

Analysis of discordance between AMH and AFC in IVF patients: a retrospective cohort study

Yannan Chen

Affiliated Hospital of Nantong University

Xiaoling Gu

Affiliated Hospital of Nantong University

Xi Guan

Affiliated Hospital of Nantong University

Minyan Yu

Affiliated Hospital of Nantong University

Shuping Zhong

Affiliated Hospital of Nantong University

Panpan Yang

Affiliated Hospital of Nantong University

Xia Wang

Affiliated Hospital of Nantong University

Di Wang (✉ wdok1972@163.com)

Nantong University Affiliated Hospital: Affiliated Hospital of Nantong University <https://orcid.org/0000-0002-1929-1798>

Research

Keywords: anti-Mullerian hormone (AMH), antral follicle count (AFC), ovarian reserve, clinical discordance, and BMI

Posted Date: August 2nd, 2021

DOI: <https://doi.org/10.21203/rs.3.rs-763045/v1>

License:  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Abstract

Background

Both anti-Mullerian hormone (AMH) and antral follicle count (AFC) predict ovarian reserve independently and have been shown to correlate well. However, it is quite common to encounter discordance between the two clinical indices. This objective of this study is to speculate which condition is favorable to patients and to decide what best to be done in cases of such discordance.

Methods

This retrospective cohort study analyzed the medical records of 714 women undergoing the first IVF cycle at the Center for Reproductive Medicine, the Affiliated Hospital of Nantong University, Nantong, Jiangsu between January 2017 and December 2020. All patients had their essential characteristics, baseline FSH, Estradiol, PRL, LH, Testosterone, CA125, AMH, and AFC recorded. Patients were classified into three groups according to whether their AMH and AFC values were in the lower quartile, upper quartile, or in between, namely Group A (Concordance, concordant in AMH and AFC), Group B (HTP, AMH concentration higher than predicted according to AFC) and Group C (LTP, AMH concentration lower than predicted according to AFC). SPSS 25.0 software package (SPSS, Inc., Chicago, IL, USA) was used for statistical analysis.

Results

The entire study population has a good concordance between AMH and AFC. 240 women (33.61%) had discordant AMH and AFC values, and the proportion in Group B(HTP) and Group C(LTP) seemed similar (16.67% VS 16.94%). The three cohorts had similar age, infertile duration, FSH, Estradiol, PRL, LH, Testosterone, and CA125 levels. BMI progressively increased from Group B to Group A and to Group C. The ratio of patients, who had completely mature oocytes (COM), rapidly decreased from Group C to Group A and to Group B. Although no significant difference in CMO rate was observed among Group A, B, and C when stratified according to BMI, CMO rate was the highest (72.31%) when BMI < 18.5 kg/m² in all BMI categories.

Conclusions

Our analysis demonstrated a high frequency of clinical discordance between AMH and AFC in women undergoing IVF therapeutic regime. Patients' BMI and completely mature oocytes ratio are significantly different among distinct grouping based on their AMH and AFC partition.

Background

The ovarian reserve is usually estimated by serum anti-Mullerian hormone (AMH) concentration(1) and antral follicle count (AFC)(2), which are also often employed in in vitro fertilization (IVF) therapeutic regime(3–5).

AMH, also known as Mullerian-inhibiting substance, is a member of the transforming growth factor β family, which is secreted by granulosa from small, growing follicles(6–9). AMH exerts its biological effects on the ovaries generally by inhibiting the initial follicle recruitment and the FSH-dependent growth of the follicles and selection of pre-antral and small antral follicles through a transmembrane serine/threonine kinase type II receptor (AMHR II)(10–12). The absence of AMH has been shown to enhance FSH-induced follicle growth in female mice(10, 13). Currently, AMH has been widely used as a golden marker for evaluating ovarian reserve in females, particularly in the field of assisted reproduction(1, 14–16).

Antral follicle count (AFC), which consists of all ultrasonographically identified and counted antral follicles measuring 2–10 mm in diameter in both ovaries, is another reliable indicator of ovarian reserve owing to its close relationship to age-related declines in follicle counts(17–21). Although AFC may be more machine and operator-dependent and could have higher inter-operator variability compared to AMH, it correlates more directly with the ovarian reserve of a single ovary than AMH(1, 3, 5, 19, 22).

Systematic reviews showed that AMH and AFC had a very good correlation with each other. In 2003, Fanchin et al. demonstrated that AFC was closely related to serum AMH level on cycle day 3 in infertile women, more strongly than other hormonal markers, such as inhibin B, E2, and FSH(23). A strong positive correlation between AMH and AFC and paralleled fluctuations of AMH and AFC have also been demonstrated throughout the regular menstrual cycle(24–26). Yet, it is not unusual for IVF specialists to encounter a serum AMH level that is higher or lower than expected according to AFC, even when the measurement of AMH and counting of antral follicles are performed during the early follicular phase of the same menstrual cycle in a single clinical center(18, 27).

In this article, we conducted a retrospective analysis to evaluate the discordance between AMH and AFC in women undergoing their first IVF treatment, the reasons hiding in this phenomenon, and maybe the clinical implications.

Methods

Patient sampling

The medical records of 714 women at the Center for Reproductive Medicine, Affiliated Hospital of Nantong University between January 2017 and December 2020 were evaluated in the analysis. Only the first cycle was included for each patient to eliminate previous failure bias.

The exclusion criteria were as follows: patients with a history of ovarian or pelvic surgery, endometriosis, chemotherapy or radiation exposure, hormonal therapy in the past 6 months, fragile X premutation, and

abnormal karyotype or other endocrine diseases, including diabetes, thyroid dysfunction, and hyperprolactinemia.

During the study period, the type of protocol was determined by the doctor according to specific characteristics, such as ovarian reserve and follicle size.

All patients who had their essential characteristics, baseline FSH, Estradiol, PRL, LH, Testosterone, CA125, AMH, and AFC recorded during their evaluation were included in the study. The Ethics Committee of Affiliated Hospital of Nantong University has authorized us to use existing patient data anonymously, waiving the requirement for written consent from individual patients. The main clinical and pathologic variables of the study patients are shown in Table 2.

Classification of patients

In the precycle analysis, the 25th percentile and 75th percentile values of serum AMH concentration were 2.05 ng/mL and 5.58 ng/mL, whereas the values of AFC were 12 and 31, respectively. Patients were classified into three groups according to whether their AMH and AFC values were in the lower quartile, upper quartile, or in between (Table 1), namely Group A (Concordant, concordance in AMH and AFC), Group B (HTP, AMH concentration higher than predicted according to AFC) and Group C (LTP, AMH concentration lower than predicted according to AFC).

Clinical investigation

Ultrasonographic examinations were performed by reproductive medicine specialists, who had at least 5 years of experience in gynecological ultrasound and have undergone uniform training. During the early follicular phase of the menstrual cycle, follicles measuring 2–10 mm in diameter were counted. The number of follicles in each ovary was assessed by using the same ultrasound equipment with a 2D transvaginal probe 6.5 MHz (Toshiba, Nemio, Japan). The total number of follicles in both ovaries was used as the AFC.

Laboratory analyses

Serum FSH, Estradiol, PRL, LH, and Testosterone were measured on day 2 or 3 of the menstrual cycle with the use of Beckman Coulter Dxl 800 (Beckman Coulter, Inc., Brea, USA). Random serum CA125 and AMH levels, unrelated to cycle day, were measured by using enzyme-linked immunosorbent assay (ELISA) on ARCHITECH I2000SI (Abbott, Chicago, IL, USA) and iFlash 3000-H (YHLO Biotechnology Co., Ltd., Shenzhen, CHINA), respectively.

The functional sensitivity, the coefficients of variation for intra-assay precision (repeatability) and the inter-assay precision (intermediate precision) of the FSH, Estradiol, PRL, LH, Testosterone, CA125 and AMH assays were as follows: 0.20 mIU/mL, 3.10%-4.30%, 4.30%-5.60%; 15 pg/mL, $\leq 10\%$, $\leq 15\%$; 0.25 ng/mL, 1.42%-1.61%, 3.32%-6.92%; 0.20 mIU/mL, 3.80%-5.40%, 4.30%-6.40%; 0.10 ng/mL, 1.67%-3.93%, 4.22%-7.08%; 1.00 U/mL, 1.50%-3.20%, 1.90%-4.70%; and 0.10 ng/mL, $\leq 10\%$, $\leq 15\%$.

Body mass index (BMI) was calculated as weight (kg)/height (m)².

Statistical analysis

SPSS 25.0 software package (SPSS, Inc., Chicago, IL, USA) was used for statistical analysis. Patient characteristics were compared between the groups by using Mann–Whitney test. Continuous variables were compared using Student’s t-test when data were normally distributed and by Mann–Whitney U when the data were not normally distributed. Chi-squared tests were used to compare proportions. All normal distribution measurement data are expressed as the mean ± standard deviation (SD). The level of significance was set at P < 0.05.

Results

In this study, 714 women, who underwent the first IVF treatment cycle, fulfilled the inclusion criteria, were included. The clinical and demographic parameters of the included subjects were listed in Table 2.

The correlation between AMH and AFC was visualized in Figure 1. A significant positive association of AMH and AFC was depicted in the entire study population (regression equation: AMH = 0.4167 × AFC; R²= 0.462, P < 0.001).

To better elucidate the relationship between AMH and AFC, the two ovarian reserve markers were stratified into the following categories. The interquartile range (IQR) value of serum AMH concentration was 2.05-5.58 ng/mL, whereas which of AFC was 12-31. Patients were grouped according to whether their AMH and AFC values were in the lower quartile, upper quartile, or in between (Table 1).

Table 1 Grouping of the entire study subjects based on the interquartile range (IQR) values of serum AMH concentration (ng/mL) and AFC.

AMH [ng/mL]	AFC			
	≤12	12-31	≥31	
≤2.05	0	0	0	
	124	49	5	178
2.05-5.58	0	0	0	
	43	249	67	359
≥5.58	0	0	0	
	0	76	101	177
	167	374	173	714

As shown in Table 1, subjects in \square , \square , and \square (474 out of 714, 66.39%) , had a concordance in AMH and AFC. In the rest 240 subjects (240 out of 714, 33.61%), those in \square , \square , and \square had AMH concentration higher than predicted(HTP) according to AFC (119 out of 714, 16.67%), while those in \square , \square , and \square had it lower than predicted(LTP) according to AFC (121 out of 714, 16.94%). Patients were then divided into three groups based on the concordance between AMH and AFC levels provided by Table 1:

Group A (Concordant): \square + \square + \square (AMH<2.05 ng/mL and AFC<12, AMH 2.05-5.58 ng/mL and AFC 12-31, AMH \square 5.58 ng/mL and AFC \square 12);

Group B (HTP): \square + \square + \square (AMH 2.05-5.58 ng/mL and AFC<12, AMH \square 5.58ng/mL and AFC \leq 31);

Group C (LTP): \square + \square + \square (AMH<2.05 ng/mL and AFC \geq 12, AMH 2.05-5.58 ng/mL and AFC \square 31).

The clinical and biochemical characteristics of subjects after grouping were presented in Table 2. As seen in Table 2, the three cohorts had similar age, infertile duration, FSH, Estradiol, PRL, LH, Testosterone, and CA125 levels.

Table 2 Comparison of clinical and biochemical characteristics of patients in different groups. P < 0.05 was considered statistically significant. Data were presented as n (%) or mean \pm standard deviation (SD).

Variables	Group A (Concordant) (N=474)	Group B (HTP) (N=119)	Group C (LTP) (N=121)	Total (N=714)	P
Age (years)	30.07±4.59	29.54±4.08	29.29±4.43	29.85±4.49	0.059
BMI (kg/m ²)	22.65±3.43	22.56±3.46	23.81±3.87	22.83±3.54	0.015 [□]
Infertile duration (years)	3.41±2.63	3.32±2.36	3.31±2.53	3.38±2.57	0.914
FSH (mIU/mL)	7.85±3.24	7.18±1.99	7.09±2.04	7.61±2.90	0.082
Estradiol (pg/mL)	51.89±32.24	52.29±20.15	50.57±22.82	51.73±29.06	0.367
PRL (pg/mL)	277.46±132.39	299.48±148.77	279.09±129.20	281.41±134.78	0.589
LH (pg/mL)	5.11±3.16	5.31±3.45	5.07±3.33	5.13±3.23	0.711
Testosterone (ng/mL)	0.54±2.52	0.68±2.72	0.43±0.15	0.54±2.33	0.512
CA125 (U/mL)	17.46±10.12	16.24±7.84	16.78±8.79	17.14±9.56	0.579
AFC	22.21±15.19	18.22±8.70	30.55±12.37	22.96±14.33	0.000 [□]
AMH (ng/mL)	4.20±3.44	5.82±2.66	3.01±1.56	4.27±3.18	0.000 [□]
Fully mature retrieved oocytes(n)	333 (70.25%)	74 (62.18%)	93 (76.86%)	500	0.045 [□]

Interestingly, BMI (P=0.015) progressively increased from Group B(HTP) to Group A(Concordant) and to Group C(LTP) (from 22.56±3.46 kg/m² to 22.65±3.43 kg/m² and to 23.81±3.87 kg/m², P=0.015). The ratio of patients, who had completely mature oocytes (COM), rapidly decreased from Group C(LTP) to Group A(Concordant) and to Group B(HTP) (from 76.86% to 70.25% and to 62.18%, P=0.045).

According to BMI, general population is classified into five categories: underweight (BMI<18.5 kg/m²), normal weight (BMI 18.5-24.9 kg/m²), class I obesity-overweight (BMI 25.0-29.9 kg/m²), class II obesity -obesity (BMI 30.0-39.9 kg/m²), and class III obesity -extreme obesity (BMI≥40 kg/m²). The range of BMI in this study sample was 14.90-36.10 kg/m². To get a better understanding of BMI-related AMH and AFC discordance, women were subdivided into the following groups on account of the above BMI classification criteria (Table 3, P=0.009). The upper numbers were the observed values, and the numbers in the brackets were the expected ones. As shown in Table 3, the observed value in Group A (concordant) was notably higher than the expected one when BMI≤24.9 kg/m² (BMI<18.5 kg/m² and BMI 18.5-24.9 kg/m²), which means in this BMI region, concordant in AMH and AFC was more common. Concurrently,

the corresponding situation in Group B (HTP) generally occurred when BMI<18.5 kg/m² and BMI 25.0-29.9 kg/m², while in Group C (LTP), widely appeared when BMI≥25.0 kg/m² (BMI 25.0-29.9 kg/m² and BMI 30.0-39.9 kg/m²).

Table 3 The number of patients in different groups sub-stratified according to BMI categories. P < 0.05 was considered statistically significant. Data were presented as n (%).

BMI classification (kg/m ²)	Group A (Concordant) (N=474)	Group B (HTP) (N=119)	Group C (LTP) (N=121)	Total (N=714)	P
Underweight ≤18.5	47 (43.2)	14 (10.8)	4 (11.0)	65	0.009 [†]
Normal weight 18.5-24.9	321 (311.4)	72 (78.2)	76 (79.5)	469	
Class I obesity [‡] overweight [‡] 25-29.9	92 (101.6)	30 (25.5)	31 (25.9)	153	
Class II obesity [‡] obesity [‡] 30-39.9	14 (17.9)	3 (4.5)	10 (4.6)	27	

Our previous studies showed that Group C (LTP) patients had the highest BMI, and meanwhile the supreme completely mature oocytes (CMO) rate. On the contrary, Group B(HTP) patients had the lowest BMI, and simultaneously the worst CMO rate. We proposed whether a lower BMI was accompanied by a worse CMO rate in different grouped specimens, and vice versa. Although no significant difference in CMO rate was observed among Group A, B, and C when stratified according to BMI (P=0.719, Table 4), CMO rate was the highest (72.31%) when BMI< 18.5 kg/m² in all BMI categories. The highest rate of CMO in Group A(Concordant) was seen when BMI< 18.5 kg/m² (72.34%), in Group B(HTP) when BMI≥ 25.0 kg/m² (100%), and in Group C(LTP) when BMI 18.5-24.9 kg/m² (81.58%).

Table 4 The ratio of completely mature oocytes in different groups sub-stratified according to BMI categories. P < 0.05 was considered statistically significant. Data were presented as n (%).

BMI classification (kg/m ²)	Group A (Concordant) (N=474)	Group B (HTP) (N=119)	Group C (LTP) (N=121)	Total (N=714)	P
Underweight ≤18.5	34/47 (72.34%)	11/14 (78.57%)	2/4 (50.00%)	47/65 (72.31%)	0.719
Normal weight 18.5-24.9	230/321 (71.65%)	41/72 (56.94%)	62/76 (81.58%)	332/469 (71.00%)	
Class I obesity/overweight 25-29.9	62/92 (67.39%)	19/30 (63.33%)	21/31 (67.74%)	102/153 (66.67%)	
Class II obesity/obesity 30-39.9	7/14 (50.00%)	3/3 (100.00%)	8/10 (80.00%)	18/27 (66.67%)	

Discussion

It is important for patients undergoing assisted reproduction treatment to evaluate ovarian reserve precisely. Both AMH and AFC predict ovarian reserve independently and have been shown to correlate well, although AFC may have a higher inter-operator variability compared to AMH. However, it is quite common for IVF specialists to encounter serum AMH level that is higher or lower than expected according to AFC. It impels us to speculate which condition is favorable to patients and to decide what best to be done in cases of such discordance.

Our initial results showed a strong correlation between AMH and AFC, and then, we classified the two measurements into three intervals based on their respective lower and upper quartiles. The cut-offs for AFC were 12 and 31, and for AMH were 2.05 and 5.58 ng/mL, respectively. There was a high level of discordance (33.61% of cases) between AMH and AFC, and the incidence of HTP and LTP seemed similar (16.67% VS 16.94%). The further division demonstrated that BMI progressively increased from Group B(HTP) to Group A(Concordant) and to Group C(LTP). The ratio of patients, who had completely mature oocytes (COM), rapidly decreased from Group C(LTP) to Group A(Concordant) and to Group B(HTP). BMI classification ulteriorly authenticated the significance of BMI in the discordance between AMH and AFC. This urged us to speculate if Group B(HTP) had the worst ratio of completely mature oocytes (COM) was due simply to its lowest BMI. We found a negative correlation, although statistically nonsignificant, between BMI and completely mature oocytes rate altogether. The highest rate of completely mature oocytes in Group A(Concordant) could be seen when BMI < 18.5 kg/m²(72.34%), in Group B(HTP) when BMI ≥ 25.0 kg/m²(100%), and in Group C(LTP) when BMI 18.5–24.9 kg/m²(81.58%).

There have been widely differing versions of the reasons for the discordance between AMH and AFC. The AFC refers to the number of follicles with a diameter of 2–10 mm in both ovaries and is a direct marker of

the recruitable follicular cohort(28). AMH is a dimeric glycoprotein produced by granulosa cells of pre-antral and small antral follicles, and it indirectly reflects the population of early growing follicles. These pre-antral and small antral follicles are not able to completely develop into maturity leading to oocyte being yielded(3). Thus, the AMH level is not exactly reflecting the population of antral follicles counted in our AFC measurement. Moreover, the intra- and inter-menstrual cycle AMH variations may contribute to discordance in some cases(29).

A negative correlation between AMH and BMI has been previously reported in late reproductive-age women(30), young women using oral contraceptive pills(31), and infertile women with (diminished ovarian reserve) DOR but not in women with normal ovarian reserve (NOR)(32). In our analysis, we have found a significant role of BMI in the discordance between AMH and AFC. Group A(Concordant) had a moderate BMI and a medium ratio of completely mature oocytes (COM). Group B(HTP) had the lowest BMI and meanwhile, the worst ratio of completely mature oocytes (COM) and Group C(LTP) was its opposite. But we could not jump to a conclusion that a patient with AMH concentration higher or lower than predicted according to AFC will have a worse or better ratio of completely mature oocytes owing to individual BMI.

Conclusions

In conclusion, approximately 33.61% of patients undergoing the first IVF therapeutic regime have discordance between AMH and AFC. Patients having a concordance in AMH and AFC have an intermediate ratio of completely mature oocytes and a moderate BMI. Those whose AMH concentration lower than predicted according to AFC have an obviously higher ratio, and their BMI are also above the average. The BMI is not an independent element that would influence the complete maturity of oocytes. This study is limited by its retrospective nature and the small number of patients, making it difficult to generalize results. Such theoretical consideration, however, is needed to be elucidated by future clinical and basic studies.

Declarations

Ethics approval and consent to participate

This study has been approved by the Ethics Committee of Affiliated Hospital of Nantong University.

Consent for publication

Not applicable.

Availability of data and materials

Datasets during the current study are available from the corresponding author upon reasonable request.

Competing interests

The authors declare no conflict of interest.

Funding

No funding was received for this study.

Authors' contributions

Yannan Chen, Xia Wang and Di Wang designed the study, conducted literature searches, selected articles for inclusion, performed all statistical analyses, and drafted the first draft of the manuscript. Xiaoling Gu, Xi Guan and Minyan Yu reviewed the study design, extracted data, planned the statistical analyses and drafted the first version of the manuscript. Panpan Yang and Shuping Zhong extracted data and assessed study quality. The manuscript has been revised and approved by all authors.

Acknowledgments

None.

Author details

¹ the Center for Reproductive Medicine, Affiliated Hospital of Nantong University, 226001 Nantong, Jiangsu, P.R. China.

² the Department of Clinical Laboratory, Affiliated Hospital of Nantong University, 226001 Nantong, Jiangsu, P.R. China.

References

1. Baker VL, Glassner MJ, Doody K, Schnell VL, Gracia C, Shin SS, et al. Validation study of the Access AMH antimullerian hormone assay for the prediction of poor ovarian response to controlled ovarian stimulation. *Fertil Steril*. 2021.
2. Al Rashid K, Taylor A, Lumsden MA, Goulding N, Lawlor DA, Nelson SM. Association of the functional ovarian reserve with serum metabolomic profiling by nuclear magnetic resonance spectroscopy: a cross-sectional study of ~ 400 women. *BMC Med*. 2020;18(1):247.
3. Li HW, Lee VC, Lau EY, Yeung WS, Ho PC, Ng EH. Ovarian response and cumulative live birth rate of women undergoing in-vitro fertilisation who had discordant anti-Mullerian hormone and antral follicle count measurements: a retrospective study. *PLoS One*. 2014;9(10):e108493.
4. Li HWR, Nelson SM. Clinical Application of AMH Measurement in Assisted Reproduction. *Front Endocrinol (Lausanne)*. 2020;11:606744.

5. Esteves SC, Yarali H, Vuong LN, Carvalho JF, Ozbek IY, Polat M, et al. Antral follicle count and anti-Mullerian hormone to classify low-prognosis women under the POSEIDON criteria: a classification agreement study of over 9000 patients. *Hum Reprod.* 2021;36(6):1530-41.
6. Baker VL, Gracia C, Glassner MJ, Schnell VL, Doody K, Coddington CC, et al. Multicenter evaluation of the Access AMH antimullerian hormone assay for the prediction of antral follicle count and poor ovarian response to controlled ovarian stimulation. *Fertil Steril.* 2018;110(3):506-13 e3.
7. Dewailly D, Andersen CY, Balen A, Broekmans F, Dilaver N, Fanchin R, et al. The physiology and clinical utility of anti-Mullerian hormone in women. *Hum Reprod Update.* 2014;20(3):370-85.
8. von Wolff M, Mitter VR, Jamir N, Stute P, Eisenhut M, Bersinger NA. The endocrine milieu in naturally matured follicles is different in women with high serum anti-Mullerian hormone concentrations. *Reprod Biomed Online.* 2021.
9. Luo E, Zhang J, Song J, Feng D, Meng Y, Jiang H, et al. Serum Anti-Mullerian Hormone Levels Were Negatively Associated With Body Fat Percentage in PCOS Patients. *Front Endocrinol (Lausanne).* 2021;12:659717.
10. Albu D, Albu A. The relationship between anti-Mullerian hormone serum level and body mass index in a large cohort of infertile patients. *Endocrine.* 2019;63(1):157-63.
11. Cui Y, Shi Y, Cui L, Han T, Gao X, Chen ZJ. Age-specific serum antimullerian hormone levels in women with and without polycystic ovary syndrome. *Fertil Steril.* 2014;102(1):230-6 e2.
12. di Clemente N, Josso N, Gouedard L, Belville C. Components of the anti-Mullerian hormone signaling pathway in gonads. *Mol Cell Endocrinol.* 2003;211(1-2):9-14.
13. Baarends WM, Uilenbroek JT, Kramer P, Hoogerbrugge JW, van Leeuwen EC, Themmen AP, et al. Anti-mullerian hormone and anti-mullerian hormone type II receptor messenger ribonucleic acid expression in rat ovaries during postnatal development, the estrous cycle, and gonadotropin-induced follicle growth. *Endocrinology.* 1995;136(11):4951-62.
14. Broer SL, Mol BW, Hendriks D, Broekmans FJ. The role of antimullerian hormone in prediction of outcome after IVF: comparison with the antral follicle count. *Fertil Steril.* 2009;91(3):705-14.
15. Weenen C, Laven JS, Von Bergh AR, Cranfield M, Groome NP, Visser JA, et al. Anti-Mullerian hormone expression pattern in the human ovary: potential implications for initial and cyclic follicle recruitment. *Mol Hum Reprod.* 2004;10(2):77-83.
16. Guo Y, Liu S, Hu S, Li F, Jin L. High Serum Anti-Mullerian Hormone Concentrations Are Associated With Poor Pregnancy Outcome in Fresh IVF/ICSI Cycle but Not Cumulative Live Birth Rate in PCOS Patients. *Front Endocrinol (Lausanne).* 2021;12:673284.

17. Iliodromiti S, Anderson RA, Nelson SM. Technical and performance characteristics of anti-Mullerian hormone and antral follicle count as biomarkers of ovarian response. *Hum Reprod Update*. 2015;21(6):698-710.
18. Nelson SM. Biomarkers of ovarian response: current and future applications. *Fertil Steril*. 2013;99(4):963-9.
19. Grisendi V, Mastellari E, La Marca A. Ovarian Reserve Markers to Identify Poor Responders in the Context of Poseidon Classification. *Front Endocrinol (Lausanne)*. 2019;10:281.
20. Esteves SC, Yarali H, Vuong LN, Carvalho JF, Ozbek IY, Polat M, et al. Cumulative delivery rate per aspiration IVF/ICSI cycle in POSEIDON patients: a real-world evidence study of 9073 patients. *Hum Reprod*. 2021.
21. Grynberg M, Labrosse J, Bennani Smires B, Sifer C, Peigne M, Sonigo C. Could hormonal and follicular rearrangements explain timely menopause in unilaterally oophorectomized women? *Hum Reprod*. 2021;36(7):1941-7.
22. Re C, Mignini Renzini M, Rodriguez A, Dal Canto M, Buccheri M, Sacchi S, et al. From a circle to a sphere: the ultrasound imaging of ovarian follicle with 2D and 3D technology. *Gynecol Endocrinol*. 2019;35(3):184-9.
23. Fanchin R, Schonauer LM, Righini C, Guibourdenche J, Frydman R, Taieb J. Serum anti-Mullerian hormone is more strongly related to ovarian follicular status than serum inhibin B, estradiol, FSH and LH on day 3. *Hum Reprod*. 2003;18(2):323-7.
24. Hadlow N, Brown SJ, Habib A, Wardrop R, Joseph J, Gillett M, et al. Quantifying the intraindividual variation of antimullerian hormone in the ovarian cycle. *Fertil Steril*. 2016;106(5):1230-7.
25. Depmann M, van Disseldorp J, Broer SL, Eijkemans MJ, Laven JS, Visser JA, et al. Fluctuations in anti-Mullerian hormone levels throughout the menstrual cycle parallel fluctuations in the antral follicle count: a cohort study. *Acta Obstet Gynecol Scand*. 2016;95(7):820-8.
26. Schiffner J, Roos J, Broomhead D, Helden JV, Godehardt E, Fehr D, et al. Relationship between anti-Mullerian hormone and antral follicle count across the menstrual cycle using the Beckman Coulter Access assay in comparison with Gen II manual assay. *Clin Chem Lab Med*. 2017;55(7):1025-33.
27. Tian Z, Zhang Y, Zhang C, Wang Y, Zhu HL. Antral follicle count is reduced in the presence of endometriosis: a systematic review and meta-analysis. *Reprod Biomed Online*. 2021;42(1):237-47.
28. Zhang Y, Xu Y, Xue Q, Shang J, Yang X, Shan X, et al. Discordance between antral follicle counts and anti-Mullerian hormone levels in women undergoing in vitro fertilization. *Reprod Biol Endocrinol*. 2019;17(1):51.

29. Alebic MS, Stojanovic N, Dewailly D. Discordance between serum anti-Müllerian hormone concentrations and antral follicle counts: not only technical issues. *Hum Reprod.* 2018;33(6):1141-8.
30. Freeman EW, Gracia CR, Sammel MD, Lin H, Lim LC, Strauss JF, 3rd. Association of anti-müllerian hormone levels with obesity in late reproductive-age women. *Fertil Steril.* 2007;87(1):101-6.
31. Steiner AZ, Stanczyk FZ, Patel S, Edelman A. Antimüllerian hormone and obesity: insights in oral contraceptive users. *Contraception.* 2010;81(3):245-8.
32. Buyuk E, Seifer DB, Illions E, Grazi RV, Lieman H. Elevated body mass index is associated with lower serum anti-müllerian hormone levels in infertile women with diminished ovarian reserve but not with normal ovarian reserve. *Fertil Steril.* 2011;95(7):2364-8.

Figures

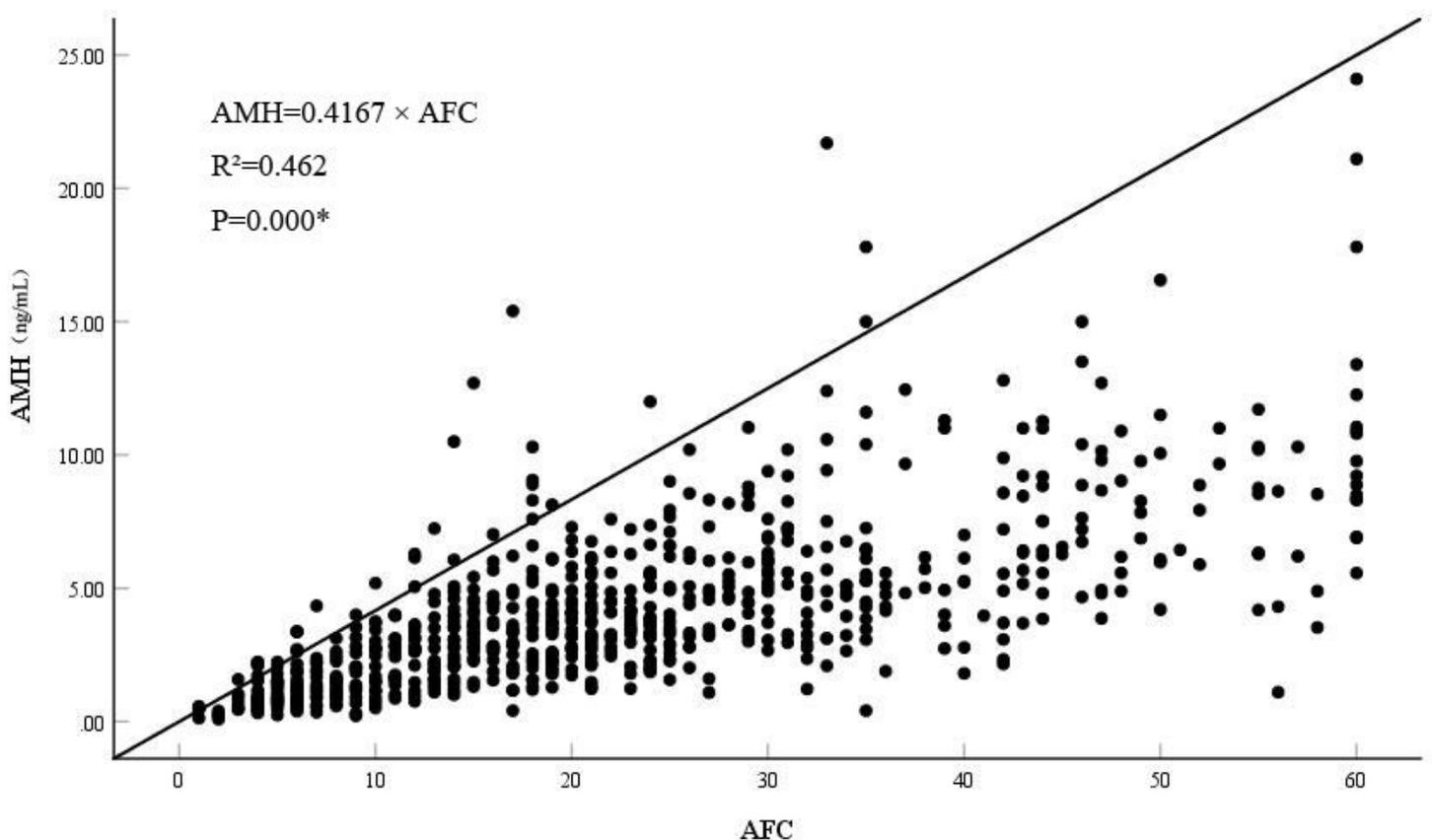


Figure 1

The relationship between serum anti-Müllerian hormone (AMH) concentration and antral follicle count (AFC) in the entire study population.