

# Distribution and Prevalence of Plasmodium Knowlesi Among Macaques In Negeri Sembilan, Malaysia

Mohd 'Ammar Ihsan Ahmad Zamzuri (✉ [rohaizat@ppukm.ukm.edu.my](mailto:rohaizat@ppukm.ukm.edu.my))

UKMMC: Pusat Perubatan Universiti Kebangsaan Malaysia <https://orcid.org/0000-0003-2381-0068>

Mohd Rohaizat Hassan

UKMMC: Pusat Perubatan Universiti Kebangsaan Malaysia

Rozita Hod

UKMMC: Pusat Perubatan Universiti Kebangsaan Malaysia

---

## Research

**Keywords:** Prevalence, Plasmodium knowlesi, zoonotic malaria, macaques

**Posted Date:** September 18th, 2020

**DOI:** <https://doi.org/10.21203/rs.3.rs-75430/v1>

**License:**  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

---

# Abstract

## Background

*Plasmodium knowlesi* infection has significant morbidity and mortality impact in Malaysia. This zoonotic malaria parasite is naturally transmissible from macaque to humans in the presence of a competent vector. Human encroachment towards the habitat of macaque has further increased the risk. Stratifying the potential risk of transmission based on the burden of parasite among macaque in a particular area can be the first step for public health intervention. Thus, the study aimed to estimate the prevalence of *Plasmodium knowlesi* parasite among macaque in Negeri Sembilan and determine its associated factors.

## METHODS

This is a cross-sectional study using a non-probabilistic sampling technique. A total of 212 blood samples from macaques in Negeri Sembilan were collected from seven districts. The *Plasmodium spp.* infecting the macaques were identified using Real-Time PCR assays on DNA extracted from these blood samples. Statistical tests were done to examine the factors associated with *Plasmodium knowlesi* infection in the macaque population.

## RESULT

The overall prevalence of *Plasmodium knowlesi* among macaque in Negeri Sembilan state was 36.3%. Both long-tailed macaque and short-tailed macaque harboured the parasite within them. Co-infection with several malaria parasites were seen in 35.5% of *Plasmodium spp* positive results. The only significant associated factors in the prevalence of *Plasmodium knowlesi* were the type of locality (rural vs. urban) and the districts where the macaques were trapped.

## CONCLUSION

The prevalence of *Plasmodium knowlesi* infection among macaques varied between districts in Negeri Sembilan. The presence of zoonotic malaria parasites among the population of macaque that live in close proximity with the community possesses a potential risk of transmission. Therefore, robust public health advocacy targeting high-risk areas is deemed necessary.

## Introduction

Malaria is a vector-borne disease that still remains a public health problem in the tropical and sub-tropical regions. A 2018 report by the World Health Organisation (WHO) estimated that around 228 million people were affected by malaria, with an estimated 405,000 mortality cases [1]. The agent responsible for the

infection is the blood parasites of the genus *Plasmodium spp.* It has been established that four species of *Plasmodium spp.* are known to infect humans, namely *Plasmodium falciparum*, *Plasmodium malariae*, *Plasmodium ovale*, and *Plasmodium vivax*. The addition of the fifth human malaria, *Plasmodium knowlesi*, only became a highlight in the last decade following notable work by Singh et. [2004] [2], despite its discovery by Knowles and Das Gupta in the laboratory way back in 1932 [3]. Since then, four other simian malaria parasites have been reported to cause malaria infection to the human population, namely *Plasmodium cynomolgi*, *Plasmodium inui*, *Plasmodium coatneyi*, and *Plasmodium fieldi*.

The natural host for *Plasmodium knowlesi* parasite to circulates are among long-tailed (*Macaca fascicularis*), pig-tailed macaques, and banded-leaf monkey (*Presbytis melalophos*) [4]. Long-tailed macaques in south-eastern Asia are broadly spread and have the third-highest regional range of all primates after human beings and rhesus macaques (*Macaca mulatta*) [5, 6]. While pig-tailed macaques are also widespread in South East Asia, morphological differences exist between those living in the northern region and those living in southern areas, namely *Macaca nemestrina* and *Macaca leonina*, respectively [7]. The majority of the malaria parasites are viewed to be host-specific, which means they can infect only one host species, though a single host can be infected by multiple *Plasmodium* species [8]. For instance, the long-tailed macaques are not only the host for *Plasmodium knowlesi* but also a natural host to the other four parasites [2, 9].

*Plasmodium knowlesi* malaria infection is considered endemic in Malaysia due to recurrent, persistent local indigenous transmission (10). This could be attributed to the dense forest and mountainous geography, which make some regions to be less accessible, especially in Borneo island that comprises the states of Sabah and Sarawak and some remote areas in peninsular Malaysia. In the year of 1965, an American traveller who returned home from peninsular Malaysia has marked as the first natural infection of *Plasmodium knowlesi* in humans [10]. To date, nearly every nation in Southeast Asia and some in Asia such as Thailand [11, 12], Singapore [13], Laos [14], Myanmar [15, 16], Philippines [17, 18], Indonesia [19–21], Vietnam [22–24] Cambodia [25], Brunei [26, 27], China [28], and India [29] have reported the presence of *Plasmodium knowlesi* infection. However, data on the actual number of *Plasmodium knowlesi* cases could still be under-reporting due to the limitation of getting an accurate diagnosis which requires more sophisticated modalities and high technology laboratory equipment.

The information regarding the burden of *Plasmodium knowlesi* and its distribution in the regional *Macaca spp* populations are scarce. The result from the available works of literature may not be applicable due to the small number of sample size for a wide coverage area [30–32]. With the increasing number of *Plasmodium knowlesi* cases and mortality cases recorded in Negeri Sembilan [33], appropriate public health measures are required. Therefore, risk stratification of the area with the highest burden of macaques harbouring *Plasmodium knowlesi* parasite is imperative to ensure targeted prevention activities can be instituted effectively. Hence the main aim of this study was to determine the prevalence of *Plasmodium knowlesi* parasites among the *Macaca spp* in Negeri Sembilan.

## Methods

## Collection of samples

A total of 254 macaque blood samples were obtained from all seven districts of Negeri Sembilan state from May to August 2018 (**Figure 1**). The procedures involved a multi-agency collaboration between Negeri Sembilan State Health Department (JKN NS), Department of Veterinary Services Negeri Sembilan (DVSNS), Department of Wildlife and National Parks Peninsular Malaysia (PERHILITAN) Negeri Sembilan, and the academic institutions.

The macaques were captured by the PERHILITAN officers following complaints made by the community about their presence and disturbance. There were approximately double digits of complaints letter being made regarding macaque disturbance in Negeri Sembilan for the year 2017 and 2018, especially in the town closed to the forest fringe and a newly developed township. The trapping sites and items used were in accordance with the standard guideline set by PERHILITAN. An average of 1 or 2 troops of macaques is usually found at a territory. The sites were revisited within 24 hours for the collection of the macaques that were trapped. Upon collection, the health and veterinary teams were informed to assemble at the PERHILITAN offices, where the macaques will be brought back.

The blood collection procedure was done by DVS officers. The macaques were tranquilised and anaesthetised intramuscularly with Ketamine (5 mg/kg body weight) before the blood samples were collected. A maximum amount of 3mls per animal of blood samples were collected using a syringe from the femoral vein of the macaques and kept into a tube with ethylenediamine- tetraacetic acid (EDTA). Three blood spots from each sample of EDTA tubes were transferred (40–50 µL each) to Whatman 3 MM filter papers in situ. The samples were kept at room temperature (20–29 °C) until they reached the Negeri Sembilan Vector laboratory on the same day. They were subsequently held at 4 °C before they were transported to the National Public Health Laboratory (MKAK) Sungai Buloh every twice a week. The macaque will be put under the care and supervision of PERHILITAN once the blood collection is completed, as per the standard operation procedure of handling complaints.

## Analysis of the Samples

DNA was extracted from blood spots in filter papers at the MKAK laboratory, Sungai Buloh. The sample was examined by using Real Time-Polymerase Chain Reaction assays with the use of genus and species-specific primers based on AITbiotech ab TESä Malaria qPCR I Kit. Positive samples were identified as one of these five malaria parasite species, namely *Plasmodium knowlesi*, *Plasmodium falciparum*, *Plasmodium vivax*, *Plasmodium ovale*, and *Plasmodium malariae*. All the samples were run using a CFX96 Touch Real-Time PCR Detection System.

## Statistical Analysis and Prevalence Estimation of Malaria Parasites

To determine whether the prevalence of *Plasmodium knowlesi*, the proportion of the positive sample with a positive RT-PCR for *Plasmodium knowlesi* were counted regardless of mono-infection or poly-infection.

Statistical analysis was done to look for the factor associated with the sample of macaques by using R software (version 3.2.4). The cut-off point for rejecting the null hypothesis was set at 0.05.

## Ethical Approval

The study was registered with NMRR (Ministry of Health) and has been approved by the Department of Wildlife and National Parks Peninsular Malaysia (PERHILITAN) with the reference letter **JPHL&TN(IP): 100-34t1.24 Jtd 12 [31]**.

## Results

In total, there were 207 number of long-tailed macaques and 5 short-tailed macaques trapped during the operation. **Table 1** described the positive RT-PCR findings for all the *Plasmodium spp.* Four species of *Plasmodium* parasites were detected; *P. knowlesi*, *P. vivax*, *P. ovale* and *P.malariae* (147/212). Of the 212 blood samples investigated, 36.3% were positive for any *Plasmodium knowlesi* infection (77/212). Macaque with mono *Plasmodium knowlesi* infection has slightly higher proportion (51.9%) than macaques with co-infection (48.1%). Among co-infection positive samples, 94.6% of them were due to *P. knowlesi* and *Plasmodium vivax*.

The prevalence of *Plasmodium knowlesi* infection among all the macaques varied by districts. District of Jelebu recorded 71.4% of macaque with positive *Plasmodium knowlesi* infection while district of Port Dickson showed zero prevalence. By using Fisher–Freeman– Halton exact test (**Table 2**), we obtained a significant association [p-value < 0.0001]. Thus, indicating that the observed districts distribution of *Plasmodium knowlesi* infection is extremely unlikely under the null hypothesis; i.e., the prevalence of *Plasmodium knowlesi* exhibits bias among different district areas.

Male macaque constituted 58.4% of the total infected macaque, although the odds being insignificant (p-value 0.588). Macaque captured in rural area constitute 92.2% from the positive samples, and thus translated into prevalence odds ratio (pOR) of 3.51 (95% CI 1.63 - 7.57, p-value < 0.0001). Although 60% of short-tailed macaque were found to be positive with *Plasmodium knowlesi* parasite in the blood, the pOR remained insignificant (p-value 0.356).

**Table 1:** Distribution of *Plasmodium spp* in macaques detected using RT-PCR multiplex.

Infection	Type of <i>Plasmodium spp</i>	Count
Mono-infection	<i>P. knowlesi</i>	40
	<i>P. vivax</i>	26
	<i>P. ovale</i>	2
	<i>P. malariae</i>	1
Co-infection	<i>P. knowlesi + P. vivax</i>	35
	<i>P. knowlesi + P.malariae</i>	1
	<i>P. knowlesi + P.ovale</i>	1
	<i>P. vivax + P. malariae</i>	1

**Table 2:** Descriptive analysis of 212 macaque blood samples.

Bil	Variables	Positive <i>P. knowlesi</i>		$\chi^2$	p-value
		Yes	No		
1	Gender			0.294	0.588
	Male	45	84		
	Female	32	51		
2	District			23.484	< 0.0001*
	Jelevu	5	2		
	Jempol	6	14		
	Kuala Pilah	14	20		
	Port Dickson	0	7		
	Rembau	14	9		
	Seremban	6	35		
	Tampin	32	48		
3	Locality type			15.223	< 0.0001
	Rural	71	93		
	Urban	6	42		
4	Types of Macaque				0.356*
	Long tailed	74	133		
	Short-tailed	3	2		

\* Fisher–Freeman– Halton exact test

## Discussion

*Plasmodium knowlesi* has been identified as the fifth human malaria, although there is still little proof of human-to-human transmission. It does, however, make a significant contribution to our attempt to eradicate malaria in this country. Nonetheless, infection with *Plasmodium knowlesi* caused considerable morbidity and raised a higher risk of death [33]. Identifying the burden of *Plasmodium knowlesi* in macaque is one of the steps to stratify the risk of potential transmission of the parasite, given that the vector also presents in some part of the country (34,35). Challenge remains as there are lot more parasite strains infecting macaque as compared to human malaria infection, making it almost difficult to conduct such a study. Nevertheless, recent literature has able to demonstrate the potential for other zoonotic parasites infecting humans besides *Plasmodium knowlesi* [10,34–36].

This study employed RT-PCR, which has been shown to be more sensitive and more specific in detecting parasite species as compared to the other modalities such as conventional PCR and nested PCR [37]. This method was necessary to overcome the problem of the false-positive result, that could make data less accurate and disputable. At the same time, the gold standard procedure of using blood film malaria parasite [BFMP] for diagnosing parasite species in the human blood sample was not utilised in this study due to the risk of a false negative result from morphological resemblance with other parasite species; namely *Plasmodium falciparum* and *Plasmodium malariae* [38].

Besides, as is apparent from the results, the state of multiple malaria parasites co-infection has further justified the use of RT-PCR over BFMP. This is because BFMP is very much operator dependent and requires a great deal of patience and perseverance, especially when dealing with blood samples with a low-density level of parasitaemia. Therefore, multiple co-infection is often more complicated to cope with and may potentially be overlooked. At the moment, there is scarce evidence to ascertain the transmissibility potential of a macaque who have been co-infected with zoonotic parasites, for example like co-infection by *Plasmodium knowlesi* and *Plasmodium cynomolgi*. This may be attributed to the need for a particular vector to achieve a specific parasite's cyclo-propagative transmission before it can start to transmit [39]. While a recent study identified the existence of several malaria parasites within the vector thorax and abdomen, no transmission danger was recorded [40].

The present study demonstrated that the prevalence of *Plasmodium knowlesi* varied between the districts. We found one common element of these impacted areas is experiencing a new developmental project involving deforestation and land clearing in particular. These areas are undergoing rapid change in the landscape to accommodate population growth and, at the same time are driven by a slower rate of expansion in the main district due to overcrowding [such as Seremban and Port Dickson]. The massive land clearing and deforestation either for the purpose of agriculture or human settlement have been identified as a significant factor to disrupt the habitat of macaque, thus explained their constant disturbance, which could potentially increase the risk of *Plasmodium knowlesi* infection [41].

Apart from that, we also found that there was a significant difference between macaque trapped in rural areas than the urban area. While Seremban and Port Dickson are considered to be the major urban cities with strong economic growth, suburbs or close forest fringe population settlements are usually regarded as rural areas for those districts. Hence, they are more prone to have a constant macaque disturbance either in the neighbourhood or the plantation. This could possibly explain the reason for a higher proportion of macaques were captured in rural areas (77.4%). Nevertheless, the competent vector for *Plasmodium knowlesi* which is *Anopheles leucospyrus* that is associated with forest fringe and dense forest might elucidate their higher risk to carry the parasite [42]. Such variation of *Plasmodium knowlesi* prevalence among macaques between rural and urban areas has also been reported in the literature [8].

Both *Macaca fascicularis* [long-tailed] and *Macaca nemestrina* [pig-tailed] could be infected by *Plasmodium knowlesi*, although the proportion was different due to sampling bias. Such bias occurred due to the segregation within the natural habitats and the attribute of *Macaca fascicularis* as a tree-

traveler while *Macaca nemestrina* travels along the ground [43]. Therefore, the PERHILITAN team faced difficulty in trapping pig-tailed macaque at the field which resulted in a significantly disproportionate number. However, our prevalence of the parasite among pig-tailed macaque remains comparable with the previously reported literature [44]. On the other side, the long-tailed macaque is an edge species [7], capable of existing in a wide variety of environments, and can quickly adapt to the new environment [45]. Hence this could be the potential reason for many studies conducted on long-tailed macaques either locally or in other regions of the world [8,31,46–49].

There is two main strength of this study, first is the number of samples which were high enough to represent the small state such as Negeri Sembilan. To the best of our knowledge, this study is the first to describe in detail the prevalence of *Plasmodium knowlesi* among macaque holistically for a single state. The second strength of this study is the collaboration received from multi agencies who are experts in their respective fields. Each of the team players provided highly skilled and specialised staffs to ensure the results obtained are accurate and have internal validity. At the same time, we humbly acknowledge the paper's main limitation, which did not integrate the entomological study. The absence of competent vector analysis which transmitted *Plasmodium knowlesi* may paint a distorted image of the potential danger. As it is the key player that connects the zoonotic infection to humans, the inclusion of its distribution and prevalence will help to stratify the potential risk of the disease better. Thus, the future study can perhaps incorporate the epidemiology of the competent vector to generate a more significant outcome.

## Conclusion

The prevalence of *Plasmodium knowlesi* parasites that infect macaques, including those transmissible to humans, differs between wild macaque populations in Negeri Sembilan. Nevertheless, co-infection with several malaria parasites was confirmed using RT-PCR assay. The presence of malaria parasites among macaque living within the vicinity of the neighbourhood emphasises the potential risk of zoonotic infections to the community.

## Abbreviations

PERHILITAN – Department of Wildlife and National Parks Peninsular Malaysia

DVS – Department of Veterinary Service

JKN NS – Negeri Sembilan State Health Department

MKAK - National Public Health Laboratory

RT – PCR – Real Time Polymerase Chain Reaction

BFMP – Blood Film for Malaria Parasite

EDTA - Ethylenediamine-Tetraacetic Acid

pOR – Prevalence Odd Ratio

## **Declarations**

### **Acknowledgement**

We would like to thank the Director General of Health of Malaysia for the approval for publication of this work. Our heartfelt gratitude towards all the staffs from Negeri Sembilan State Health Department, Negeri Sembilan Department of Veterinary Services Malaysia (DVS), and Negeri Sembilan branch of Department of Wildlife And National Parks (PERHILITAN) who have contributed a lot in the study. We also would like to take the opportunity to acknowledge the significant contribution from all laboratory officers from Negeri Sembilan Vector Unit and National Public Health Laboratory (MKAK Sungai Buloh). We dedicated a special thank you to Dr Nur Azlina Zabani [the former Negeri Sembilan DVS veterinary officer] for her significant contribution in blood sampling procedures.

### **Competing interests**

The authors have declared that no competing interests exist.

### **Financial Funding**

The authors received no specific funding for this work

### **List of Abbreviation**

PERHILITAN – Department of Wildlife and National Parks Peninsular Malaysia

DVS – Department of Veterinary Service

JKN NS – Negeri Sembilan State Health Department

MKAK - National Public Health Laboratory

RT – PCR – Real Time Polymerase Chain Reaction

BFMP – Blood Film for Malaria Parasite

EDTA - Ethylenediamine-Tetraacetic Acid

pOR – Prevalence Odd Ratio

### **Authors' Contribution**

MAI, MFMY, and WMWH conducted the field work.

MAI, NDMD, RD, LR, MFAR conceptualised the study and participate in study design.

MAI, RD and WMWH worked on securing the permits and clearance from PERHILITAN

RD, RNM, ZH and HMH provide support and assistant to develop protocol and networking.

ZH and HMH conducted the molecular analysis of the blood samples in MKAK.

MAI, RH, and MRH interpret the data and wrote the paper.

All authors read and approved the final manuscript.

### **Consent for publication**

All authors have seen and approved the manuscript and its contents thus agreed to submit for publication.

### **Ethics Approval and Consent to Participate**

Approval for this study was provided by the Department of Wildlife and National Parks Peninsular Malaysia [PERHILITAN] with the reference letter **JPHL&TN[IP]: 100-34t1.24 Jtd 12 [31]**.

### **Availability of data and materials**

All relevant data are within the paper..

## **References**

1. The “World malaria report 2019” at a glance [Internet]. [cited 2020 Sep 7]. Available from: <https://www.who.int/news-room/feature-stories/detail/world-malaria-report-2019>
2. Singh B, Sung LK, Matusop A, Radhakrishnan A, Shamsul SSG, Cox-Singh J, et al. A large focus of naturally acquired Plasmodium knowlesi infections in human beings. *Lancet* [Internet]. 2004 Mar 27 [cited 2020 Sep 7];363(9414):1017–24. Available from: <https://pubmed.ncbi.nlm.nih.gov/15051281/>
3. Knowles R, Gupta BM Das. A Study of Monkey-Malaria, and Its Experimental Transmission to Man. *Ind Med Gaz* [Internet]. 1932 Jun [cited 2020 Sep 7];67(6):301–20. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/29010910>
4. Vythilingam I, Hii J. Simian Malaria Parasites: Special Emphasis on Plasmodium knowlesi and Their Anopheles Vectors in Southeast Asia. In: *Anopheles mosquitoes - New insights into malaria vectors* [Internet]. InTech; 2013 [cited 2020 Sep 8]. Available from: <http://dx.doi.org/10.5772/54491>
5. Fooden J. Systematic Review of Southeast Asian Longtail Macaques, *Macaca fascicularis* (Raffles, [1821]). *Syst Rev Southeast Asian longtail macaques, Macaca Fasc* (Raffles, [1821]) [Internet]. 1985 [cited 2020 Sep 8];81:1–206. Available from: <https://www.biodiversitylibrary.org/part/76074>

6. Eudey AA. The Crab-Eating Macaque ( *Macaca fascicularis* ): Widespread and Rapidly Declining . *Primate Conserv*. 2008 Nov;23(1):129–32.
7. Gumert MD, Kluck M, Malaivijitnond S. The physical characteristics and usage patterns of stone axe and pounding hammers used by long-tailed macaques in the Andaman sea region of Thailand. *Am J Primatol* [Internet]. 2009 Jul [cited 2020 Sep 7];71(7):594–608. Available from: <https://pubmed.ncbi.nlm.nih.gov/19405083/>
8. Gamalo LE, Dimalibot J, Kadir KA, Singh B, Paller VG. *Plasmodium knowlesi* and other malaria parasites in long-tailed macaques from the Philippines. *Malar J* [Internet]. 2019;18(1):1–7. Available from: <https://doi.org/10.1186/s12936-019-2780-4>
9. Jeslyn WPS, Huat TC, Vernon L, Irene LMZ, Sung LK, Jarrod LP, et al. Molecular epidemiological investigation of *Plasmodium knowlesi* in humans and macaques in Singapore. *Vector-Borne Zoonotic Dis* [Internet]. 2011 Feb 1 [cited 2020 Sep 8];11(2):131–5. Available from: </pmc/articles/PMC3033207/?report=abstract>
10. Hussin N, Lim YAL, Goh PP, William T, Jelip J, Mudin RN. Updates on malaria incidence and profile in Malaysia from 2013 to 2017. *Malar J* [Internet]. 2020 Jan 31 [cited 2020 Sep 8];19(1):55. Available from: </pmc/articles/PMC6995112/?report=abstract>
11. Chin W, Contacos PG, Coatney GR, Kimball HR. A naturally acquired quotidian-type malaria in man transferable to monkeys. *Science* (80- ) [Internet]. 1965 [cited 2020 Sep 8];149(3686):865. Available from: <https://pubmed.ncbi.nlm.nih.gov/14332847/>
12. Saritapirak N, Sookkasem K, Lekcharoen P, Kanjanasombut H, Buathong R. Emergence of *Plasmodium knowlesi* at Thai-Myanmar border , Ratchaburi , Thailand ( March - September 2018 ). 2018;(September):2018–9.
13. Jongwutiwes S, Putaporntip C, Iwasaki T, Sata T, Kanbara H. Naturally acquired *Plasmodium knowlesi* malaria in human, Thailand. *Emerg Infect Dis* [Internet]. 2004 [cited 2020 Sep 8];10(12):2211–3. Available from: <https://pubmed.ncbi.nlm.nih.gov/15663864/>
14. Ng OT, Eng EO, Cheng CL, Piao JL, Lee CN, Pei SW, et al. Naturally acquired human *Plasmodium knowlesi* infection, Singapore. *Emerg Infect Dis* [Internet]. 2008 [cited 2020 Sep 8];14(5):814–6. Available from: <https://pubmed.ncbi.nlm.nih.gov/18439370/>
15. Iwagami M, Nakatsu M, Khattignavong P, Soundala P, Lorphachan L, Keomalaphet S, et al. First case of human infection with *Plasmodium knowlesi* in Laos. Diemert DJ, editor. *PLoS Negl Trop Dis* [Internet]. 2018 Mar 22 [cited 2020 Sep 8];12(3):e0006244. Available from: <https://dx.plos.org/10.1371/journal.pntd.0006244>
16. Jiang N, Chang Q, Sun X, Lu H, Yin J, Zhang Z, et al. Co-infections with *Plasmodium knowlesi* and other Malaria parasites, Myanmar. *Emerg Infect Dis* [Internet]. 2010 [cited 2020 Sep 8];16(9):1476–8. Available from: <https://pubmed.ncbi.nlm.nih.gov/20735938/>
17. Ghinai I, Cook J, Hla TTW, Htet HMT, Hall T, Lubis IN, et al. Malaria epidemiology in central Myanmar: identification of a multi-species asymptomatic reservoir of infection. *Malar J* [Internet]. 2017 Jan 5

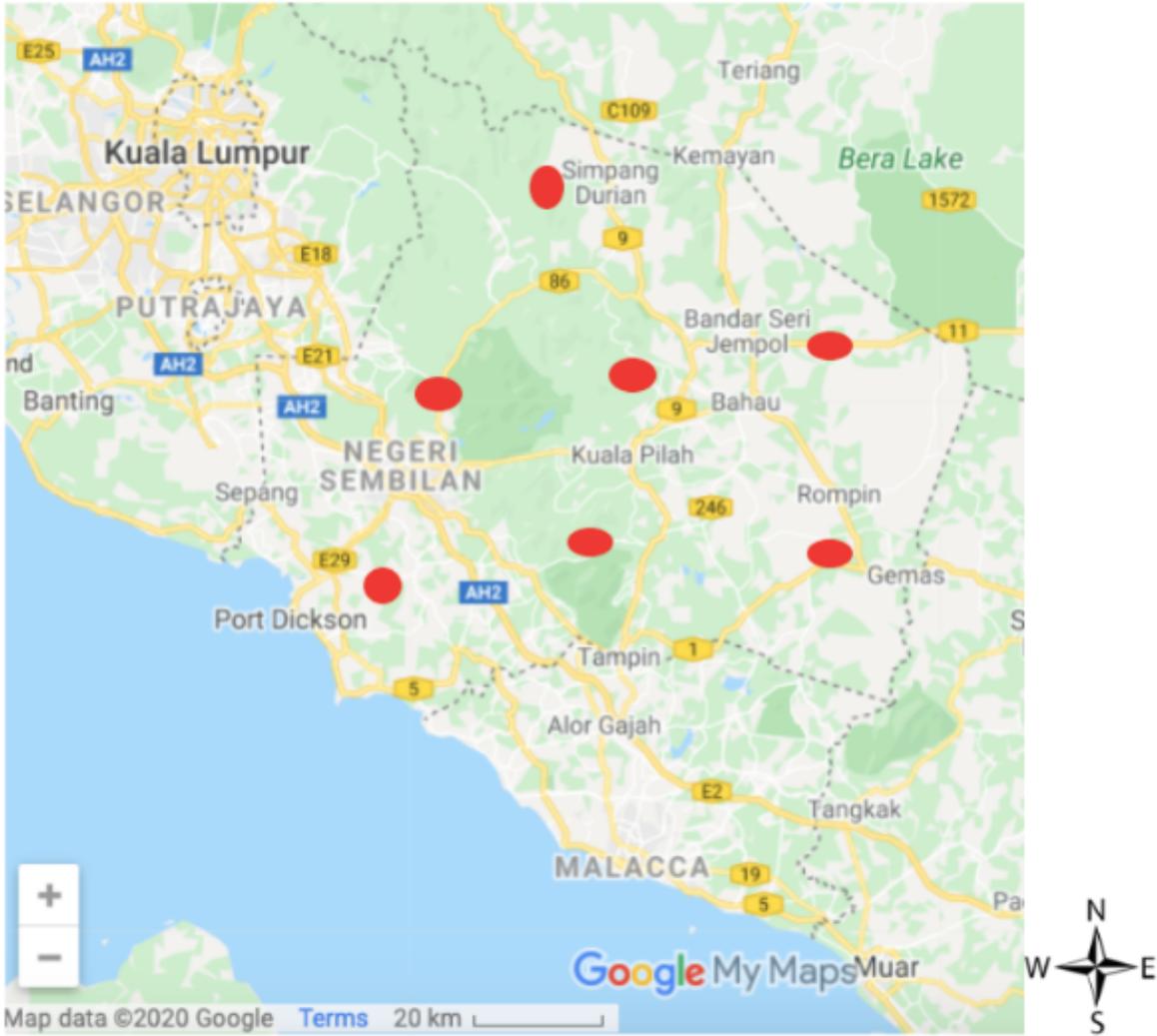
- [cited 2020 Sep 8];16(1):1–10. Available from:  
<http://malariajournal.biomedcentral.com/articles/10.1186/s12936-016-1651-5>
18. Luchavez J, Espino F, Curameng P, Espina R, Bell D, Chiodini P, et al. Human infections with *Plasmodium knowlesi*, the Philippines. *Emerg Infect Dis*. 2008;14(5):811–3.
  19. Fornace KM, Herman LS, Abidin TR, Chua TH, Daim S, Lorenzo PJ, et al. Exposure and infection to *Plasmodium knowlesi* in case study communities in Northern Sabah, Malaysia and Palawan, The Philippines. Barry AE, editor. *PLoS Negl Trop Dis* [Internet]. 2018 Jun 14 [cited 2020 Sep 8];12(6):e0006432. Available from: <https://dx.plos.org/10.1371/journal.pntd.0006432>
  20. Setiadi W, Sudoyo H, Trimarsanto H, Sihite BA, Saragih RJ, Juliawaty R, et al. A zoonotic human infection with simian malaria, *Plasmodium knowlesi*, in Central Kalimantan, Indonesia. *Malar J* [Internet]. 2016 Apr 16 [cited 2020 Sep 8];15(1):218. Available from:  
<http://malariajournal.biomedcentral.com/articles/10.1186/s12936-016-1272-z>
  21. Herdiana H, Iriawati I, Coutrier FN, Munthe A, Mardiaty M, Yuniarti T, et al. Two clusters of *Plasmodium knowlesi* cases in a malaria elimination area, Sabang Municipality, Aceh, Indonesia. *Malar J* [Internet]. 2018 May 2 [cited 2020 Sep 8];17(1):186. Available from:  
<https://malariajournal.biomedcentral.com/articles/10.1186/s12936-018-2334-1>
  22. Coutrier FN, Tirta YK, Cotter C, Zarlinda I, González IJ, Schwartz A, et al. Laboratory challenges of *Plasmodium* species identification in Aceh Province, Indonesia, a malaria elimination setting with newly discovered *P. knowlesi*. Barry AE, editor. *PLoS Negl Trop Dis* [Internet]. 2018 Nov 30 [cited 2020 Sep 8];12(11):e0006924. Available from: <https://dx.plos.org/10.1371/journal.pntd.0006924>
  23. Maeno Y, Culleton R, Quang NT, Kawai S, Marchand RP, Nakazawa S. *Plasmodium knowlesi* and human malaria parasites in Khan Phu, Vietnam: Gametocyte production in humans and frequent co-infection of mosquitoes. *Parasitology*. 2017 Apr 1;144(4):527–35.
  24. Pongvongsa T, Culleton R, Ha H, Thanh L, Phongmany P, Marchand RP, et al. Human infection with *Plasmodium knowlesi* on the Laos-Vietnam border. *Trop Med Health* [Internet]. 2018 Sep 18 [cited 2020 Sep 8];46(1):33. Available from:  
<https://tropmedhealth.biomedcentral.com/articles/10.1186/s41182-018-0116-7>
  25. Eede P Van Den, Van HN, Van Overmeir C, Vythilingam I, Duc TN, Hung LX, et al. Human *Plasmodium knowlesi* infections in young children in central Vietnam. *Malar J* [Internet]. 2009 Dec 30 [cited 2020 Sep 8];8(1):249. Available from: <https://malariajournal.biomedcentral.com/articles/10.1186/1475-2875-8-249>
  26. Khim N, Siv S, Kim S, Mueller T, Fleischmann E, Singh B, et al. *Plasmodium knowlesi* infection in humans, Cambodia, 2007-2010. *Emerg Infect Dis* [Internet]. 2011 [cited 2020 Sep 8];17(10):1900–2. Available from: [/pmc/articles/PMC3310675/?report=abstract](http://pmc/articles/PMC3310675/?report=abstract)
  27. Ninan T, Nalees K, Newin M, Sultan Q, Than MM, Shinde S, et al. *Plasmodium knowlesi* malaria infection in human.
  28. Koh GJ, Ismail PK, Koh D. Occupationally Acquired *Plasmodium knowlesi* Malaria in Brunei Darussalam. *Saf Health Work* [Internet]. 2019 Mar 1 [cited 2020 Sep 8];10(1):122–4. Available from:

/pmc/articles/PMC6429035/?report=abstract

29. Zhu H min, Li J, Zheng H. Human natural infection of *Plasmodium knowlesi*. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi* [Internet]. 2006 [cited 2020 Sep 8];24(1):70–1. Available from: <https://pubmed.ncbi.nlm.nih.gov/16866152/>
30. Tyagi RK, Das MK, Singh SS, Sharma YD. Discordance in drug resistance-associated mutation patterns in marker genes of *Plasmodium falciparum* and *Plasmodium knowlesi* during coinfections. *J Antimicrob Chemother* [Internet]. 2013 May 1;68(5):1081–8. Available from: <http://dx.doi.org/10.1093/jac/dks508>
31. Akter R, Vythilingam I, Khaw LT, Qvist R, Lim YA-L, Sitam FT, et al. Simian malaria in wild macaques: first report from Hulu Selangor district, Selangor, Malaysia. 2015 Dec 13;14(1):386. Available from: <https://app.dimensions.ai/details/publication/pub.1045175907>
32. Zhang X, Kadir KA, Quintanilla-Zariñan LF, Villano J, Houghton P, Du H, et al. Distribution and prevalence of malaria parasites among long-tailed macaques (*Macaca fascicularis*) in regional populations across Southeast Asia. *Malar J*. 2016;15(1).
33. Lee K-S, Divis PCS, Zakaria SK, Matusop A, Julin RA, Conway DJ, et al. *Plasmodium knowlesi*: Reservoir Hosts and Tracking the Emergence in Humans and Macaques. Kazura JW, editor. *PLoS Pathog* [Internet]. 2011 Apr 7 [cited 2020 Sep 8];7(4):e1002015. Available from: <https://dx.plos.org/10.1371/journal.ppat.1002015>
34. Hawkes FM, Manin BO, Cooper A, Daim S, Homathevi R, Jelip J, et al. Vector compositions change across forested to deforested ecotones in emerging areas of zoonotic malaria transmission in Malaysia. *Sci Rep*. 2019 Dec 1;9(1).
35. Chua TH, Manin BO, Vythilingam I, Fornace K, Drakeley CJ. Effect of different habitat types on abundance and biting times of *Anopheles balabacensis* Baisas (Diptera: Culicidae) in Kudat district of Sabah, Malaysia. *Parasites and Vectors* [Internet]. 2019 Jul 25 [cited 2020 Sep 10];12(1):364. Available from: <https://parasitesandvectors.biomedcentral.com/articles/10.1186/s13071-019-3627-0>
36. Grignard L, Shah S, Chua TH, William T, Drakeley CJ, Fornace KM. Natural Human Infections With *Plasmodium cynomolgi* and Other Malaria Species in an Elimination Setting in Sabah, Malaysia. *J Infect Dis* [Internet]. 2019 Nov 6 [cited 2020 Sep 8];220(12):1946–9. Available from: <https://academic.oup.com/jid/article/220/12/1946/5550405>
37. Ta TH, Hisam S, Lanza M, Jiram AI, Ismail N, Rubio JM. First case of a naturally acquired human infection with *Plasmodium cynomolgi*. *Malar J* [Internet]. 2014 Feb 24 [cited 2020 Sep 8];13(1):68. Available from: /pmc/articles/PMC3937822/?report=abstract
38. Raja TN, Hu TH, Kadir KA, Mohamad DSA, Rosli N, Wong LL, et al. Naturally Acquired Human *Plasmodium cynomolgi* and *P. knowlesi* Infections, Malaysian Borneo. *Emerg Infect Dis* [Internet]. 2020 Aug 1 [cited 2020 Sep 8];26(8):1801–9. Available from: <https://doi.org/10.3201/eid2608.200343>

39. Thura Zaw M, Lin Z. Methods for Detection and Identification of Plasmodium knowlesi: A Review Article. Vol. 6, International Journal of Collaborative Research on Internal Medicine & Public Health. 2014.
40. Silva JR De. Plasmodium knowlesi malaria : current research perspectives. 2018;1145–55.
41. Lane RS. Zoonotic Agents, Arthropod-Borne. In: Encyclopedia of Insects. Elsevier Inc.; 2009. p. 1065–8.
42. Chinh VD, Masuda G, Hung VV, Takagi H, Kawai S, Annoura T, et al. Prevalence of human and non-human primate Plasmodium parasites in anopheline mosquitoes: a cross-sectional epidemiological study in Southern Vietnam. Trop Med Health [Internet]. 2019 Dec 23 [cited 2020 Sep 7];47(1):9. Available from: <https://tropmedhealth.biomedcentral.com/articles/10.1186/s41182-019-0139-8>
43. Fornace KM, Abidin TR, Alexander N, Brock P, Grigg MJ, Murphy A, et al. Association between landscape factors and spatial patterns of Plasmodium knowlesi infections in Sabah, Malaysia. Emerg Infect Dis [Internet]. 2016 Feb 1 [cited 2020 Sep 7];22(2):201–8. Available from: </pmc/articles/PMC4734530/?report=abstract>
44. Sallum MAM, Peyton EL, Wilkerson RC. Six new species of the Anopheles leucosphyrus group, reinterpretation of An. elegans and vector implications. Med Vet Entomol [Internet]. 2005 Jun [cited 2020 Sep 9];19(2):158–99. Available from: <https://pubmed.ncbi.nlm.nih.gov/15958025/>
45. Davidson G, Chua TH, Cook A, Speldewinde P, Weinstein P. Defining the ecological and evolutionary drivers of Plasmodium knowlesi transmission within a multi-scale framework [Internet]. Vol. 18, Malaria Journal. BioMed Central Ltd.; 2019 [cited 2020 Sep 7]. p. 66. Available from: </pmc/articles/PMC6408765/?report=abstract>
46. Singh B, Daneshvar C. Human infections and detection of plasmodium knowlesi. Clin Microbiol Rev. 2013;26(2):165–84.
47. Lucas PW, Corlett RT. Relationship between the Diet of *Macaca fascicularis* and Forest Phenology. Folia Primatol. 2008 Sep 15;57(4):201–15.
48. Hassim NA, Hambali K, Idris NSU, Amir A, Ismail A, Zulkifli SZ, et al. Lead concentration in long-tailed macaque (*Macaca fascicularis*) hair in kuala Selangor, Malaysia. Trop Life Sci Res [Internet]. 2018 [cited 2020 Sep 7];29(2):175–86. Available from: </pmc/articles/PMC6072728/?report=abstract>
49. Stark DJ, Fornace KM, Brock PM, Abidin TR, Gilhooly L, Jalius C, et al. Long-Tailed Macaque Response to Deforestation in a Plasmodium knowlesi-Endemic Area. Ecohealth [Internet]. 2019 Dec 1 [cited 2020 Sep 7];16(4):638–46. Available from: <https://doi.org/10.1007/s10393-019-01403-9>
50. Saleh Huddin A, Md Yusuf NA, Razak MRMA, Ogu Salim N, Hisam S. Genetic diversity of Plasmodium knowlesi among human and long-tailed macaque populations in Peninsular Malaysia: The utility of microsatellite markers. Infect Genet Evol. 2019 Nov 1;75:103952.
51. Fungfuang W, Pimpraphai W, Sontiphun S. Survey of Plasmodium knowlesi in long-tailed macaque (*Macaca fascicularis*) at Khao Chongkrachok, Prachuap Khirikhan Province. undefined. 2015.

## Figures



**Fig 1:** Locations of macaques sampling

Figure 1

Locations of macaques sampling