

The Neurotrophic and Anti-Inflammatory Effect of β -asarone on the DA-Induced the Pathology of Minimal Hepatic Encephalopathy

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Abstract

Background: The mechanism underlying the impaired cognitive function and memory loss in Minimal hepatic encephalopathy (MHE) remains unclear. Dopamine (DA) is reported to be associated with dementia.

Methods: In this study, we investigated mechanism underlying DA-induced MHE pathology by immunoblotting, ELISA, FM4-64 and fluorescence staining.

Results: We observed that MHE brains showed the increased content of DA, after administration of anti-DA antibody, and cognitive loss in MHE rats was recovered to the normal level, indicating the involvement of DA in the pathogenesis of MHE. Moreover, DA (10 μ M) treatment obviously induced the decrease in the production of GDNF/NGF and the increase in TNF α levels in primary cultured neurons, which were blocked by addition of β -asarone (β ASA). We also demonstrated that DA stimulated the activation of ASK1/JNK1 pathway. and the addition of anti-TNF α antibody reversed the inactivation of Notch signaling, the downregulation of neurotrophins and synaptic loss.

Conclusions: Overall, we suggested that DA stimulated abundant production and secretion of neuronal TNF α , which elicited progressive loss of neurotrophic factors, leading to cognitive disorder of MHE.

1. Introduction

Brain dopamine (DA) has been implicated in cognitive processes such as working memory and cognitive flexibility [1] and the DA system is involved in cognitive decline [2]. Our previous studies have confirmed that the pathogenesis of MHE was caused by the abnormal elevation of intracranial DA [3, 4, 5, 6, 7, 8]. It has been demonstrated as a novel regulatory mechanism that DA-induced astrocytic TNF- α release triggers progressive neurodegeneration, which leads to learning and memory decline in MHE [9]. Tumor necrosis factor- α (TNF α) signaling has been shown to regulate neuroinflammation, and plays roles in modulation of microglial activation [10], control of glutamatergic transmission and regulation of synaptic strength [11] in the central nervous system (CNS) [12]. Ample evidence suggests that elevated levels of TNF α have been implicated in some neurodegenerative disorders including Alzheimer's disease [13], Parkinson's disease [14] and HIV-associated dementia [15]. Hence, Neuroinflammatory conditions are often related with memory dysfunction [16, 17, 18], which suggests that DA may be influenced on brain function involved with inflammatory factor. We further proposed that DA may regulate synaptic formation underlying long-term cognition with involvement of TNF α in MHE. Therefore, we hypothesized that the induction of cognitive decline by accumulation of DA levels in the MHE rats is associated with its ability to produce TNF α .

Here, we investigated (1) whether neuroinflammation was induced by DA in neurons in MHE; (2) whether neurons interference in disruption of synaptic formation triggered by DA in MHE. Moreover, β -asarone increased the expression of neurotrophin such as brain-derived neurotrophic factor (BDNF), glial cell-derived neurotrophic factor, and ciliary neurotrophic factor in RSC96 cells [19]. β -asarone treatment also

improved the spatial learning and memory in A β (1–42)-induced memory-impaired rats [20]; in transgenic model of Alzheimer's disease such as Senescence-accelerated prone-8 (SAMP-8) mice [21]; and A β PP/PS1 double transgenic mice [22, 23], also in chronic exogenous corticosterone-induced memory-impaired rats [24, 25]. Then, we describe that the attribution of pathogenesis of MHE is upregulation of DA, followed by the production of TNF α from neurons, leading to the disruption of synaptic formation, which was diminished by β ASA treatment.

2. Materials And Methods

2.1 MHE models and treatment

A total of 40 Sprague-Dawley (SD) rats (220–250 g) were used, and all guidelines of the care and use of animals for experimental procedures were laid down by the Ethics Committees of the Affiliated Hospital of Wenzhou Medical University regarding [26].

Before experimenting, all animals were subjected to behavioural tests: Y-maze (YM) and water-finding task (WFT). Normal values of YM and WFT these behavioural tests were obtained. Rats were then intraperitoneally (i.p.) injected with TAA (200 mg/kg in normal saline, Sigma-aldrich) to induce liver cirrhosis twice per week for 8 weeks. TAA-treated rats with symptoms of decreased movement, lethargy, and coma were diagnosed as HE. Then behavioral tests were conducted for TAA-treated rats with no HE symptoms. If values of YM were decreased or values of WFT were increased compared to mean \pm 1.96·SD of normal values, rats were included in the MHE group.

MHE Rats were anaesthetized with pentobarbital (40 mg/kg, i.p.) and subjected to stereotaxic surgery. Two 23-gauge, stainless-steel, thin-wall cannulae were implanted to the right lateral ventricles at the following coordinates: 0.4 mm posterior to the bregma, 0.8 mm lateral to the midline, and 2.0 mm from the surface of the skull. MHE rats were injected with DA (10 μ g / 3 μ l) 3 times per week for 1 week through guide cannula, following by the intraperitoneal injection with β -asarone (β ASA, 20 μ g) 3 times per week for 1 week. Then Rats were conducted with a YM and a WFT test.

2.2 Behavioral test

Rats were individually placed at the end of one of arm of the three arms in the apparatus for YM, and were allowed to find the arm entries freely for 8 min. Total arm entries and spontaneous alternation percentage (SA%, a ratio of successful choices to total choices minus two) were measured [27].

Rats were deprived of water for 24 h and individually placed at the near-right corner of the testing apparatus for WFT and allowed to explore the tube and drink the water in the cubic alcove freely for 3 min. The latency times of the first entry into the alcove (entry latency, EL), of the first touching/sniffing/licking of the water tube (contacting latency, CL) and of the initiation of drinking from the water tube (drinking latency, DL) were measured [28].

2.3 Primary rats neurons culture and treatments

Primary hippocampal rats neurons (PHNs) were obtained from freshly dissected hippocampus and cerebral cortex from 1-day-old SD rat pups by mechanical disruption in the presence of trypsin and DNase and then seeded in poly-L-lysine-precoated six-well plates at a density of 2×10^6 cells/well in Neurobasal® Medium (1X) supplemented with 0.5 mM GlutaMAX™-I, B-27®. Then neurons remained untreated or were stimulated with DA (1, 10 or 20 $\mu\text{mol/l}$) alone or after preincubation with JNK1 inhibitor SP600125, ASK1 inhibitor K811, 50 μM Notch inhibitor MK-0752, 100 ng/ml Notch activator Jagged-1 or anti-TNF α antibody (ATAB), or treated with β ASA in the presence or absence of DA together with MK-0752/TNF α .

2.4 PC12 cells culture and treatments

The rat pheochromocytoma PC12 cell line were achieved from the American Type Culture Collection (ATCC, Manassas, VA). PC12 cells were routinely cultured in DMEM with 2 mM glutamine, 10% horse serum, and 5% heat-inactivated fetal bovine serum (FBS) and passaged one time per week. Then PC12 cells remained untreated or were stimulated with DA (1, 10 or 20 $\mu\text{mol/l}$) alone or after preincubation with JNK1 inhibitor SP600125, ASK1 inhibitor K811, 50 μM Notch inhibitor MK-0752, 100 ng/ml Notch activator Jagged-1 or anti-TNF α antibody (ATAB), or treated with β ASA in the presence or absence of DA together with MK-0752/TNF α .

2.5 Transfection for PC12 cells

PC12 cells were transfected with Notch1/JNK1/ASK1/TNF α / the Silencer Negative Control siRNA (Scrambled siRNA) (Santa Cruz, CA, USA) or Notch cDNA (activated) /empty vector plasmid (Millipore) using Lipofectamine 2000 (Invitrogen, Carlsbad, USA) according to the instructions suggested by the manufacture.

2.6 Measurement of TNF α release

TNF α level in the culture medium of neurons was measured using enzyme-linked immunosorbent assay (ELISA) kit (R&D systems) in the light of the manufacturer's protocols and a Thermo-Fisher Multiskan MCC plate reader.

2.7 Immunoblotting (IB) analysis

The total amount of protein in the lysates were separated by 10% SDS-PAGE and electroblotted to PVDF membrane, the membrane was blocked with 5% non-fat dry milk dissolved in TBS-T (150 mM NaCl, 50 mM Tris, 0.05% Tween 20), and incubated with following primary antibodies: DA, TNF α , GDNF, NGF,

Apoptosis signal-regulating kinase1 (ASK1), c-Jun N-Terminal Kinase1 (JNK1), pASK1, pJNK1, Notch Intracellular Domain1 (NICD1), Hairy and enhancer of split1 (HES1), Homer, synaptotagmin, β -actin (Abcam) overnight at 4°C, then probed with horseradish peroxidase-conjugated secondary antibody for 1 h. After extensive washing, membrane was visualized by ECL reagent (Thermo) and exposed on Kodak BioMax film (Kodak). Intensities of protein bands were analysed with Quantity One software.

Densitometry was indicated as ratio of proteins to β -actin levels.

2.8 Functional labeling of presynaptic boutons with FM4-64

FM4-64 staining (Invitrogen) was followed by the manufacturer's instructions. Briefly, neurons were incubated with 5 mg/ml FM4-64 and 50 mM KCl-contained Hank's balanced salt solution for 1 minute at 4 °C.

2.9 Double-labeled fluorescent staining

Frozen brain sections or neurons cultured on glass coverslips were fixed with 4% paraformaldehyde for 30 min and treated with 0.1% Triton X-100 for 10 min, then blocked with bovine serum albumin (BSA) for 1 h. Sections were then incubated with the following primary antibodies: DA, TNF α , GDNF, NGF, pASK1, pJNK1, NICD1, HES1, MAP2 (Abcam) overnight at 4 °C, following with FITC (green)/Alexa Fluor 594 (red) conjugated secondary antibody for 30 min. A Leica TCS SP2 confocal laser scanning microscope was used for imaging.

2.10 Dendritic spine density analysis in primary neurons

After fixation, neurons cultured on glass coverslips were incubated with primary antibodies: microtubule-associated protein 2B (MAP2B; 1:200; BD Transduction Laboratories, San Jose, CA, USA), and vesicular glutamate transporter 1 (vGlut1; 1:100; Neuromab, Davis, CA, USA), then with 1-hour secondary antibody-conjugated AlexaFluor (1:500; Life Technologies, Waltham, MA, USA). A z stack of optical section was imaged on a confocal laser scanning microscope (FV10i-W, Olympus, Japan).

2.11 Statistical analysis

All of the data were expressed as mean \pm SD. Data comparisons were analyzed using one-way analysis of variance (ANOVA). All of the data were detected in normal distribution and equal variances, when Back-of-the-envelope test was used to verify the normal distribution, and F-test was applied for determining the equality of variances. Dunnett's post hoc multiple comparison test was applied when significant differences were detected by the ANOVA model. Then P values were made for adjustment by Bonferroni correction. The level of significance was determined for $P < 0.05$ or $P < 0.01$. All analyses were performed with SPSS 18.0 (PASW Statistics 18.0).

3. Results

3.1 DA inhibits the production of neurotrophic factors in neurons

First, we assessed the effect of DA on the production of neurotrophic factors in neurons. As indicated by the IB analysis, the decreased trend on the expressions of glial cell line-derived neurotrophic factor (GDNF)/Nerve growth factor (NGF) were observed after lower DA treatment, higher DA treatment significantly decreased the two proteins levels in PHNs (Fig. 1A and B). PC12 cells in response to DA showed that the expressions of GDNF/NGF were also reduced in a dose-dependent fashion (Fig. 1C and D). Time course analyses via IB analysis of PHNs treated with high dose of DA revealed steady decreases in GDNF/NGF expression up to 48 h (Fig. 1E and F). Double IF staining of PHNs using GDNF antibody combined with MAP2 antibody confirmed that high DA treatment induced the weak expression of GDNF (Fig. 1G). As analyzed by immunostaining, PC12 in response to DA displayed significant decrease in NGF expression (Fig. 1H). These results suggested that DA dramatically triggered the loss of neurotrophins.

3.2 DA stimulated TNF α secretion from neurons

Recent studies have reported that DA is involved in the process of inflammation [29, 30, 31, 32]. Therefore, we examined the effect of DA on the production of proinflammatory factor TNF α in neurons. Significant upregulation of TNF α was induced by higher DA treatment, but not by lower DA treatment in PHNs using IB analysis (Fig. 2A and B). Otherwise, IB analysis of PC12 cells showed that DA also significantly triggered TNF α expression in a dose-dependent manner (Fig. 2C and D). As determined by ELISA, DA significantly stimulated the release of TNF α with dose-dependence in PHNs (Fig. 2E). Parallely, as shown by IB analysis, PHNs exhibited high TNF α levels in incubation of DA with a time-dependent manner (Fig. 2F and G). By ELISA, DA significantly elevated release of TNF α with time-dependence from PHNs (Fig. 2H). Double IF staining using TNF α antibody with MAP2 antibody confirmed that the expression of TNF α was obviously increased in PHNs after DA treatment (Fig. 2I). Immunostaining of PC12 cells in response to DA displayed the strong expression of TNF α (Fig. 2J). These results suggested that DA induced the production of proinflammatory factor TNF α in neurons in vitro.

3.3 DA modulates production of TNF α by ASK1/JNK1 pathway in neurons

In this experiment, we studied the mechanism underlying DA-induced TNF α production. ASK1 upregulates c-Jun N-terminal kinase (JNK1) signaling [33]. Inflammatory responses are frequently associated with c-Jun NH2-terminal kinase (JNK1) signaling pathway [34]. Thus, we examined the association of DA-induced production of TNF α with ASK1/JNK1 pathway in neurons. As determined by IB analysis, the phosphorylation of ASK1 was significantly increased in PHNs in response to DA with dose-dependent manner (Fig. 3A and B). DA treatment significantly elevated the phosphorylation of JNK1 in PHNs, which was reversed by ASK1 inhibitor K811 by IB analysis (Fig. 3C and D). DA-mediated the increase in TNF α expression in PHNs was significantly diminished by addition of K811 or JNK1 inhibitor SP600125 by IB analysis (Fig. 3E and F). The efficiency of ASK1 (Fig. 3G) or JNK1 siRNA (Fig. 3H) plasmid transfection

was confirmed by qPCR in PC12 cells. Then we found that the increase in TNF α expression in response to DA were significantly diminished by ASK1 or JNK1 silencing in PC12 cells by IB analysis (Fig. 3I and J). ELISA of supernatants from PC12 cells exhibited that DA-induced release of TNF α was blocked by knockdown of ASK1 or JNK1 (Fig. 3K).

3.4 DA induced downregulation of neurotrophins depending on TNF α in neurons

In this experiment, we tried to confirm whether DA-induced synaptic loss was associated with ASK1/JNK1/TNF α pathway. Parallely, as shown by IB analysis, PHNs exhibited lower GDNF/NGF levels after DA treatment, addition of K811, SP600125 or ATAB blocked the decrease in GDNF/NGF expression (Fig. 4A and B). The efficiency of TNF α siRNA plasmid transfection into PC12 cells was confirmed by qPCR (Fig. 4C). Parallely, as shown by IB analysis, PC12 cells exhibited the increase in GDNF/NGF levels after knockdown of ASK1/JNK1/TNF α (Fig. 4D and E). Furthermore, by IB analysis, exposure of PHNs to DA also displayed the decreased expression of Homer/synaptotagmin which were blocked by addition of K811, SP600125 or ATAB (Fig. 4F and G). Furthermore, by immunostaining, GDNF expression in PC12 cells with DA treatment was markedly decreased, while K811, SP600125 or ATAB significantly increased the protein expression (Fig. 4H). Double IF staining of PHNs with anti-vGluT1 (for staining dendritic spines) and anti-MAP2 (for staining microtubules) antibodies for probing dendritic spine density showed that vGluT1-positive signals in PHNs was markedly increased after DA treatment DA, whereas ATAB administration diminished the decrease in vGluT1-positive signals (Fig. 4I and J). These results suggested that DA impaired the production of neurotrophins and synaptogenesis through the activation of ASK1/JNK1/TNF α signaling pathway.

3.5 DA induced inactivation of Notch pathway in neurons via TNF α

Notch is also required for memory consolidation, a process believed to require remodeling of existing neurons in adults[35]. Several studies have shown that Notch also plays an important role in memory formation in adults, both in mammals and *Drosophila*[36, 37]. It is reported that Notch also plays an important role in memory formation in adults [36, 37]. Then we addressed whether DA impacted on notch signaling via the involvement of TNF α . The expressions of NICD1/HES1 were markedly decreased in PHNs in response to higher dose of DA, whereas lower dose of DA had no significant effect on the two proteins by IB analysis (Fig. 5A and B). Parallely, as shown by IB analysis, PHNs exposed to DA exhibited lower NICD1/HES1 levels, addition of ATAB blocked the decrease in NICD1/HES1 expression (Fig. 5C and D). ATAB PC12 cells after exposure to DA also displayed the decreased expression of NICD1/HES1, while knockdown of TNF α blocked the decrease in NICD1/HES1 expression PHNs (Fig. 5E and F). As determined by double IF staining using NICD1 antibody with MAP2 antibody, we confirmed that the decrease in NICD1 expression in PHNs in response to DA was diminished by ATAB (Fig. 5G). Immunostaining confirmed that addition of ATAB diminished the decrease in NICD1 expression in

response to DA in PC12 cells (Fig. 5H). Parallely, these data indicated that DA inactivated notch signaling depending on stimulation of TNF α secretion.

3.6 DA impairs the production of neurotrophins by the inactivation of Notch pathway in vitro

Hence, we examined whether DA had effect on the expressions of GDNF/NGF through notch signaling in vitro. As indicated by IB analysis, we also found that DA-induced the decreases in the expressions of GDNF/NGF in PHNs were significantly enhanced by addition of Notch inhibitor MK-0752 and reversed by addition of Notch activator Jagged-1 (Fig. 6A and B). Notch overexpression plasmid transfection into PC12 cells was further confirmed by western blot using the anti-Flag antibody (Fig. 6C). IB analysis revealed that the decreases in GDNF/NGF levels in PC12 cells in response to TNF α were abolished by Notch overexpression (Fig. 6D and E). Notch siRNA transfection into PC12 cells was further confirmed by western blot using the anti-Flag antibody (Fig. 6F). Furthermore, DA -mediated the decreases in Homer/synaptotagmin expression were enhanced by Notch silencing in PC12 cells (Fig. 6G and H). These data suggested that DA impaired the production of neurotrophins and synaptic formation through the inactivation of notch signaling in vitro.

3.7 β ASA inhibits the DA-induced the loss of neurotrophins in vitro

β ASA has been reported to have neuroprotective and synaptoprotective effects on MHE [38]. We therefore examined the effect of β ASA on the DA-mediated downregulation of GDNF/NGF. Treatment with β ASA abolished DA-mediated the increase in the expression of TNF α in a dose-dependent manner in PHNs (Fig. 7A and B). As determined by ELISA, DA-stimulated the release of TNF α was abated after β ASA treatment from PHNs with dose-dependence (Fig. 7C). And the addition of β ASA abrogated DA-induced the decrease in the notch expression with dose-dependence in PHNs (Fig. 7D and E). We then examined whether the effect of β ASA on the DA-induced the loss of neurotrophins was associated with TNF α . IB analysis indicated that DA-induced the downregulation of GDNF/ synaptotagmin levels were reversed by the addition of β ASA in PHNs, whereas, the effect of β ASA was then inhibited by addition of TNF α or MK-0752 (Fig. 7F and G). Using FM4-64 dye to probe activity-dependent synaptic vesicle recycling in PHNs revealed that synaptic activity displayed obvious reduction in response to DA, which was blocked by addition of β ASA, however, the effect of β ASA was reversed by treatment of TNF α or MK-0752 (Fig. 7H and I). These data suggested that β ASA diminished DA-mediated the loss of neurotrophins required for downregulation of TNF α and activation of notch signaling.

3.8 β ASA improves DA-induced memory impairment in MHE rats

Then, we confirmed the effect of β ASA on TNF α level, neurotrophic expression, and synaptic formation in vivo. It is clear from IF staining of cortices of MHE rats using TNF α combined with MAP2, pronounced increased levels in neurons in cortices of MHE rats were blocked by administration of β ASA (Fig. 8A), we

confirmed that the administration of β ASA significantly diminished the decrease in expression of GDNF in neurons in cortices (Fig. 8B) and in expression of synaptotagmin in neurons in hippocampi (Fig. 8C) in MHE rats. Furthermore, the decreases in the NGF/synaptotagmin expressions were detected in hippocampus of MHE rats, which were amplified by DA and recovered to the normal level by administration of β ASA, furthermore, the enhancement of DA was blocked by β ASA addition (Fig. 8D and E). Then we addressed whether β ASA impacted on the memory decline. In YM test, SA% in MHE rats, which were significantly lower ($P < 0.01$) than that of control rats, was enhanced by DA and reversed by β ASA administration, furthermore, the enhancement of DA on the SA% of MHE rats was blocked by β ASA addition (Fig. 8F). These data suggested that β ASA inhibited the impairment of synaptic formation and memory function in vivo.

4. Discussion

Neurons regulate synaptic transmission via monitoring neuronal signaling [39], and they release inflammatory cytokines, such as TNF α [40]. TNF α is one of the major inflammatory cytokines [41] and a vital acute injury associated cytokine. Elevated baseline and systemic TNF α may have roles in lower hippocampal volume [42] and hippocampal neurodegeneration [43] that triggers cognitive changes [44]. DA directly changed the synaptic formation and elevated the expression and release of TNF α in neurons in vitro. TNF α was reported to impair memory function. Neuron with DA treatment, displayed the elevation of TNF α level in media, the impairment of synaptic formation and activity and the decrease of dendrite spine, and addition of ATAB reversed synaptic loss. MHE rats also showed the increase in the expression of TNF α , neutralization of DA blocked expression of TNF α , indicating the induction of TNF α by DA. And neutralization of TNF α diminished the synaptic formation. Furthermore, neurons exposed to DA exhibited increased production and release of TNF α levels, suggesting that neuronal TNF α in turn induced disruption of synaptogenesis. Our findings strongly suggest that the increase in DA could be responsible for enhancing synaptic loss and cognitive decline by increasing TNF α expression and secretion.

Apoptosis signal-regulating kinase1 (ASK1) is one of the mitogen-activated protein kinase (MAPK) kinase family members. c-Jun N-terminal kinase (JNK) is a member of MAPK family that is upregulated by MAPK kinase family members. ASK plays an essential role in determining cell fate, including death, survival, differentiation, and mitosis [33]. The ASK1 signaling pathway is involved in the modulation of expression of pro-inflammatory cytokines through JNK1 signaling pathway [34, 45]. Inflammatory responses are frequently mediated by the JNK signaling pathway that causes production of pro-inflammatory cytokines TNF α , and IL-6 [46]. DA which appears to be implicated in inflammatory response is its ability to induce the activation of the ASK1/JNK1 signaling pathway. Notably, evidence from DA-treated primary cultured neurons suggested that DA-induced TNF α production might be due to activation of ASK1/JNK1 pathway. Findings from TNF α -treated primary cultured neurons showed that TNF α had decreased activity of Notch signaling leading to the decreased expression of GDNF and NGF. The inhibition of ASK1/JNK1 pathway, blocked DA-induced the increase in TNF α release from neurons, inactivation of Notch pathway and impairment of synaptogenesis in neurons.

Notch is a 320 kDa polypeptide that is cleaved in the trans-Golgi network by a furin-like convertase to generate a functionally active receptor consisting of a 200 kDa extracellular ligand-binding domain and a 120 kDa transmembrane domain [47, 48]. Activation of the Notch-1 receptor by the Jagged or Delta membrane bound ligands results in cleavage of the Notch-1 transmembrane domain to yield the Notch intracellular signaling domain (NICD). This 110 kDa domain of Notch translocates to the nucleus and induces expression of basic helix-loop-helix transcription repressors, such as hairy/enhancer of split (Hes), in particular Hes-1 [49, 50]. Notch is also required for memory consolidation, a process believed to require remodeling of existing neurons in adults [35]. Several studies have shown that Notch also plays an important role in memory formation in adults, both in mammals and *Drosophila* [36, 37]. Moreover, mice with antisense-reduced hippocampal Notch-1 mRNA and protein levels fail to sustain long-term potentiation but exhibit no defect in paired-pulse facilitation, a short-term form of synaptic plasticity [51]. Dysregulation of the Notch signaling pathway has been shown to contribute to a number of neurological diseases including Alzheimer's disease (AD) [52], cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy (CADASIL) [53], and cortical dysplasia [54]. Our study suggests that DA induced the dysregulation of the Notch to inhibit synaptic formation through TNF α , which was responsible for the memory impairment of MHE. Counteracting of TNF α , abated DA-induced inactivation of Notch pathway and impairment of synaptogenesis in neurons. Remarkably, DA-induced impairment of synaptogenesis was reversed by activation of Notch pathway and was enhanced by the inhibition of Notch pathway in neuron. These findings suggested that DA elicited synaptic loss via ASK1/JNK1/TNF α /Notch pathway in neuron, which suggested that this cellular pathway was associated with pathogenesis of MHE.

5. Conclusion

In this study, we are the first demonstrating that DA levels are increased under MHE conditions and DA can exert the learning impairment. the data in vitro presents show that DA plays a role in directly triggering TNF α production and accumulation in neurons, and resulting in synaptic loss, thus reflecting that DA burden modulates dendritic spine generation and synaptic formation via TNF α release. And the signals from DA mediated via ASK1/JNK1 pathway to production of TNF α operate linked to the inactivation of Notch pathway that are sensitive to the inhibition of synaptic formation in neurons. Notably, the increase in DA was targeted to study the possible therapeutic effects of β ASA on the associated increase in synaptic function and decrease in inflammation in MHE.

Abbreviations

AD: Alzheimer's disease; ANOVA: analysis of variance; ASK1: Apoptosis signal-regulating kinase1; ATAB: anti-TNF α antibody; ATCC: American Type Culture Collection; A β : amyloid β -protein; β ASA: β -asarone; BD: Becton, Dickinson and Company; BDNF: brain-derived neurotrophic factor; BSA: bovine serum albumin; CA: California; CADASIL: cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy; cDNA: complementary Deoxyribonucleic acid; CL: contacting latency; CNS: central

nervous system; DA: Dopamine; DL: drinking latency; DMEM: dulbecco's modified eagle medium; DNase: deoxyribonuclease; ECL: electrochemiluminescence; EL: entry latency; ELISA: enzyme-linked immunosorbent assay; FBS: fetal bovine serum ; Fig: figure; FITC: fluorescein isothiocyanate; FM4-64: N-(3-triethylammom iumpropyl)-4-(p-diethylam inophenylhexatrienyl) ; GDNF: glialcellline-derived neurotrophicfactor; HE: hepatic encephalopathy; Hes: enhancer of split; HES1: Hairy and enhancer of split1; HIV: human immunodeficiency virus; i.p. : intraperitoneally; IB: Immunoblotting; IF: Immunofluorescence; IL-6: Interleukin- 6; JNK: c-Jun N-terminal kinase; JNK1: c-Jun N-Terminal Kinase1; MA: Massachusetts; MAP2: microtubule-associated protein 2; MAP2B: microtubule-associated protein 2B; MAPK: mitogen-activated protein kinase; MHE: Minimal hepatic encephalopathy; mRNA: Messenger Ribonucleic Acid; NGF: Nerve Growth Factor; NICD1: Notch Intracellular Domain 1; PASW: Predictive Analytics Software; PHNs: Primary hippocampal rats neurons; PVDF: Polyvinylidene fluoride; qPCR: quantitative real-time PCR; RSC96: Rat Schwann Cells; SA%: spontaneous alternation percentage; SAMP-8: Senescence-accelerated prone-8; SD: Sprague-Dawley; SD: standard deviation; SDS-PAGE: sodium dodecyl sulfate polyacrylamide gel electrophoresis; siRNA: Small interfering RNA; SPSS: Statistical Product and Service Solutions; TAA: Thioacetamide; TNF α : tumor necrosis factor- α ; USA: the United States of America; VA: Virginia; vGluT1: vesicular glutamate transporter 1; WFT: water-finding task; YM: Y-maze

Declarations

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Authors' contributions

Saidan Ding supervised the entire project and designed the research. Jian wang conceived and designed the experiments, performed the research interpreted, and analysed the data, and wrote the paper. Xuebao Wang conceived and designed the experiments, interpreted and analysed the data, and supervised all the experimental procedure. Leping Liu and He Yu performed the research and analysed the data. Xiaoai Lu, Ruimin You and Saidan Ding analysed the data and critically revised the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets generated and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval

All experiments were conducted in accordance with the guidelines for the Care and Use of Laboratory Animals of the National Institutes of Health and Institutional Animal Care and Use Committee of Wenzhou Medical University.

Consent for publication

Not applicable

Competing interests

The authors declare that they have no competing interests.

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Figures

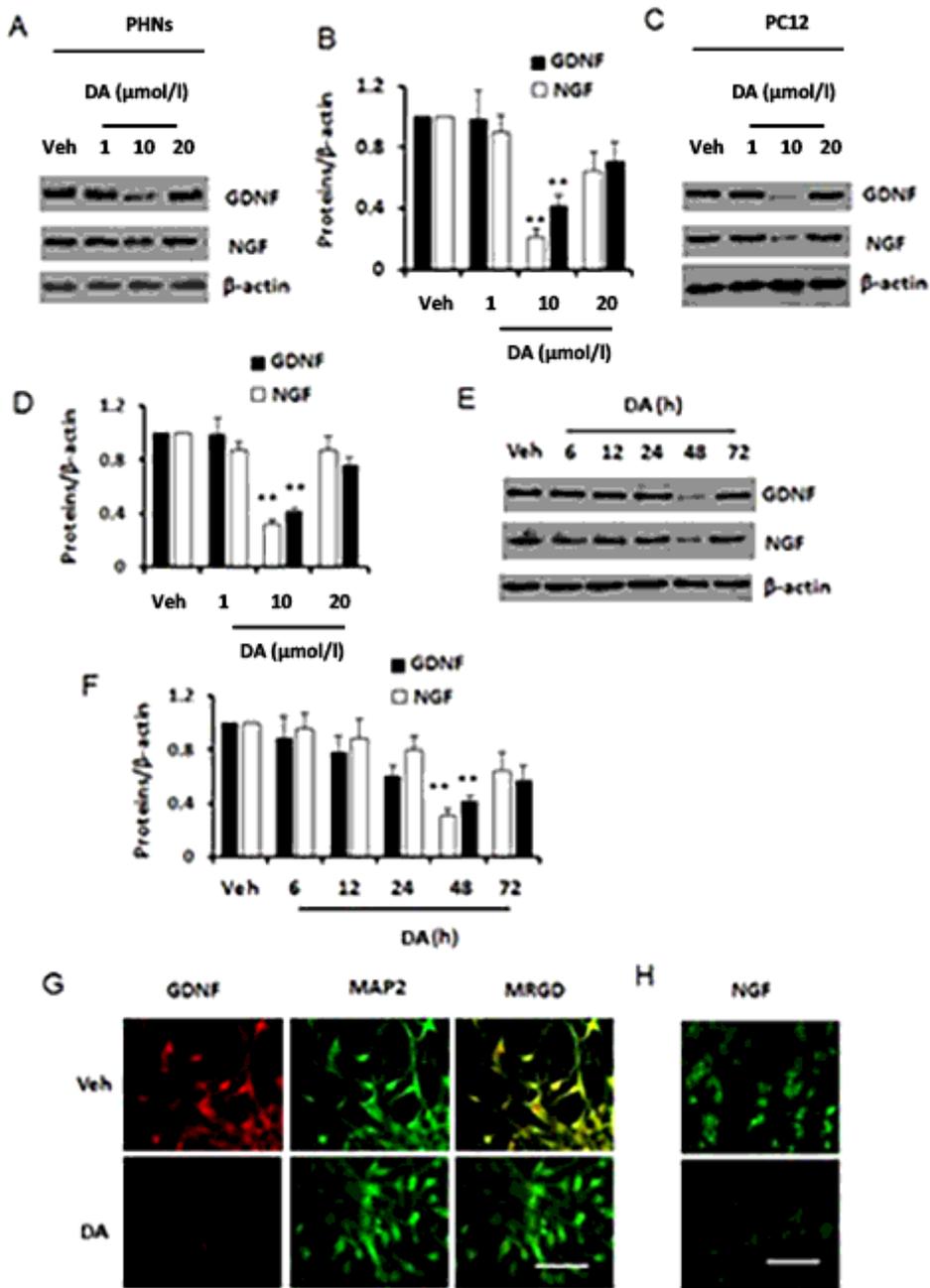


Figure 1

DA impairs synaptic formation in vitro. (A-D) IB analysis and subsequent densitometry of lysate from PHNs (A,B) and PC12 cells. (C,D) stimulated with various concentrations of DA using anti-GDNF/NGF and anti- β -actin antibodies. (E,F) IB analysis of lysate from PHNs stimulated with 2.5 mmol/L DA for various time points using anti-GDNF/NGF and anti- β -actin antibodies and subsequent densitometry. (G) Immunostaining of PHNs stimulated with DA using antibodies against GDNF (red), MAP2 (green). (H) Immunostaining of PC12 cells stimulated with DA using antibodies against NGF. Data are shown as mean \pm SD. * $P < 0.05$, ** $P < 0.01$ vs vehicle-treated group. veh, vehicle. Scale bar, 25 μm . MRGD, merged image.

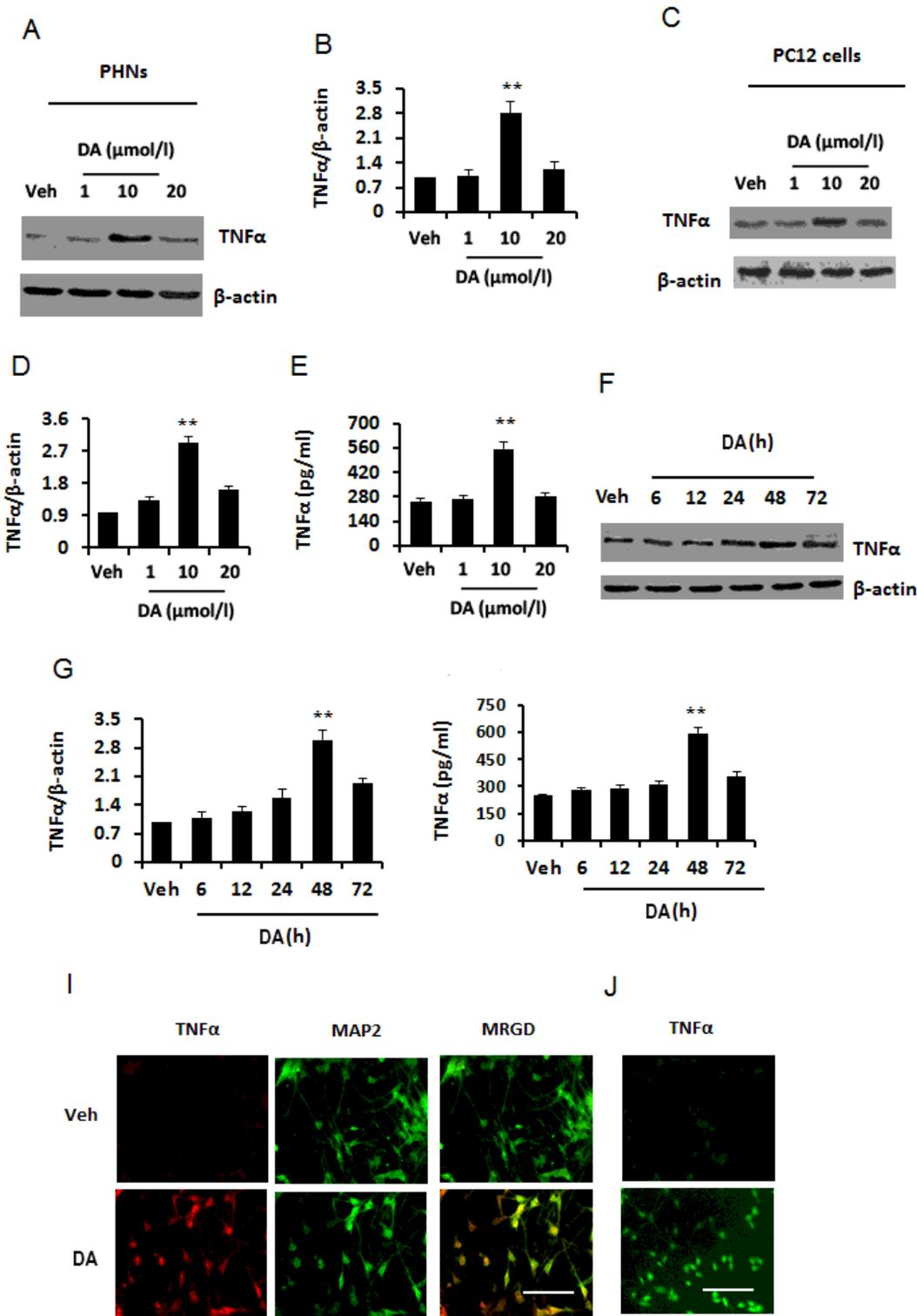


Figure 2

DA elicits the production of TNF α in neurons in vitro. (A-D) IB analysis of lysate from PHNs (A,B) and PC12 cells (C,D) stimulated with various concentrations of DA using anti-TNF α and anti- β -actin antibodies and subsequent densitometry. (E) ELISA assay for TNF α level of Supernatants from PHNs stimulated with various concentrations of DA. (F,G) IB analysis of lysate from PHNs stimulated with 2.5 mmol/L DA for various time points using anti- TNF α and anti- β -actin antibodies and subsequent

densitometry. (H) ELISA assay for TNF α level of Supernatants from PHNs stimulated with 2.5 mmol/L DA for various time points. (I) Immunostaining of PHNs stimulated with DA using antibodies against TNF α (red), MAP2 (green). (J) Immunostaining of PC12 cells stimulated with DA using antibodies against TNF α . Data are shown as mean \pm SD. *P < 0.05, **P < 0.01 vs vehicle-treated group. veh, vehicle. Scale bar, 25 μ m. MRGD, merged image.

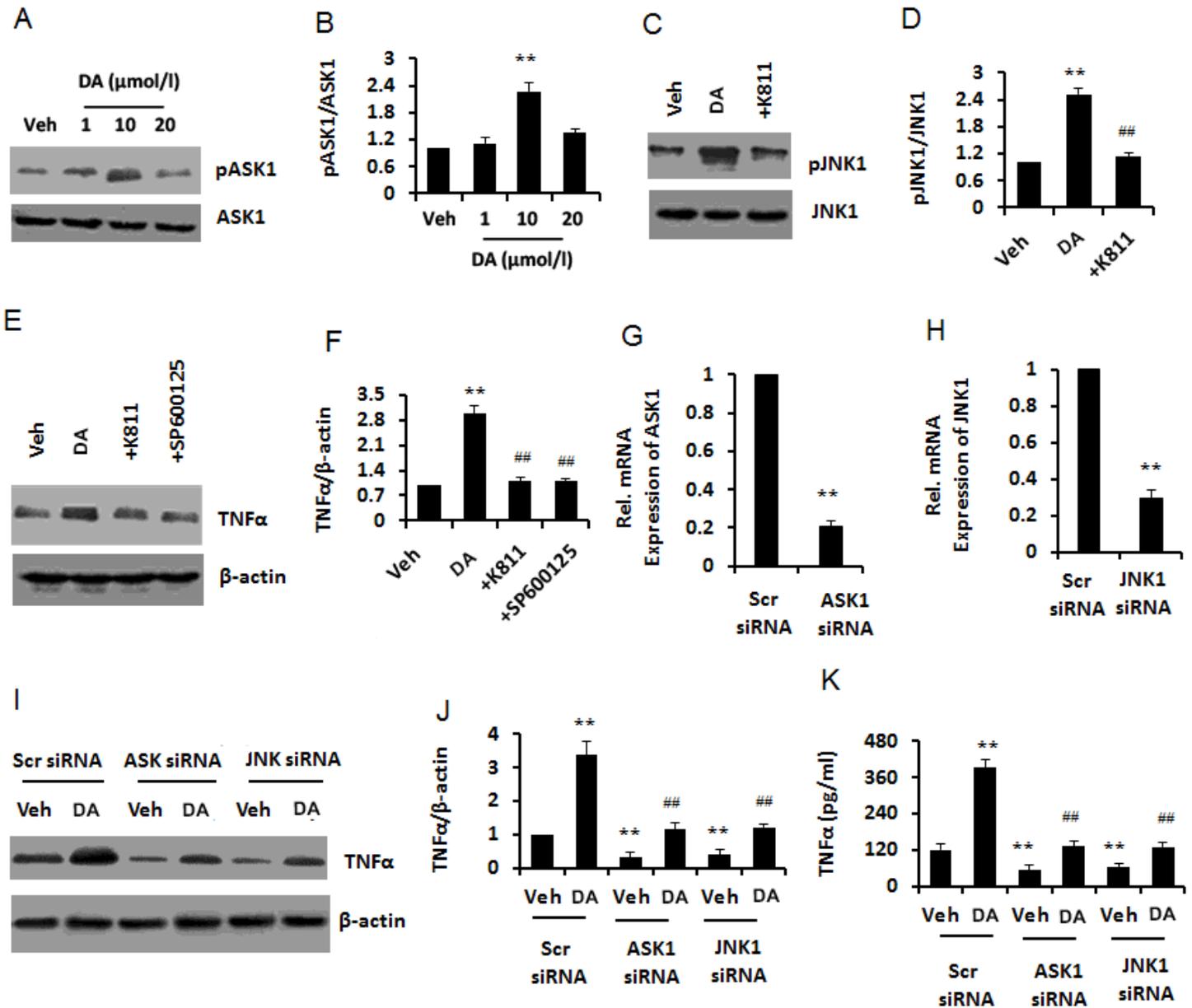


Figure 3

DA triggers TNF α production via activation of ASK1/JNK1 signaling in neurons. (A,B) IB analysis of PHNs stimulated with various concentrations of DA (10, 50, 100nmol/l) using anti- pASK1 and anti-ASK1 antibodies and subsequent densitometry. (C,D) IB analysis of PHNs stimulated with 100nmol/l DA alone or after preincubation with K811 using antibodies against pJNK1 and JNK1 and subsequent densitometry. (E,F) IB analysis of lysate from PHNs stimulated with 100nmol/l DA in the presence of K811 or SP600125 using anti-TNF α and anti- β -actin antibodies and subsequent densitometry. (G,H) RT-

PCR analysis of ASK1(G)/JNK1(H) mRNAs of PC12 cells transfected with ASK1 siRNA or JNK1 siRNA. (I,J) IB analysis of lysate from PC12 cells stimulated with 100nmol/l DA after transfection with ASK1 siRNA or JNK1 siRNA using anti-TNF α and anti- β -actin antibodies and subsequent densitometry. (K) ELISA assay for TNF α level of Supernatants from PC12 cells stimulated with 100nmol/l DA after transfection with ASK1 siRNA or JNK1 siRNA. Data are shown as mean \pm SD. *P <0.05, **P <0.01 vs vehicle-treated group. #P <0.05, ##P <0.01 vs DA-treated group. Scr, scrambled. veh, vehicle. n.s., not significant vs vehicle-treated group. Scale bar, 25 μ m. MRGD, merged image.

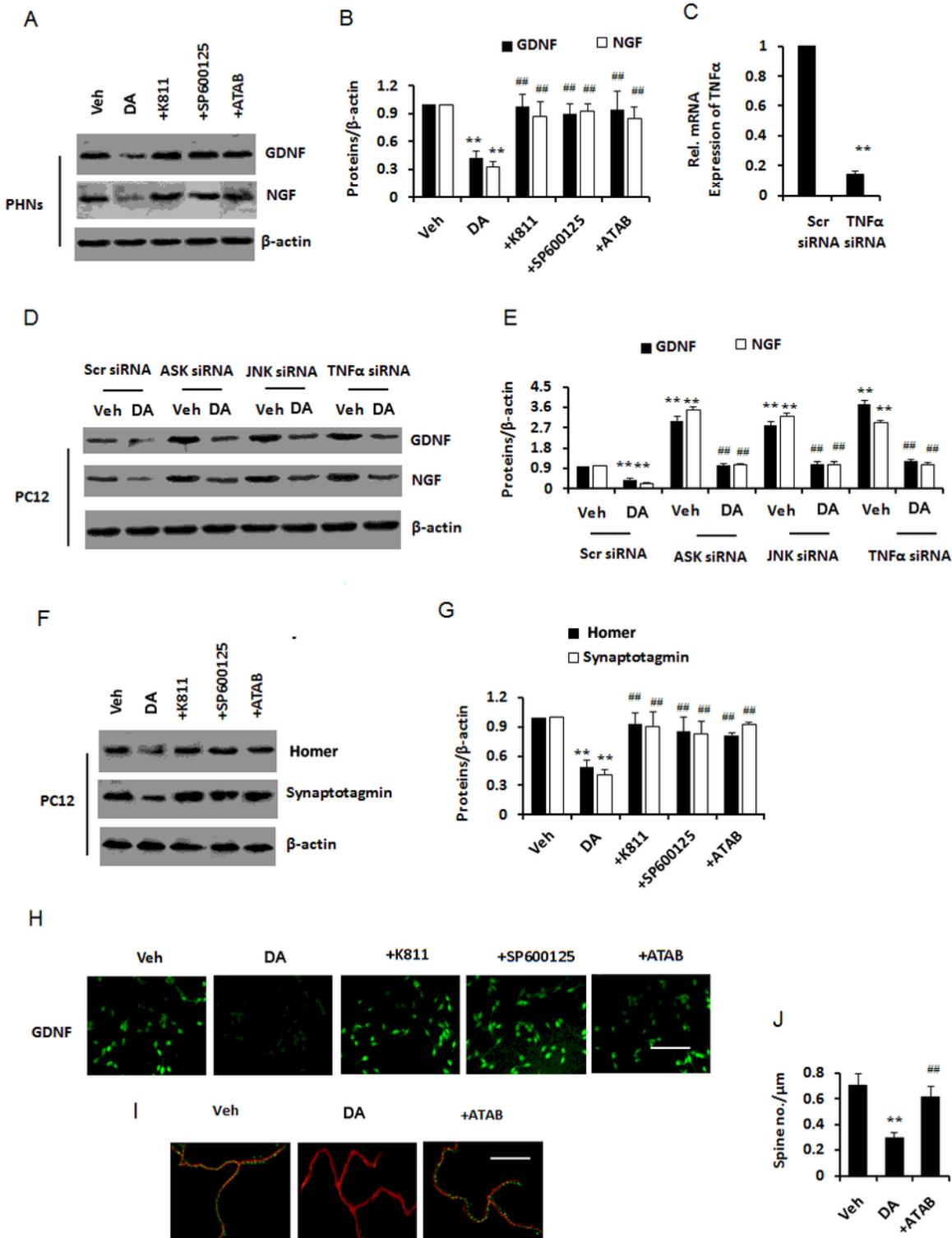


Figure 4

DA induces synaptic loss via upregulation of TNF α in vitro. (A,B) IB analysis of lysates from PHNs stimulated with DA after preincubation with K811, SP600125 or ATAB using anti- GDNF/NGF and anti- β -actin antibodies and subsequent densitometry. (C) RT-PCR analysis of TNF α mRNAs of PC12 cells transfected with TNF α siRNA. (D,E) IB analysis of lysates from PC12 cells stimulated with 100nmol/l DA after transfection with ASK1/JNK1/TNF α siRNA using anti- GDNF/NGF and anti- β -actin antibodies and subsequent densitometry. (F,G) IB analysis of lysates from PHNs stimulated with DA after preincubation with K811, SP600125 or ATAB using anti-Homer/synaptotagmin and anti- β -actin antibodies and subsequent densitometry. (H) Immunostaining of PC12 cells stimulated with 100nmol/l DA in the presence of K811, SP600125 or ATAB using antibodies against GDNF. (I,J) Immunostaining of PHNs stimulated with 100nmol/l DA after preincubation with ATAB against MAP2B (red), vGluT1 (green). Red signals indicate MAP2B for microtubule staining and green signals indicate vGluT1 for detecting excitatory synapses. Synaptic density was analyzed by counting green signals (vGluT1-positive dendritic spines) using ImageJ, and expressed per 1 μ m of apical dendrite. Data are shown as mean \pm SD. *P <0.05, **P <0.01 vs vehicle-treated group. #P <0.05, ##P <0.01 vs DA-treated group. Scr, scrambled. veh, vehicle. Scale bar, 25 μ m. MRGD, merged image.

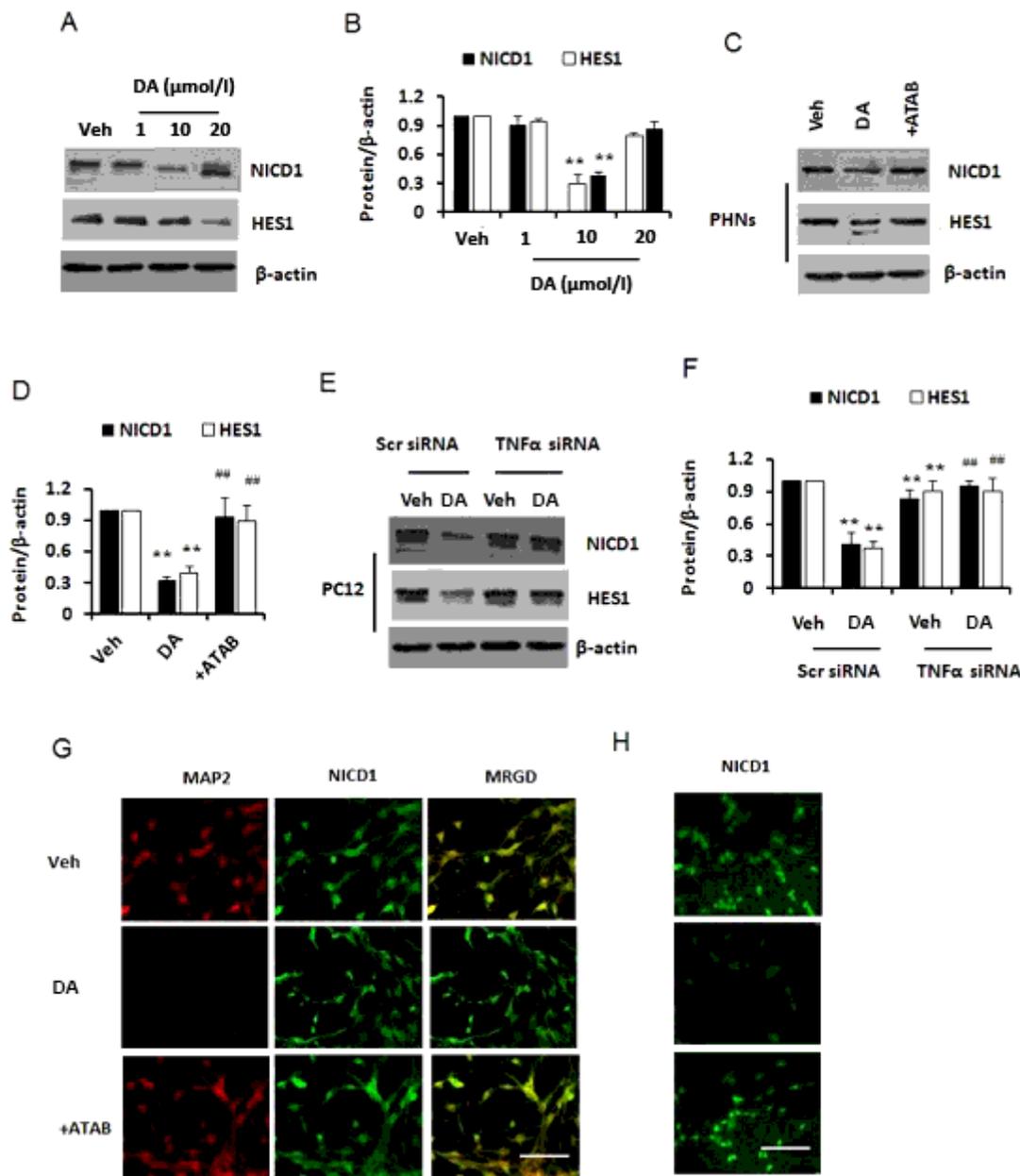


Figure 5

DA inactivates notch signaling pathway via upregulation of TNFα (A,B) IB analysis of lysate from PHNs stimulated with various concentrations of DA using anti-NICD1/HES1 and anti-β-actin antibodies and subsequent densitometry. (C,D) IB analysis of lysates from PHNs stimulated with 100nmol/l DA after preincubation with ATAB using anti- NICD1/HES1 and anti-β-actin antibodies and subsequent densitometry. (E,F) IB analysis of lysates from PC12 cells stimulated with 100nmol/l DA after transfection with TNFα siRNA using anti- NICD1/ HES1 and anti-β-actin antibodies and subsequent densitometry. (G) Immunostaining of PHNs stimulated with 100nmol/l DA in the presence of ATAB using antibodies against NICD1 (red), MAP2 (green). (H) Immunostaining of PC12 cells stimulated with 100nmol/l DA in the presence of ATAB using antibodies against NICD1. Data are shown as mean± SD. *P <0.05, **P <0.01 vs vehicle-treated group. #P <0.05, ##P <0.01 vs DA-treated group. Scr, scrambled. veh, vehicle. Scale bar, 25 μm. MRGD, merged image.

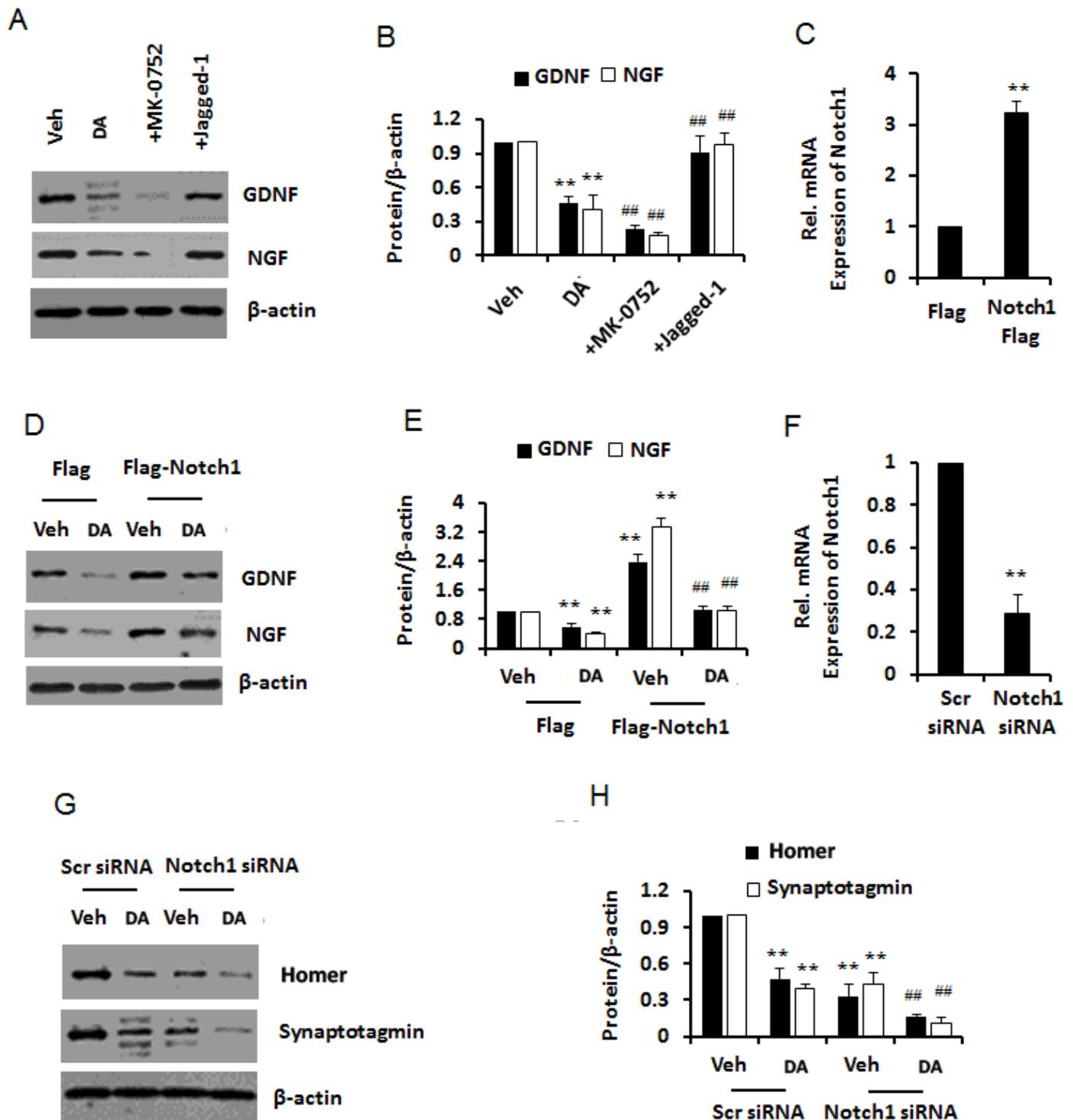


Figure 6

DA induces synaptic loss via inactivation of Notch signaling pathway in vitro. (A,B) IB analysis of lysate from PHNs stimulated with 100nmol/l DA in the presence of MK-0752 or Jagged-1 using anti-GDNF/NGF and anti-β-actin antibodies and subsequent densitometry. (C) RT-PCR analysis of notch mRNA of PC12 cells transfected with notch overexpression plasmid. (D,E) IB analysis of lysate from PC12 cells stimulated with 100nmol/l DA after transfection with Notch1 overexpression plasmid using anti-

GDNF/NGF and anti- β -actin antibodies and subsequent densitometry. (F) RT-PCR analysis of notch mRNA of PC12 cells transfected with notch siRNA. (G,H) IB analysis of lysate from PC12 cells stimulated with 100nmol/l DA after transfection with Notch1 siRNA using anti- Homer/synaptotagmin and anti- β -actin antibodies and subsequent densitometry. Data are shown as mean \pm SD. *P < 0.05, **P < 0.01 vs vehicle-treated group. #P < 0.05, ##P < 0.01 vs TNF α -treated group. Scr, scrambled. veh, vehicle. MRGD, merged image.

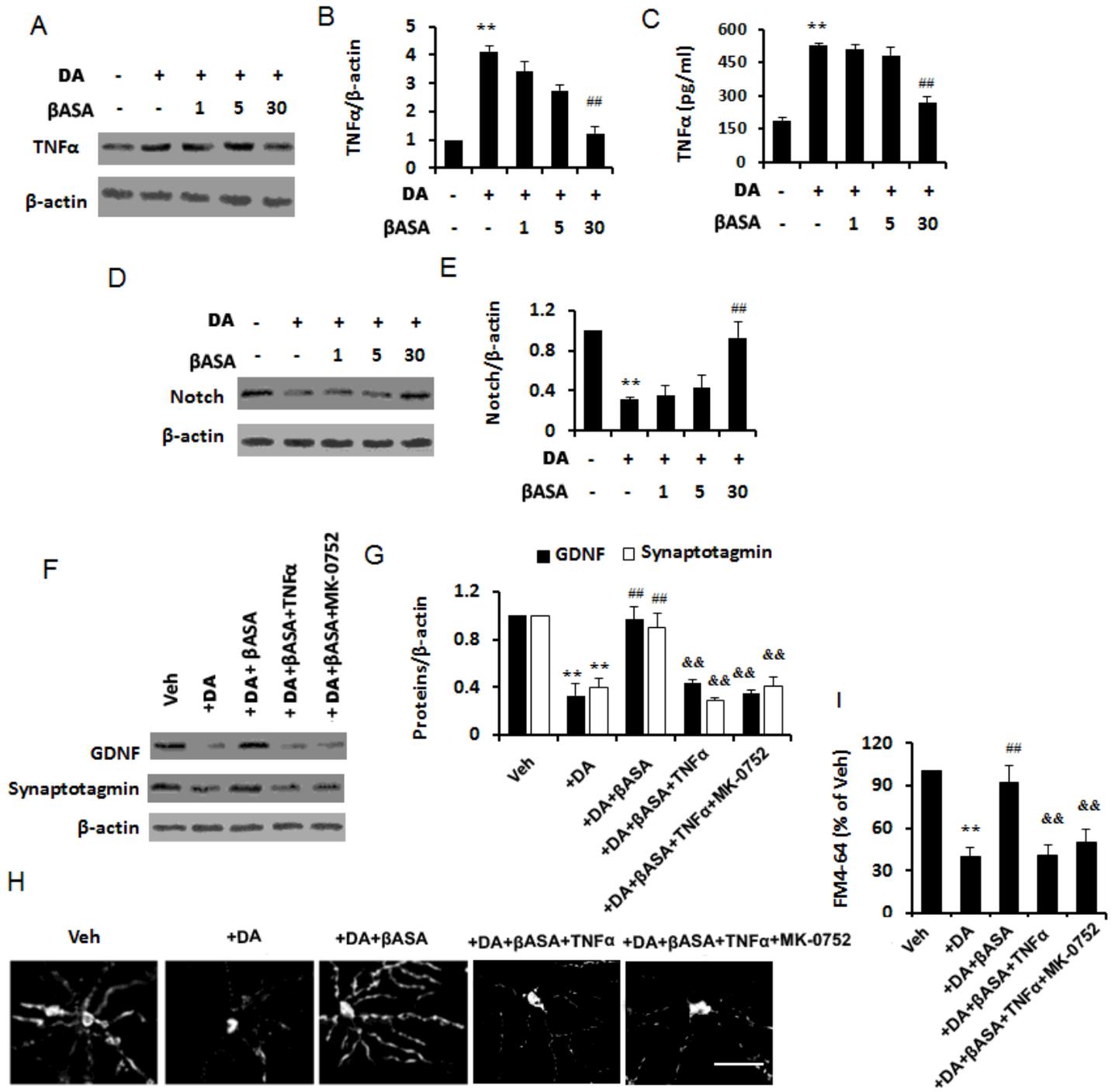


Figure 7

β ASA attenuates DA-induced synaptic loss via downregulation of TNF α (A,B) IB analysis of lysate from PHNs stimulated with 100nmol/l DA in the presence of various concentration of β ASA using anti- TNF α and anti- β -actin antibodies and subsequent densitometry. (C) ELISA assay for TNF α level of Supernatants from PHNs stimulated with 100nmol/l DA in the presence of β ASA. (D,E) IB analysis of lysate from PHNs stimulated with 100nmol/l DA in the presence of various concentration of β ASA using anti- Notch and anti- β -actin antibodies and subsequent densitometry. (F,G) IB analysis of lysate from PHNs stimulated with 100nmol/l DA in the presence of β ASA together with TNF α or MK-0752 using anti-GDNF/synaptotagmin and anti- β -actin antibodies and subsequent densitometry. (H,I) Representative image of FM4-64 staining of functional presynaptic terminals in PHNs stimulated with 100nmol/l DA in the presence of β ASA together with TNF α or MK-0752. The panel H indicates that quantitative analysis of changes on average in FM4-64 puncta intensity. Data are shown as mean \pm SD. *P <0.05, **P <0.01 vs vehicle-treated group. #P <0.05, ##P <0.01 vs DA-treated group, &P <0.05, &&P <0.01 vs DA+ β ASA-treated group. Scr, scrambled. veh, vehicle. MRGD, merged image.

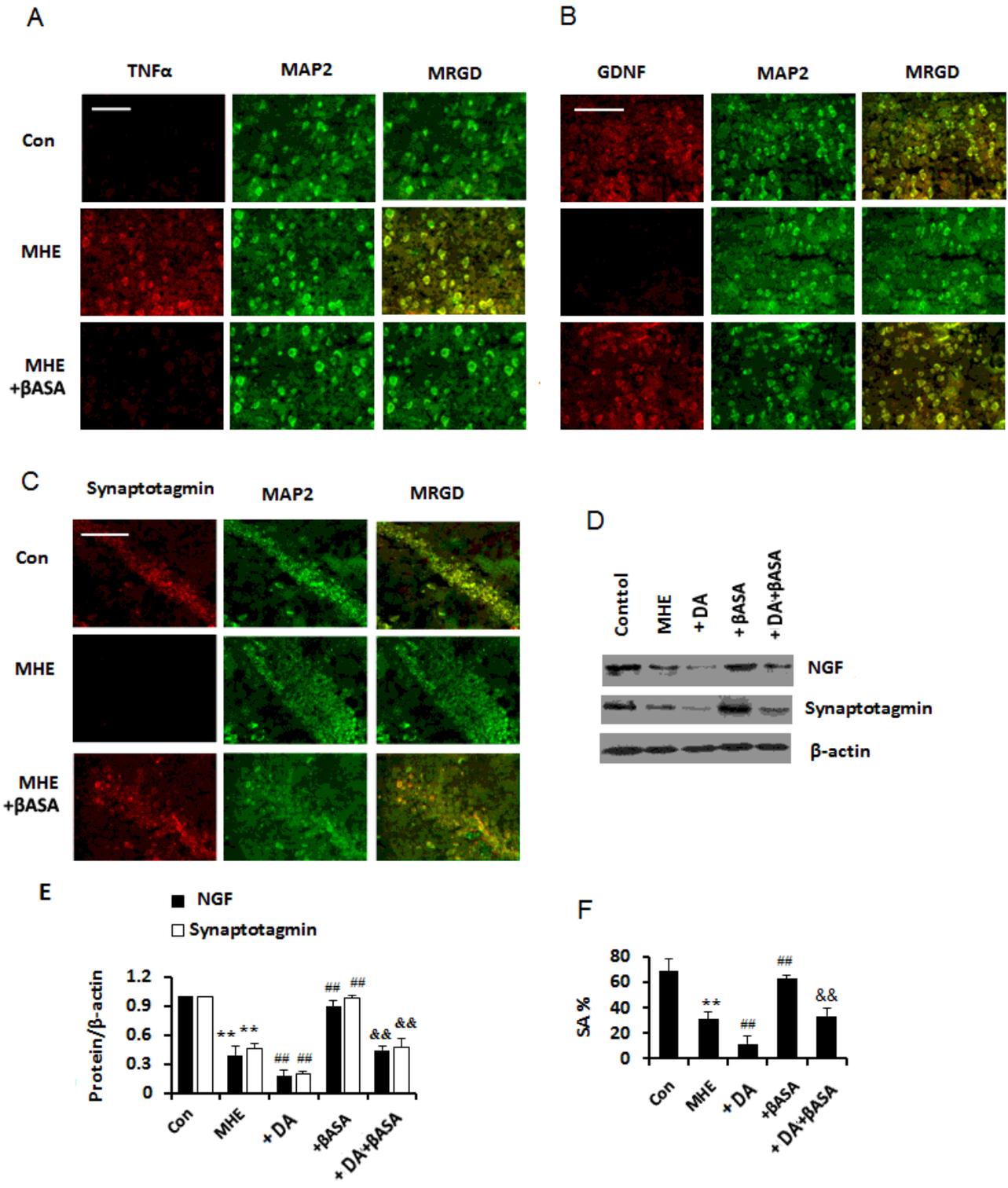


Figure 8

β BASA improves inflammation and DA-induced synaptic loss in MHE rats. (A,B) Immunostaining of free floating cortical sections from MHE rats after administration with β BASA using antibodies against TNF α /GDNF (red), GFAP (green). (C) Immunostaining of free floating hippocampal sections from MHE rats after administration with β BASA using antibodies against synaptotagmin (red), GFAP (green). (D,E) IB analysis of lysate from MHE rats after administration with DA, ATAB, or DA+ATAB using anti-

NGF/synaptotagmin and anti- β -actin antibodies and subsequent densitometry. (F) Spontaneous alternation percentage (SA%) in YM of MHE rats after administration with DA, ATAB, or DA+ATAB, Data are shown as mean \pm SD. *P <0.05, **P <0.01 vs vehicle-treated group. #P <0.05, ##P <0.01 vs control group, &P <0.05, &&P <0.01 vs MHE+DA-treated group. Scr, scrambled. con, control. Scale bar, 25 μ m. MRGD, merged image.