

The Green Toad Example: A Comparison of Pattern Recognition Software

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Short report

Keywords: Amphibia, picture comparison, population statistics, recapture study

Posted Date: September 30th, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-76239/v1>

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1 **The green toad example: a comparison of pattern**

2 **recognition software**

3

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11

12 **Abstract**

13 **Background**

14 Individual identification of animals is important for assessing the size and status of
15 populations. Photo-based approaches, where animals are recognized by naturally
16 occurring and visually identifiable features, such as color patterns, are cost-effective
17 methods for this purpose. We compared five available programs for their power to
18 semi-automatically identify dorsal patterns of the European green toad (*Bufo*
19 *viridis*).

20

21 **Method**

22 We created a data set of 200 pictures of known identity, two pictures for each
23 individual, and analyzed the percentage of correctly identified animals for each
24 software. Furthermore, we employed a generalized linear mixed model to identify
25 important factors contributing to correct identifications. We used these results to
26 estimate the population size of our hypothetical population.

27

28 **Conclusions**

29 The freely available HotSpotter application was the software which performed by far
30 the best for our green toad example, identifying close to 100% of the photos
31 correctly. The animals' sex highly significantly influenced detection probability,
32 presumably because of sex-specific differences in the pattern contrast. Population
33 estimates were close to the expected 100 for HotSpotter, but for the other
34 applications population size was highly overestimated. Given the clarity of our results
35 we strongly recommend the HotSpotter software, which is a highly efficient tool for
36 individual pattern recognition.

37

38 **Keywords** Amphibia, picture comparison, population statistics, recapture study,

39

40 **Background**

41 One of the most basic but also most important tasks in conservation biology is to
42 assess size and status of animal populations. Therefore, mark-recapture techniques
43 in combination with population statistics are used to estimate demographic
44 parameters. In many cases such data are collected in long-term studies and allow
45 prediction of population trajectories. Certain mark-recapture techniques require to tag
46 animals with more or less invasive methods ranging from toe clipping in amphibians
47 (1), inserting PIT tags (2) to adding visual markings (3). Alternatively, genetic
48 identification can be used as a non-invasive method (4).

49

50 The most cost-effective method, however, is to make use of naturally occurring body
51 patterns (5–7). Unfortunately, visually matching pictures of such patterns can be very
52 time consuming and, especially for long-term studies, decreases efficiency (8).
53 Therefore, a number of pattern-matching algorithms have been developed which can
54 cut time of photo comparisons, but in many instances, they can also lead to decreased
55 accuracy (5,9,10). Decreased accuracy, in turn, can cause considerable errors when
56 estimating demographic parameters (11,12). Consequently, deciding on the right
57 algorithm to use for the respective data set is crucial.

58

59 Differences between data sets could include distance to the animals (e.g. photos of
60 giraffes from the distance (13) and close-up pictures of tree frogs (14)), background of
61 photos and quality of cameras used. In long-term data sets picture quality and

62 attributes might vary drastically, therefore, in many instances, scientists prepare the
63 photos before comparing the patterns, for example by cropping the pictures to only
64 show the region of interest (ROI). This can be a quite elaborate task, although it is
65 arguably less time consuming than comparing pictures visually. Matthé et al. (15)
66 compared the performance of four pattern recognition programs with various
67 amphibian species with high contrast patterns. In our case study we extended the
68 selection of software packages and used different picture preparations, where
69 applicable. Furthermore, we chose the green toad as an example of the programs'
70 effectiveness on species with patterns of highly variable and changing contrast. We
71 compared five different pattern recognition programs, which are freely available to
72 researchers, with the exception of AmphIdent. Our results can help population
73 ecologists to gain efficiency when comparing a variety of animal patterns for individual
74 identification.

75

76 **Methods**

77 We used pictures from a long-term capture-recapture study of green toads (*Bufo*
78 *viridis*) at the Rudolf-Bednar-Park in Vienna (48° 13' 35" N, 16° 23' 47" E). Green toads
79 are suitable for automated pattern matching as they have striking dorsal patterns,
80 although the contrast of this pattern can change over time (16). Pictures of the dorsal
81 patterns of green toads were taken in the field with a variety of cameras. The photo
82 background was mostly uniformly white or grey (sheet, cup etc.). A few photos were
83 taken on patchy background (e.g. gravel) or contain parts of a researcher's hand. We
84 classified each camera as one of three types: smart phone cameras, compact
85 cameras and reflex cameras. For the purpose of this study we selected 200 pictures
86 of known animal IDs (two per ID, 100 'reference' and 100 'unknown') of varying quality

87 and different sexes (females: 18, males: 58, unidentifiable: 24, this resembles naturally
88 occurring ratios). We used five different software applications (Wild-ID (13), HotSpotter
89 (17), AmphIdent (application for fire bellied toads) (18), I3S Pattern+ (19) and APHIS
90 (20)) to compare the pictures. Furthermore, for Wild-ID and HotSpotter we chose three
91 different ways to pre-process the pictures, where we used different cropping extents.
92 The cropping types are called a, b and c in this paper, and refer to 'whole animal',
93 'head', 'ROI on head', respectively (Figure S1). In AmphIdent, APHIS and I3SP+ it is
94 required to define a region of interest for every image after loading. For I3SP+ we
95 defined a ROI similar to the shape of cropping type c. For AmphIdent and APHIS we
96 had to adhere to the ROI selection of the respective software. These are squares and
97 rectangles, respectively, which we placed in the same part of the head as in cropping
98 type c. (see Figure S2 for details).

99

100 Pictures were always compared in the same order. The picture comparisons followed
101 the applications' manuals; however, the general procedure was the same throughout.
102 We first added 100 'reference' pictures for each of the used individuals to the
103 application's database, which simulates that we already encountered each of the
104 individuals (once) before. We did this to ensure that each of the new pictures could
105 find a match in the previous record. We then compared 100 pictures of 'unknown' ID
106 to the application database. Usually the user decides whether one of the first few
107 automatically selected pictures is a match or not, but for the purpose of this analysis
108 we only considered the picture ranked first. If this picture was a true match, we
109 recorded a true detection (1), otherwise a false detection (0). AmphIdent and I3SP+
110 have functions for manual input to improve the performance, we used them to the
111 following extent. In I3SP+ pixels can be chosen to discern the pattern's fore- and

112 background. Our time effort to discern fore- from background did never exceed three
113 minutes. AmphIdent has the option to only compare subareas of the ROI. If comparing
114 the whole ROI did not yield a true detection, this function was used up to two times.
115 We bootstrapped a 95% confidence interval for the mean detection success for each
116 software using the function `boot.ci` in the package `boot` (21) in R (22).

117

118 To statistically analyze the performance of the different software we used a
119 generalized linear mixed model (GLMM) with a binomial error distribution employing
120 the function `glmmTMB` from the package `glmmTMB` (23). As the response variable we
121 used the level of detection (0/1) and predictors were the software application, sex of
122 the animals, camera type of the first picture and the second picture as well as
123 interaction between the camera types. In addition, we used individual as a random
124 factor in the model.

125

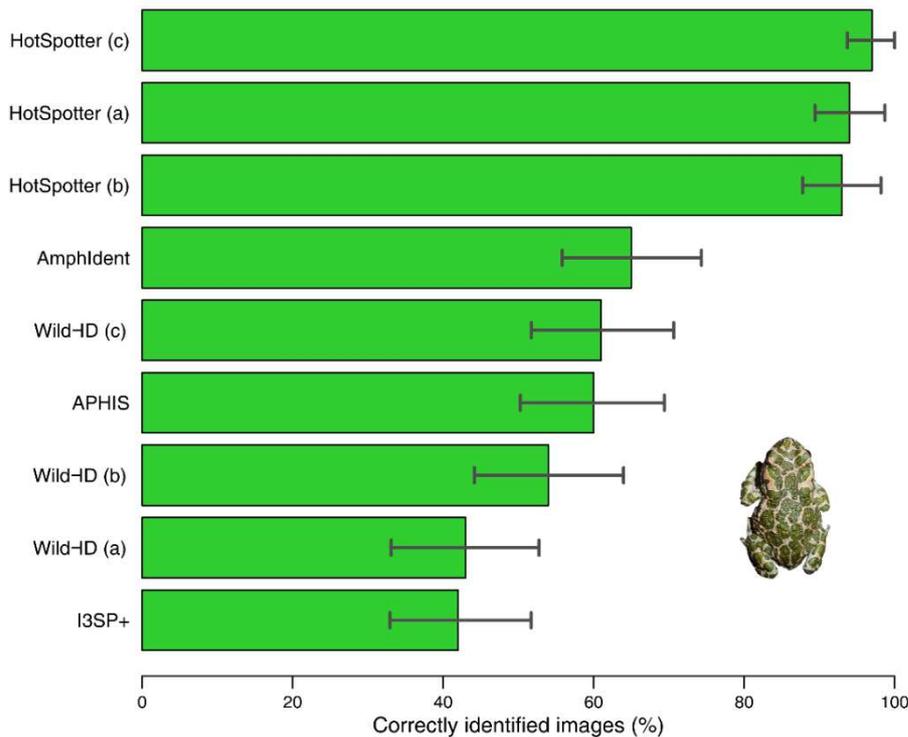
126 We used the Petersen index (24) to estimate abundance assuming that we compared
127 two sampling occasions (the first occasion was the set of the first 100 pictures of 100
128 individuals, the second occasion was the second set of 100 pictures). The true
129 'population size' therefore was 100. Raw data and R code to reproduce our analysis
130 are available in the supplementary information.

131

132 **Results**

133 From the evaluated applications HotSpotter performed the best with detection rates
134 close to 100%, there was only little variation accounted for by image cropping for this
135 software (Figure 1). AmphIdent, Wild-ID (c), and APHIS identified just over 60% of the
136 images correctly. Wild-ID (b), Wild-ID (a) and I3SP+ all identified less than 60% of the

137 images correctly. Cropping was important for Wild-ID, but not for HotSpotter,
138 presumably because this application contains a well-developed background
139 subtraction algorithm incorporated in the software. It should be noted that in all cases
140 results would have been substantially better, if we had included more than just the first
141 hit for an image.



142

143 **Figure 1: Results of comparison showing the percentage of correctly identified pictures in our**
144 **data set.**

145

146 Apart of the highly significant effect of the software also the sex of the animals
147 influenced probability of correct identification (Table 1). This might be expected as the
148 patterns of females have higher contrasts between the green and white patches than
149 the male patterns (16). Nonetheless, it highlights potential confounding factors when
150 performing capture-recapture analyses based on pattern recognition. The camera type
151 used for the pictures only showed weakly significant effects or no effects at all, we did
152 not find any interaction effect between the camera used for the first and the second

153 photo. Switching cameras from one to another occasion might not be an important
154 factor for successful identification.

155

156 Table 1: ANOVA table showing the results for the GLMM

	<i>Chisq</i>	<i>df</i>	<i>p</i>
Sex	28.38	2	<0.01
Software	124.62	8	<0.01
Cam 1	6.02	2	0.049
Cam 2	2.30	1	0.129
Cam 1 : Cam 2	4.17	2	0.124

157

158 If we used solely the first hit from the respective software application and no additional
159 manual comparison, the population sizes would be overestimated to different extent
160 (Table 2). For the HotSpotter software, the estimated numbers are still close to the
161 actual N, and only a few animals would be added. However, for I3SP+ the estimated
162 size would be more than doubled, in comparison to the real population size.

163

164 Table 2: Population size estimates for the different applications

<i>Software</i>	<i>Petersen index estimate</i>
AmphIdent	153.85
APHIS	166.67
HotSpotter (a)	106.38
HotSpotter (b)	107.53
HotSpotter (c)	103.09
I3SP+	238.10
Wild-ID (a)	232.56
Wild-ID (b)	185.19
Wild-ID (c)	163.93

165

166 **Discussion**

167 We show that using automatic pattern recognition applications can be a highly efficient
168 tool to identify individual animals. The software which worked best for our example
169 was the freely available HotSpotter (17). Using this software, we identified the majority

170 of the individuals correctly, and this even with our stringent detection criterium.
171 Applying the same criteria for all other tested applications yielded detection rates
172 around 60% and below. HotSpotter also allows to process the pictures without any
173 time-consuming pre-processing, i.e. cropping the pictures did not significantly change
174 the detection probability. Note that for animals lacking well-defined color patterns
175 these applications might not be an efficient solution for individual identification (10).
176 Focusing on other distinct features (e.g. arrangement of warts or the pores of the
177 parotoid gland) might be a viable alternative in these cases, possibly demanding
178 different cropping techniques. Furthermore, we showed that differences in the
179 detection probability between individuals and sexes could bias population estimations.
180 Knowing such potential biases (e.g. detection probability for a certain group) and/or
181 error rates could be used to inform and improve population analyses (25).

182

183 Recently, it has been shown that HotSpotter can be used for identification of animals
184 that were photographed from the distance (26). Therefore, apart from the general use
185 in traditional population analyses, matching individuals from photos could also be used
186 in citizen science projects. In cases where citizens are asked to take pictures of certain
187 animals in a specific area, scientists might be able to determine the individual identity
188 from such photos (27).

189 In conclusion, at least for our green toad example and possibly species with similar
190 patterns the freely available HotSpotter software constitutes a major improvement to
191 other more widely used pattern recognition software, such as APHIS and Wild-ID.

192

193

194 **Declarations**

195 **Ethics approval**

196 Toad pictures are taken from an ongoing effort in monitoring green toad populations
197 in Vienna. All pictures were taken under a permit issued by the municipal department
198 for environmental protection (MA 22 – 984143-2015-5).

199 **Consent for publication**

200 All authors consent for publication of this manuscript.

201 **Availability of data and materials**

202 Raw data are available in the supplementary of this manuscript.

203 **Competing interests**

204 The authors declare to have no competing interests.

205 **Funding**

206 The authors received no funding for this study.

207 **Authors' contributions**

208 SB and LL conceived the study. SB performed the picture comparisons. SB and LL
209 did the statistical analyses. SB, LL, and GG interpreted the data and wrote the paper.

210 **Acknowledgements**

211 We thank Christoph Leeb, Eileen Heyer, Christian Wappl and the students who
212 participated in the field courses “Populationsbiologie heimischer Amphibien” and
213 ”Freilandpraktikum Amphibienökologie” at the University of Vienna from 2015 to 2019,
214 who helped in collecting the pictures in ongoing green toad monitoring efforts in
215 Vienna.

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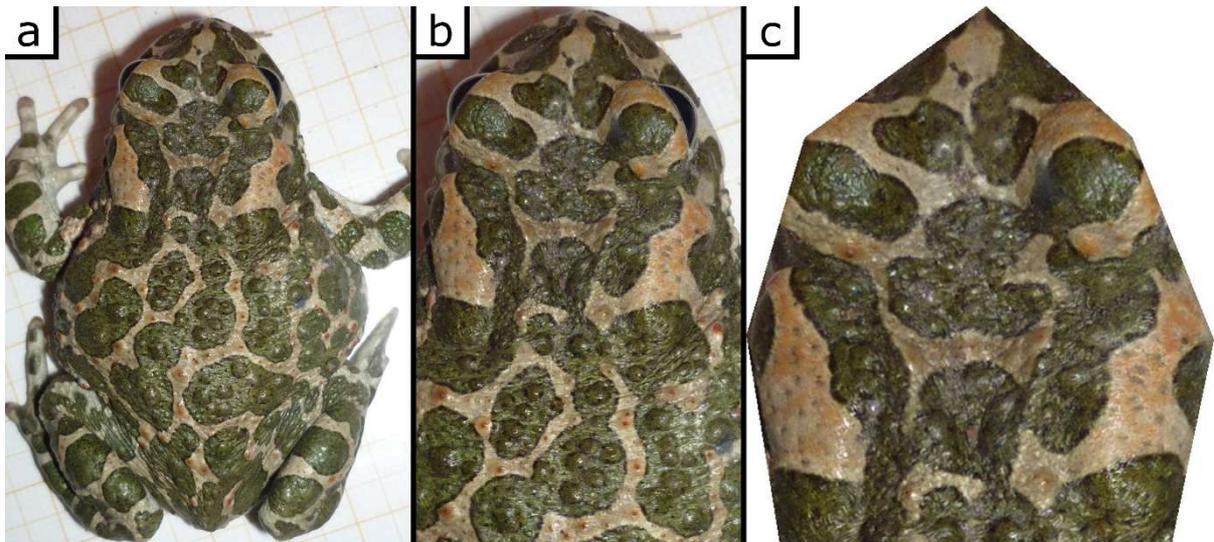
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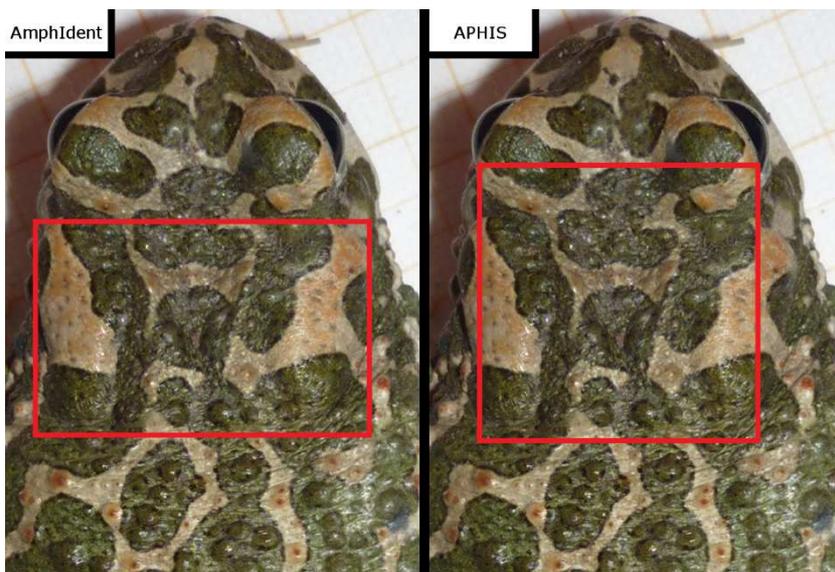
304 **Supplementary figure**



305

306 **Figure S1: The three different cropping types used in the study for HotSpotter and Wild-ID: whole**
307 **animal (a), head (b), region of interest (c).**

308



309

310 **Figure S2: The shapes of the ROI for AmphIdent (left) and APHIS (right). The width to height**
311 **ratio for APHIS is fixed at 1:1. We chose to locate the upper two corners of the ROI at the lower**
312 **end of the visible eyeballs. For AmphIdent the ROI ratio can be freely chosen but the selected**
313 **area will be stretched to a 10:7 ratio, so in order not to distort the original picture too much we**
314 **selected an area with this approximate ratio. The ROI extends from the upper to the lower, as**
315 **well as from the left to the right edges of the parotoid glands.**

Figures

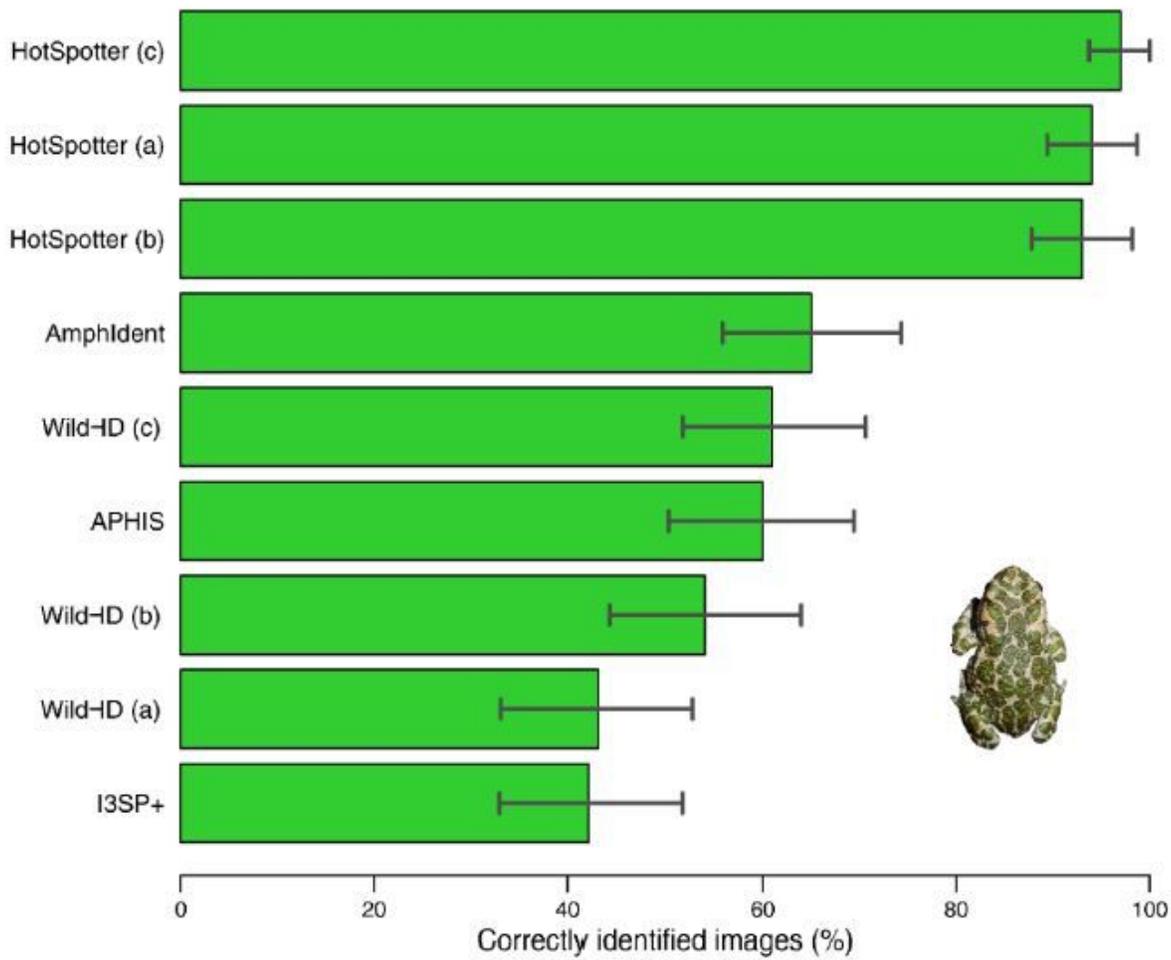


Figure 1

Results of comparison showing the percentage of correctly identified pictures in our data set.

Supplementary Files

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- [FigS1.JPG](#)
- [FigS2.JPG](#)