	Laminar			Economic importance	Grade allotted		Assessment
					Size	Texture	Hardiness		Total plus	% scoring	
<b>Mining</b>											
<i>Ailanthus excelsa</i>	+	+	+		+	+		+	6	37.5	Very poor
<i>Bixa orellana</i>	+	+	+	+	+			+	6	37.5	Very poor
<i>Callicarpa tomentosa</i>	+	+	+		++	+	+	+	8	50	Poor
<i>Combretum roxburghii</i>	++	+	+	+	+		+	+	8	50	Poor
<i>Ficus hispida</i>	+++	+	+		++	+	+	++	11	68.75	Good
<i>Holarrhena pubescens</i>	+++++	+	+	+	++			++	12	75	Very good
<i>Macaranga peltata</i>	++++	++	++	+	++	+		+	13	81.25	Excellent
<i>Terminalia tomentosa</i>	++	++	++		+			++	9	56.25	Moderate
<i>Trema orientalis</i>	++++	+	+	+	-	+		+	9	56.25	Moderate
<i>Woodfordia fruticosa</i>	+			+		+		+	4	25	Not recommended
<b>Control</b>											
<i>Ailanthus excelsa</i>		+	+		+	+		+	5	31.25	Very poor
<i>Bixa orellana</i>	+	+	+	+	+			+	6	37.5	Very poor
<i>Callicarpa tomentosa</i>	+	+	+		++	+	+	+	8	50	Poor
<i>Combretum roxburghii</i>		+	+	+	+		+	+	6	37.5	Very poor
<i>Ficus hispida</i>	++	+	+		++	+	+	++	10	62.5	Good
<i>Holarrhena pubescens</i>	+++	+	+	+	++			++	10	62.5	Good
<i>Macaranga peltata</i>	++	++	++	+	++	+		+	11	68.75	Good
<i>Terminalia arjuna</i>	++	++	++		+			++	9	56.25	Moderate
<i>Trema orientalis</i>	+	+	+	+	-	+		+	6	37.5	Very poor
<i>Woodfordia fruticosa</i>	+			+		+		+	4	25	Not recommended

Table 3

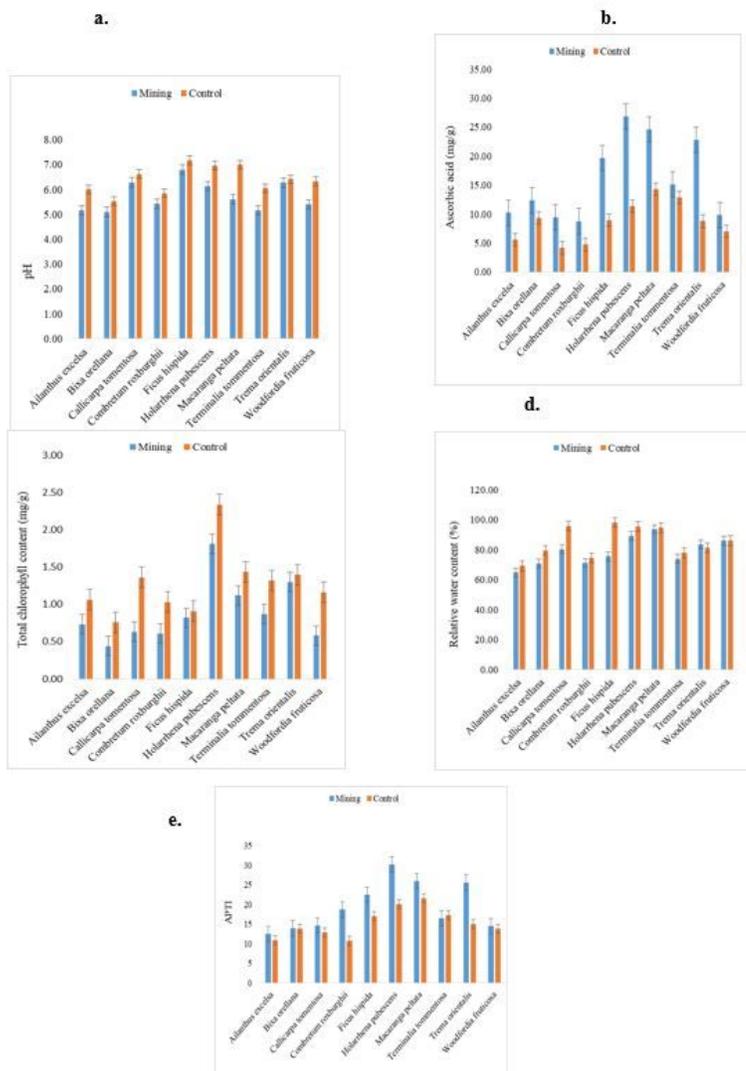
DCC of plants (mg/cm<sup>2</sup>) in mining and control area

<b>Species</b>	<b>Mining</b>	<b>Control</b>
Ailanthus excelsa	1.4 ± 0.26	0.17 ± .02
Bixa orellana	1.07 ± 0.19	0.19 ± 0.1
Callicarpa tomentosa	4.26 ± 0.27	0.37 ± .07
Combretum roxburghii	1.25 ± 0.30	0.16 ± 0.14
Ficus hispida	5.94 ± 0.43	0.36 ± 0.2
Holarrhena pubescens	3.61 ± 0.43	0.23 ± 0.04
Macaranga peltata	3.52 ± 0.32	0.39 ± 0.1
Terminalia arjuna	2.54 ± 0.55	0.33 ± 0.2
Trema orientalis	3.55 ± 0.36	0.28 ± 0.07
Woodfordia fruticosa	1.03 ± 0.11	0.14 ± 0.06

Table 4  
Heavy metal in plant leaves (mg/kg) and their respective MAI

Species	Al	Cr	Fe	Ni	Pb	Zn	MAI
<b>Mining</b>							
<i>Ailanthus excelsa</i>	389.76 ± 20.06	195.7 ± 18.74	2570 ± 188.47	38.06 ± 2.8	11.17 ± 1.26	35.6 ± 2.95	7.46
<i>Bixa orellana</i>	241.16 ± 36.26	52.58 ± 2.8	3105.33 ± 374.78	37.83 ± 6.4	2.06 ± 0.69	18.86 ± 1.62	5.17
<i>Callicarpa tomentosa</i>	239.56 ± 28.35	48.96 ± 2.58	3072 ± 30.99	19.66 ± 2.43	1.8 ± 0.53	18.65 ± 0.44	17.29
<i>Combretum roxburghii</i>	430.76 ± 23.7	151.71 ± 24.56	3870.3 ± 695.79	64.56 ± 7.66	2.03 ± 0.2	24.23 ± 2.65	5.52
<i>Ficus hispida</i>	1358.76 ± 23.7	432.26 ± 24.16	6793.93 ± 797.15	32.06 ± 1.21	1.06 ± 0.07	15.86 ± 2.25	10.31
<i>Holarrhena pubescens</i>	404.42 ± 13.47	110.4 ± 5.57	1891.93 ± 67.55	45.13 ± 4.22	1.23 ± 0.26	19.13 ± 2.37	9.74
<i>Macaranga peltata</i>	833.06 ± 25.15	328.03 ± 14.1	5239 ± 253.49	62.56 ± 4.33	2.76 ± 0.61	37.93 ± 3.92	10.16
<i>Terminalia tomentosa</i>	326.16 ± 11.2	99.36 ± 12.26	1677.53 ± 170.98	33.06 ± 2.6	6.8 ± 0.79	30.06 ± 6.03	7.05
<i>Trema orientalis</i>	183.2 ± 10.42	72.23 ± 6.37	1028.86 ± 55.69	19.26 ± 1.85	3.13 ± 0.9	20.93 ± 2.01	6.89
<i>Woodfordia fruticosa</i>	159.43 ± 14.29	162.13 ± 31.5	2138.16 ± 232.16	27.5 ± 3.01	2.18 ± 0.32	17.2 ± 1.62	4.99
<b>Control</b>							
<i>Ailanthus excelsa</i>	1.44 ± 0.33	1.85 ± 0.19	74 ± 21.41	1.82 ± 0.38	0.24 ± 0.07	4.23 ± 1.05	2.82
<i>Bixa orellana</i>	2.37 ± 0.67	1.29 ± 0.32	193.33 ± 54.07	2.11 ± 0.8	0.05 ± 0.01	3.58 ± 1.37	1.94
<i>Callicarpa tomentosa</i>	2.52 ± 0.48	1.69 ± 0.41	245.89 ± 36.75	1.09 ± 0.25	0.06 ± 0.01	4.95 ± 1.27	2.88
<i>Combretum roxburghii</i>	4.43 ± 0.72	2.56 ± 0.45	184 ± 43.18	1.3 ± 0.8	0.07 ± 0.006	0.94 ± 0.2	3.4
<i>Ficus hispida</i>	5.23 ± 1.36	3.1 ± 0.08	240.9 ± 26.78	1.88 ± 0.22	0.05 ± 0.009	2.07 ± 0.82	6.38
<i>Holarrhena pubescens</i>	0.89 ± 0.22	2.01 ± 0.46	61.55 ± 7.94	2.51 ± 0.43	0	2.01 ± 0.82	2.34
<i>Macaranga peltata</i>	1.69 ± 0.53	1.49 ± 0.26	215.11 ± 31.46	2.03 ± 0.51	0.08 ± 0.009	5.2 ± 1.22	3.15
<i>Terminalia tomentosa</i>	0.76 ± 0.17	1.76 ± 0.15	117.47 ± 18.9	1.39 ± 0.42	0.12 ± 0.02	5.85 ± 1.75	3.17
<i>Trema orientalis</i>	0.62 ± 0.28	0.53 ± 0.26	57.03 ± 15.18	1.03 ± 0.23	0.07 ± 0.02	2.46 ± 0.32	2.26
<i>Woodfordia fruticosa</i>	0.91 ± 0.3	1.35 ± 0.16	121.96 ± 28.66	1.6 ± 0.26	0.43 ± 0.01	3.37 ± 1.16	2.57

## Figures



**Figure 1**

Area wise difference in a. pH, b. Ascorbic acid, c. total chlorophyll content, d. relative water content, and e. APTI of selected plant. Error bars refer to standard error