

Mechanism of Electric Field Assisted Screening for *Candida Tropicalis*

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1 **Mechanism of electric field assisted screening for *Candida tropicalis***

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8 **Abstract:**

9 Electric field assisted technology has been widely used in many fields. *Candida tropicalis*

10 degrading thiophene has been successfully screened by electric field assisted screening (EFAS).

11 However, the mechanism of EFAS is yet unclear. In this study, the cell concentration of *C.*

12 *tropicalis* at different locations in the electric field was studied. In the electric field, *C. tropicalis*

13 cells migrate towards the cathode plate. When the loading electric field intensity was 0.6 V /cm,

14 the concentration of cells near the cathode plate reached the maximum and it was nearly stable at

15 15 minutes of loading time. Analysis of the cell morphology at different locations showed that

16 only the budding cells could migrate in the electric field. The charged characteristic of the budding

17 cells in the electric field explained the screening mechanism of thiophene-degrading *C. tropicalis*

18 by EFAS. The results can provide a theoretical foundation for EFAS in screening other

19 biodegradable microorganisms.

20 **Keywords:** *Candida tropicalis*; Electric field; Migration; Budding cell; mechanism

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23 **1. Introduction**

24 Electric and magnetic fields can convert to each other, and they are closely related to the growth
25 and metabolism of living things[1]. Almost all living creature has some changes in properties and
26 physiological activities under a certain electric field(EF)[2]. Therefore, in recent years,
27 electric-field assisted (EFA) technology has been gradually applied in various researches[3,4].
28 Through application of EFA technology, the efficiency of soil remediation was improved [5,6], the
29 growth and trace metal content of aquatic plants are affected [2], and the harmful contaminants,
30 which strictly limited the land application of the sludge, were reduced from sludge [7]. The EFA
31 technology was also being used in drug delivery research[8]. In order to improve the activity and
32 degradation efficiency of microorganisms, the EFA technology has been applied more and more in
33 biodegradation[9,3,6].

34 In our previous studies[10], the EFA technology was used to screen functional microorganisms,
35 and a *Candida tropicalis* biodegrading thiophene was obtained. The EFA screening (EFAS)
36 method is simple, timesaving, and can also improve the activity of *C. tropicalis* at the same time.
37 The screened strains showed excellent biodegradation efficiency in study of thiophene
38 biodegradation. However, the mechanism that EFAS can screen for the microbes remains unclear.
39 Single molecule electrophoresis under the EF has been proved to be a feasible method for DNA
40 sequencing[11]. At low voltages (1.5-3.0V), the EF could provide electron donors and receptors
41 for microbial degradation of PCBs[12]. These researches explained the microbial aggregation and
42 migration in the EF.

43 Therefore, the objective of this study was to investigate the migratory and aggregate behavior of
44 *C. tropicalis* in the EF by examining the distribution of *C. tropicalis* cells under different EF. The

45 mechanism of EFAS was explored by the migratory behavior of the cells under different EF.

46 **2. Materials and methods**

47 **2.1 Strain and medium**

48 The strain used in the experiment was *Candida tropicalis* TCCC30004, which was stored in the
49 Tianjin University of Science and Technology's center of culture collection (TCCC).

50 The yeast extract peptone dextrose (YEPD) medium was prepared under the following
51 conditions: yeast extract 10 g/L, peptone 20 g/L, glucose 20 g/L, pH 7.0, at 115°C for 20 min.

52 **2.2 Yeast counting**

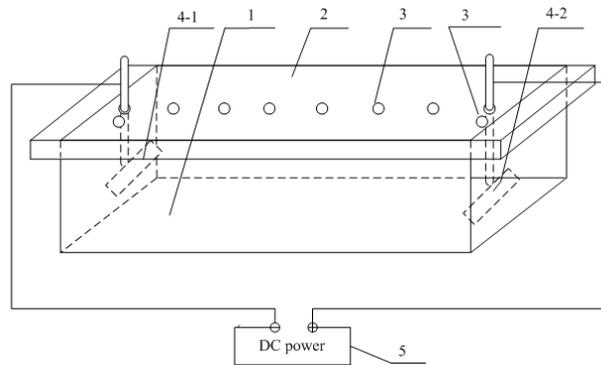
53 *C. tropicalis* cultured on the inclined surface was transferred to 5 mL YEPD medium in a 30°C
54 shaker at 180 rpm for 12h, and then 5% (v/v) of broth was inoculated into 50 mL YEPD medium.
55 After shaking culture at 30°C and 180 rpm for 12 h, the yeast were collected by centrifugation at
56 4°C and 5000 r/min. They were suspended in 50mL YEPD medium. YEPD was used to dilute the
57 suspension into samples with different concentrations of yeast, and OD₆₀₀ value of each sample
58 was determined. Yeast concentrations in samples were counted by hemocytometer. Taking OD₆₀₀
59 as the abscissa and yeast concentration as the ordinate, a curve was drawn to fit the relation curve
60 between OD₆₀₀ and yeast concentration. When counting yeast, 200 μL yeast suspension was
61 placed in a 96-well plate and OD₆₀₀ value was measured by a microplate reader. The results were
62 repeated for three times and the mean value was obtained.

63 **2.3 Experimental device**

64 An EFA experiment device was set up (Fig. 1). A cube with a length of 25cm, a width of 8cm
65 and a height of 6cm was used as the culture tank, with a cover on it. There were eight sampling
66 holes on the cover with an interval of 3cm between the holes. During the experiment, a sampling

67 needle could be used to sample through the sampling hole. The anode plate and the cathode plate
68 were two platinum plates located at the ends of the culture tank and connected to a direct current
69 power supply with adjustable voltage and current.

70



71

72 **Fig. 1 The EF experimental device (1, culture tank; 2, lid; 3, Sampling hole; 4-1, cathode**
73 **plate; 4-2, anode plate; 5, direct current power)**

74 **2.4 Experiment of aggregation or/and migration in the EF**

75 The experimental device was placed in the clean workbench, the inside and electrode of the
76 device were cleaned with alcohol, and then ultraviolet sterilization was conducted for 30min.
77 Afterwards, 1000mL of yeast suspension of a certain concentration was added to the EFA culture
78 tank. Then at 0.3 V/cm of EF strength, power was applied for 5min, and 1mL *C. tropicalis*
79 suspensions at different sampling holes(0, 3, 6, 9, 12, 15, 18, and 21cm distance from the anode
80 plate) were quickly collected into the sampling tube with a pipette. The OD₆₀₀ value of 200 μL *C.*
81 *tropicalis* suspension was measured by a microplate reader in a 96-well plate. The OD₆₀₀ value
82 was repeated three times and the mean value was obtained. The above operations were repeated
83 when the loading EF strength was changed to 0.4, 0.5, 0.6 and 0.7 V/cm, respectively. The
84 suspension must be shaken well before the loading voltage was changed each time. According to

85 the measured OD₆₀₀ value, the cell concentrations at different location of the culture tank under
86 different loading voltages were calculated. The loading EF that were most conducive to *C.*
87 *tropicalis* migration or/and aggregation were obtained.

88 After the loading EF was determined, the loading time was explored under the condition of the
89 optimum EF. The selected loading time was 5, 10, 15, 20, 25 and 30min, respectively. The cell
90 concentration at different location of the culture tank under different loading times was obtained
91 according to the above mentioned method. The loading time to facilitate *C. tropicalis* migration
92 or/and aggregation was determined.

93 **3. Results and discussion**

94 **3.1 Verification of counting method**

95 In this study, the number of samples was large, and the count of *C. tropicalis* cell needed to be
96 fast and accurate, so the hemocytometer method could not meet the requirements. The microplate
97 reader that could simultaneously determine 96 samples at a high speed was selected for counting
98 *C. tropicalis* cell in solution. The amount of cell in the solution corresponding to different OD₆₀₀
99 was measured using hemocytometer. The results were shown in Fig.2. The mathematical equation
100 of the corresponding curve was $Y = 0.9235 X + 5.5193$. Y was the cell concentration. X stood for
101 OD₆₀₀. The linear correlation coefficient was greater than 0.98.

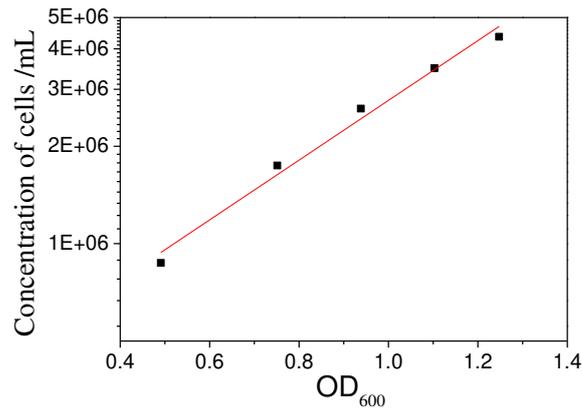


Fig. 2 The counting curve of *C. tropicalis*

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103

104 3.2The behavior under different EF

105 The loading voltage at both ends of the culture tank was changed, and the concentration of *C.*
 106 *tropicalis* at different locations was sampled and analyzed. The results were shown in Fig. 3a.

107 Under a certain loading voltages(in the test range), the cell concentration of *C. tropicalis*
 108 fluctuated slightly with the distance (distance from anode plate) in the culture tank. However, at

109 the loading EF of 0.3-0.6 V/cm, the concentration of *C. tropicalis* at 21cm distance(near the
 110 cathode plate) was significantly higher than that at other distance. And the concentration of *C.*

111 *tropicalis* at 21 cm distance increased with the increase of the loading EF. When the loading EF
 112 reached 0.6 V/cm, the cell concentration at 21 cm distance reached its maximum value. When the

113 loading EF reached 0.7 V/cm, the cell concentration at 21 cm distance decreased again. The
 114 overloading EF, whose strenght was 0.7 V/cm, might cause damage to the *C. tropicalis* cells.

115 Therefore, the loading EF strength of 0.6 V/cm was the most suitable for the migration of the cells
 116 to cathode plate. The physiological activities of cells are different in different living

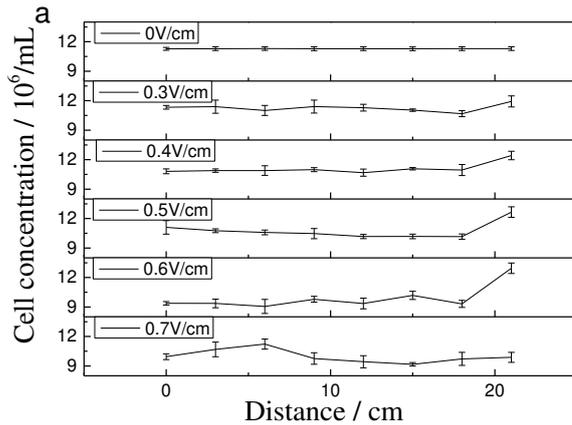
117 environments[14]. Subsequently, the above experiments were carried out with saline instead of

118 YEPD medium (Fig. 3b). The results showed that the movement trend of *C. tropicalis* cells in

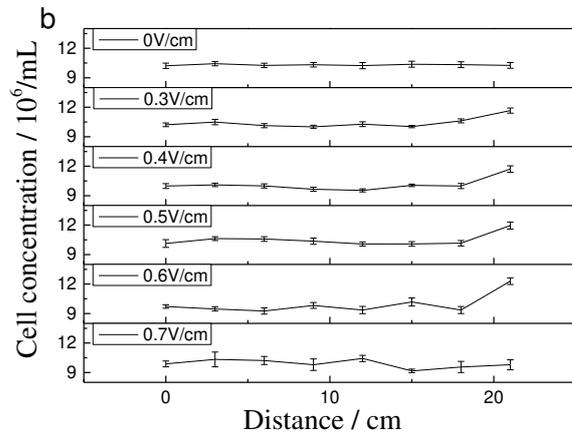
119 saline was consistent with that in YEPD medium. However, the cell concentration of *C. tropicalis*

120 at 21cm distance(near the cathode plate) in saline was lower than that in the YEPD under the same
121 condition.

122 Stimulated by an EF, cell growth, metabolism, enzyme activity, membrane permeability and
123 other aspects were affected accordingly[13]. Previous studies have shown that the biomass in
124 fermentation broth increased after direct current stimulation[10]. However, *C. tropicalis* did not
125 reproduce for less than five minutes. It just proved that *C. tropicalis* could migration and
126 aggregation to cathode plate in a certain EF. Samples taken at different distances were examined
127 by microscope. Cells in the samples taken from 21cm distance(near of cathode plate) were
128 predominantly budding cells, whereas cells in the sample taken from other distances were not so.
129 The external energy can also acted in some form as the metabolic regulator of cells[15]. The cell
130 activity of *C. tropicalis* was enhanced under the EF. The migration and aggregation of *C.*
131 *tropicalis* cells to the cathode plate may be related to the physiological activity of the cells. Under
132 a certain EF, the centers of negative charge and positive charge in the budding cell were shifted.
133 This caused the cells to become positively charged and move towards the cathode plate.
134 Therefore, the following conclusions were drawn: in the budding stage, *C. tropicalis* cells were
135 positively charged and move towards the negative pole (cathode plate) in the EF, whereas cells at
136 other growth stages do not have this property.



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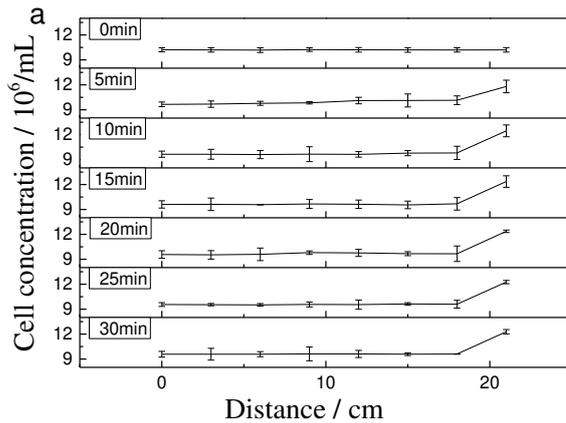
139 **Fig. 3 Effect of loading EF on cell migration (Distance, the distance from the anode plate;**
 140 **5min of loading time; a-YEPD, b-Saline)**

141 **3.3 The behavior under different loading time**

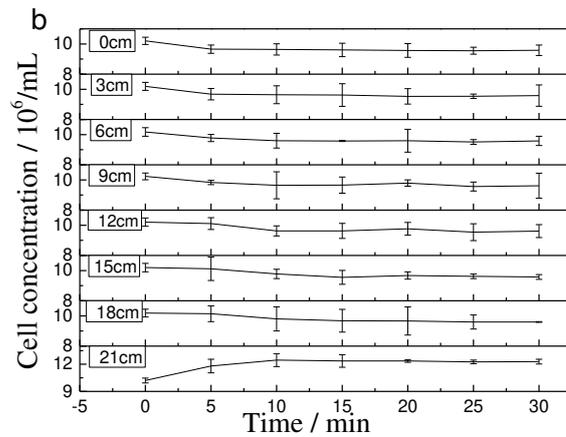
142 Under the loading EF of 0.6 V/cm, the cell concentrations at different locations were analyzed
 143 by changing the loading time. The results were shown in Fig. 4. The *C. tropicalis* cells became
 144 active in an EF. After loading the EF, the cell concentration began to decrease at almost all
 145 locations, except the cell concentration near the cathode plate(21cm of distance) was
 146 increased(Fig. 4a). The cell concentration near the anode plate(0cm of distance) was the lowest
 147 after 5min of EF loading. As the cells migrated, the concentration of cells near the anode plate did
 148 not change. And then at a distance of 12cm, the concentration of the cells didn't change when the
 149 EF was applied for 10min(Fig.4b). After loading the EF for 15min, the cell concentration at all

150 positions did not change. Except near the cathode plate(21cm of distance), the cell concentration
151 at all positions was almost the same and lower than the initial value. This was the concentration of
152 non-budding cells in the solution. At this time, almost all the budding cells gathered near the
153 cathode plate. And the cell concentration at a distance of 21cm was significantly higher than that
154 at other distance. With the extension of the loading time, the cell concentration near the cathode
155 plate became higher and higher, however the increasing trend slowed down (Fig.4 b). It might be
156 due to a decrease in the number of budding cells in the solution other than near the cathode plate.

157 After loading the EF for about 20min, the plaque that proved the attachment of bacteria to the
158 cathode plate began to appear. Therefore, in the experiment to investigate the migration behavior
159 of *C. tropicalis* in the EF, the loading time of EF should be decreased as much as possible to
160 reduce the influence of cell aggregation in the cathode plate on experimental results. The *C.*
161 *tropicalis* capable of biodegrading thiophene was screened by EFAS method[10] because only the
162 budding cells could aggregate in the cathode plate. The aggregation behavior of *C. tropicalis* cells
163 on the cathode plate is the key to the successful screening of it. Only the cells that can degrade
164 thiophene could grow and germinate in the corresponding enrichment medium and the stronger
165 the capacity to degrade thiophene, the stronger the capacity to grow and germinate.



166



167

168 **Fig.4 Effect of loading time on cell migration (a- Different distances at the same time, b-**
 169 **Same distance at different times)**

170 **4. Conclusions**

171 In this study, the migration of *C. tropicalis* cells in the direct current field were observed.
 172 Under different EF, the migration rate of cells was different. Budding *C. tropicalis* cells with a
 173 positive charge were more likely to move towards the cathode plate in the EF. Plaque attachment
 174 to the cathode plate can be observed in a short period of time (about 20 minutes). The mechanism
 175 by which thiophene-degrading *C. tropicalis* was screened by an EFAS method was explained by
 176 the characteristic of budding cells being positively charged in EF.

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182 **Compliance with ethical standards**

183 **Conflict of interest**

184 All authors declare that there are no conflict of interests.

185 **Ethical guidelines**

186 All applicable international, national, and/or institutional guidelines for the care and use of
187 animals were followed.

188

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