

# A Novel Minimally Invasive OFM Technique Combined Orthotopic Transplantation of hUC-MSCs with in vivo Monitoring of Liver Metabolic Microenvironment in Liver Fibrosis Treatment

**Hui Yang**

The Affiliated Drum Tower Hospital of Nanjing University Medical School

**Yuanyuan Xie**

The Affiliated Drum Tower Hospital of Nanjing University Medical School

**Tuo Li**

Peking Union Medical College Hospital

**Shuo Liu**

The Affiliated Drum Tower Hospital of Nanjing University of Medical School

**Sheng Zeng**

The Affiliated Drum Tower Hospital of Nanjing University Medical School

**Bin Wang** (✉ [wangbin022800@126.com](mailto:wangbin022800@126.com))

The Affiliated Drum Tower Hospital of Nanjing University Medical School <https://orcid.org/0000-0003-3981-8849>

---

## Research Article

**Keywords:** Umbilical cord mesenchymal stem cells, liver fibrosis, orthotopic transplantation, open-flow microperfusion, microenvironment, metabolomics

**Posted Date:** July 28th, 2021

**DOI:** <https://doi.org/10.21203/rs.3.rs-744958/v1>

**License:**   This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

---

# Abstract

## Background

Mesenchymal stromal cells (MSCs) transplantation showed promising therapeutic results in liver fibrosis. However, efficient cell delivery method is urgently needed and the therapeutic mechanism remains unclear. This study focused on developing a minimally invasive open-flow microperfusion (OFM) technique, which combined orthotopic transplantation of human umbilical cord-derived (hUC)-MSCs to liver and *in vivo* monitoring of liver microenvironment in mice with CCl<sub>4</sub>-induced liver fibrosis.

## Methods

The therapeutic potential of OFM route was evaluated by comparing OFM with intravenous (IV) injection route in terms of hUC-MSCs engraftment at the fibrosis liver, liver histopathological features, liver function and fibrotic markers expression after hUC-MSCs administration. OFM was also used to sample liver interstitial fluid *in vivo*, following metabolomic analysis was performed to investigate metabolic changes in liver microenvironment.

## Results

OFM route caused more hUC-MSCs accumulation in the liver and was more effective in improving the remodeling of liver structure and reducing collagen deposition in fibrotic liver than IV. OFM transplantation of hUC-MSCs reduced blood ALT and AST levels, to a greater extent than IV route. And OFM route appeared to have a more pronounced effect on ameliorating the CCl<sub>4</sub>-induced up-regulation of the fibrotic markers, such as  $\alpha$ -SMA, collagen I and TGF- $\beta$ . *In vivo* monitoring of liver microenvironment demonstrated the metabolic perturbations induced by pathological condition and treatment intervention. Two metabolites and eight metabolic pathways which were most likely to be associated with the liver fibrosis progression, were regulated by hUC-MSCs administration.

## Conclusion

The results demonstrated that the novel OFM technique would be useful for hUC-MSCs transplantation in liver fibrosis treatment and for monitoring of the liver metabolic microenvironment to explore the underlying therapeutic mechanisms.

## Background

Liver fibrosis remains a serious health problem which affects a significant number of people all over the world [1]. The onset of liver fibrosis is typically caused by a variety of pathogenic factors, including viral

hepatitis, alcohol abuse, drug toxicity, autoimmunity, and so on [2]. The chronic injuries lead to a wound-healing response, which is characterized by a switch of hepatic stellate cells (HSCs) from quiescent to an activated myofibroblast-like phenotype [3]. As the central mediators of fibrogenesis, activated HSCs release profibrogenic factors and cytokines, such as transforming growth factor beta (TGF- $\beta$ ) and alpha-smooth muscle actin ( $\alpha$ -SMA) [4–6]. Moreover, liver fibrogenesis accompanies excessive production of extracellular matrix (ECM) to reconstruct the intrahepatic structure, which would further contribute to liver cirrhosis and hepatocellular carcinoma (HCC) [7]. The reversal of liver fibrosis is crucially important for reducing the mortality relevant to liver cirrhosis and HCC. Thus far, there currently is no effective therapy for treatment of liver fibrosis. Although liver transplantation has become a preferred strategy, there remain a number of challenges in this method, including shortage of donor organs, immune rejection response and surgery complications [8, 9]. Hence, it is urgent to search for new treatment strategy.

Currently, mesenchymal stromal cell (MSC) therapy is regarded as a promising strategy for liver fibrosis treatment [10] because MSCs possess many advantageous characteristics, such as continuous self-renewal, strong proliferative ability, immunomodulatory activities, and multidifferentiation potential [11, 12]. The human umbilical cord-derived (hUC)-MSCs exhibit not only the general characters of MSCs but also relatively easy accessibility, abundant source, no substantial ethical issues and more stable biological properties, making the use of hUC-MSCs a superior choice for liver fibrosis treatment [13, 14]. Many studies have demonstrated the efficacy of hUC-MSCs therapy in liver fibrosis [15].

Up to now, the translation of many MSC therapies has not fully realized their clinical application potential. One critical but often overlooked challenge has been the administration route of MSCs; indeed, many studies have shown when the same MSC therapy is introduced through different routes of administration, the engraftment of MSCs within the liver and retention of MSCs after transplantation are different, which results in different therapeutic outcomes [16, 17]. Compared with the common method of clinical systemic administration route: intravenous (IV) injection, which resulted in “lung entrapment” [18, 19], directly delivering cells to a target tissue such as intraportal and intrahepatic injection is increasingly considered to be able to increase the therapeutic effects [12]. Therefore, the development of an orthotopic injection route is urgently required to improve MSC delivery to preclinical liver fibrosis models.

The mechanisms of MSC-based liver fibrosis therapy have not been fully delineated [20], which is another reason why the clinical application of MSC-based therapy have not been fully exploited. Growing researches suggest that a more comprehensive assessment of the change of the tissue microenvironment components during the progression of the disease and healing process could give new clues about the mechanism studies [21, 22]. However, there is no report about monitoring of liver microenvironment during liver fibrosis progression and MSCs treatment process. To achieve this, a minimally-invasive *in vivo* sampling technique that allows continuous sampling of liver microenvironment components is required.

In this study, a novel minimally-invasive open-flow microperfusion (OFM) route was developed to improve hUC-MSCs delivery efficiency to the liver for liver fibrosis treatment. MSCs entrapment in liver and

therapeutic effects of OFM route were compared with IV route. In addition, OFM was simultaneously used to sample the liver interstitial fluid *in vivo* during fibrosis progression and healing of liver fibrosis, and then combined with metabolomic analysis. This allowed for the investigation of the potential therapeutic mechanisms.

## Methods

### Isolation and culture of hUC-MSCs

With informed consent, the hUC-MSCs were isolated from fresh umbilical cord of a full-term delivery donor. The entire procedure was approved by the Medical Ethics Committee of The Affiliated Hospital of Nanjing University Medical School. The umbilical cord was washed with sterile PBS and was cut into 3-5-mm long pieces, suspended in Dulbecco's modified Eagle's medium-low glucose (DMEM-LG, gibco, USA) supplemented with 10% FBS (Gibco, USA) and 1% penicillin/streptomycin (Gibco, USA). Then the isolated cells were cultured in an atmosphere of 5% CO<sub>2</sub> at 37 °C and the medium was replaced every 3-4 days until well-developed colonies of fibroblast-like cells appeared. The cells were then digested with 0.25% trypsin and seeded in new culture bottles for further expansion. The cells of fourth generation were harvested as purified hUC-MSCs and taken for further studies. The adhesiveness properties were identified and the morphological characteristics of hUC-MSCs were detected by an inverted microscope (Olympus, Japan). Flow cytometry (BD facsaria<sup>tm</sup>, USA) was employed for hUC-MSCs characterization, data analysis was performed using FACS software.

### Liver fibrosis model

C57BL/6 mice (7–8 weeks old, weighing 19-23g) were purchased from Nanjing Medical University and housed in a temperature/humidity-controlled environment with light illumination cycles of 12 h/day and had free access to diet and water. The animal experiments performed were approved by the Institutional Animal Care and Use Committee of *Nanjing University*. To induce liver fibrosis, CCl<sub>4</sub> was dissolved in olive oil at the volume ratio of 1:3. Mice were intraperitoneally injected with 150 µL of CCl<sub>4</sub> solution twice a week for eight weeks. The normal group received 150 µL olive oil twice a week for eight weeks.

### Fabrication of OFM probe

A polyimide capillary (O.D. 0.38 mm, I.D. 0.28 mm) was used for the fabrication of the OFM probe. The capillary was perforated with 60 holes at intervals of about 50 µm on a laser ablation platform (NWR-213 system, Electro Scientific Industries, USA). The diameter of the hole was 100µm.

### Transplantation of hUC-MSCs

After induction of liver fibrosis, CCl<sub>4</sub>-exposed mice were randomly divided into a PBS model group, a hUC-MSCs OFM-treated group, and a hUC-MSCs IV-treated group. In hUC-MSCs OFM-treated group, the mouse was anesthetized and the abdominal cavity of the mouse was opened. OFM probe with 60 holes (100 µm

diameter) was implanted into the left lobe of liver under the traction of catheter. One end of the probe was plugged with an empty syringe, another end of the probe was connected to a push pump and  $1 \times 10^6$  hUC-MSCs in 300  $\mu$ L PBS were delivered at flow rate of 25  $\mu$ L/min. In IV group, same amount of hUC-MSCs was injected to the caudal vein of the mouse.

### **Cell labeling and homing experiments *in vivo***

hUC-MSCs were labeled by dyechloromethylbenzamido-1,1'-dioctadecyl-3,3',3'-tetramethyl indocarbocyanine perchlorate (CM-Dil, Sigma-Aldrich, USA) in accordance with the manufacturer's guidance. After digestion and centrifugation,  $6 \times 10^6$  hUC-MSCs were resuspended in 400  $\mu$ L CM-Dil solution with concentration of 20  $\mu$ g/mL. The mixed suspension was incubated at 37 °C for 15 min, then at 4 °C for 15 min. Then, the labeled-cells were washed with PBS 3 times to eliminate free CM-Dil residual. Finally, the labeled hUC-MSCs suspension was prepared with a concentration of  $3.3 \times 10^6$  cells/mL. To evaluate the distribution of hUC-MSCs delivered by OFM or IV route in liver fibrosis model at days 3, 7, 14 and 21, the same amount of CM-Dil-labeled hUC-MSCs were administrated using transplantation method of OFM or IV as described above. Mice were sacrificed 3, 7, 14 or 21 days after cell transplantation. Livers, lungs, spleens, hearts and kidneys were collected and fixed in 4 % (v/v) paraformaldehyde (PFA) overnights. Then the tissues were transferred to 20% (w/v) sugar for 12 h and 30 % (w/v) sugar overnight for dehydration. After dehydration, the tissues were embedded in chilled OCT and frozen at -80 °C. Cryosections of liver, lung, spleen, heart and kidney samples were prepared with a thickness of 12- $\mu$ m (three slides each sample) and incubated with DAPI dye (Sigma-Aldrich, USA) in the dark. The distribution of CM-Dil-labeled cells in different tissues was observed by confocal microscope (Leica Microsystems, Germany).

### **Serum transaminase levels**

Serum samples were collected and serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) levels were measured on the same day by a chemistry analyzer (VITROS 5600, USA).

### **Histopathological and immunohistochemical evaluation**

Liver tissues were removed from mice instantly after euthanization and divided into sections. One section was washed with PBS, fixed in 4% PFA and embedded in paraffin. Then the liver samples were cut into 4  $\mu$ m-thickness slices. To illustrate the histological details, hematoxylin&eosin (H&E) staining was performed. To evaluate the collagen deposition, masson's trichrome and sirius-red was performed. Immunohistochemistry (IHC) staining was also performed with antibodies against  $\alpha$ -SMA (abcam, UK), collagen I (abcam, UK) or TGF- $\beta$  (abcam, UK). The staining results were detected with an optical microscope (Leica, Germany).

### **Evaluation of liver fibrosis gene expression markers**

Total RNA was harvested from another unmanipulated liver tissue using TRIzol<sup>®</sup> reagent (Invitrogen, USA) and was reverse-transcribed into cDNA with HiScript<sup>®</sup> RT SuperMix for qPCR (Vazyme Biotech Co., Ltd, China) using 10 µg RNA. ChamQ Universal SYBR qPCR Master Mix (Vazyme Biotech Co., Ltd, China) was used for template amplification with a primer for each of the transcripts examined. PCR reagents were assessed by a three color real-time PCR machine (Applied Biosystems, Carlsbad, CA). All reactions repeated three times. Relative quantification of gene expression was performed through normalizing to the expression of β-actin as an internal control. The mRNA expression levels of α-SMA and collagen I in model group and hUC-MSCs-treated groups were compared with the normal group. The primer sequences including α-SMA, collagen I, and β-actin were listed in table 1.

**Table 1.** Primer sequences

Gene name	Primer sequences(5'-3')
α-SMA	GAACACGGCATCATCACCAAC
	CTCCAGAGTCCAGCACAATACC GCTCCTCTTAGGGGCCACT
Collagen I	
	CCACGTCTCACCATTGGGG
β-Actin	GGCTGTATTCCCCTCCATCG
	CCAGTTGGTAACAATGCCATGT

### OFM sampling

OFM sampling of liver interstitial fluid was performed in normal group, model group and hUC-MSCs OFM-treated group. The operation of OFM sampling was similar to OFM transplantation of hUC-MSCs described above with a little modification. In brief, one end of the OFM probe was connected to a push pump (Cole-Parmer, USA) and another end of the OFM probe was connected to a pull pump (Harvard Apparatus, USA) after implantation into the liver. The perfusion fluid was PBS. The flow rate of the push pump and the pull pump was 2 µL/min. The probe was perfused for 30 min for equilibration, then 80µL dialysate sample was obtained from each mouse. The dialysates were stored at -80 °C until the time of analysis.

### UPLC-ESI HR MS/MS

The liver dialysates obtained by OFM sampling were analyzed using ultra performance liquid chromatography-electrospray high resolution MS/MS (UPLC-ESI HR MS/MS). In brief, 80 µL dialysate sample was dried with a vacuum dryer, and then re-dissolved in 50 µL of ethanol containing 0.25 µg/mL isoprenaline (Sigma-Aldrich, USA) as internal standard (IS). The mixture was centrifuged for 15 min at a rate of 10000 g at 4 °C. The supernatant was collected for UPLC-ESI HR MS/MS analysis. Samples were loaded into the UPLC system (Ultimate 3000, Thermo Fisher Scientific, USA) equipped with a BEH amide column (1.7µm, 2.1 mm ID × 20 mm, Waters, USA). The mobile phase A was water containing 20 mM ammonium acetate and the mobile phase B was acetonitrile containing 0.1% formic acid. The LC

separations were 30 min per sample with a flow rate at 0.3 mL/min using LC gradient reported in the previous literature [23]. The metabolite profile was acquired using Orbitrap Fusion Lumos MS (Thermo Fisher Scientific, USA) with positive-ion mode. Major operating parameters were as follows: electrospray voltage +3000 V, *m/z* range 150–1000, ion transfer tube temperature 325 °C, vaporizer temperature 275 °C, sheath gas flow 30 Arb, auxiliary gas flow 10 Arb. Metabolite fragments were obtained under high energy collisional dissociation (HCD) mode with a collision energy at 20 eV.

## Metabolite identification

The high resolution MS spectra and MS/MS spectra was loaded into Compound Discovery (Thermo Fisher Scientific, USA). Database search was performed after peak alignment and peak area integration. The putative annotation of the metabolites was achieved by precisely matching mass with mzCloud, ChemSpider and MassList databases. Further structural confirmation was performed by MS/MS fragmentations obtained in a data dependent acquisition mode. The concentration data of the hUC-MSCs OFM-treated group, the model group and the normal group were acquired by comparing the peak area of each metabolite with that of IS. Principal component analysis (PCA) was performed using SIMCA-P 14.1 (Umetrics AB, Sweden) with the concentration data. Volcano plot and heat map were obtained using Graphpad Prism5.0 (GraphPad Software, USA) with the concentration data. Metabolic pathway analysis was performed on MetaboAnalyst website (<https://www.metaboanalyst.ca>).

## Statistical analysis

The data generated by the experiments described above was presented as mean  $\pm$  standard error of the mean (SEM). Graphpad Prism 5.0 was used to generate graphs. Statistical significance of differences between groups was evaluated using a standard one-way analysis of variance (ANOVA). A *p* value of less than 0.05 was considered statistically significant.

# Results

## Characterization of isolated huc-MSCs

The established hUC-MSCs cell lines exhibited typical fibroblastic-like morphology (Fig. 1A). The immune phenotypes of hUC-MSCs were characterized by flow cytometry. As shown in Fig. 1B, the cells positively expressed CD73, CD90 and CD105, which are MSC-specific surface antigens. However, they negatively expressed hematopoietic stem cell-specific markers, such as CD34 and CD45.

## OFM route allowed a greater number of transplanted cells to seed into the fibrosis liver

OFM is a minimally invasive, universal and continuous *in vivo* sampling technique, which is widely used in sampling interstitial fluid components via macroscopic holes without nominal cut-off value [24]. In this study, to our knowledge, OFM was firstly employed for orthotopic transplantation of hUC-MSCs into the fibrosis liver. As shown in Fig. 2A and D, OFM probe was inserted in the liver and perfused with hUC-MSCs

suspension at a constant speed. hUC-MSCs were delivered into the liver through macroscopic holes on the OFM probe (Fig. 2C). The number of transplanted hUC-MSCs that can seed at the injury site is one of the essential prerequisite for successful cell therapy. In order to investigate the homing ability of hUC-MSCs transplanted by OFM route, fluorescent CM-Dil was introduced to label hUC-MSCs. CM-Dil labeling had no distinct effect on cell morphology (Fig. 1C). CM-Dil-labeled hUC-MSCs were infused into mice with liver fibrosis by OFM and IV route. As shown in Fig. 3, in the OFM group, most of hUC-MSCs were trapped in the liver, the fluorescence signal intensity did not change significantly from day 3 to day 14 after infusion and on day 21, the fluorescence signals decreased obviously. In addition, red fluorescence was weak in lung and spleen though the signal intensity increased slowly as time progressed. However, in the IV group, hUC-MSCs accumulated mainly in the lung while the cell concentration was much lower in the liver. The IV group showed higher hUC-MSCs existence in the spleen than OFM group. There was no CM-Dil staining within the kidney and heart in both OFM and IV group, which indicated hUC-MSCs could not migrate to the kidney and heart.

### **OFM makes the cell therapy more effective for liver fibrosis**

Since the OFM route allowed more cells to reside in the fibrosis liver, we further explored the effect of OFM route in the treatment of liver fibrosis and compared the difference in therapeutic efficacy between OFM and IV routes. Mice were euthanized and livers were collected. The macroscopic condition of the livers was evaluated firstly. As shown in Fig. 4A, the livers obtained from the model group were dark red and characterized by a rough and fractured surface with raised speckle and blunt edge, while livers of the normal group were bright-red and distinguished by a soft and smooth surface with sharp edges. Both OFM and IV routes improved the overall appearance of livers as compared to the model group. In the OFM group, the number of the raised speckle was less than that in the IV group.

H&E staining results (Fig. 4B) showed that the structure of liver lobules in normal group was regular, and there was no portal inflammation. In model group, the structure of liver lobules was destroyed, the arrangement of liver cell cords was disordered, the liver cells were denatured and partially necrotic, accompanied by obvious inflammatory cell infiltration in the portal area and around the central vein. Both OFM and IV transplantation significantly reduced portal inflammation and improve the lobular structural pattern. In addition, from the H&E staining results, the degree of liver injury in OFM group was less than that in IV group.

Masson's trichrome and sirius-red staining results (Fig. 4B) showed that small amounts of collagen fibers were distributed in portal area in normal group, while fibrous septa were detected in model group. OFM and IV transplantation of hUC-MSCs could reduce fibrous expansion around the portal area in different degrees. In comparison with IV group, masson's trichrome staining of livers from OFM group appeared more similar to normal group.

Liver function was evaluated through measurements of serum biomarkers, including serum ALT and AST. As shown in Fig. 5, after 8 weeks of CCl<sub>4</sub> administration, remarkable increase of serum ALT and AST were

detected in model group. hUC-MSCs transplantation via OFM or IV route significantly reduced ALT and AST levels in comparison with model group. Whereas, it is worth noting that the serum ALT and AST levels in OFM group were noticeably lower than that in IV group, and showed no significant difference comparison to normal group, which indicated an approximate return of these liver function biomarkers to normal level.

The mRNA expression levels of  $\alpha$ -SMA and collagen I that were the main ECM components of the fibrotic liver and widely used as fibrotic markers, were evaluated through qRT-PCR. Model group showed significant elevation of  $\alpha$ -SMA and collagen I mRNA expression as compared to normal group. After hUC-MSCs transplantation via OFM or IV route, the mRNA expression levels of  $\alpha$ -SMA and collagen I decreased significantly. The relative expression of  $\alpha$ -SMA mRNA and collagen I mRNA in OFM group showed no difference with normal group, whereas there was significant difference between IV group and normal group, indicating that OFM route appeared to have a more pronounced effect on  $\alpha$ -SMA and collagen I mRNA expression (Fig. 6A and B).

The protein levels of  $\alpha$ -SMA and collagen I were evaluated through IHC technique. As shown in Fig. 6C, similar to mRNA expression,  $\text{CCl}_4$  exposure induced an excessive accumulation of  $\alpha$ -SMA and collagen I, and hUC-MSCs transplantation via OFM or IV route prevented this effect in different degrees. The protein expression of  $\alpha$ -SMA and collagen I in OFM group was lower than that in IV group. The presence of cytokine TGF- $\beta$ , which was known as a strong inducer of HSCs to produce excessive ECM components, was also investigated by IHC. Intense expression in the entire liver sections of model group was observed while TGF- $\beta$  existed in few specific regions of liver sections in normal group. hUC-MSCs treatment decreased the expression of TGF- $\beta$ , and protein levels of TGF- $\beta$  in OFM group was lower than that in IV group, which indicating that OFM route was more effective on ameliorating the induced up-regulation of TGF- $\beta$  in liver fibrosis.

### In vivo liver metabolomic analysis

As an *in vivo* and *in situ* sampling technique, OFM was employed to monitor metabolic changes in liver microenvironment during progression and treatment process of hepatic fibrosis. As shown in Fig. 1B and D, OFM probe was inserted in the liver and perfused with PBS at a constant speed. Liver dialysates obtained by OFM sampling were analyzed using mass spectrometry. PCA was applied to the metabolite concentration data set (shown in Fig. 7A). The normal group (N), the model group (M) and the hUC-MSCs OFM-treated group (H) could be well separated on PCA score plots, indicating that pathological condition and treatment intervention induced metabolic perturbations in liver microenvironment. Hierarchical clustering based on metabolite patterns of liver dialysates samples was also performed and the result was presented as heat maps (Fig. 7B). It's shown that even though model group and hUC-MSCs OFM-treated group overlapped slightly, most samples clearly grouped into three differentiated clusters, which was consistent with PCA analysis result. Individual metabolites in model group vs. normal group or hUC-MSCs OFM-treated group vs. model group was plot in volcano plot (Fig. 7C&D). The specific metabolites that changed significantly ( $p$  value  $< 0.05$  and fold change (FC)  $> 2.0$  or  $< 0.5$ ) during progression and

treatment process of liver fibrosis were highlighted. As shown in Fig. 7C, it was found that the levels of 9 metabolites in liver dialysates of model group were significantly higher compared with that in the normal group, and the levels of 10 metabolites were significantly down-regulated. Figure 7D showed that 12 metabolites were significantly up-regulated in the hUC-MSCs OFM-treated group compared to the model group. It is worth noting that the levels of 9-cis-retinoic acid and dehydroretinaldehyde were significantly decreased in model group compared to the normal group, which had the opposite trend in hUC-MSCs OFM-treated group compared to the model group. The findings indicate a potential role for hUC-MSCs in regulating the levels of 9-cis-retinoic acid and dehydroretinaldehyde.

The metabolite concentration data set was imported into MetaboAnalyst website to explore the significantly disturbed metabolic pathways under the conditions of this study. Figure 7E and F showed the overview of metabolic pathway analysis. Pathway with impact factor > 0.10 and *p* value < 0.05 was considered as potential target pathway. It was found that there were 10 metabolic pathways that were most likely to be associated with the development of liver fibrosis. 8 of these metabolic pathways were regulated by treatment with hUC-MSCs, including alanine, aspartate and glutamate metabolism, D-glutamine and D-glutamate metabolism, arginine biosynthesis, aminoacyl-tRNA biosynthesis, taurine and hypotaurine metabolism, histidine metabolism, arginine and proline metabolism, and glycine, serine and threonine metabolism.

## Discussion

The aims of this study were to improve hUC-MSCs administration routes in liver fibrosis models by developing a minimally-invasive OFM, which was simultaneously employed as an *in vivo* and *in situ* sampling technique to monitor liver metabolic microenvironment during fibrosis progression and healing of liver fibrosis. To evaluate the therapeutic potential of OFM transplantation of hUC-MSCs in liver fibrosis, OFM route was compared with IV route in terms of hUC-MSCs engraftment at the fibrosis liver, liver histopathological features, liver function and fibrotic markers expression after hUC-MSCs administration. Metabolomic analysis of liver dialysates obtained by OFM sampling was performed to investigate metabolic changes in liver microenvironment. Our results showed that compared with IV, OFM route caused more hUC-MSCs accumulation in the liver. Histopathological data revealed that OFM route was more effective in reducing collagen deposition in fibrotic liver and improving liver structure than IV. OFM transplantation of hUC-MSCs significantly reduced blood ALT and AST levels, to a greater extent than IV route. Consistent with the histopathological and biochemical results, OFM route appeared to have a more pronounced effect on ameliorating the CCl<sub>4</sub>-induced upregulation of the fibrotic markers, such as  $\alpha$ -SMA, collagen I and TGF- $\beta$ . Metabolomic analysis of liver dialysates showed that pathological condition and treatment intervention induced metabolic perturbations in liver microenvironment. The levels of 9-cis-retinoic acid and dehydroretinaldehyde were significantly decreased in model group compared to the normal group, which had the opposite trend in hUC-MSCs OFM-treated group compared to the model group. 8 metabolic pathways, which were most likely to be associated with the liver fibrosis progression, were regulated by treatment with hUC-MSCs. In summary, the results of our study strongly

suggested the high therapeutic potential of utilizing OFM route in the administration of hUC-MSCs for treatment of liver fibrosis and monitoring of the liver metabolic microenvironment *in vivo*.

Considering the complexity of liver fibrosis progression, efficient therapy options are severely limited. Cell therapy, particularly involving MSCs, has gained great interest for the treatment of liver diseases [25]. It has been confirmed that MSCs administration could inhibit CCl<sub>4</sub>-induced liver fibrosis in mice [26, 27]. The optimum route of administration is an important clinical issue for stem cell therapy. The accessibility of the transplanted MSCs to their expectable target tissue and the efficacy of therapy are strongly dependent on the administration route [28]. Thus the transplantation route adopted in the stem cell therapy is supposed to be tailored to the lesion type and customized according to the mechanism of action of MSCs. IV is historically most common route of administration adopted by current studies and the benefits of IV delivery in liver disease treatment were demonstrated. Sun et al reported that an obvious improvement of liver function was observed in rats acute liver failure (ALF) following transplantation of bone marrow stromal cells (BMSCs) via IV route [3]. Xuan et al proved that IV delivery of hUC-MSCs can evidently inhibited liver fibrosis [29]. However, it has been well documented that MSCs transplanted via IV route mainly accumulated in the lung and spleen, thereby showed a low presence in the injured liver tissue [30]. As expected, similar scenario was observed in our study: although mononuclear phagocytic system, the liver and spleen showed MSCs presence, IV route indeed caused more cell entrapment in the lung, which may be a hindrance for MSCs to fully display its therapeutic effects. In our study, the therapeutic effect of hUC-MSCs therapy with IV injection was modest and transient. To increase the number of cells that reach to the injury site, increased initial cell dose is required, which could lead to tremendous cell loss. Wang and others found a dose as high as  $5 \times 10^6$  cells/mouse to observe any effect in colitis treatment with IV transplantation of MSCs [31]. Wang also found that for IV injection, when the cell dose increase, most mice will lead increased mortality because of potential pulmonary cell embolus. Hoogduijn et al reported that myocardial infarction (MI) in healthy vasculature was induced even though IV injection dose was  $0.5 \times 10^6$  MSCs per kg body weight, and a high dose was also more likely to induce a severe systemic immune response [32]. So the IV route of administration needs several injections, such huge demand of MSCs brings a cell source issue, which is an ongoing technical and operational challenge.

To bypass cell entrapment in the lung, alternative administration routes, such as hepatic artery injection, portal vein injection, intrahepatic injection and intraperitoneal injection have been investigated in several studies. Sang et al found that MSCs transplantation through intraportal injection was superior to hepatic intra-arterial injection, intrahepatic injection and peripheral intravenous injection for treatment of ALF in swine, which could prolong the life span of swine with ALF, inhibit apoptosis and improve liver function [28]. In our study, OFM was employed for orthotopic transplantation of hUC-MSCs into the fibrosis liver. OFM is widely reported as an *in vivo* sampling technology based on microdialysis (MD) with advantages of minor invasiveness, sampling continuity [33]. However, unlike MD probes, OFM probes are free of semi-permeable membrane, which is replaced by macroscopic holes. Therefore, OFM probes allow direct access to the tissue milieu and sampling of interstitial components is not limited by molecular weight or

lipophilicity. For example, OFM has been repeatedly reported as an alternative to MD in sampling of large molecules such as albumin and insulin or lipophilic drugs like ketoprofen [34–36]. However, it has been rarely reported that OFM can be used for orthotopic administration of substances into different tissues *in vivo*. To our knowledge, the potential of OFM transplantation of hUC-MSCs for treatment of liver fibrosis was firstly explored. Compared with IV route, OFM route allowed a greater number of transplanted cells to seed into the fibrotic liver, so hUC-MSCs did not immediately enter to the blood flow and could fully display its therapeutic effects. Additionally, OFM route didn't suffer from pulmonary embolism issue. In our study, OFM transplantation of  $1 \times 10^7$  cell/mouse was even tried, but no embolism-related death occurred. The histopathological and biochemical data and the results of levels of fibrotic markers presented in our study suggested OFM transplantation of hUC-MSCs into livers was a promising technique for treatment of liver fibrosis.

It is a great challenge to completely understand the action mechanisms of liver fibrosis and hUC-MSCs treatment for the disease. Metabolomics is a powerful tool to reveal the potential targets for therapeutic intervention by identifying significantly altered metabolites relevant to the progression of the disease and the healing process [37]. However, metabolomics strategy is commonly performed with collected tissues as subjects, which may give rise to contamination of samples and bias of the metabolite concentrations, resulting in misinterpretation of the results [38–40]. Hence, in our study, the simultaneously *in vivo* and *in situ* sampling of liver interstitial fluid via OFM, and then combined with metabolomic analysis, will help to obtain solid information on metabolome alterations. It was found the levels of 9-cis-retinoic acid and dehydroretinaldehyde, which significantly increased with the development of CCl<sub>4</sub>-induced hepatic fibrosis, were reversed with transplantation of hUC-MSCs. 9-cis-retinoic acid and dehydroretinaldehyde belongs to the class of organic compounds known as retinoids. 9-cis-retinoic acid is a ligand of retinoic acid X receptor (RXR). Previous study found that the 9-cis-retinoic acid and synthetic RXR agonists had an inhibition effect on synthesis of fibronectin and collagen I and HSC proliferation [41]. Lee reported that 9-cis-retinoic acid treatment decreased the TGF- $\beta$ 1 mRNA level in L929 fibroblasts [42]. Further study revealed that the activation of PPAR gamma-RXR heterodimer represses the TGF- $\beta$ 1 gene. Therefore 9-cis-retinoic acid and dehydroretinaldehyde possibly were the potential hUC-MSCs-targeted metabolites in treatment of liver fibrosis.

Using metabolic pathway analysis, we identified 8 metabolic pathways, including alanine, aspartate and glutamate metabolism, D-glutamine and D-glutamate metabolism, arginine biosynthesis, aminoacyl-tRNA biosynthesis, taurine and hypotaurine metabolism, histidine metabolism, arginine and proline metabolism, and glycine, serine and threonine metabolism, which were significantly regulated in hUC-MSCs-treated group versus the model group. These metabolic pathways may play a crucial role in underlying therapeutical mechanism of hUC-MSCs in liver fibrosis. Du et al recently reported that the transdifferentiation of HSCs to myofibroblastic (MF)-HSCs was strongly dependent on glutamine [43].  $\alpha$ -ketoglutarate, the end-product of D-glutamine and D-glutamate metabolism, which helped to replenish the TCA cycle to satisfy the biosynthetic demands of highly proliferative MF-HSCs [44]. Glutamine metabolism was mediated by liver fibrosis related signal transduction pathway, such as TGF- $\beta$ , hedgehog

(Hh), hypoxia-inducible factor 1 $\alpha$  (HIF-1 $\alpha$ ) and wnt signaling pathways[45]. Arginine metabolism generates nitric oxide (NO) by NO synthase. It's reported that in acute liver injury, NO has the ability to oxidase lipids, proteins and DNA, promoting cell injury and death [46]. It was found that the activation of arginase located in arginine biosynthesis pathway accelerated the resolution of inflammation and promoted tissue repair [47, 48]. Taurine plays important roles in reducing reactive oxygen species (ROS) formation and restoring mitochondrial function. Taurine administration to fibrotic rats protects the liver from oxidative damage [49]. However, Wu et al observed elevated level of taurine in CCl<sub>4</sub>-induced chronic liver injury [50]. The over activation of the conversion from cysteine to taurine may result in GSH depletion, which in turn, exacerbated liver damage. Shi et al reported that in MSC significantly altered aminoacyl-tRNA biosynthesis of liver-resident immune cells in acute liver injury model [26]. Gloria et al found that alterations in blood metabolites participating in glycine, serine and threonine metabolism coexisted with active profibrotic transcriptomic programs such as TGF- $\beta$  signaling pathways, ECM-receptor interaction and cell adhesion molecules pathway. Even though the roles of these key metabolic pathways needed to be further validated in our study, our results gave new insights into the mechanisms of MSC-based liver fibrosis therapy.

The results presented in this study showed that OFM transplantation of hUC-MSCs could possibly be further investigated as an alternative route in clinical therapy of liver fibrosis and *in vivo* OFM sampling of liver interstitial fluid combined with metabolomic analysis increased understanding of liver fibrosis progression and therapeutic mechanism in hUC-MSCs therapy, which could lead to improved clinical therapeutic outcomes for liver fibrosis. However, there are still a few limitations in the study. The comparison between OFM route and other common local delivery routes such as intraportal injection and intrahepatic injection in terms of stem cell entrapment in liver and therapeutic effect is not performed. Furthermore, as it is widely believed that the liver repair effect of MSCs mainly depend on their paracrine manner by which MSCs secrete various soluble factors, the monitoring of cytokines in liver environment can be included in addition to small molecular metabolites by OFM sampling, which would promote mechanism studies of stem cell therapy.

## Conclusion

In summary, a minimally invasive OFM technique, which can be used not only to deliver hUC-MSCs to the liver but also to sample liver interstitial fluid *in vivo*, was successfully developed in this study. It was demonstrated that OFM transplantation of hUC-MSCs is superior to IV route in treatment of liver fibrosis due to its advantages of increasing hUC-MSCs delivery to the target tissue, inhibiting collagen deposition, improving liver function and reducing expression of fibrotic markers. *In vivo* monitoring of liver microenvironment demonstrated the metabolic perturbations induced by pathological condition and treatment intervention. Following metabolomic analysis of liver dialysates revealed that hUC-MSCs treatment reversed the levels of 9-cis-retinoic acid and dehydroretinaldehyde that significantly increased with the development of CCl<sub>4</sub>-induced hepatic fibrosis. 8 metabolic pathways were significantly regulated after hUC-MSCs treatment in liver microenvironment, including alanine, aspartate and glutamate

metabolism, D-glutamine and D-glutamate metabolism, arginine biosynthesis, aminoacyl-tRNA biosynthesis, taurine and hypotaurine metabolism, histidine metabolism, arginine and proline metabolism, and glycine, serine and threonine metabolism. Our results indicated that OFM transplantation of hUC-MSCs and OFM sampling of interstitial fluid could possibly be applied in clinical therapy and therapeutic mechanisms study in the future.

## Abbreviations

**HSCs**: hepatic stellate cells; **TGF- $\beta$** : transforming growth factor beta;  **$\alpha$ -SMA**: alpha-smooth muscle actin; **ECM**: extracellular matrix; **HCC**: hepatocellular carcinoma; **MSC**: mesenchymal stromal cell; **hUC**: human umbilical cord-derived; **IV**: intravenous; **OFM**: open-flow microperfusion; **CM-Dil**: dyechloromethylbenzamido-1,1'-dioctadecyl-3,3',3'-tetramethyl indocarbocyanine perchlorate; **PFA**: paraformaldehyde; **ALT**: aminotransferase; **AST**: aspartate aminotransferase; **H&E**: hematoxylin&eosin; **IHC**: immunohistochemistry; **UPLC-ESI HR MS/MS**: ultra performance liquid chromatography-electrospray high resolution MS/MS; **IS**: internal standard; **HCD**: high energy collisional dissociation; **PCA**: principal component analysis; **SEM**: standard error of the mean; **ANOVA**: analysis of variance; **MI**: myocardial infarction; **MD**: microdialysis; **MF**: myofibroblastic; **Hh**: hedgehog; **HIF-1 $\alpha$** : hypoxia-inducible factor 1 $\alpha$ ; **NO**: nitric oxide; **ROS**: reactive oxygen species.

## Declarations

### Ethics approval and consent to participate

All umbilical cord samples were taken after informed and written consent, and the study was approved by the Research Ethics Board of Nanjing Drum Tower Hospital (permit number: 2017–161-01). The animal experiments performed were approved by the Institutional Animal Care and Use Committee of *Nanjing University*.

### Consent for publication

Not applicable.

### Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

### Competing interests

The authors declare that they have no competing interests.

### Funding

This study was supported by the National Natural Science Foundation of China (NSFC) [82070459 and 81571213 (Bin Wang), and 81800583 (YuanyuanXie)], National Key Research and Development Program of China (2017YFA0104304), Nanjing Medical Science and Technique Development Foundation (YKK20071, QRX17006, QRX17057, and ZKX20016), Nanjing Department of Health (201803024).

### Authors' contributions

BW and HY designed the study. YyX and SL carried out the isolation and culture of HUCMSCs. HY, YyX, TL and SZ performed animal experiments and analyzed the data. HY and TL performed metabolomic analysis. BW, HY and YyX gave financial support. HY wrote the first draft of the manuscript and all authors read, edited, and approved the final manuscript.

### Acknowledgments

The authors thank all of the umbilical cord donors in this study for their collaborative participation.

## References

1. Asrani SK, Devarbhavi H, Eaton J, et al. Burden of liver diseases in the world. *J Hepatol.* 2019;70:151–71.
2. Poelstra K. Crucial steps towards any effective treatment. *Nat Rev Gastroenterol Hepatol.* 2016;13:67–8.
3. Seki E, Schwabe RF. Hepatic Inflammation and Fibrosis: Functional Links and Key Pathways. *Hepatology.* 2015;61:1066–79.
4. Ghavami S, Cunnington RH, Gupta S, et al. Autophagy is a regulator of TGF-beta(1)-induced fibrogenesis in primary human atrial myofibroblasts. *Cell Death Dis.* 2015;6:e1696.
5. Higashi T, Friedman SL, Hoshida Y. Hepatic stellate cells as key target in liver fibrosis. *Adv Drug Delivery Rev.* 2017;121:27–42.
6. Ma JJ, Qiu YZ, Wang M, et al. Locostatin Alleviates Liver Fibrosis Induced by Carbon Tetrachloride in Mice. *Dig Dis Sci.* 2019;64:2570–80.
7. Lee YA, Wallace MC, Friedman SL. Pathobiology of liver fibrosis: a translational success story. *Gut.* 2015;64:830–41.
8. Dutkowski P, Linecker M, DeOliveira ML, et al. Challenges to Liver Transplantation and Strategies to Improve Outcomes. *Gastroenterology.* 2015;148:307–23.
9. McGlynn KA, Petrick JL, London WT. Global Epidemiology of Hepatocellular Carcinoma An Emphasis on Demographic and Regional Variability. *Clin Liver Dis.* 2015;19:223–38.
10. Zhang D. A clinical study of bone mesenchymal stem cells for the treatment of hepatic fibrosis induced by hepatolenticular degeneration. *Genet Mol Res.* 2017;16:16019352.
11. Liu WH, Song FQ, Ren LN, et al. The multiple functional roles of mesenchymal stem cells in participating in treating liver diseases. *J Cell Mol Med.* 2015;19:511–20.

12. Zhao L, Chen SQ, Shi XW, et al. A pooled analysis of mesenchymal stem cell-based therapy for liver disease. *Stem Cell Res Ther.* 2018;9:72.
13. Wu KH, Liu YL, Zhou B, et al. Cellular therapy and myocardial tissue engineering: the role of adult stem and progenitor cells. *Eur J Cardiothorac Surg.* 2006;30:770–81.
14. Jo CH, Kim OS, Park EY, et al. Fetal mesenchymal stem cells derived from human umbilical cord sustain primitive characteristics during extensive expansion. *Cell Tissue Res.* 2008;334:423–33.
15. Zhang LT, Peng XB, Fang XQ, et al. Human umbilical cord mesenchymal stem cells inhibit proliferation of hepatic stellate cells in vitro. *Int J Mol Med.* 2018;41:2545–52.
16. Li ZR, Hu XJ, Mao JJ, et al. Optimization of Mesenchymal Stem Cells (MSCs) Delivery Dose and Route in Mice with Acute Liver Injury by Bioluminescence Imaging. *Mol Imaging Biol.* 2015;17:185–94.
17. Giri J, Galipeau J. Mesenchymal stromal cell therapeutic potency is dependent upon viability, route of delivery, and immune match. *Blood Adv.* 2020;4:1987–97.
18. Spriet M, Hunt GB, Walker NJ, et al. Scintigraphic tracking of mesenchymal stem cells after portal, systemic intravenous and splenic administration in healthy beagle dogs. *Vet Radiol Ultrasound.* 2015;56:327–34.
19. Liu ZW, Mikrani R, Zubair HM, et al. Systemic and local delivery of mesenchymal stem cells for heart renovation: Challenges and innovations. *Eur J Pharmacol.* 2020;876:173049.
20. Yin F, Wang WY, Jiang WH. Human umbilical cord mesenchymal stem cells ameliorate liver fibrosis in vitro and in vivo: From biological characteristics to therapeutic mechanisms. *World J Stem Cells.* 2019;11:548–64.
21. Hadrévi J, Ghafouri B, Sjörs A, et al. Comparative metabolomics of muscle interstitium fluid in human trapezius myalgia: an in vivo microdialysis study. *Eur J Appl Physiol.* 2013;113:2977–89.
22. Wang L, Pi Z, Liu S, et al. Targeted metabolome profiling by dual-probe microdialysis sampling and treatment using *Gardenia jasminoides* for rats with type 2 diabetes. *Sci Rep.* 2017;7:10105.
23. Li N, Li SM, Li T, et al. Co-Incorporated Mesoporous Carbon Material-Assisted Laser Desorption/Ionization Ion Source as an Online Interface of In Vivo Microdialysis Coupled with Mass Spectrometry. *Anal Chem.* 2020;92:5482–91.
24. Li T, Yang H, Li X, et al. Open-flow microperfusion combined with mass spectrometry for in vivo liver lipidomic analysis. *Analyst.* 2021;146:1915–23.
25. Ng NN, Thakor AS. Locoregional delivery of stem cell-based therapies. *Sci Transl Med.* 2020;12:eaba4564.
26. Shi XW, Liu JQ, Chen DY, et al. MSC-triggered metabolomic alterations in liver-resident immune cells isolated from CCl<sub>4</sub>-induced mouse ALI model. *Exp Cell Res.* 2019;383:111511.
27. Mazhari S, Gitiara A, Baghaei K, et al. Therapeutic potential of bone marrow-derived mesenchymal stem cells and imatinib in a rat model of liver fibrosis. *Eur J Pharmacol.* 2020;882:173263.

28. Sang JF, Shi XL, Han B, et al. Intraportal mesenchymal stem cell transplantation prevents acute liver failure through promoting cell proliferation and inhibiting apoptosis. *Hepatobiliary Pancreatic Dis Int.* 2016;15:602–11.
29. Xuan J, Feng W, An ZT, et al. Anti-TGF $\beta$ -1 receptor inhibitor mediates the efficacy of the human umbilical cord mesenchymal stem cells against liver fibrosis through TGF $\beta$ -1/Smad pathway. *Mol Cell Biochem.* 2017;429:113–22.
30. Gholamrezanezhad A, Mirpour S, Bagheri M, et al. In vivo tracking of In-111-oxine labeled mesenchymal stem cells following infusion in patients with advanced cirrhosis. *Nucl Med Biol.* 2011;38:961–7.
31. Wang M, Liang C, Hu H, et al. Intraperitoneal injection (IP), Intravenous injection (IV) or anal injection (AI)? Best way for mesenchymal stem cells transplantation for colitis. *Sci Rep.* 2016;6:30696.
32. Hoogduijn MJ, Roemeling-van Rhijn M, Engela AU, et al. Mesenchymal Stem Cells Induce an Inflammatory Response After Intravenous Infusion. *Stem Cells Dev.* 2013;22:2825–35.
33. Jadhav SB, Khaowroongrueng V, Derendorf H. Microdialysis of Large Molecules. *J Pharm Sci.* 2016;105:3233–42.
34. Bodenlenz M, Ellmerer M, Schaupp L, et al. Bioavailability of insulin detemir and human insulin at the level of peripheral interstitial fluid in humans, assessed by open-flow microperfusion. *Diabetes Obesity Metabolism.* 2015;17:1166–72.
35. Pickl KE, Magnes C, Bodenlenz M, et al. Rapid online-SPE-MS/MS method for ketoprofen determination in dermal interstitial fluid samples from rats obtained by microdialysis or open-flow microperfusion. *J Chromatogr B.* 2007;850:432–9.
36. Ellmerer M, Schaupp L, Brunner GA, et al. Measurement of interstitial albumin in human skeletal muscle and adipose tissue by open-flow microperfusion. *Am J Physiol Endocrinol Metab.* 2000;278:E352-6.
37. Tan G, Zhou Q, Liu K, et al. Cross-platform metabolic profiling deciphering the potential targets of Shenfu injection against acute viral myocarditis in mice. *J Pharm Biomed Anal.* 2018;160:1–11.
38. Scalbert A, Brennan L, Fiehn O, et al. Mass-spectrometry-based metabolomics: limitations and recommendations for future progress with particular focus on nutrition research. *Metabolomics.* 2009;5:435–58.
39. Pawar AK, Samhitha T, Prasanna B, et al. Metabolomics: current technologies and future trends. *Indo Am J Pharm Sci.* 2018;5:5114–21.
40. Wang L, Pi ZF, Liu S, et al. Targeted metabolome profiling by dual-probe microdialysis sampling and treatment using *Gardenia jasminoides* for rats with type 2 diabetes. *Sci Rep.* 2017;7:10105.
41. Hellems K, Verbuyst P, Quartier E, et al. Differential modulation of rat hepatic stellate phenotype by natural and synthetic retinoids. *Hepatology.* 2004;39:97–108.
42. Lee SJ, Yang EK, Kim SG. Peroxisome proliferator-activated receptor-gamma and retinoic acid X receptor alpha represses the TGF beta 1 gene via PTEN-mediated p70 ribosomal S6 kinase-1 inhibition: Role for Zf9 dephosphorylation. *Mol Pharmacol.* 2006;70:415–25.

43. Du K, Hyun J, Premont RT, et al. Hedgehog-YAP Signaling Pathway Regulates Glutaminolysis to Control Activation of Hepatic Stellate Cells. *Gastroenterology*. 2018;154(5):1465–79.
44. Hou W, Syn WK. Role of Metabolism in Hepatic Stellate Cell Activation and Fibrogenesis. *Frontiers in Cell Developmental Biology*. 2018;6:150.
45. Khomich O, Ivanov AV, Bartosch B. Metabolic Hallmarks of Hepatic Stellate Cells in Liver Fibrosis. *Cells*. 2020;9:24.
46. Ganz T, Wainstein J, Gilad S, et al. Serum asymmetric dimethylarginine and arginine levels predict microvascular and macrovascular complications in type 2 diabetes mellitus. *Diabetes Metab Res Rev*. 2017;33:e2836.
47. Martinez FO, Helming L, Gordon S. Alternative Activation of Macrophages: An Immunologic Functional Perspective. *Annu Rev Immunol*. 2009;27:451–83.
48. Bogdan C. Nitric oxide and the immune response. *Nat Immunol*. 2001;2:907–16.
49. Devi SL, Anuradha CV. Mitochondrial damage, cytotoxicity and apoptosis in iron-potentiated alcoholic liver fibrosis: amelioration by taurine. *Amino Acids*. 2010;38:869–79.
50. Wu F, Zheng H, Yang ZT, et al. Urinary metabonomics study of the hepatoprotective effects of total alkaloids from *Corydalis saxicola* Bunting on carbon tetrachloride-induced chronic hepatotoxicity in rats using H-1 NMR analysis. *J Pharm Biomed Anal*. 2017;140:199–209.

## Figures

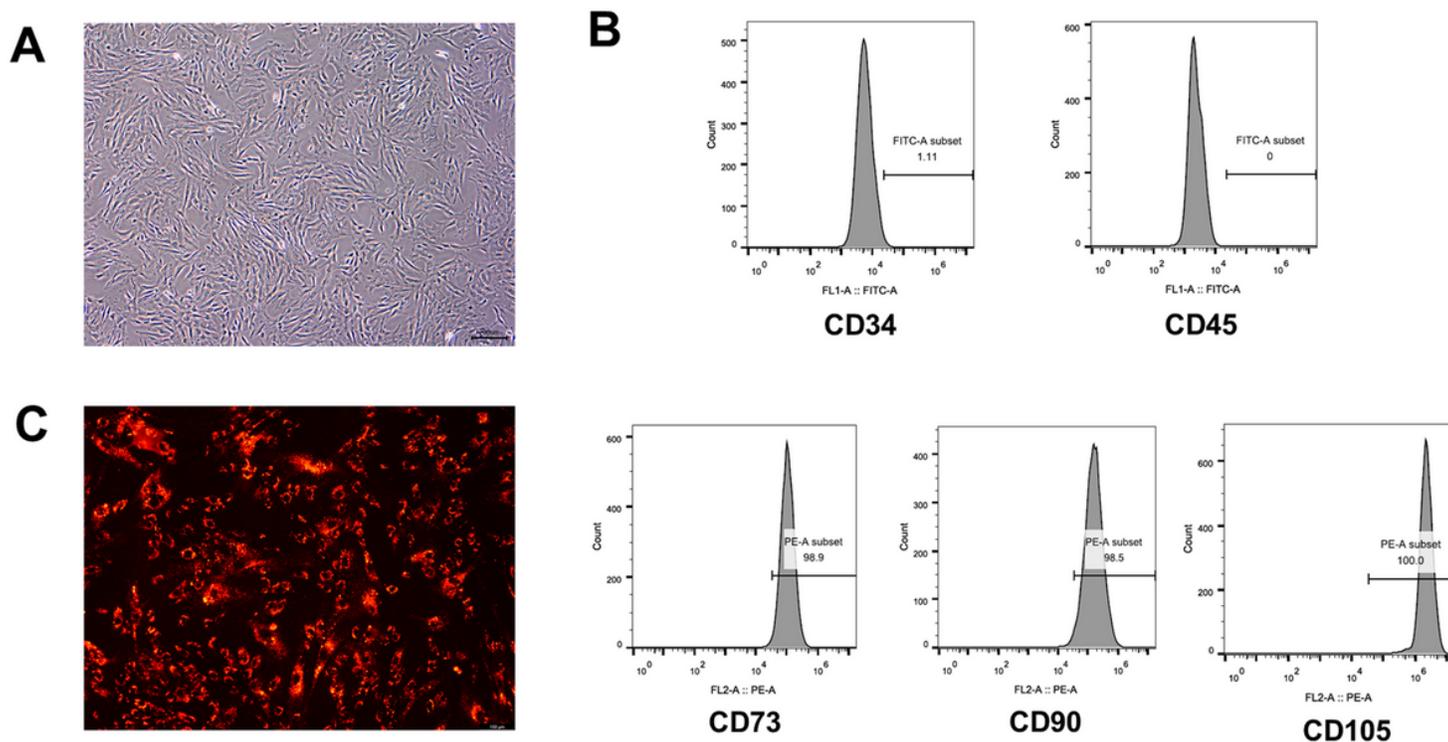
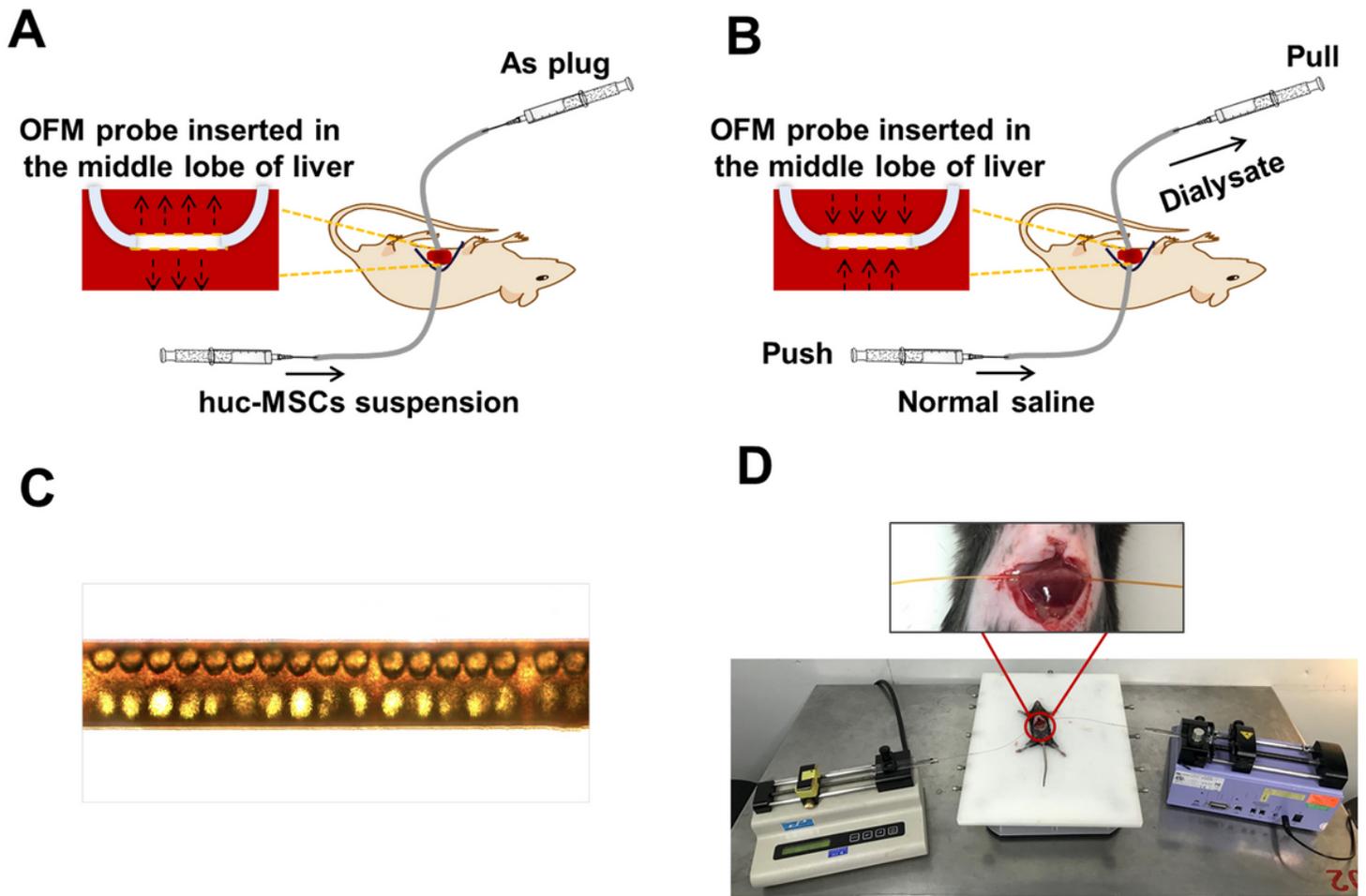


Figure 1

The characteristics of hUC-MSCs. A. Morphology of hUC-MSCs at passage 4. B. Surface markers of the isolated and cultured hUC-MSCs were detected using flow cytometry. More than 95% hUC-MSCs expressed CD73, CD90 and CD105, but not CD34 and CD45. C. Fluorescence image of CM-Dil-labeled hUC-MSCs.



**Figure 2**

The schematic diagram of OFM. A. Scheme of hUC MSCs transplantation via OFM route. B. Scheme of OFM as in vivo and in situ sampling of liver interstitial fluid. C. Microscope image of OFM probe. D. Photograph of OFM probe inserted in liver of anesthetized mice.

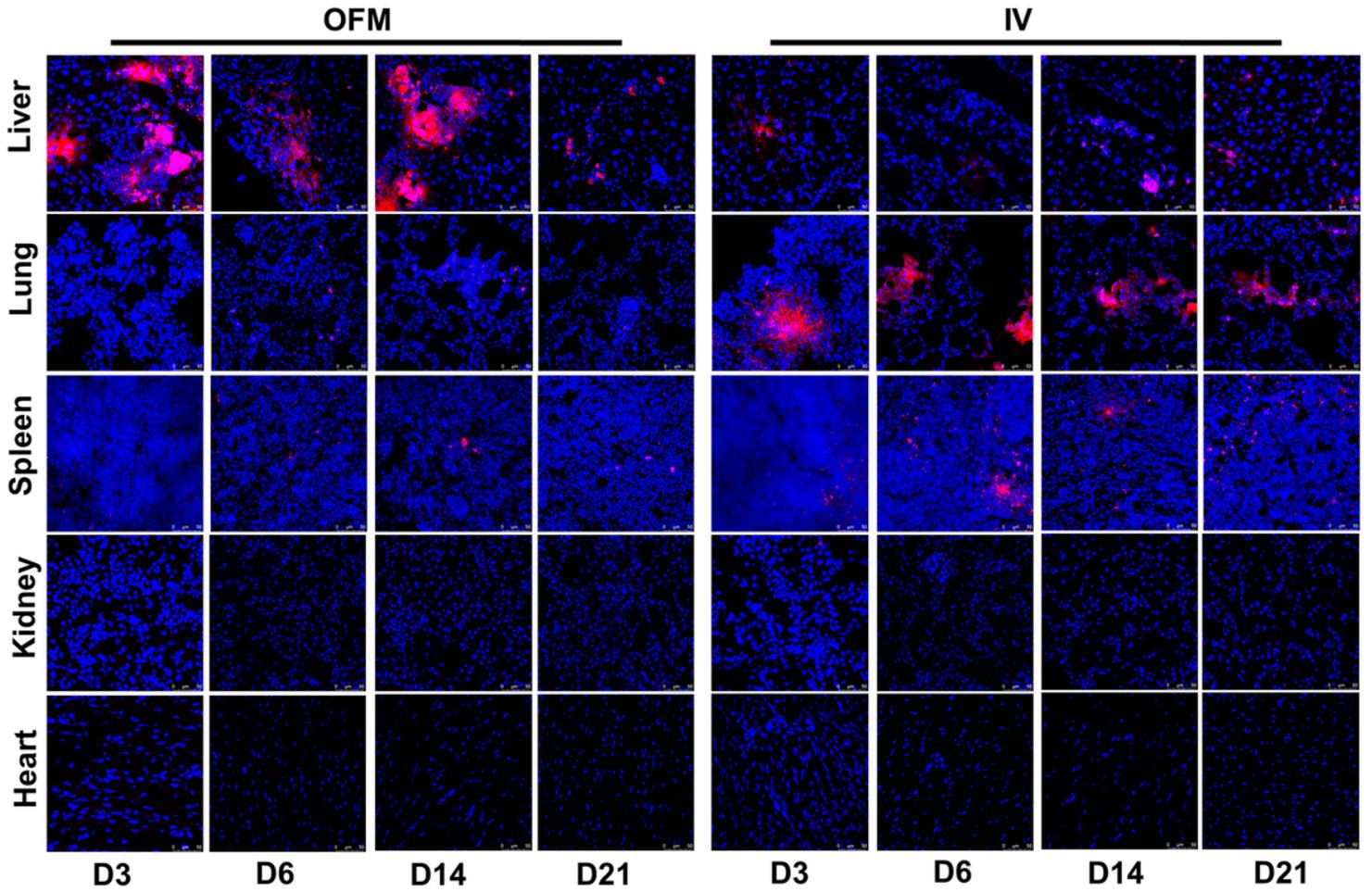
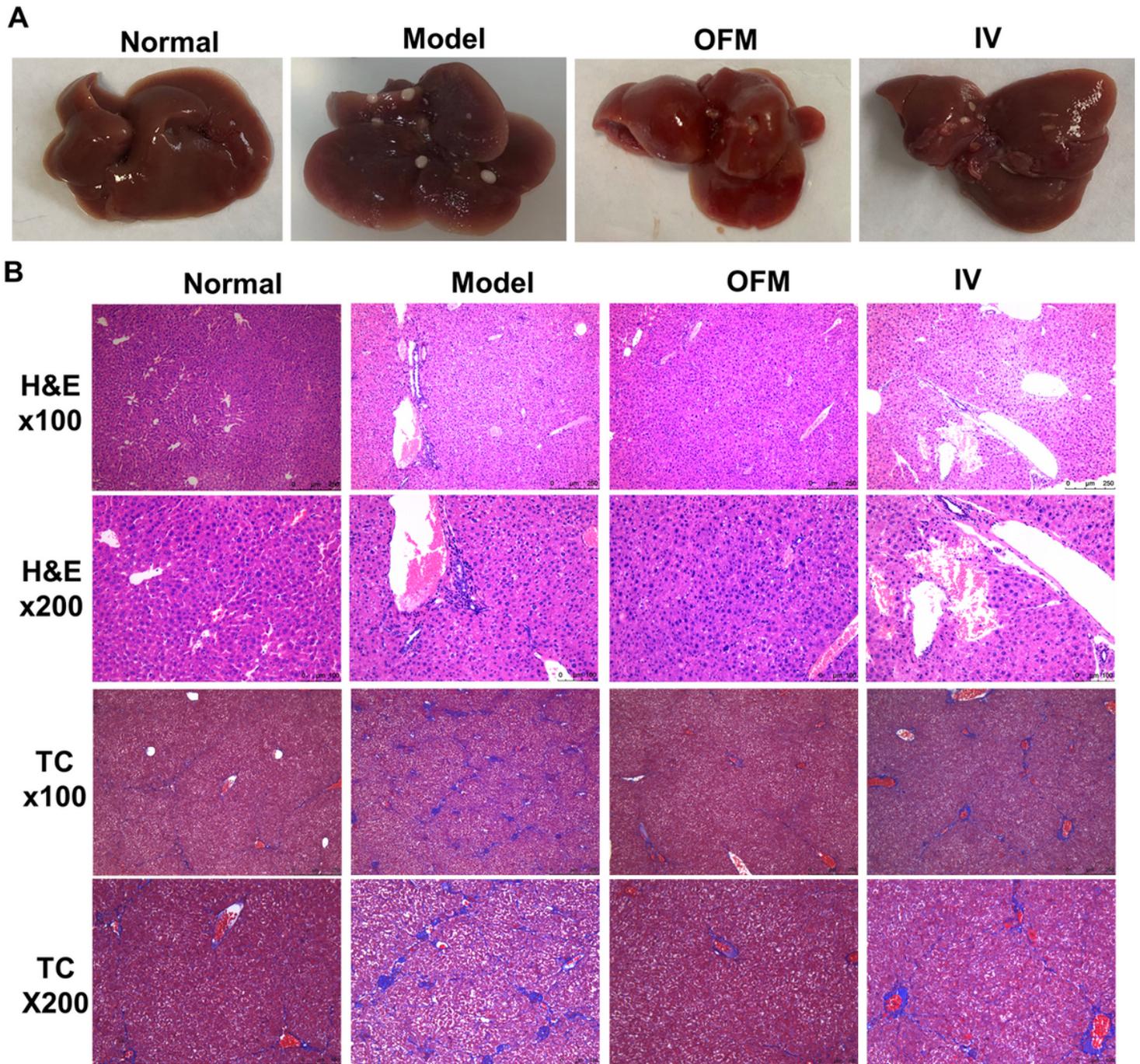


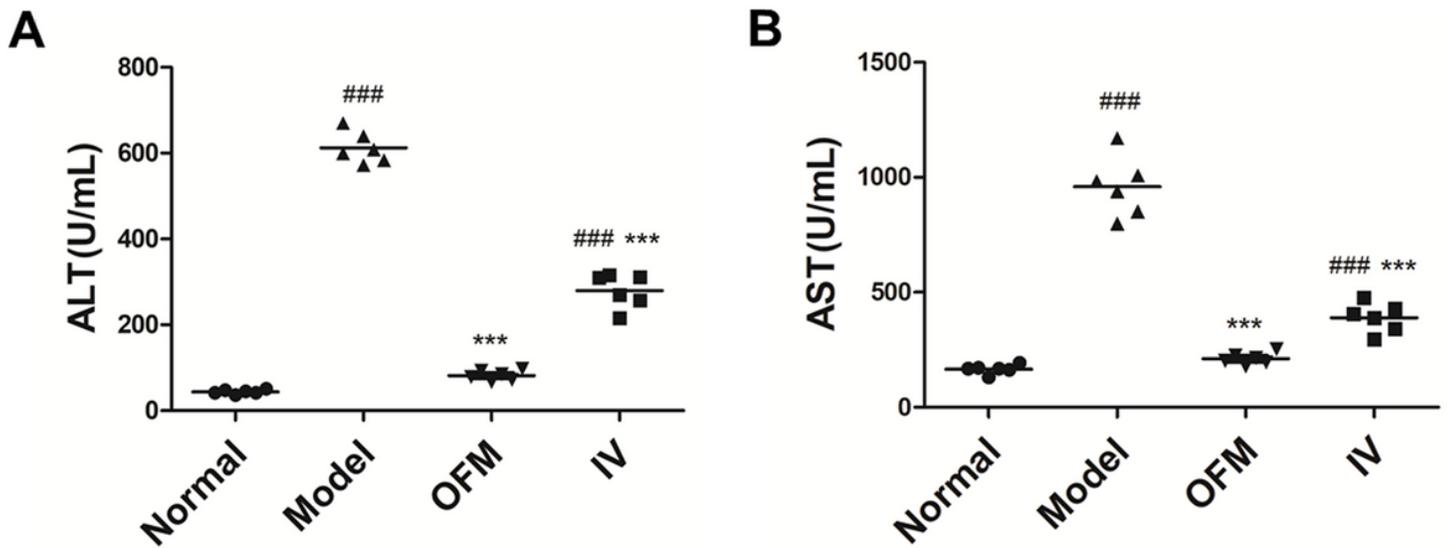
Figure 3

CM-DiI-labeled hUC-MSCs distribution analysis on liver, lung, spleen, kidney and heart at day 3, 6, 14 and 21 post transplantation via OFM and IV, respectively.



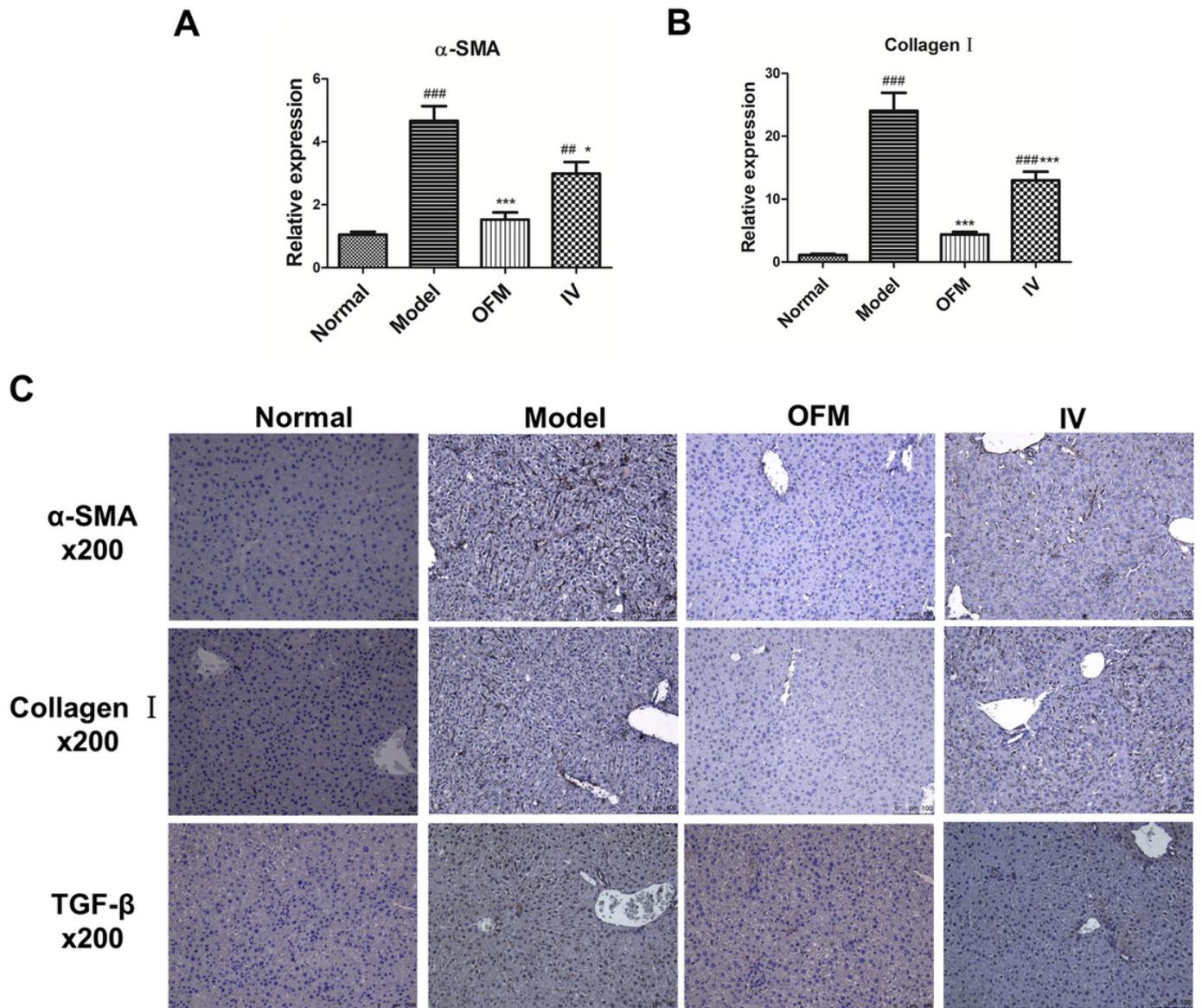
**Figure 4**

The therapeutic effects on liver fibrosis after hUC-MSCs transplantation via IV or OFM route. A. Macroscopic observation of the liver condition. B. Histopathological improvement of liver tissues at 3 weeks after hUC-MSCs transplantation was evaluated by H&E staining (100x, 200x) and masson's trichrome staining (TC, 100x, 200x).



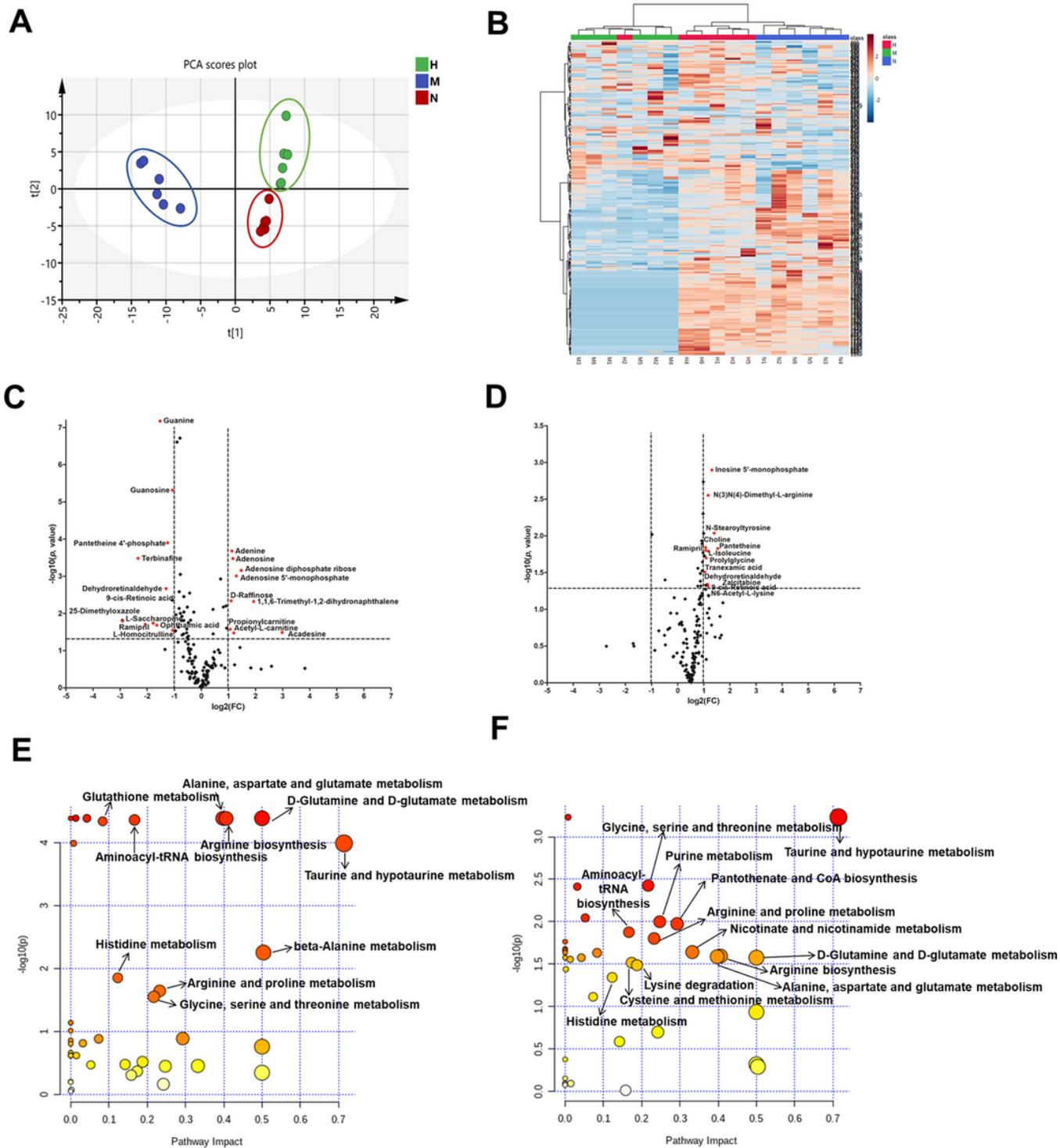
**Figure 5**

The recovery of liver function of mice with liver fibrosis after hUC-MSCs transplantation was assessed by measuring the levels of serum enzymatic biomarkers. A. ALT levels in normal group, model group, OFM group and IV group. B. AST levels in each group. ANOVA were used for statistical analysis (\*\* $p < 0.001$ , compared with model group; ### $p < 0.001$ , compared with normal group).



**Figure 6**

Alteration in fibrotic marker expressions after hUC-MSCs transplantation. A.  $\alpha$ -SMA and B. Collagen-I were measured in OFM group and IV group and then compared to the normal group and model group. (Data are represented as mean  $\pm$  SEM, ### $p$  < 0.001, ## $p$  < 0.01, compared with normal group, \*\*\* $p$  < 0.001, \* $p$  < 0.05, compared with model group;). C. Alteration in protein levels of  $\alpha$ -SMA, collagen I and TGF- $\beta$  in all groups.



**Figure 7**

Metabolomic analysis of liver interstitial fluid via OFM sampling combining mass spectrometry detection. A. PCA analysis of liver interstitial fluid in normal group (N), model group (M) and hUC-MSCs OFM-treated group (H) based on metabolomic analysis. B. Heat maps of normalized metabolite concentrations in liver dialysates. Columns represent the samples, and rows represent the metabolites. The data were already normalized by using mean-centered. C. A volcano plot based on the metabolomic data in model group vs. normal group. D. A volcano plot based on the metabolomic data in hUC-MSCs OFM-treated group vs.

model group. Metabolites with a p value  $< 0.05$  and a fold change (FC)  $> 2.0$  or  $< 0.5$  are highlighted in the volcano plot. E. Metabolic pathway analysis between normal group and model group. F. Metabolic pathway analysis between hUC-MSCs OFM-treated group and model group.