

Cost-Effectiveness of Screening With Polymerase Chain Reaction For *Helicobacter Pylori* To Prevent Gastric Cancer And Peptic Ulcers

Aaron Oh

Columbia University Medical Center

Han Truong

Columbia University Medical Center

Judith Kim

Columbia University Medical Center

Sheila D. Rustgi

Icahn School of Medicine at Mount Sinai

Julian A. Abrams

Columbia University Medical Center

Chin Hur (✉ ch447@cumc.columbia.edu)

Columbia University Medical Center

Research Article

Keywords: Helicobacter pylori, Gastric Cancer, Peptic Ulcer Disease, Polymerase Chain Reaction

Posted Date: July 13th, 2021

DOI: <https://doi.org/10.21203/rs.3.rs-657186/v1>

License: © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Abstract

Background: *Helicobacter pylori* is a major risk factor for gastric cancer. Screening and treatment of *H. pylori* may reduce the risk of gastric cancer and peptic ulcer disease. Polymerase chain reaction (PCR) of gastric biopsies provides superior sensitivity and specificity for the detection of *H. pylori*. This study explores whether population-based *H. pylori* screening with PCR is cost-effective in the US.

Methods: A Markov cohort state-transition model was developed to compare three strategies: no screening with opportunistic eradication, ¹³C-UBT population screening and treating of *H. pylori*, and PCR population screening and treating of *H. pylori*. Estimates of risks and costs were obtained from published literature. Since the efficacy of *H. pylori* therapy in gastric cancer prevention is not certain, we broadly varied the benefit 30-100% in sensitivity analysis.

Results: PCR screening was cost-effective and had an incremental-cost effectiveness ratio per quality adjusted life-year (QALY) of \$38,591.89 when compared to ¹³C-UBT strategy with an ICER of \$2373.43 per QALY. When compared to no screening, PCR population screening reduced cumulative gastric cancer incidence from 0.84% to 0.74% and reduced peptic ulcer disease risk from 14.8% to 6.0%. The cost-effectiveness of PCR screening was robust to most parameters in the model.

Conclusion: Our modeling study finds PCR screening and treating of *H. pylori* to be cost-effective in the prevention of gastric cancer and peptic ulcer disease. However, the potential negative consequences of *H. pylori* eradication such as antibiotic resistance could change the balance of benefits of population screening.

Background

Helicobacter pylori is a known risk factor for gastric cancer, which is the third leading cause of cancer death worldwide [1]. Though more than half of the global population is infected with *H. pylori*, rates of colonization vary greatly, with higher rates in lower income nations generally related to socioeconomic status and hygiene levels [2]. While most people infected with *H. pylori* are asymptomatic, approximately 10% develop peptic ulcer disease (PUD) and 1 to 3% develop gastric adenocarcinoma [3]. *H. pylori* promotes gastric carcinogenesis through chronic gastric inflammation which progresses through the stages of atrophic gastritis, intestinal metaplasia, dysplasia, to gastric adenocarcinoma [4]. Eradication of *H. pylori* infection has been shown to reduce gastric cancer incidence [5]. With more than 60% of gastric cancers attributable to *H. pylori*, early detection and eradication of infection is important in reducing the risk of cancer [6].

H. pylori infection fulfills many of the criteria for population screening [7]. It can be detected through both noninvasive and invasive methods, each with its own advantages and limitations [8]. There are a variety of diagnostic tests available yet no single gold standard has been established in clinical practice. However, diagnostic tests with sensitivity and specificity exceeding 90% are necessary for accurate diagnosis of *H. pylori* infection. American College of Gastroenterology guidelines recommend

noninvasive ^{13}C -UBT in populations with low probability of *H. pylori* infection due to its inexpensive costs and quick results [9]. Compared to this, polymerase chain reaction (PCR) of gastric biopsies has superior sensitivity and specificity for the detection of *H. pylori* [10]. However, use of PCR testing of gastric biopsies is limited by accessibility, its inherent invasive nature, and expertise level of laboratories. Though screening and treating the population for *H. pylori* infection may reduce gastric cancer morbidity and mortality, such screening programs can be costly and difficult to implement. In this study, we aimed to evaluate the cost-effectiveness of ^{13}C -UBT and PCR population screening strategies of *H. Pylori* for the prevention of peptic ulcer disease and gastric cancer in the United States.

Methods

Model Design

A Markov state-transition cohort model was constructed in TreeAge Pro (TreeAge 2020, Williamstown, Massachusetts). The model compared three strategies: (1) no screening with opportunistic eradication, (2) single population screening for *H. pylori* using ^{13}C -UBT and treating those who tested positive with eradication therapy, and (3) single population screening for *H. pylori* with upper endoscopy and PCR of gastric biopsies and treating those who tested positive with eradication therapy. The hypothetical patient for this analysis is a 40-year-old individual from the U.S. general population. A 40-year-old individual was chosen as the target population of our model due to the sharp increase in gastric cancer incidence after 40 years of age [11]. Additionally, in countries such as Japan and Korea where screening programs exist, the recommended start age is also 40 years old [12]. The model follows patients for 60 years or until death and has a cycle length of one year.

Management Strategies

The management strategies in our analysis consisted of no screening, population based ^{13}C -UBT with eradication therapy, and population-based *H. pylori* PCR screening with eradication therapy. The health states in our model included *H. pylori* positive, *H. pylori* negative, gastric cancer, and death (Fig. 1). In addition, both *H. pylori* positive and negative individuals had ongoing risks of peptic ulcer disease throughout the model. Patients could move from any health state to death due to all-cause mortality, bleeding from peptic ulcers, or gastric cancer.

In the no screening strategy, the distribution of the cohort at the beginning of the simulation (i.e. cycle 0) was based on U.S. *H. pylori* prevalence rates. Though none of the patients initially received screening, those who developed peptic ulcer disease were tested with UBT and given eradication therapy.

For the screening strategies, the entire cohort began with screening and eradication therapy was given to those who tested positive (i.e. true-positive and false-positive). Both first-line triple therapy (proton pump inhibitors (PPI), clarithromycin, and amoxicillin) and second-line quadruple therapy (PPI, bismuth, tetracycline, and metronidazole) were modeled. Individuals who failed both lines of eradication therapy

were considered permanently *H. pylori* positive in the model. Risk of reinfection was restricted to the first five years after successful eradication.

Outcomes

The primary outcomes of interest were total cost, life expectancy, quality-adjusted life years (QALYs), and incremental cost-effectiveness ratios (ICERS). QALYs are a composite measure of the value of health outcomes that can be used to compare medical interventions where a value of 0 represents death and a value of 1.0 represents a year of perfect health. To calculate a QALY for a patient, we multiplied the utility value associated with a given health state multiplied by the cycle length of 1 year. Life expectancy was calculated as the total number of years the hypothetical patient was alive throughout the model duration without applying the health state utility values that reflected disease morbidity. A willingness to pay (WTP) threshold of \$100,000/QALY was used to determine cost-effectiveness. Secondary clinical outcomes of interest included total lifetime gastric cancer incidence. In the model, total lifetime gastric cancer incidence was determined by aggregating the proportion of patients who were in the gastric cancer health state with those who died of gastric cancer by the end of the simulation duration.

Parameter Estimates and Model Assumptions

Model parameters were based on estimates from the literature. Base-case values and ranges used in sensitivity analyses are summarized in Table 1. *H. pylori* prevalence in the U.S. was obtained from a meta-analysis by Hooi *et al* [2].

Table 1. Model inputs

CMS Centers for Medicare & Medicaid services, *GC* gastric cancer, *HP helicobacter pylori*, *PCR* polymerase chain reaction, *PPI* proton pump inhibitor, *PUD* peptic ulcer disease, *UBT* urea breath test

Risks of PUD and gastric cancer varied based on *H. pylori* status. The incidence of PUD in *H. pylori* positive individuals was 6 to 10-fold higher than for uninfected individuals [13, 14]. For patients who were *H. pylori* negative, they had a 0.66 relative risk of gastric cancer compared to those who were *H. pylori* positive [15]. Due to scarce data regarding gastric cancer risk for individuals who were never infected compared to those who were successfully eradicated of *H. pylori*, the 0.66 relative risk was applied for both groups. Though the risk of gastric cancer in individuals successfully treated of *H. pylori* could still be higher than the risk for those never infected due to developed gastric atrophy, we assumed the two risks were the same in the principal analysis.

Costs

The present study was performed from a third-party payer perspective. The model included direct medical costs of ¹³C-UBT, upper endoscopy and PCR of gastric biopsies, and eradication therapies. Gastric cancer treatment costs varied with age and were divided into first year, continuing care, and final year of death [16]. Costs were accrued from time of screening until death. Published cost estimates from prior years

Parameter	Base-Case Estimate	Range Used in Sensitivity Analysis	Distribution for PSA	Sources
Start Age	40			
Probabilities				
All-Cause Mortality - General	life table			[25]
UBT Sensitivity	0.96	(0.92–1)	β	[26]
UBT Specificity	0.93	(0.86–1)	β	[26]
PCR Sensitivity	1	(0.96–1)	β	[27]
PCR Specificity	0.98	(0.96–1)	β	[27]
HP Prevalence	0.356	(0.267–0.445)	β	[2]
HP Recrudescence Rate	0.0267	(0.0200–0.0334)	β	[28]
HP Reinfection Rate	0.0145	(0.0108–0.0181)	β	[28]
1 st Line Eradication Success Rate	0.80	(0.6–1)	β	[29, 30]
2 nd Line Eradication Success Rate	0.81	(0.61–1)	β	[31]
Risk of PUD – HP Negative	0.00125	(0.000938–0.00156)	β	[32]
Risk of PUD – HP Positive	0.01	(0.0075–0.0125)	β	[14]
PUD Mortality	0.031	(0.023–0.039)	β	[33, 34]
Risk of Gastric Cancer – HP Negative	0.000183	(0.000137–0.000229)	β	[11, 15, 35, 36]
Risk of Gastric Cancer – HP Positive	0.000277	(0.000208–0.000346)	β	[11, 21]
Excess Gastric Cancer Risk Reduction Attributable to HP eradication	100%	(30-100)	β	
GC 5 Year Mortality Ages < 45	0.66	(0.495–0.825)	β	[11]
GC 5 Year Mortality Ages 45-54	0.646	(0.485–0.808)	β	[11]
GC 5 Year Mortality Ages 55-64	0.653	(0.490–0.816)	β	[11]
GC 5 Year Mortality Ages 65-74	0.655	(0.491–0.819)	β	[11]
GC 5 Year Mortality Ages 75+	0.761	(0.561–0.951)	β	

Utilities				
Healthy	1			[37]
HP Infection	0.90	(0.80–1)	β	[38]
Gastric Cancer	0.68	(0.58–1)	β	[38]
Disutility of Upper Endoscopy	-0.0012	(-0.0015– -0.0009)	β	[37]
Disutility of PUD	-0.11	(-0.1375– -0.0825)	β	[39]
Costs				
Cost of UBT	75.56	(37.78–151.12)	γ	cms HCPCS 83013
Cost of PCR	603.66	(301.83–1207.32)	γ	cms HCPCS 0008U
Cost of Upper Endoscopy with Biopsy	614.80	(307.40–1229.60)	γ	[40]
Cost of 1 st Line Eradication Therapy	425.61	(212.81–851.22)	γ	[40]
Cost of 2 nd Line Eradication Therapy	118.64	(59.32–237.28)	γ	[41]
Cost of PPI	48.00	(24–96)	γ	[40]
First Year Gastric Cancer Costs < 65 years old	106199.47	(53099.74– 212398.94)	γ	[16]
First Year Gastric Cancer Costs > 65 years old	88499.46	(44249.73– 176998.92)	γ	[16]
Final Year Gastric Cancer Costs < 65 years old	187222.03	(93611.02– 374444.06)	γ	[16]
Final Year Gastric Cancer Treatment > 65 years old	124815.08	(62407.54– 249630.16)	γ	[16]
Continuing Gastric Cancer Care	4,888.13	(2444.07– 9776.26)	γ	[16]

were converted to 2020 dollars using the Consumer Price Index (U.S Bureau of Labor Statistics), and all costs were discounted an annual rate of 3%.

Utilities

Quality of life utility values relating to healthy, *H. pylori* positive, and gastric cancer states were incorporated in our model. Utility decrements due to upper endoscopy and PUD were also applied. Quality adjusted life years were discounted at an annual rate of 3%.

Sensitivity Analyses

We performed one-way deterministic sensitivity analyses by altering individual variables across a range of values to investigate the key parameters that most impacted the outcomes of the model. Due to uncertainty in the true clinical efficacy of *H. pylori* therapy for reducing excess gastric cancer risk, we varied efficacy rates with sensitivity analysis from 30–100%. In addition, a probabilistic sensitivity analysis (PSA) was performed to address parameter uncertainty. β distributions were fitted for transition probabilities and utilities, while γ distributions were fitted for cost parameters. The PSA was performed using Monte Carlo simulations with 100,000 reiterations.

Results

Our base case analysis demonstrated that both ¹³C-UBT and PCR were cost-effective treatment strategies (Table 2). The no-screening strategy yielded the lowest costs at \$1146.55 and the lowest QALY with 21.99. The ¹³C-UBT strategy had a total cost of \$1201.19 and resulted in 22.45 QALY. The PCR strategy was the most expensive strategy at \$2329.69 and yielded 22.48 QALY. Compared to the ¹³C-UBT strategy, in the context of an efficiency frontier, the ICER for the PCR strategy was \$38,591.89 per QALY. Compared with the no-screening strategy, the ICER was \$2373.43 per QALY for the PCR strategy and \$116.46 per QALY for the ¹³C-UBT strategy.

Table 2
Model outputs

Strategy	Total Cost	QALYs	ICER	GC Incidence	PUD Incidence
No Screening	\$1146.55	21.99	Reference	0.84%	14.8%
¹³ C-UBT	\$1201.19	22.45	\$116.46	Reference	0.75%
PCR + Biopsy	\$2329.69	22.48	\$2373.43	\$38591.89	0.74%

ICER incremental cost-effectiveness ratio, *GC* gastric cancer, *PUD* peptic ulcer disease, *QALYs* quality adjusted life years

The cumulative gastric cancer incidence in the no-screening strategy was 0.84% and was calibrated to reflect published SEER life gastric cancer risks for an average 40-year old in the US.[11] The lifetime gastric cancer incidence for the PCR strategy and ¹³C-UBT strategy was 0.74% and 0.75% respectively. In

the no-screening strategy, there was a 14.8% lifetime risk of PUD. In the PCR and UBT strategies, the lifetime risk of PUD dropped to 6.0% and 6.4%, respectively.

When each *H. pylori* eradication strategy was compared to the no-screening strategy, the results of the one-way sensitivity analysis showed that the ¹³C-UBT and PCR strategies were most sensitive to the risk of gastric cancer if *H. pylori* positive, the risk of gastric cancer after *H. pylori* eradication, *H. pylori* prevalence, and costs of screening (Fig. 2). However, even within the prescribed ranges of the one-way sensitivity analysis, the ¹³C-UBT and PCR strategies remained more cost effective than the no-screening strategy. When varying the efficacy of *H. pylori* eradication from 30%-100%, PCR remained the most cost-effective strategy at a WTP of \$100,000 per QALY (Fig. 3).

Cost-effectiveness acceptability curves were used to present the results of the PSA and determine the probability of any strategy being the cost effective at a given WTP (Fig. 4). When the WTP was <\$5000 per QALY, the no screening strategy was the most cost-effective strategy majority of the time. As the WTP exceeded \$60,000 per QALY, PCR became the cost-effective the majority of times.

Discussion

In this study, we compared the lifetime cost-effectiveness of different population-based *H. pylori* screening strategies for the prevention of gastric cancer for average 40-year-old Americans. To our knowledge, our study is the first to determine the cost-effectiveness of PCR as a potential screening strategy. Based on our results, the PCR strategy was the most cost-effective at the prescribed WTP threshold of \$100,000 per QALY as it resulted in the lowest lifetime risks of gastric cancer at 0.74% and peptic ulcer disease at 6.0%.

Our study findings are similar to the conclusions of prior cost-effectiveness analyses that suggest population-based screening and eradication treatment can be a viable option for the reduction of gastric cancer [17, 18]. While previous cost-effectiveness studies focused mostly on gastric cancer reduction, we also considered the additional benefits of *H. pylori* screening and eradication on peptic ulcer disease. Relative to gastric cancer, the incidence of peptic ulcer disease is much higher and can confer significant physical and financial burden in patients due to hospitalization costs, bothersome symptoms, and chronic PPI use. Our model determined that screening and eradicating *H. pylori* decreased the lifetime incidence of peptic ulcer disease from 14.8–6%. The inclusion of peptic ulcer disease allowed us to incorporate opportunistic eradication within the no screening strategy and provide a more clinically accurate representation within our model to mitigate any bias towards the intervention strategies.

Due to its superior sensitivity and specificity, PCR screening and treatment of *H. pylori* can achieve significant benefits towards long term gastric cancer reduction. However, implementation of population PCR screening is challenged by unequal availability of necessary diagnostic facilities. Furthermore, PCR of gastric biopsies requires endoscopy, an invasive procedure that comes with risk of complications such as bleeding. As determined by our model, compared to no screening, ¹³C-UBT is a cost-effective and

noninvasive alternative that also reduces gastric cancer and peptic ulcer disease. While the results of our model showed only marginal benefits towards gastric cancer reduction with PCR compared ¹³C-UBT, there can be cases where the most accurate measures are required. High risk patients for gastric cancer, such as in the context of hereditary diffuse gastric cancer or Lynch syndrome, often have earlier onset and worst prognoses [19]. In these higher risk populations, PCR screening and treating of *H. pylori* could be recommended due to its superior sensitivity and specificity to reduce as much excess risk of gastric cancer as possible.

Our study does have several limitations. Data were combined from multiple sources and some values for variables were imprecise. The relative risk of gastric cancer for *H. pylori* positive patients in our model was more conservative than what other studies had published [20, 21]. However, this conservative measure would bias the results of our model against screening and treatment. Adherence to eradication therapy is critical to the effectiveness of screening programs, and within our model we assumed 100% adherence. Compliance to a treatment program is a multifactorial process which can include factors such as complexity of treatment, trust in doctor-patient relations, and side effects of treatment. Incorporating such factors into the model was not possible and therefore a limitation in our study. While 100% adherence is not feasible in a real-world context, we assumed a perfect scenario for model simplicity and function. The exact efficacy of *H. pylori* therapy toward gastric cancer reduction is unknown. We assumed 100% efficacy in our principal analysis but varied the efficacy from 100%-30% within the sensitivity analysis to account for this limitation. While development of resistance to existing eradication therapies is an important consideration, and significant concern to public health, it is difficult to quantify the specific clinical harm it would confer to an individual patient and therefore was not included in our model. Risk of antibiotic resistance is a significant concern to public health and a considerable counterargument to a population wide test and treat strategy. Though implementation of population screening can be challenged by risks of antibiotic resistance, PCR screening can confer an additional advantage by determining which strains of *H. pylori* could be carrying genes for antibiotic resistance. This information could potentially decrease costs by informing which antibiotic regimens are most likely to succeed, reducing the need for re-tests. PCR could also provide similar benefits when testing strains for cytotoxin-associated gene A (CagA) status. As CagA positive strains are associated with higher risk of gastric cancer development, patients that are determined CagA negative by PCR could potentially withhold or delay eradication treatment for surveillance. While our study focused on the attributable risk of gastric cancer due to *H. pylori* infection, we do not account for inherited predispositions to gastric cancers. Though inherited gastric cancers make up only small fraction of all gastric cancers, the subset of the population with these heightened risk factors can benefit from a PCR screening strategy as cancer onset often occurs at younger ages for them. Furthermore, we did not consider the potential adverse effects of curing *H. pylori*, such as the development of reflux esophagitis, esophageal cancer, and autoimmune diseases [22, 23]. Rubenstein *et al.* have observed infection with *H. pylori* was inversely associated with esophagitis and Barrett's esophagus, while Lin *et al.* have noted increased risk of developing inflammatory bowel disease after receiving *H. pylori* treatment [23, 24].

Model uncertainty is an inevitable challenge faced in cost-effectiveness analyses. While the true efficacy of *H. pylori* eradication toward gastric cancer reduction is controversial, our model remained robust despite varying efficacy rates. Although our study considers the benefits of screening specifically within the United States, other countries with higher prevalence of *H. pylori* and gastric cancer, screening and treating could be even more cost-effective, or possibly cost-saving. In the US context, there is a wide ethnic and racial disparity in *H. pylori* and gastric cancer prevalence and incidence. While our model does not take into account specific racial and ethnic differences in gastric cancer risk, cost-effectiveness studies targeted for immigrants and people from higher risk regions and people groups could show increased effectiveness of screening programs.

Conclusions

In summary, our modeling analysis finds that *H. pylori* testing with upper endoscopy and PCR testing of gastric biopsies is a cost-effective option to prevent gastric cancer and peptic ulcer disease in the US population. Additional clinical studies to affirm some of our model assumptions, especially those regarding the differences in risk of gastric cancer between individuals cured of *H. pylori* and those were never infected with *H. pylori*, are needed.

Abbreviations

CagA: cytotoxin-associated gene A; ¹³C-UBT: carbon 13 urea breath test; GC: gastric cancer; HP: *H. pylori*; ICER: incremental cost-effectiveness ratio; PCR: polymerase chain reaction; PPI: proton pump inhibitor; PSA: probabilistic sensitivity analysis; PUD: peptic ulcer disease; QALY: quality adjusted life year; WTP: willingness to pay

Declarations

Ethics approval and consent to participate

Not applicable

Consent for publication

Not applicable

Availability of data and materials

All data generated or analyzed during this study are included in this article

Competing interests

The authors declare that they have no competing interests

Funding

NIH P30 CA0136969

Authors' contributions

Conceptualization: AO, HT, JK, CH

Data curation: AO, HT, JK, CH

Validation: JK, CH

Writing – review & editing: AO, HT, JK, SDR, JAA, CH

All authors read and approved the final manuscript.

References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: A Cancer Journal for Clinicians*. 2018;68(6):394-424.
2. Hooi JKY, Lai WY, Ng WK, Suen MMY, Underwood FE, Tanyingoh D, et al. Global Prevalence of *Helicobacter pylori* Infection: Systematic Review and Meta-Analysis. *Gastroenterology*. 2017;153(2):420-9.
3. Wroblewski LE, Peek RM, Jr., Wilson KT. *Helicobacter pylori* and gastric cancer: factors that modulate disease risk. *Clin Microbiol Rev*. 2010;23(4):713-39.
4. Yao X, Smolka AJ. Gastric Parietal Cell Physiology and *Helicobacter pylori*-Induced Disease. *Gastroenterology*. 2019;156(8):2158-73.
5. Chen HN, Wang Z, Li X, Zhou ZG. *Helicobacter pylori* eradication cannot reduce the risk of gastric cancer in patients with intestinal metaplasia and dysplasia: evidence from a meta-analysis. *Gastric cancer : official journal of the International Gastric Cancer Association and the Japanese Gastric Cancer Association*. 2016;19(1):166-75.
6. Parkin DM. The global health burden of infection-associated cancers in the year 2002. *International Journal of Cancer*. 2006;118(12):3030-44.
7. Wilson JMG, Jungner G, World Health O. Principles and practice of screening for disease / J. M. G. Wilson, G. Jungner. Geneva: World Health Organization; 1968.
8. Wang Y-K, Kuo F-C, Liu C-J, Wu M-C, Shih H-Y, Wang SSW, et al. Diagnosis of *Helicobacter pylori* infection: Current options and developments. *World J Gastroenterol*. 2015;21(40):11221-35.

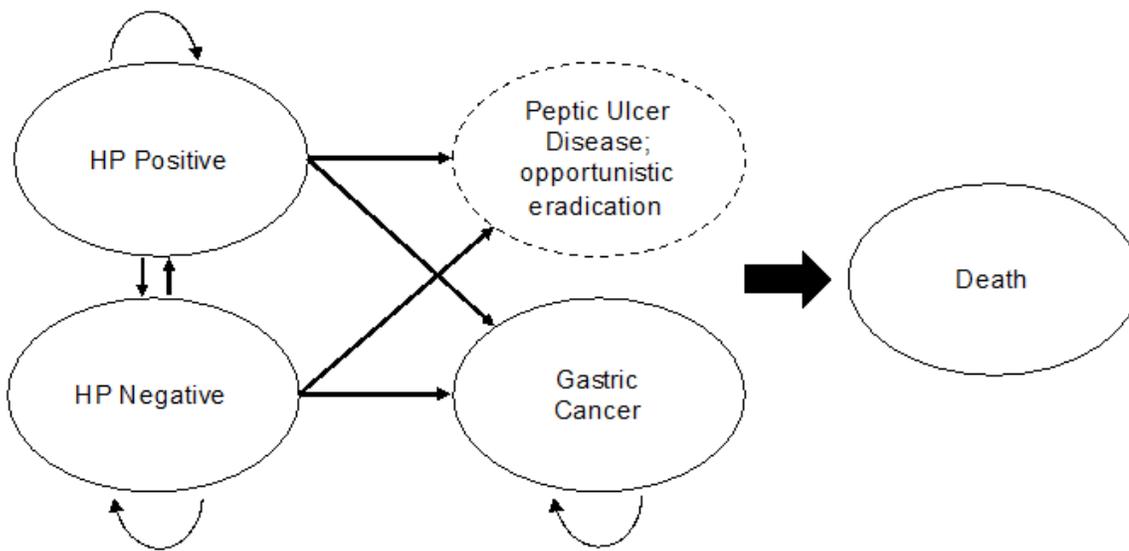
9. Chey WD, Wong BC. American College of Gastroenterology guideline on the management of *Helicobacter pylori* infection. *Am J Gastroenterol*. 2007;102(8):1808-25.
10. Patel SK, Pratap CB, Jain AK, Gulati AK, Nath G. Diagnosis of *Helicobacter pylori*: what should be the gold standard? *World J Gastroenterol*. 2014;20(36):12847-59.
11. SEER Cancer Statistics Review, 1975-2016 [Internet]. National Cancer Institute. 2019. Available from: https://seer.cancer.gov/csr/1975_2016/.
12. Lin JT. Screening of gastric cancer: who, when, and how. *Clin Gastroenterol Hepatol*. 2014;12(1):135-8.
13. Kuipers EJ, Thijs JC, Festen HP. The prevalence of *Helicobacter pylori* in peptic ulcer disease. *Aliment Pharmacol Ther*. 1995;9 Suppl 2:59-69.
14. Sipponen P, Varis K, Fraki O, Korri UM, Seppala K, Siurala M. Cumulative 10-year risk of symptomatic duodenal and gastric ulcer in patients with or without chronic gastritis. A clinical follow-up study of 454 outpatients. *Scand J Gastroenterol*. 1990;25(10):966-73.
15. Ford AC, Forman D, Hunt R, Yuan Y, Moayyedi P. *Helicobacter pylori* eradication for the prevention of gastric neoplasia. *The Cochrane database of systematic reviews*. 2015(7):Cd005583.
16. Mariotto AB, Yabroff KR, Shao Y, Feuer EJ, Brown ML. Projections of the cost of cancer care in the United States: 2010-2020. *Journal of the National Cancer Institute*. 2011;103(2):117-28.
17. Fendrick AM, Chernew ME, Hirth RA, Bloom BS, Bandekar RR, Scheiman JM. Clinical and Economic Effects of Population-Based *Helicobacter pylori* Screening to Prevent Gastric Cancer. *Archives of Internal Medicine*. 1999;159(2):142-8.
18. Xie F, Luo N, Lee HP. Cost effectiveness analysis of population-based serology screening and (13)C-Urea breath test for *Helicobacter pylori* to prevent gastric cancer: a markov model. *World J Gastroenterol*. 2008;14(19):3021-7.
19. Hansford S, Kaurah P, Li-Chang H, Woo M, Senz J, Pinheiro H, et al. Hereditary Diffuse Gastric Cancer Syndrome: CDH1 Mutations and Beyond. *JAMA Oncology*. 2015;1(1):23-32.
20. Helicobacter CCG. Gastric cancer and *Helicobacter pylori*: a combined analysis of 12 case control studies nested within prospective cohorts. *Gut*. 2001;49(3):347-53.
21. Vohlonen I, Pukkala E, Malila N, Härkönen M, Hakama M, Koistinen V, et al. Risk of gastric cancer in *Helicobacter pylori* infection in a 15-year follow-up. *Scandinavian journal of gastroenterology*. 2016;51(10):1159-64.

22. Labenz J, Blum AL, Bayerdorffer E, Meining A, Stolte M, Borsch G. Curing *Helicobacter pylori* infection in patients with duodenal ulcer may provoke reflux esophagitis. *Gastroenterology*. 1997;112(5):1442-7.
23. Lin K-D, Chiu G-F, Waljee AK, Owyang SY, El-Zaatari M, Bishu S, et al. Effects of Anti-*Helicobacter pylori* Therapy on Incidence of Autoimmune Diseases, Including Inflammatory Bowel Diseases. *Clinical Gastroenterology and Hepatology*. 2019;17(10):1991-9.
24. Rubenstein JH, Inadomi JM, Scheiman J, Schoenfeld P, Appelman H, Zhang M, et al. Association between *Helicobacter pylori* and Barrett's esophagus, erosive esophagitis, and gastroesophageal reflux symptoms. *Clin Gastroenterol Hepatol*. 2014;12(2):239-45.
25. Arias E, Xu J. United States Life Tables, 2017. National vital statistics reports : from the Centers for Disease Control and Prevention, National Center for Health Statistics, National Vital Statistics System. 2019;68(7):1-65.
26. Ferwana M, Abdulmajeed I, Alhajiahmed A, Madani W, Firwana B, Hasan R, et al. Accuracy of urea breath test in *Helicobacter pylori* infection: meta-analysis. *World J Gastroenterol*. 2015;21(4):1305-14.
27. Schabereiter-Gurtner C, Hirschl AM, Dragosics B, Hufnagl P, Puz S, Kovách Z, et al. Novel Real-Time PCR Assay for Detection of *Helicobacter pylori* Infection and Simultaneous Clarithromycin Susceptibility Testing of Stool and Biopsy Specimens. *Journal of Clinical Microbiology*. 2004;42(10):4512.
28. Niv Y, Hazazi R. *Helicobacter pylori* Recurrence in Developed and Developing Countries: Meta-Analysis of 13C-Urea Breath Test Follow-Up after Eradication. *Helicobacter*. 2008;13(1):56-61.
29. Gisbert JP, Calvet X. Review article: the effectiveness of standard triple therapy for *Helicobacter pylori* has not changed over the last decade, but it is not good enough. *Alimentary Pharmacology & Therapeutics*. 2011;34(11-12):1255-68.
30. Kowada A. Cost-effectiveness of *Helicobacter pylori* screening followed by eradication treatment for employees in Japan. *Epidemiology and Infection*. 2018;146(14):1834-40.
31. Gisbert JP. Second-line rescue therapy of *Helicobacter pylori* infection. *Therap Adv Gastroenterol*. 2009;2(6):331-56.
32. Sung JJY, Kuipers EJ, El-Serag HB. Systematic review: the global incidence and prevalence of peptic ulcer disease. *Alimentary Pharmacology & Therapeutics*. 2009;29(9):938-46.
33. Lanas A, García-Rodríguez LA, Polo-Tomás M, Ponce M, Quintero E, Perez-Aisa MA, et al. The changing face of hospitalisation due to gastrointestinal bleeding and perforation. *Alimentary Pharmacology & Therapeutics*. 2011;33(5):585-91.

34. Zhao Y, Encinosa W. Hospitalizations for Gastrointestinal Bleeding in 1998 and 2006: Statistical Brief #65. Healthcare Cost and Utilization Project (HCUP) Statistical Briefs. Rockville (MD): Agency for Healthcare Research and Quality (US); 2006.
35. Lee Y-C, Chiang T-H, Chou C-K, Tu Y-K, Liao W-C, Wu M-S, et al. Association Between Helicobacter pylori Eradication and Gastric Cancer Incidence: A Systematic Review and Meta-analysis. *Gastroenterology*. 2016;150(5):1113-24.e5.
36. Sugano K. Effect of Helicobacter pylori eradication on the incidence of gastric cancer: a systematic review and meta-analysis. *Gastric cancer : official journal of the International Gastric Cancer Association and the Japanese Gastric Cancer Association*. 2019;22(3):435-45.
37. Tengs TO, Wallace A. One Thousand Health-Related Quality-of-Life Estimates. *Medical Care*. 2000;38(6):583-637.
38. Areia M, Alves S, Brito D, Cadime AT, Carvalho R, Saraiva S, et al. Health-related quality of life and utilities in gastric premalignant conditions and malignant lesions: a multicentre study in a high prevalence country. *Journal of gastrointestinal and liver diseases : JGLD*. 2014;23(4):371-8.
39. Groeneveld PW, Lieu TA, Fendrick AM, Hurley LB, Ackerson LM, Levin TR, et al. Quality of life measurement clarifies the cost-effectiveness of Helicobacter pylori eradication in peptic ulcer disease and uninvestigated dyspepsia¹¹This work was conducted at the Division of Research, Kaiser Permanente Medical Care Program, Oakland, CA; University of California, San Francisco; and VA Medical Center, San Francisco, CA. *The American Journal of Gastroenterology*. 2001;96(2):338-47.
40. Holmes KP, Fang JC, Jackson BR. Cost-effectiveness of six strategies for Helicobacter pylori diagnosis and management in uninvestigated dyspepsia assuming a high resource intensity practice pattern. *BMC Health Services Research*. 2010;10(1):344.
41. Boklage SH, Mangel AW, Ramamohan V, Mladi D, Wang T. Cost-effectiveness analysis of universal noninvasive testing for post-treatment confirmation of Helicobacter pylori eradication and the impact of patient adherence. *Patient Prefer Adherence*. 2016;10:1025-35.

Figures

(a)



(b)

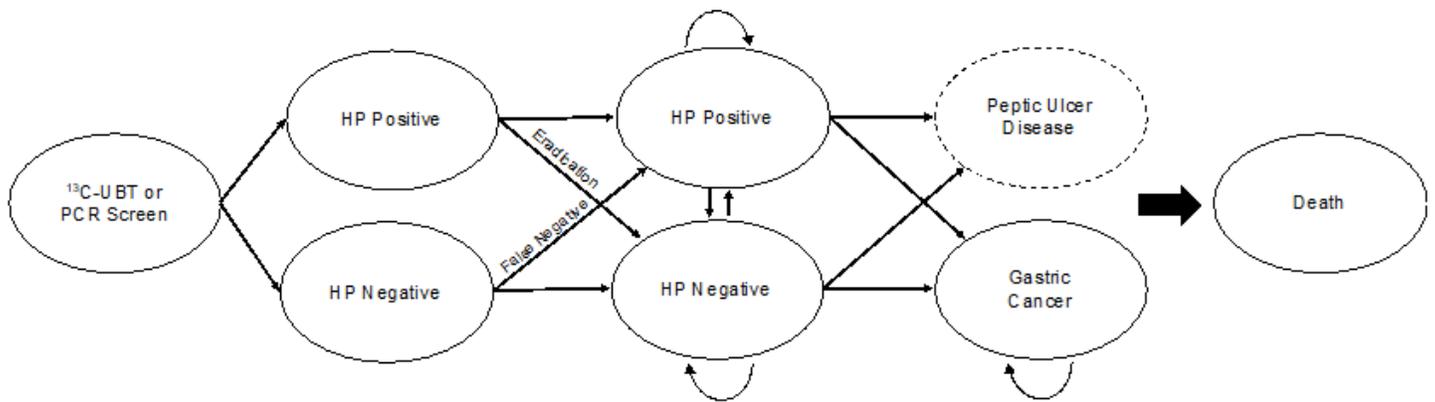
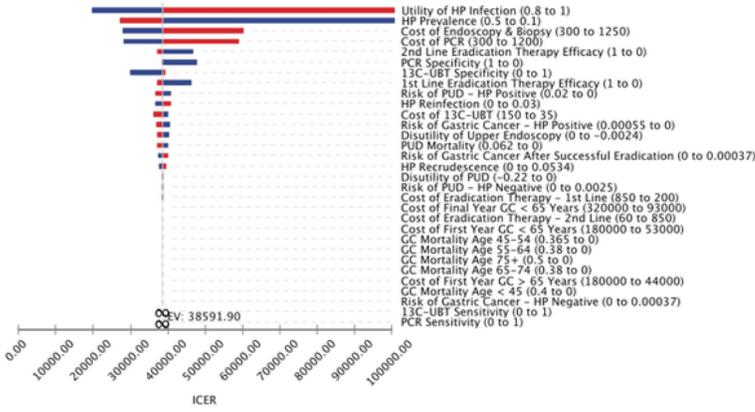


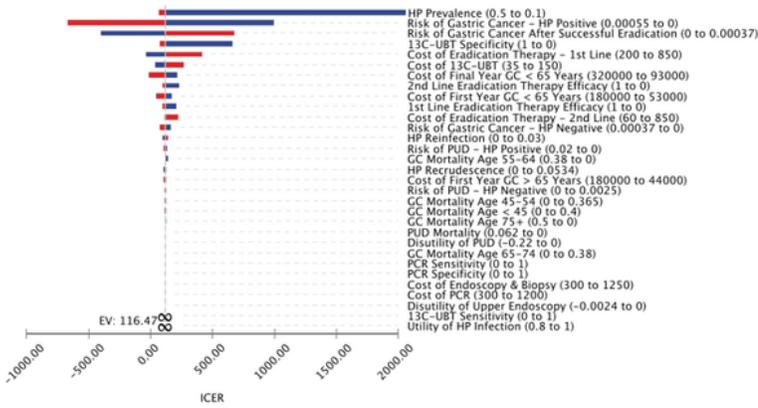
Figure 1

a. Model Schematic for No Screening Strategy. Patients can move from any health state to death. b. Model Schematic for Screening Strategies. Patients can move from any health state to death.

Tornado Diagram – ICER 13C-UBT vs. PCR



Tornado Diagram – ICER 13C-UBT vs. No Screening



Tornado Diagram – ICER PCR vs. No Screening

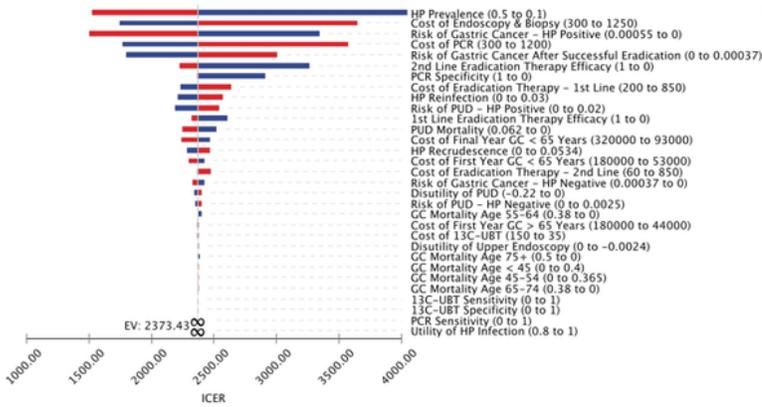


Figure 2

Tornado diagrams.

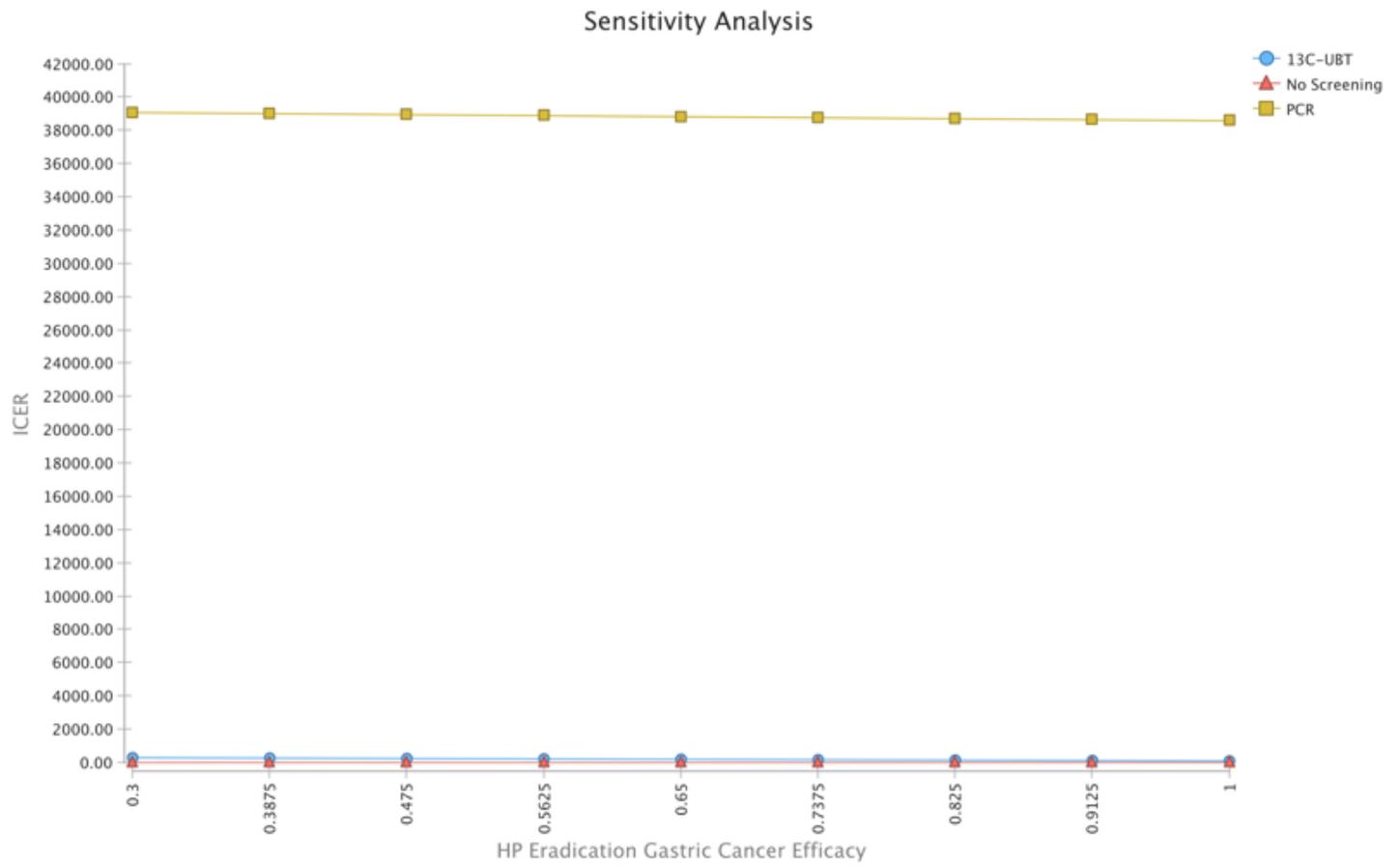


Figure 3

Sensitivity analysis of *H. pylori* eradication gastric cancer efficacy.

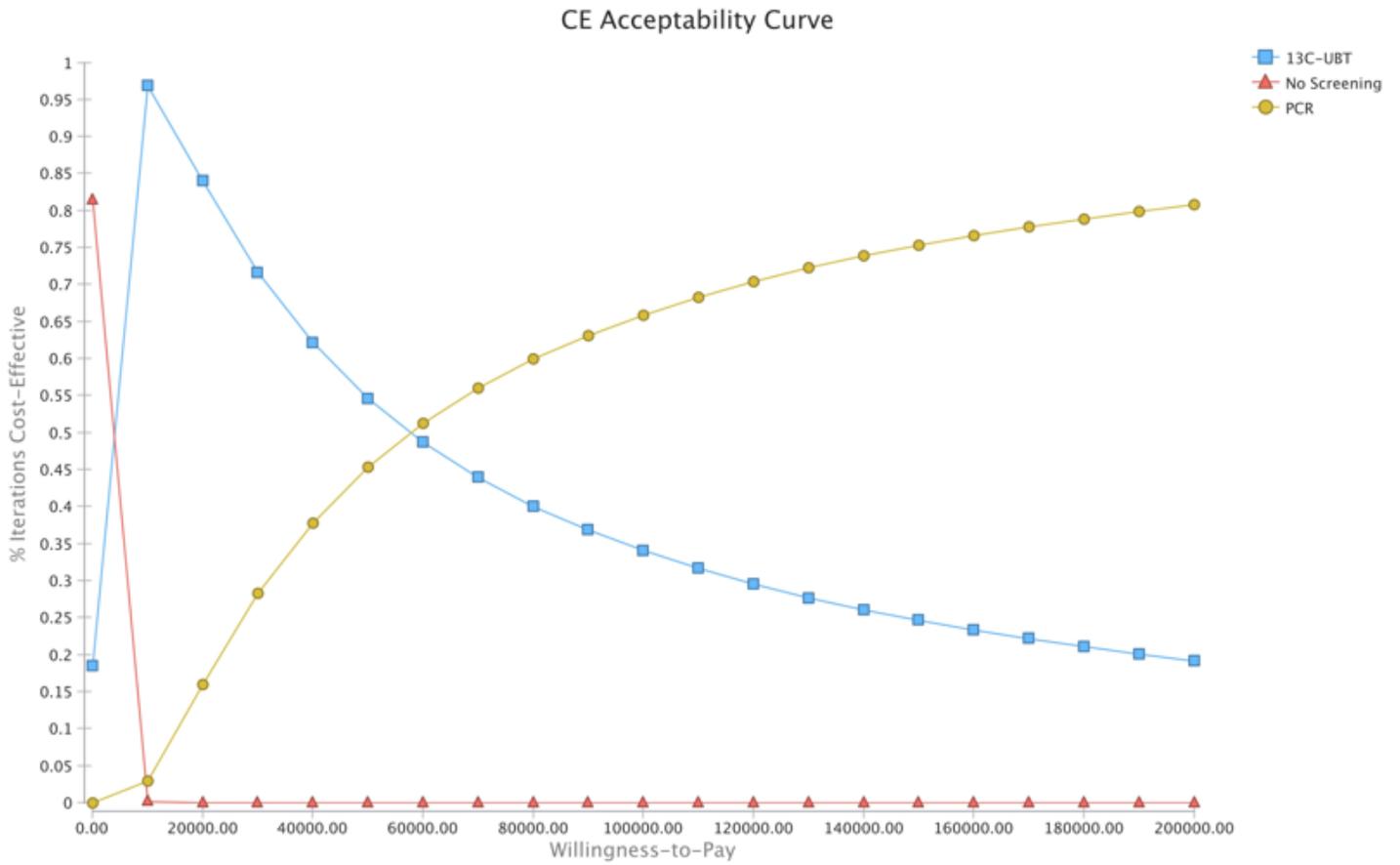


Figure 4

PSA cost-effectiveness acceptability curve.