

The Effect of Malaria on Haemoglobin Concentrations in Extreme Poverty: A Nationally Representative Household Fixed-Effects Study of 17 599 Children under 5 Years of Age in Burkina Faso.

Tim Starck (✉ tim.starck@uni-heidelberg.de)

Universitätsklinikum Heidelberg Institut für Global Health <https://orcid.org/0000-0001-5912-6415>

Caroline A. Bulstra

Universitätsklinikum Heidelberg Institut für Global Health

Halidou Tinto

Clinical Research Unit of Nanoro, Institut de Recherche en Sciences de la Santé, Nanoro, Burkina Faso

Toussaint Rouamba

Clinical Research Unit of Nanoro, Institut de Recherche en Sciences de la Santé, Nanoro, Burkina Faso

Ali Sie

National Institute of Public Health (INSP), Nouna Health Research Centre (CRSN), Nouna, Burkina Faso

Thomas Jaenisch

Colorado School of Public Health

Till Bärnighausen

Universitätsklinikum Heidelberg Institut für Global Health

Research

Keywords: Malaria, Anaemia, Haemoglobin, Poverty, Household fixed-effects, Burkina Faso, Rapid Diagnostic Tests, Microscopy

Posted Date: June 29th, 2021

DOI: <https://doi.org/10.21203/rs.3.rs-657462/v1>

License:  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Abstract

Background: Although the association between malaria and anaemia is widely studied in patient cohorts, the population-representative causal effects of malaria on anaemia remain unknown. We estimated the malaria-induced decrease in haemoglobin levels among young children in malaria-endemic Burkina Faso. Many children in Burkina Faso live in extreme poverty and are thus particularly vulnerable to suffer from the consequences of malaria-induced anaemia.

Methods: We pooled individual-level nationally representative health survey data (2010-11, 2014, 2017-18) from 17 599 children under 5 years of age. We estimated the effects of malaria on haemoglobin concentration, controlling for household fixed-effects, age, and sex in a series of regression analyses. This allowed us to control for observed and unobserved confounding on the household level and to determine the impact of malaria infection status on haemoglobin levels and anaemia prevalence. We further leveraged diagnostic results from microscopy and rapid diagnostic tests to provide a quasi-longitudinal perspective of acute and prolonged effects after malaria infection.

Results: Both malaria (survey prevalence ranging from 17.4% to 65.2%) and anaemia prevalence (survey prevalence ranging from 74% to 88.2%) were very high in the included surveys. Malaria is estimated to significantly reduce haemoglobin levels, with an overall effect of -7.5g/dL (95% CI -8.5, -6.5). Acute malaria resulted in a -7.7 g/dL (95% CI -8.8, -6.6) decrease in haemoglobin levels. Recent malaria without current parasitaemia decreased haemoglobin concentration by -7.1 g/dL (95% CI -8.3, -5.9). The in-sample predicted prevalence of severe anaemia was 9.4% among malaria positives, but only 2.2% among children without malaria.

Conclusion: Malaria infection has a strong detrimental effect on haemoglobin levels among young children in Burkina Faso. This effect seems to carry over even after acute infection, indicating prolonged haemoglobin reductions even after successful parasite-elimination. The quasi-experimental fixed-effect approach adds a population level perspective to existing clinical evidence.

Background

Burkina Faso has, according to WHO data, by far the highest prevalence of anaemia worldwide among young children (86% in 2016) [1]. It is also among the countries in sub-Saharan Africa that are burdened the most by malaria [2]. A large portion (43.8%) of Burkinabe people live in conditions marked by extreme poverty (less than 1.90 USD a day) and they and their children are thus particularly vulnerable to anaemia, malaria, and the associated, serious long term health consequences [3, 4]. However, Burkina Faso was also the setting of a recent, highly promising malaria-vaccine trial [5].

Malaria is a tropical disease, caused by the *plasmodium spp.* parasites that replicate in human tissues and erythrocytes [6]. In 2019, malaria accounted for the deaths of 384 000 children younger than 5 years in in sub-Saharan Africa, 4% of which occurred in Burkina Faso [2]. These deaths are frequently caused by acute haemolytic anaemia, one of malaria's most prominent complications. However, malarial

anaemia can also become chronic through pathways of persistent inflammation and bone marrow suppression leading to reduced production of erythrocytes [7–9]. Since anaemic children have less capacity for oxygen transport, they are more susceptible to opportunistic infections, more tired, and less resilient than healthy children[10]. These symptoms ultimately add up to a higher risk of cognitive and physical development deficits in anaemic young children [3, 10–14].

Anaemia is screened for by measuring blood haemoglobin (Hb [g/L]), the erythrocytic protein that binds oxygen. Although the effect of malaria on haemoglobin has been the subject of extensive clinical research, epidemiological investigations on the population impact are severely lacking. The difficulty with determining the malaria attributable effect to population-wide haemoglobin reductions lies in the complex aetiology of anaemia. Aside from malaria, anaemia in children can be caused by genetic predisposition, other infectious diseases and wider socio-economic factors, especially nutritional deficits [10]. These contributors to anaemia are difficult to measure and may vary substantially between and even within given countries, thus making it hard to causally determine the impact of any one particular contributor to anaemia on the population level.

Most epidemiologic studies that try to causally link malaria to population haemoglobin are conducted within relatively small local communities within highly endemic areas [9, 15–18]. They consistently report a detrimental effect over time of acute and repeated malaria infections on haemoglobin levels among children and adults [19–21]. While this important research confirms the causal effect of malaria on anaemia, external validity of these studies is often low due to regional restrictions and comparatively small or non-representative study populations. We aim to build on this base by using an econometric modelling approach, fixed-effect analysis, that makes it possible to approximate causal effects by implicitly controlling for known and unknown confounders at the household level [22–25]. The household fixed-effect analysis is an extension on the analytic concept of repeated measures data, where every individual within a household is considered a measurement of the same entity, *i.e.*, the household. We apply this method to a large cross-sectional and nationally representative dataset of young children in Burkina Faso, one of the poorest countries world-wide. Burkina Faso is burdened by extreme prevalence levels of malaria and anaemia alike and is therefore a highly relevant target in the effort to combat both (Fig. 1).

We wish to contribute to the fight against the harmful effects of anaemia in Burkina Faso by providing a better understanding of its causes. Our quasi-experimental study design makes it possible to estimate the population-wide effect of malaria infection on haemoglobin levels in children, while avoiding ethically questionable randomized controlled study designs. This will also underline the importance of eradicating malaria and illustrates the potential gains from the recent vaccine candidate in Burkina Faso [5].

Methods

Data

We pooled the data from the 2010 Demographic and Health Survey (DHS) and 2014 and 2017-18 Malaria Indicator Surveys (MISs) from Burkina Faso. The DHSs and MISs are nationally representative household surveys that include demographic, health, and nutrition data, including recognized malaria indicators. The combined datasets contained data from 17 599 children from 11 886 households divided over 572 sample clusters. Field operators collected the survey data from May 2010 to January 2011 (DHS 2010-11), from September to October 2014 (MIS 2014) and from November 2017 to March 2018 (MIS 2017-18). Malaria transmission in Burkina Faso is highest from July to November, therefore the pooled surveys reflect the annual average alongside changes between years, *e.g.*, successful implementation of malaria programs [26–28]. New households are selected for each survey; therefore, our study does not offer a true longitudinal perspective on the individual households and the repeated measures are only for one given household per survey. During the surveys, field workers gather data for two parallel methods of malaria diagnostics: rapid diagnostic antigen tests (RDT) for immediate screening results and thick smear microscopy (henceforth referred to as microscopy) to be analysed later in central laboratories. The RDT model for MIS 2014 and 2017-18 was “*SD Bioline Pan/Pf*”, a combined *HRP-2/pLDH*-test. The MIS 2010 final report did not specify the RDT model or type. Haemoglobin concentrations ([g/L], Hb) were collected with the survey and measured by HemoCue test. More information on the data and collection process can be found at www.dhsprogram.com.

We extracted the following variables from the surveys for analysis: RDT results, microscopy test result, haemoglobin blood levels, age in months, sex, and household identifier. We applied the WHO cut-off values for anaemia among children 59 and younger: any haemoglobin concentrations of less than 110 g/L are considered anaemia, further classified as mild (90 g/L – 109 g/L), moderate (70 g/L – 89 g/L) or severe anaemia (< 70 g/L) [29]. We converted age in months to age groups by completed years (6–12; 13–24; 25–36; 37–48; 49–59 months).

Plasmodium presence in the bloodstream was detected with thick smear microscopy, the current gold standard. Formally, ruling out malaria with microscopy requires three consecutive negative samples. Therefore, since survey samples are only collected once, DHS microscopy results are at risk of false negatives. Rapid diagnostic tests leverage *plasmodium* antigens in the blood stream to indirectly diagnose malaria infections. However, these antigens have been reported to remain positive for up to 30 days even after an acute infection is under medical- or immune-system control and parasites are cleared from the blood stream [30–32]. This creates a time-lag, where positive microscopy represents acute cases and positive RDTs with negative microscopy indicate post-infection status [33]. We exploited this time-lag to create a quasi-longitudinal perspective in the cross-sectional data to illustrate acute and prolonged effects of malaria on haemoglobin.

Statistical methods

In our primary analysis, we estimated the population-level effect of malaria on haemoglobin concentrations (g/L) in a series of nested linear regression models. The first model, the overall malaria model, used any positive malaria-test result (RDT or microscopy) as indicator of malaria infection. The second, the stratified model, used the two malaria measurements, microscopy and RDT, to create three

malaria-status groups: malaria negative (if microscopy and RDT negative), acute malaria (if microscopy positive), post-malaria (if microscopy negative, but RDT positive). We included (econometric) household fixed-effects in all primary analyses to control for observed and unobserved confounders that are shared between all children within one given household [22, 23]. These confounders include known factors such as socioeconomic, temporal, and spatial differences that vary between households but not within. Unknown factors could, amongst others, include nutritional or socio-economic traits which are likely shared between family members within one household but not across households [10].

The final model was stratified for malaria status, age, sex and household fixed-effect. For the mathematical formula of the final model please refer to **Supplement S30**. We did not apply weights because the within-survey weights are not representative for multiple-survey analyses.

To provide a more tangible perspective on the malaria-attributable effect on anaemia prevalence in the observed population, we generated in-sample predictions from the overall malaria and stratified models to calculate predicted haemoglobin values for every child in the data. We stratified anaemia into three severity groups (any anaemia if Hb < 110 g/L; moderate or worse anaemia if Hb < 90 g/L; severe anaemia if Hb < 70 g/L) to assess the influence of malaria on anaemia prevalence by severity.

Finally, we also performed an additional series of sensitivity analyses on the stratified model to further validate our modelling approach. Firstly, we repeated the nested series to check for possible interaction between malaria, age, and sex, respectively and combined. Secondly, we added subgroup analyses based on sex, survey, and malaria season. We concluded with an additional nested series with household as random rather than fixed effect. We expected a model-dependent difference with the random effects model showing larger effects than the fixed-effects model.

All analyses were done in R version 4.0.2 or higher (“plm” package version 2.2-5, “fixest” package version 0.8.2), maps were generated with ArcGIS Pro version 2.3.

Results

A summary of the population and survey characteristics are provided in Table 1. Our final sample included 17 599 children from 11 816 households in Burkina Faso, aged 6 months to 5 years. Figure 2 and Table 1 present a more detailed description of malaria and anaemia in the study populations by survey. The prevalence of malaria varied between the consecutive surveys and averaged 44% for the pooled data (17.4–65.2%, Table 1). Anaemia (haemoglobin < 110 g/L) prevalence showed less variation between surveys but also declined over the years. The overall anaemia prevalence for the pooled data was 83.2%, 31.1% for moderate anaemia and 9.2% for severe anaemia. Regional distributions of malaria prevalence and average haemoglobin levels across the pooled surveys are shown in **Fig. 1**. We added an additional plot of the haemoglobin levels by malaria status, age, and sex to the supplements (**S29**). The differences in mean haemoglobin for the subgroups of negative malaria and acute malaria [$p < 0.001$], negative and prolonged malaria [$p < 0.001$] and acute and chronic malaria [$p < 0.001$] were statistically significant in unpaired T-tests.

Table 1. Characteristics of study population and subgroups.

	Overall (pooled data sample)	Stratified by survey			Stratified by sex	
		2010-11	2014	2017-18	Males	Females
Sample Size	17 599 (100)	5926	6115	5563	8962 (100)	8637 (100)
Sex (%)						
Female	8637 (49.1)	2882 (48.6)	3003 (49.1)	2752 (49.5)	-	-
Male	8962 (50.9)	3044 (51.4)	3107 (50.9)	2811 (50.5)	-	-
Age [months] (%)						
6 – 12	1918 (10.9)	651 (11)	657 (10.8)	610 (11)	984 (11)	934 (10.8)
13 - 24	3750 (21.3)	1301 (22)	1297 (21.2)	1152 (20.7)	1945 (21.7)	1805 (20.9)
25 – 36	3942 (22.4)	1328 (22.4)	1404 (23)	1210 (21.8)	2032 (22.7)	1910 (22.1)
37 – 48	4091 (23.2)	1363 (23)	1402 (22.9)	1322 (23.8)	2020 (22.5)	2071 (24)
49 - 59	3898 (22.1)	1283 (21.7)	1350 (22.1)	1265 (22.7)	1981 (22.1)	1917 (22.2)
Malaria status (%)						
Negative	6954 (39.5)	1045 (17.6)	1787 (29.2)	4122 (74.1)	3558 (39.7)	3396 (39.3)
Acute	7740 (44)	3861 (65.2)	2913 (47.7)	966 (17.4)	38876 (43.2)	3864 (44.7)
Prolonged	2905 (16.5)	1020 (17.2)	1410 (23.1)	475 (8.5)	1528 (17)	1377 (15.9)
Anaemia						
Mean Hb (g/L) (SD)	93 (17)	90 (17)	90 (17)	99 (17)	92 (17)	94 (17)
Anaemic	14 639 (83.2)	5229 (88.2)	5291 (86.6)	4119 (74)	7576 (84.5)	7063 (81.8)
Not anaemic	2960 (16.8)	697 (11.8)	819 (13.4)	1444 (26)	1386 (15.2)	1574 (18.2)
Mild	7535 (42.8)	2464	2382	2689	3802	3733

		(41.6)	(39)	(48.3)	(42.4)	(43.2)
Moderate	5482 (31.1)	2135 (36)	2112 (34.6)	1235 (22.2)	2856 (31.9)	2626 (30.4)
Severe	1622 (9.2)	630 (10.6)	797 (13.0)	195 (3.5)	918 (10.2)	704 (8.2)
Survey Period						
Duration	-	May – Jan ^a	Sept – Dec ^a	Dec – March	-	-

Hb = Haemoglobin; SD = Standard Deviation; TSM = Thick smear microscopy; RDT = Rapid Diagnostic Test;

^a Time frames covering malaria peak season in Burkina Faso from July to November.

Anaemia categories: Not anaemic (Hb \geq 110 g/L); Mild anaemia (Hb < 110 g/L); Moderate anaemia (Hb < 90 g/L); Severe anaemia (Hb < 70 g/L). Due to rounding the percent might not add up to 100.

The outcomes of our overall malaria and stratified models and the respective reductions in haemoglobin are summarised in Table 2. In the overall model, a positive malaria test (RDT or microscopy) reduced haemoglobin by -7.5g/L [95% CI -8.5; -6.5]. In the stratified model (by malaria infection duration), acute malaria resulted in a -7.7 g/L [95% CI -8.8; -6.6] decrease in haemoglobin concentration after controlling for age and sex. The prolonged effect post-infection was - 7.1 g/L [95% CI -8.3; -5.9] (**S31**). Older age had an increasingly beneficial effect on haemoglobin levels, except for the 12–24 months age-group, that conversely had reduced haemoglobin levels of -2 g/L [95% CI -3.3; -0.7]. Female sex had a protective effect of 2 g/L [95% CI 1.3; 2.7]. We appended the results for the remaining models of the nested series to the supplements (**S1 – S8**).

Table 2

Main household fixed-effect regression models of malaria status adjusted for sex and age. The overall effect of all positive malaria tests is -7.5 g/L (LM overall). Children with acute malaria have - 7.7 g/L less haemoglobin when compared to the healthy population. The prolonged effect after infections is -7.1 g/L when compared to the healthy population (LM stratified).

Malaria effect on haemoglobin		
Coefficients	LM overall	LM stratified^β
Overall malaria status		
Reference: negative		
Malaria	Estimate	-7.5
	p-value	< 0.001
	CI95	(-8.5; -6.5)
Stratified malaria status		
Reference: negative		
Acute		-7.7
		< 0.001
		(-8.8; -6.6)
Prolonged		-7.1
		< 0.001
		(-8.3; -5.9)
Age group		
Reference: < 1 year old		
1 year old	-2	-2
	0.002	0.002
	(-3.3; -0.7)	(-3.3; -0.7)
2 years old	2.8	2.8
	< 0.001	< 0.001
	(1.5; 4.1)	(1.5; 4.1)
3 years old	8.3	8.3
	< 0.001	< 0.001

Malaria effect on haemoglobin		
	(7.1; 9.4)	(7.1; 9.4)
4 years old	13.2	13.3
	< 0.001	< 0.001
	(12.1; 14.4)	(12.1; 14.5)
Sex		
Reference: male		
Female	0.2	0.2
	< 0.001	< 0.001
	(0.13; 0.27)	(0.13; 0.27)

CI = confidence interval; α = linear regression of all malaria positive cases on haemoglobin; β = linear regression of acute and prolonged malaria cases on haemoglobin

The in-sample predictions from the overall malaria model indicate a 92.5% prevalence of anaemia among malaria positive children, compared to 77.9% among malaria negative children. The absolute difference was largest for moderate or worse anaemia (malaria positives: 51.5%, malaria negatives: 24.6%) and the relative difference was largest for severe anaemia (malaria positives: 9.4%, malaria negatives: 2.2%). The results from the predictions are illustrated in **Fig. 3**.

The random effects models showed generally larger effects for the acute and prolonged effects (**S9 – S15**) but remained consistent with the results of the main fixed-effect models. The larger effects are attributable to the reduced control for confounding on the household level due to the random effect assumption and are thus likely the result of bias which is eliminated in the household fixed-effect analysis. Similarly, the subset analysis for male participants (**S16 – S18**) and female participants (**S19 – S21**), seasonality (**S22, S23**) and survey (**S24 - S26**) remained consistent with the main outcomes.

Discussion

Both malaria prevalence (44% in the pooled data) and anaemia prevalence (83% in the pooled data) were high among young children in Burkina Faso between 2010 and 2018. We estimated a malaria-attributable haemoglobin decrease of -7.7 g/L during acute infection and of -7.1g/L in the time post-infection. Older children had higher haemoglobin levels than younger children and female sex improved haemoglobin levels by 2 g/L.

The malaria-induced haemoglobin changes can have large clinical implications. For instance, it has been shown that an increase of 10 g/L haemoglobin is associated with a 0.78 relative risk of mental retardation in young children [34]. This implies that a malaria-attributable haemoglobin reduction as

shown in our data might pose substantial threat of cognitive development disorders in affected children. Especially the predictions of anaemia prevalence illustrate the severity of the burden of malarial anaemia in Burkina Faso. Our analyses indicate that most cases of severe anaemia and a sizeable portion of moderate anaemia could be avoided if malaria were successfully eradicated.

Several studies have previously reported on the malaria-associated decrease in haemoglobin concentrations in clinical and national settings using different analytic methods [35–37]. The age group and sex dependent variation in haemoglobin values, as observed in our study, have been described previously. The observed differences in magnitude of the effect by age and sex are typical for early childhood development and in line with current research[38].

Our study is unique in that we could estimate the close-to-causal association between malaria and anaemia at the population-level, using a household fixed-effect approach controlling for all confounding that is constant within a given household. Furthermore, it is representative not only in its sampling design, but also in its seasonal composition, given that surveys were conducted on- and off malaria season. Finally, the study is based on a very large and nationally representative sample of 17 599 children and thus offers enough power to inspire confidence in our results as they are consistent even in the reduced subset analyses.

Our study is influenced by several limitations. Firstly, a large number of children had a positive rapid test, but no corresponding positive microscopy test result. Thick smear microscopy is considered the gold standard but has varying sensitivity (from 55–98%) and specificity (from 81% to >98%), depending on the experience of the diagnostician and the slide quality [39–41]. To rule out malaria it is required to repeat the microscopy test over the course of several days, which has not been done in the surveys and thus likely results in an underestimation of the malaria prevalence in our data [32]. The other method, RDTs, produce a comparatively high rate of false positives where *plasmodium* antigens are present on their gametocytes, even when the disease itself is controlled by the immune system or medical treatment. This can cause microscopy-negative cases to show RDT positive results for up to thirty days even after parasite elimination and clinical remission [30, 33]. We leveraged this effect to create a quasi-longitudinal perspective, where positive RDTs with negative microscopy results represent children that are currently recovering from malaria. Biologically, this prolonged effect might be a mix of several contributing factors, such as persistent bone-marrow suppression, delayed haemolysis, delayed recovery and false-negative microscopy tests [42].

A second limitation is the way in which the pooled cross-sectional data reflects the patterns of malaria and changes between survey years in Burkina Faso. Since we pooled several years and seasons of surveys, our study population is not representative of any malaria point-prevalence in Burkina Faso and thus our analysis neither reflects malaria's seasonal pattern, nor does it reflect progress made in the fight against malaria between 2010 and 2018. It is, however, still comparable to the extremes of poverty, anaemia burden and malaria transmission intensity found in West African countries [2, 43, 44].

Thirdly, the fixed-effect method itself also comes with a caveat: It controls for all confounders above the household level but lacks control for the within household confounders, particularly anaemia risk factors that vary between children in a household. These risk factors include nutritional (e.g., iron deficiency) and genetical traits (e.g., sickle-cell anaemia), other infectious diseases (e.g., helminths) and other, frequently interacted factors[45–47]. For our model we assumed that these unmeasured confounders are reasonably similar for all children within the household.

Conclusions

In summary, we propose a strong estimate of the population-wide effect of malaria on haemoglobin among young children in Burkina Faso, a setting marked by extreme poverty, high malaria burden and extremely high anaemia prevalence. Our findings shed light on the acute and prolonged effects of infection and the potential gains against severe anaemia by eradicating malaria. Hopefully, these effects will soon be diminished through the successful vaccine candidate. The household fixed-effects analysis has proven to be a suitable design to quantify these effects in a quasi-experimental setup and makes it possible to draw conclusions approaching causality. Further research should attempt to clarify how genetic relationship and other factors contribute to the risk of acquiring malaria infection and decreased haemoglobin levels. There is a large knowledge gap on the longitudinal change in haemoglobin levels during an acute malaria infection.

Abbreviations

DHS: demographic and health surveys

Hb: haemoglobin

MIS: malaria indicator surveys

RDT: rapid diagnostic test

SSA: Sub-saharan Africa

Declarations

Ethics approval: Not applicable. Since we used only anonymized and untraceable data, this study did not require ethical review from our home institution (EKHD).

Consent: Not applicable

Availability of data and material: All data is available from the DHS website (<https://dhsprogram.com/>). Codes will be made available upon request.

Competing interests: The authors declare that they have no competing interests.

Funding: No funding was received for conducting this study.

Author Contributions: TB and TS designed the study. TS, CB, and TB performed the data analysis. TS and CB drafted the manuscript. All co-authors contributed significantly to the revision of the manuscript and provided scientific guidance.

Acknowledgments: We would like to thank ICF and DHS for providing the data. Our special thanks are owed to the thousands of Burkinabe that participated in the extensive surveys and the field workers that collected the data.

References

1. **Prevalence of anaemia in children under 5 years (%)**
[https://www.who.int/data/gho/data/indicators/indicator-details/GHO/prevalence-of-anaemia-in-children-under-5-years-\(-\)](https://www.who.int/data/gho/data/indicators/indicator-details/GHO/prevalence-of-anaemia-in-children-under-5-years-(-)).
2. WHO. World malaria report 2020: 20 years of global progress and challenges. Geneva: World Health Organization; 2020.
3. Sachs J, Malaney P. The economic and social burden of malaria. *Nature*. 2002;415:680–5.
4. The World Bank. Burkina Faso. In: *Macro poverty outlook: sub-Saharan Africa*. Washington DC: The World Bank; 2020. pp. 208–9 208–209.
5. Dattoo MS, Natama MH, Somé A, Traoré O, Rouamba T, Bellamy D, Yameogo P, Valia D, Tegneri M, Ouedraogo F, et al. Efficacy of a low-dose candidate malaria vaccine, R21 in adjuvant Matrix-M, with seasonal administration to children in Burkina Faso: a randomised controlled trial. *Lancet*. 2021;397:1809–18.
6. Phillips MA, Burrows JN, Manyando C, van Huijsduijnen RH, Van Voorhis WC, Wells TNC. Malaria. *Nature Reviews Disease Primers*. 2017;3:17050–0.
7. White NJ. Anaemia and malaria. *Malar J*. 2018;17:371.
8. Ekvall H. Malaria and anemia. *Curr Opin Hematol*. 2003;10:108–14.
9. Hedberg K, Hightower A, Shaffer N, Paluku KM, Lyamba B, Davachi F, Breman JG, Nguyen-Dinh P. Plasmodium falciparum-associated anemia in children at a large urban hospital in Zaire. *The American Journal of Tropical Medicine Hygiene*. 1993;48:365–71.
10. Balarajan Y, Ramakrishnan U, Özaltın E, Shankar AH, Subramanian SV. Anaemia in low-income and middle-income countries. *The Lancet*. 2011;378:2123–35.
11. Yang F, Liu X, Zha P. Trends in socioeconomic inequalities and prevalence of anemia among children and nonpregnant women in low- and middle-income countries. *JAMA Network Open*. 2018;1:e182899–9.
12. Plessow R, Arora NK, Brunner B, Tzogiou C, Eichler K, Brügger U, Wieser S. **Social costs of iron deficiency anemia in 6-59-month-old children in India.** *PLoS ONE* 2015, 10.

13. Tusting LS, Willey B, Lucas H, Thompson J, Kafy HT, Smith R, Lindsay SW. Socioeconomic development as an intervention against malaria: A systematic review and meta-analysis. *The Lancet*. 2013;382:963–72.
14. Holding PA, Snow RW. Impact of Plasmodium falciparum malaria on performance and learning: review of the evidence. *Am J Trop Med Hyg*. 2001;64:68–75.
15. Kotepui M, Piwkham D, PhunPhuech B, Phiwklam N, Chupeerach C, Duangmano S. Effects of malaria parasite density on blood cell parameters. *PLoS ONE*. 2015;10:1–11.
16. Premji Z, Hamisi Y, Shiff C, Minjas J, Lubega P, Makwaya C. Anaemia and Plasmodium falciparum infections among young children in an holoendemic area, Bagamoyo, Tanzania. *Acta Trop*. 1995;59:55–64.
17. Ekvall H, Premji Z, Bennett S, Bjorkman A. Hemoglobin concentration in children in a malaria holoendemic area is determined by cumulated Plasmodium falciparum parasite densities. *Am J Trop Med Hyg*. 2001;64:58–66.
18. McElroy PD, Ter Kuile FO, Lal AA, Bloland PB, Hawley WA, Oloo AJ, Monto AS, Meshnick SR, Nahlen BL. Effect of Plasmodium falciparum parasitemia density on hemoglobin concentrations among full-term, normal birth weight children in western Kenya, IV. The Asembo Bay Cohort Project. *Am J Trop Med Hyg*. 2000;62:504–12.
19. Menendez C, Fleming AF, Alonso PL. Malaria-related anaemia. *Parasitol Today*. 2000;16:469–76.
20. Means RT. The anaemia of infection. *Bailliere's Best Practice Research in Clinical Haematology*. 2000;13:151–62.
21. Calis JCJ, Phiri KS, Faragher EB, Brabin BJ, Bates I, Cuevas LE, de Haan RJ, Phiri AI, Malange P, Khoka M, et al. Severe anemia in Malawian children. *Malawi Medical Journal*. 2016;28:99–107.
22. Barnighausen T, Oldenburg C, Tugwell P, Bommer C, Ebert C, Barreto M, Djimeu E, Haber N, Waddington H, Rockers P, et al. Quasi-experimental study designs series-paper 7: Assessing the assumptions. *J Clin Epidemiol*. 2017;89:53–66.
23. Gunasekara FI, Richardson K, Carter K, Blakely T. Fixed effects analysis of repeated measures data. *Int J Epidemiol*. 2014;43:264–9.
24. Anekwe TD, Newell ML, Tanser F, Pillay D, Barnighausen T. The causal effect of childhood measles vaccination on educational attainment: A mother fixed-effects study in rural South Africa. *Vaccine*. 2015;33:5020–6.
25. McGovern ME, Herbst K, Tanser F, Mutevedzi T, Canning D, Gareta D, Pillay D, Barnighausen T. Do gifts increase consent to home-based HIV testing? A difference-in-differences study in rural KwaZulu-Natal, South Africa. *Int J Epidemiol*. 2016;45:2100–9.
26. Ouédraogo M, Rouamba T, Samadoulougou S, Kirakoya-Samadoulougou F. Effect of free healthcare policy for children under five years old on the incidence of reported malaria cases in Burkina Faso by bayesian modelling: “Not only the ears but also the head of the hippopotamus”. *International Journal of Environmental Research Public Health*. 2020;17:417–7.

27. Rouamba T, Nakanabo-Diallo S, Derra K, Rouamba E, Kazienga A, Inoue Y, Ouédraogo EK, Waongo M, Dieng S, Guindo A, et al. Socioeconomic and environmental factors associated with malaria hotspots in the Nanoro demographic surveillance area, Burkina Faso. *BMC Public Health*. 2019;19:249–9.
28. Ouedraogo B, Inoue Y, Kambiré A, Sallah K, Dieng S, Tine R, Rouamba T, Herbreteau V, Sawadogo Y, Ouedraogo LSLW, et al. Spatio-temporal dynamic of malaria in Ouagadougou, Burkina Faso, 2011–2015. *Malaria Journal*. 2018;17:1–12.
29. **Haemoglobin concentrations for the diagnosis of anaemia and assessment of severity**
<http://www.who.int/vmnis/indicators/haemoglobin.pdf>.
30. Dalrymple U, Arambepola R, Gething PW, Cameron E. How long do rapid diagnostic tests remain positive after anti-malarial treatment? *Malar J*. 2018;17:228.
31. Mathison BA, Pritt BS. Update on malaria diagnostics and test utilization. *J Clin Microbiol*. 2017;55:2009–17.
32. Ashley EA, Pyae Phyo A, Woodrow CJ. Malaria. *The Lancet*. 2018;391:1608–21.
33. Ndour PA, Larréché S, Mouri O, Argy N, Gay F, Roussel C, Jauréguiberry S, Perillaud C, Langui D, Biligui S, et al. Measuring the Plasmodium falciparum HRP2 protein in blood from artesunate-treated malaria patients predicts post-artesunate delayed hemolysis. *Sci Transl Med*. 2017;9:eaaf9377–7.
34. Stoltzfus RJ, Mullany L, Black RE: Chap. 3: **Iron deficiency anemia**. In *Comparative quantification of health risks: global and regional burden of disease attributable to selected major risk factors. Volume 1*. Edited by Ezzati M, Lopez AD, Rodgers AA, Murray CJL. Geneva: World Health Organization; 2004: 163–210.
35. Kotepui M, Phunphuech B, Phiwklam N, Chupeerach C, Duangmano S. Effect of malarial infection on haematological parameters in population near Thailand-Myanmar border. *Malar J*. 2014;13:218.
36. Bickerton Blackburn CR. Observations on the development of resistance to vivax malaria. *Trans R Soc Trop Med Hyg*. 1948;42:117–62.
37. Fanello C, Onyamboko M, Lee SJ, Woodrow C, Setaphan S, Chotivanich K, Buffet P, Jauréguiberry S, Rockett K, Stepniewska K, et al. Post-treatment haemolysis in African children with hyperparasitaemic falciparum malaria; a randomized comparison of artesunate and quinine. *BMC Infect Dis*. 2017;17:1–8.
38. Fulgoni VL 3rd, Agarwal S, Kellogg MD, Lieberman HR. Establishing pediatric and adult RBC reference intervals with NHANES data using piecewise regression. *Am J Clin Pathol*. 2019;151:128–42.
39. Berzosa P, de Lucio A, Romay-Barja M, Herrador Z, González V, García L, Fernández-Martínez A, Santana-Morales M, Ncogo P, Valladares B, et al. Comparison of three diagnostic methods (microscopy, RDT, and PCR) for the detection of malaria parasites in representative samples from Equatorial Guinea. *Malaria Journal*. 2018;17:333–3.
40. Rodulfo H, De Donato M, Mora R, González L, Contreras CE. Comparison of the diagnosis of malaria by microscopy, immunochromatography and PCR in endemic areas of Venezuela. *Braz J Med Biol Res*. 2007;40:535–43.

41. Wangai LN, Karau MG, Njiruh PN, Sabah O, Kimani FT, Magoma G, Kiambo N. Sensitivity of microscopy compared to molecular diagnosis of *P. falciparum*: Implications on malaria treatment in epidemic areas in Kenya. *African Journal of Infectious Diseases*. 2011;5:1–6.
42. Helleberg M, Goka BQ, Akanmori BD, Obeng-Adjei G, Rodriques O, Kurtzhals JAL. Bone marrow suppression and severe anaemia associated with persistent *Plasmodium falciparum* infection in African children with microscopically undetectable parasitaemia. *Malaria Journal*. 2005;4:1–7.
43. Assele V, Ndoh GE, Nkoghe D, Fandeur T. No evidence of decline in malaria burden from 2006 to 2013 in a rural province of Gabon: Implications for public health policy. *BMC Public Health*. 2015;15:1–8.
44. Ouédraogo A, Tiono AB, Diarra A, Sanon S, Yaro JB, Ouedraogo E, Bougouma EC, Soulama I, Gansané A, Ouedraogo A, et al. Malaria morbidity in high and seasonal malaria transmission area of Burkina Faso. *PLoS ONE*. 2013;8:e50036–6.
45. Elguero E, Delicat-Loembet LM, Rougeron V, Arnathau C, Roche B, Becquart P, Gonzalez JP, Nkoghe D, Sica L, Leroy EM, et al. Malaria continues to select for sickle cell trait in Central Africa. *Proc Natl Acad Sci U S A*. 2015;112:7051–4.
46. Taylor SM, Parobek CM, Fairhurst RM. Haemoglobinopathies and the clinical epidemiology of malaria: A systematic review and meta-analysis. *Lancet Infect Dis*. 2012;12:457–68.
47. Petry N, Olofin I, Hurrell RF, Boy E, Wirth JP, Moursi M, Donahue Angel M, Rohner F. The proportion of anemia associated with iron deficiency in low, medium, and high human development index countries: A systematic analysis of national surveys. *Nutrients*. 2016;8:1–17.

Figures

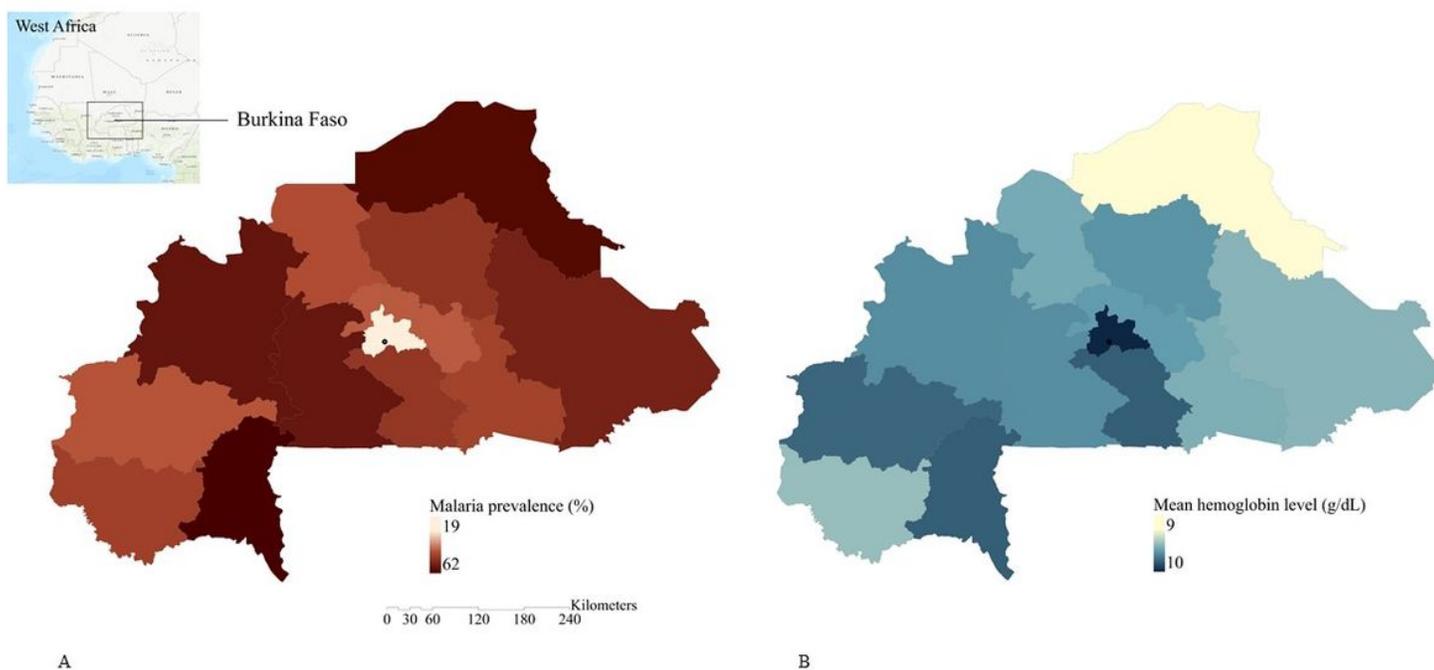


Figure 1

Map of the prevalence of malaria (a) and mean haemoglobin (b) in Burkina Faso. Panel a indicates the regional prevalence of acute malaria as diagnosed by thick smear microscopy. Darker colours represent higher prevalence of anaemia. Panel b indicates mean regional haemoglobin values across all three surveys. Lighter colours represent lower average haemoglobin values and thus higher prevalence of anaemia.

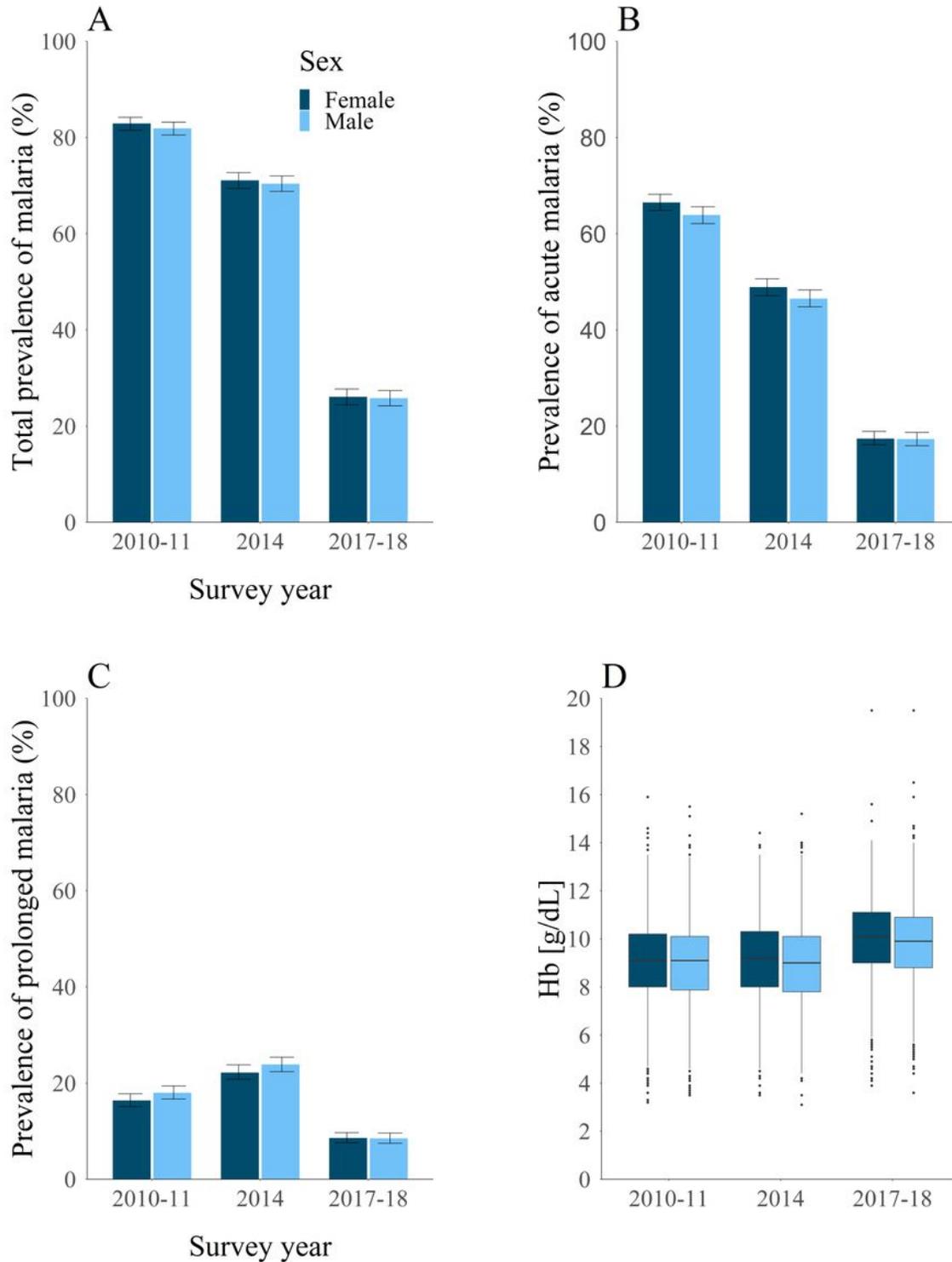


Figure 2

Prevalence of overall (a), acute (b) and prolonged malaria (c); Haemoglobin concentrations [g/L] (d). The plots demonstrate the change in the study populations from the successive surveys based on sex. The error bars mark the 95% confidence intervals in panels A, B and C. Dark blue (dark grey, left column) represents females, light blue (grey, right column) represents males.

Main regression results Malaria induced change in hemoglobin

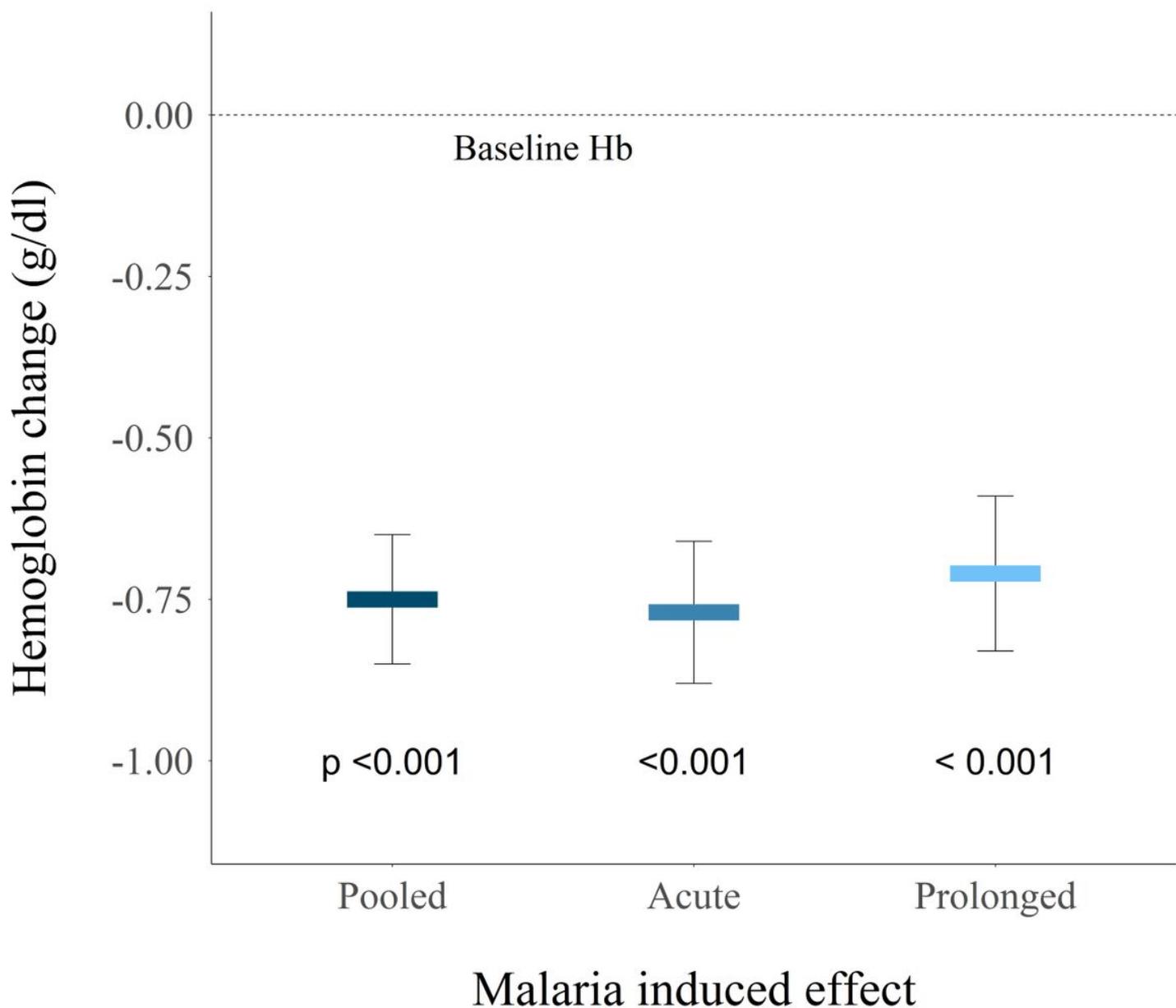


Figure 3

Predicted prevalence of any, moderate or worse and severe anaemia by malaria status. In-sample predicted prevalence of any anaemia (Hb < 110 g/L), moderate or worse anaemia (Hb < 90 g/L) and

severe anaemia (Hb < 70 g/L) by malaria status for all malaria positive cases (a), acute cases (b) and prolonged cases (c). Eliminating malaria from the study population would substantially reduce the total prevalence of malaria, moderate anaemia and almost eliminate severe anaemia. The stratification by malaria status affected the results only marginally. Red shades (lighter colours, left bars) indicate malaria negative cases and blue shades (darker colours, right bars) malaria positive cases.

Poisson regression results

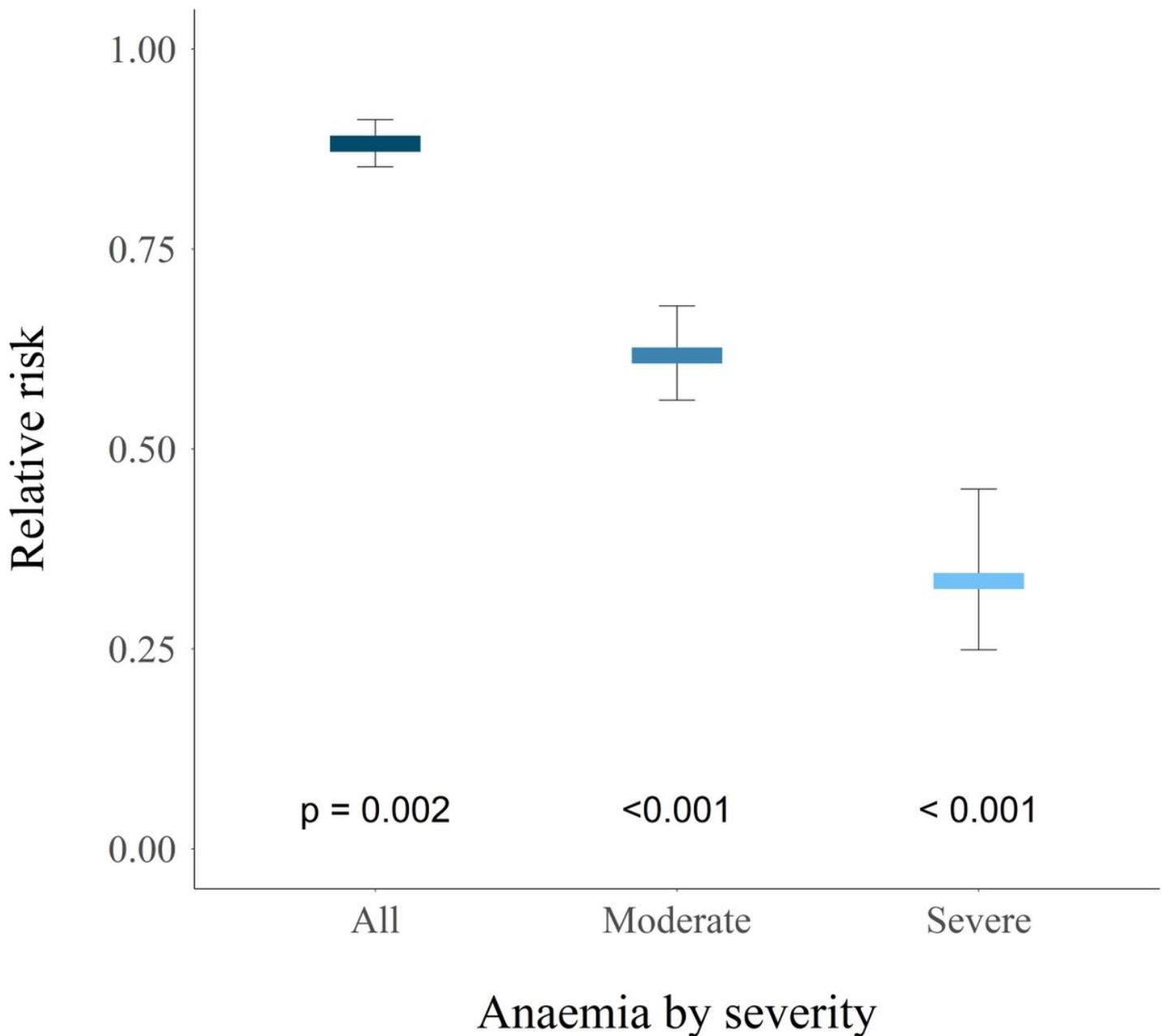


Figure 4

Poisson regression results

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [supplementSIUnits.docx](#)