

# The 'Un-Shrunk' Partial Correlation in Gaussian Graphical Models

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## Research article

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# Title: The ‘un-shrunk’ partial correlation in Gaussian graphical models

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## Abstract

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**Background:** In systems biology, it is important to reconstruct regulatory networks from quantitative molecular profiles. Gaussian graphical models (GGMs) are one of the most popular methods to this end. A GGM consists of nodes (representing the transcripts, metabolites or proteins) inter-connected by edges (reflecting their partial correlations). Learning the edges from quantitative molecular profiles is statistically challenging, as there are usually fewer samples than nodes (‘high dimensional problem’). Shrinkage methods address this issue by learning a regularized GGM. However, it is an open question how the shrinkage affects the final result and its interpretation.

**Results:** We show that the shrinkage biases the partial correlation in a non-linear way. This bias does not only change the magnitudes of the partial correlations but also affects their order. Furthermore, it makes networks obtained from different experiments incomparable and hinders their biological interpretation. We propose a method, referred to as the ‘un-shrunk’ partial correlation, which corrects for this non-linear bias. Unlike traditional methods, which use a fixed shrinkage value, the new approach provides partial correlations that are closer to the actual (population) values and that are easier to interpret. We apply the ‘un-shrunk’ method to two gene expression datasets from *Escherichia coli* and *Mus musculus*.

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1 **Conclusions:** GGMs are popular undirected graphical models based on partial correlations. The application of  
2 GGMs to reconstruct regulatory networks is commonly performed using shrinkage to overcome the “high-  
3 dimensional” problem. Besides its advantages, we have identified that the shrinkage introduces a non-linear bias  
4 in the partial correlations. Ignoring this type of effects caused by the shrinkage can obscure the interpretation of  
5 the network, and impede the validation of earlier reported results.

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6

7 **Keywords:** Gaussian graphical models, Partial correlations, Shrinkage, Gene regulatory networks.

8 **Supplementary information:** Supplementary data are available at *BMC Bioinformatics* online.

9

## 1 **1 Background**

2 An important goal in systems biology is to elucidate gene regulatory and protein interaction patterns. To this  
3 end, in the literature many network models have been proposed, such as Relevance networks (RNs) [1],  
4 Bayesian networks (BNs) [2], and Gaussian graphical models (GGMs) [3]. For more details, see [4].

5 A GGM consists of a network structure of nodes representing transcripts, metabolites, or proteins, which  
6 are inter-connected by edges reflecting significant partial correlations. Partial correlations measure linear  
7 associations between pairs of random variables, where the contribution from the remaining variables is ad-  
8 justed for. Unlike RNs, which are based on Pearson's correlation, GGMs remove the spurious patterns  
9 caused by confounded variables (e.g. when genes share a common regulator). Compared to BNs, GGMs  
10 scale up more efficiently to large network analyses (i.e. to omics' data) and often yield comparable network  
11 reconstruction accuracies [5]. Although the edges in a GGM are undirected, there are various methods to  
12 learn their directions [6, 7]. These aforementioned features made GGMs a popular tool in bioinformatics and  
13 biomedical studies of colon cancer [8], immunological diseases [9], diabetes [10], respiratory diseases [11],  
14 functional connectivity between brain regions [12], and chronic mental disorders [13].

15 Partial correlations can be computed from the (standardized) inverse of the covariance matrix (i.e. the pre-  
16 cision matrix). In principle, the covariance matrix is unknown and has to be estimated from data. The esti-  
17 mated covariance matrix must be well-conditioned to ensure that its inverse exists and that numerical (or  
18 estimation) errors are not magnified during its computation. The sample covariance, as obtained from a da-  
19 taset of  $n$  samples and  $p$  variables, is (i) invertible and well-conditioned when  $n$  is greater than  $p$ , (ii) inverti-  
20 ble but ill-conditioned when  $n$  is comparable to  $p$ , and (iii) not invertible when  $n$  is smaller than  $p$  [14]. The  
21 last case is known as a 'high-dimensional problem', 'small  $n$ , large  $p$ ', or ' $n \ll p$ '. This scenario is common  
22 in omics' studies, where often a large set of genes, proteins or metabolites is quantified in relatively few  
23 samples.

24 Shrinkage approaches are widely applied to deal with the 'high-dimensional problem'. They produce a  
25 more stable estimator at the cost of some bias. The most popular shrinkage approaches for estimating GGMs  
26 are Glasso [15] and the Ledoit-Wolf (LW) shrinkage [14, 16]. Glasso estimates a 'shrunk' precision matrix  
27 using L1 regularization, which forces some entries to be equal to zero. The LW-shrinkage estimates a  
28 'shrunk' covariance (or 'shrunk' correlation) matrix, which is invertible and henceforth allows the indirect

1 computation of the partial correlations. Although both approaches have been successfully applied in bioin-  
2 formatics, to the best of our knowledge, it has not been studied in the literature yet how the shrinkage af-  
3 fects/biases the estimated partial correlations.

4 In this paper, we present an improvement for the ‘shrunk’ partial correlations inferred with the LW-  
5 shrinkage [17]. In our previous work we focused on the estimation of shrinkage-based p-values [18], and  
6 here we study a fundamental source of bias, namely the effect of the shrinkage value on the ‘shrunk’ partial  
7 correlation. Most importantly, we show that this effect is non-linear, so that not only the magnitudes but also  
8 the order of the estimated partial correlations changes with the shrinkage value. This has unexpected conse-  
9 quences on the results, as GGMs learnt from different experiments (e.g. differing in their sample sizes or  
10 number of genes) have unequal shrinkage values, and thus are affected differently. Therefore, to compare  
11 studies, or to decide whether partial correlations are relevant associations, the users require (i) a ‘shrunk’ test  
12 of significance, and (ii) an accurate assessment of the ‘shrunk’ partial correlation coefficients. The first was  
13 addressed in our recent study [18], and the second is the focus of this article.

## 14 **2 Methods**

15 In this section, we review Gaussian graphical models (GGMs) with the Ledoit-Wolf (LW) shrinkage [14,  
16 16]. We illustrate how the shrinkage modifies the network structure (i.e. the magnitudes and orders of the  
17 partial correlations) in a non-linear way. To overcome this pitfall, we propose the ‘un-shrunk’ partial correla-  
18 tion and discuss its properties. Throughout the text, uppercase bold letters are used for matrices (e.g.  $\mathbf{P}$  is the  
19 matrix of partial correlations) and the *hat* symbol denotes statistical estimators (e.g.  $\hat{\mathbf{P}}$  is an estimator of  $\mathbf{P}$ ).

### 20 **2.1 The ‘shrunk’ partial correlation**

21 The partial correlation is a measure of (linear) association between Gaussian variables, where confounding  
22 effects coming from the other variables are removed (i.e. a full-conditional correlation). A GGM is repre-  
23 sented by a matrix  $\mathbf{P}$  of partial correlations (Whittaker, 1990). The matrix element  $\mathbf{P}_{ij}$  is the partial correla-  
24 tion between the variables  $i$  and  $j$  and can be computed via the relationship

$$25 \mathbf{P}_{ij} = \frac{-\Omega_{ij}}{\sqrt{\Omega_{ii}\Omega_{jj}}} \quad (1)$$

1 where  $\mathbf{\Omega}$  is the inverse of the covariance matrix  $\mathbf{C}$  (or equivalently, the inverse of the correlation matrix  $\mathbf{R}$ ,  
 2 see **Supplementary material S1**). For a dataset  $\mathbf{D}$  that consists of  $p$  variables and  $n$  samples,  $\mathbf{C}$  is a  $p \times p$   
 3 matrix that can be estimated from data, e.g. by the sample covariance matrix  $\hat{\mathbf{C}}^{\text{SM}}$ . However, estimating  $\mathbf{C}$  is  
 4 challenging when  $n$  is comparable to or smaller than  $p$ , as the estimator then becomes ill-conditioned (numer-  
 5 ically unstable) or, even worse, non-invertible.

6

7 The LW-shrinkage estimator  $\hat{\mathbf{C}}^{[\lambda]}$  consists of a convex linear combination of  $\hat{\mathbf{C}}^{\text{SM}}$  and a target matrix  $\mathbf{T}$   
 8 (e.g. a diagonal matrix of variances), and it is defined as

9

$$\hat{\mathbf{C}}^{[\lambda]} := (1 - \lambda)\hat{\mathbf{C}}^{\text{SM}} + \lambda\mathbf{T} \quad (2)$$

10

11 where  $\lambda \in (0, 1)$  represents the weight allocated to  $\mathbf{T}$ . The inverse of  $\hat{\mathbf{C}}^{[\lambda]}$ , denoted by  $\hat{\mathbf{\Omega}}^{[\lambda]}$ , can then be  
 12 plugged into **Equation (1)**, yielding

13

$$\hat{\mathbf{P}}^{[\lambda]}_{ij} = \frac{-\hat{\mathbf{\Omega}}^{[\lambda]}_{ij}}{\sqrt{\hat{\mathbf{\Omega}}^{[\lambda]}_{ii}\hat{\mathbf{\Omega}}^{[\lambda]}_{jj}}} \quad (3)$$

14

15 This is the ‘shrunk’ partial correlation [17], and an edge in the GGM structure is selected according to its  
 16 magnitude and/or statistical significance. The operations involved in **Equation (3)** (i.e. matrix inversion,  
 17 square roots, and standardization) suggest that  $\hat{\mathbf{P}}^{[\lambda]}_{ij}$  is a non-linear function of  $\lambda$ . In addition, the value of  $\lambda$  is  
 18 usually optimized by minimizing the mean square error between  $\hat{\mathbf{C}}^{[\lambda]}$  and  $\mathbf{C}$ , but this  $\lambda$  does not necessarily  
 19 minimize the MSE between  $\hat{\mathbf{\Omega}}^{[\lambda]}$  and  $\mathbf{\Omega}$ .

## 20 **Pitfalls of the ‘shrunk’ partial correlation**

21 Let us consider the following covariance matrix  $\mathbf{C}$  and its inverse  $\mathbf{\Omega}$ ,

22

$$\mathbf{C} = \begin{pmatrix} 1 & \frac{1}{2} & \frac{-1}{4} & \frac{-1}{8} \\ \frac{1}{2} & 1 & \frac{-3}{4} & \frac{-3}{4} \\ \frac{-1}{4} & \frac{-3}{4} & 1 & \frac{3}{4} \\ \frac{-1}{8} & \frac{-3}{4} & \frac{3}{4} & 1 \end{pmatrix}, \quad \mathbf{\Omega} = \begin{pmatrix} \frac{160}{97} & \frac{-152}{97} & \frac{-8}{97} & \frac{-88}{97} \\ \frac{-152}{97} & \frac{416}{97} & \frac{124}{97} & \frac{200}{97} \\ \frac{-8}{97} & \frac{124}{97} & \frac{272}{97} & \frac{-112}{97} \\ \frac{-88}{97} & \frac{200}{97} & \frac{-112}{97} & \frac{320}{97} \end{pmatrix} \quad (4)$$

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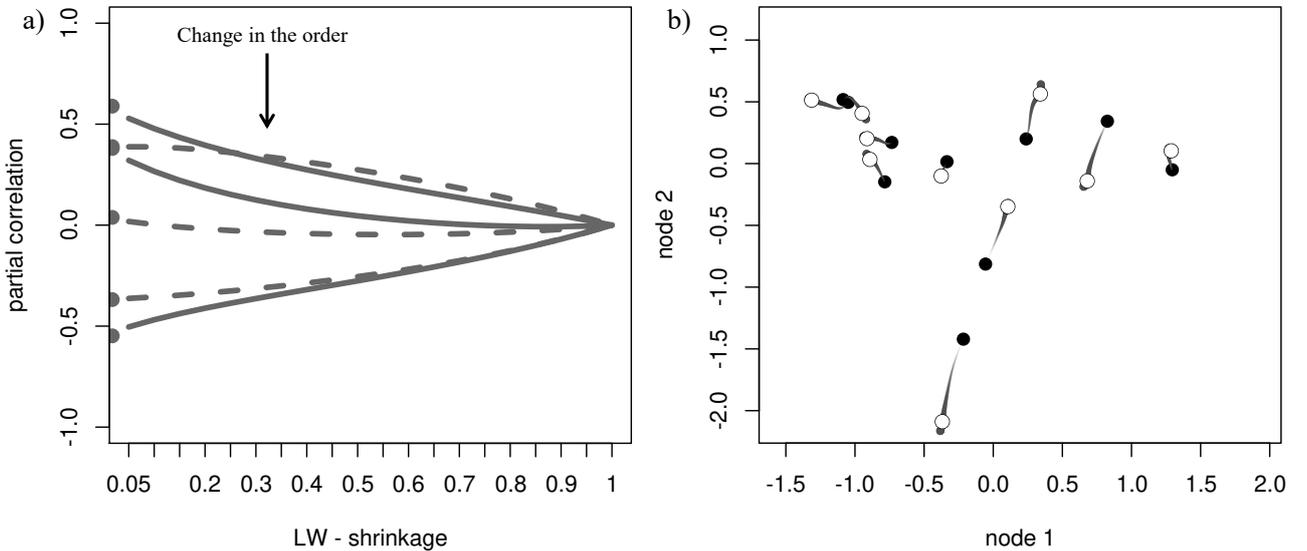
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The matrix  $\mathbf{C}$  is invertible, its eigenvalues are 2.66, 0.94, 0.25, and 0.15, its determinant is 0.09, and its condition number 16.89. From **Equation (1)** we find the partial correlations  $\mathbf{P}_{12} = 152/\sqrt{160}\sqrt{416} = 0.63$  and  $\mathbf{P}_{34} = 112/\sqrt{272}\sqrt{320} = 0.37$ , concluding that  $\mathbf{P}_{12}$  is *stronger* than  $\mathbf{P}_{34}$ . On the other hand, computing  $\hat{\mathbf{C}}^{[\lambda]}$  with **Equation (2)** and substituting it into **Equation (3)** gives the ‘shrunk’ partial correlations. As  $\lambda$  increases, the value of the two ‘shrunk’ partial correlations change. **Figure 1a** shows that for  $\lambda$  greater than 0.3,  $\hat{\mathbf{P}}^{[\lambda]}_{12}$  gets *weaker* than  $\hat{\mathbf{P}}^{[\lambda]}_{34}$  and their relative order reverses. Although **Equation (2)** is a linear shrinkage of  $\hat{\mathbf{C}}^{[\lambda]}$ , the matrix inversion and the standardization in **Equation (3)** propagate the effect from  $\lambda$  to  $\mathbf{P}^{[\lambda]}$  in a non-linear way.



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**Fig. 1.** Non-linear effects of the shrinkage. Panel a) partial correlations obtained from **Equations (3-4)** while varying the shrinkage value. Lines represent ‘shrunk’ partial correlations, and their intersection reflects the changes in their order. Panel b) scatter plot of ‘shrunk’ data points (for two variables) that change as the shrinkage increases. We simulated data ( $n = 10$ ) from a random network ( $p = 8$ ), and the effect of the shrinkage at the data level is obtained via the Singular Value Decomposition of the data matrix, see **Supplementary material S3**. In black: the original data points. In grey: the data points changing their positions for  $\lambda \in (0, 1)$ . The dots’ sizes are proportional to  $\lambda$ . In white: the ‘shrunk’ data points for the optimal shrinkage of 0.65.

## 1 2.2 Additional properties

2 Without loss of generality, we now switch to the correlation matrix  $\mathbf{R}$  (the standardized  $\mathbf{C}$ ). Using **Equation**  
3 **(2)**,  $\mathbf{R}$  (instead of  $\mathbf{C}$ ) can be ‘shrunk’ towards its diagonal ( $\mathbf{T}$  is the identity matrix). For small samples sizes,  
4 the sample correlation  $\widehat{\mathbf{R}}^{\text{SM}}$  (the standardized  $\widehat{\mathbf{C}}^{\text{SM}}$ ) is not positive definite. Some eigenvalues of  $\widehat{\mathbf{R}}^{\text{SM}}$  can be  
5 (i) near to zero, (ii) equal to zero, or (iii) slightly negative. This translates into  $\widehat{\mathbf{R}}^{\text{SM}}$  being (i) ill-conditioned,  
6 (ii) singular, or (iii) indefinite, respectively. The case of an indefinite matrix arises from a numerical inaccu-  
7 racy that can make zero eigenvalues slightly negative.

8

9 Let  $\alpha_k$  ( $k = 1, 2, \dots, p$ ) denote the eigenvalues of  $\widehat{\mathbf{R}}^{\text{SM}}$ . **Equation (2)** yields  $\widehat{\mathbf{R}}^{[\lambda]}$ , which has eigenvalues:

10

$$\alpha_k^{[\lambda]} = (1 - \lambda)\alpha_k + \lambda \quad (k = 1, 2, \dots, p) \quad (5)$$

11

12 and the shrinkage  $\lambda \in (0, 1)$  transforms each  $\alpha_k$  into a positive  $\alpha_k^{[\lambda]}$ , so that  $\widehat{\mathbf{R}}^{[\lambda]}$  becomes positive definite  
13 (see **Supplementary material S2**). Accordingly, the eigenvalues of  $\widehat{\mathbf{R}}^{[\lambda]-1}$  that are obtained by inversion:

14

$$\frac{1}{\alpha_k^{[\lambda]}} = \frac{1}{(1 - \lambda)\alpha_k + \lambda} \quad (k = 1, 2, \dots, p) \quad (6)$$

15 are positive as well. This ensures that  $\widehat{\mathbf{R}}^{[\lambda]-1}$  is a symmetric positive definite precision matrix (i.e. the covar-  
16 iance matrix of the full-conditioned random variables).

### 17 **Shrinking the data**

18 Traditionally, the shrinkage is interpreted as a modification of the statistical model (a ‘shrunk’ covari-  
19 ance/correlation/partial correlation), where the data remains unchanged. However, most research questions  
20 need to be interpreted in terms of the dataset. We therefore propose to discuss the shrinkage from a different  
21 perspective, namely from the data level. To this end, we use the Singular Value Decomposition (SVD) of the  
22 data matrix  $\mathbf{D}$  and the fact that the shrinkage only modifies the eigenvalues, while the eigenvectors stay iden-  
23 tical, see **Supplementary material S2**. As singular values are the positive square roots of the eigenvalues  
24  $\alpha^{[\lambda]}$  given in **Equation (5)**, we can derive the SVD of the ‘shrunk’ data matrix  $\mathbf{D}^{[\lambda]}$  as

1

$$\mathbf{D}^{[\lambda]} = \mathbf{U} \text{diag} \left( \sqrt{(n-1)\alpha^{[\lambda]}} \right) \mathbf{V}^t \quad (7)$$

2

3 Here  $\mathbf{U}$  and  $\mathbf{V}$  are the matrices of (left and right) singular vectors of  $\mathbf{D}$ , and the singular values are replaced  
 4 by their ‘shrunk’ counterparts. This relationship allows us to study the shrinkage effect at the data level. That  
 5 is, analyzing the original dataset  $\mathbf{D}$  with a ‘shrunk’ model  $\hat{\mathbf{C}}^{[\lambda]}$  is equivalent to analyzing  $\mathbf{D}^{[\lambda]}$  with the classi-  
 6 cal model  $\hat{\mathbf{C}}^{\text{SM}}$ . To illustrate this, we generate data from a network ( $p = 8, n = 10$ ) and investigate what hap-  
 7 pens to the original data points as the shrinkage increases; see **Figure 1b** and **Supplementary material S3**  
 8 for more details.

### 9 2.3 The ‘un-shrunk’ partial correlations

10 Here we propose the concept of ‘un-shrunk’ partial correlation, which we define as the limit of  $\hat{\mathbf{P}}^{[\lambda]}_{ij}$  as  $\lambda$   
 11 approaches zero,

12

$$\mathbf{P}^{[0]}_{ij} := \lim_{\lambda \rightarrow 0} \mathbf{P}^{[\lambda]}_{ij} \quad (8)$$

13

14 where  $\mathbf{P}^{[\lambda]}_{ij}$  is a continuous and bounded function of  $\lambda$ , see **Supplementary material S4**. For a general proof  
 15 of the existence of this limit, see **Supplementary material S5**. The key idea is that there is no divergence in  
 16 **Equation (8)**, and to illustrate this we consider the eigen-decomposition,

17

$$\hat{\mathbf{R}}^{[\lambda]-1} = \mathbf{V} \text{diag} \left( \frac{1}{\alpha^{[\lambda]}} \right) \mathbf{V}^t \quad (9)$$

18

19 where  $\mathbf{V}$  is a matrix, whose columns are the eigenvectors of  $\hat{\mathbf{R}}^{[\lambda]}$ , and  $\text{diag}(1/\alpha^{[\lambda]})$  is the diagonal matrix of  
 20 eigenvalues  $1/\alpha^{[\lambda]}_k$  ( $k = 1, 2, \dots, p$ ).

21

22 Let us assume that  $\hat{\mathbf{R}}^{\text{SM}}$  is singular, e.g.  $\alpha_k = 0$ , and recall two facts from the previous subsection. First,  
 23 any  $\alpha_k = 0$  is transformed into  $\alpha^{[\lambda]}_k = \lambda$  in **Equation (5)**, and the corresponding eigenvalue of  $\hat{\mathbf{R}}^{[\lambda]-1}$  is

1  $1/\alpha_k^{[\lambda]} = 1/\lambda$  by **Equation (6)**. Second,  $\widehat{\mathbf{R}}^{\text{SM}}$  and  $\widehat{\mathbf{R}}^{[\lambda]}$  have the same eigenvectors, because the shrinkage  
 2 only changes the eigenvalues (see **Supplementary material S2**). Substituting **Equation (9)** in **Equation (3)**,  
 3 and factorizing out the term  $1/\lambda$  gives

$$\mathbf{P}^{[\lambda]}_{ij} = \frac{-\frac{1}{\lambda} [\mathbf{V} \text{diag}(\frac{\lambda}{\alpha^{[\lambda]}}) \mathbf{V}^t]_{ij}}{\frac{1}{\lambda} \sqrt{[\mathbf{V} \text{diag}(\frac{\lambda}{\alpha^{[\lambda]}}) \mathbf{V}^t]_{ii} [\mathbf{V} \text{diag}(\frac{\lambda}{\alpha^{[\lambda]}}) \mathbf{V}^t]_{jj}}} \quad (10)$$

5  
 6 Any singularity disappears by cancelling the term  $1/\lambda$ . As  $\alpha_k^{[\lambda]} > \lambda$ , the diagonal elements  $\lambda/\alpha_k^{[\lambda]}$  in **Equa-**  
 7 **tion (10)** have limits equal to (i) zero for  $\alpha_k \neq 0$ , or (ii) one for  $\alpha_k = 0$ , see **Equations (S43, S48, S52, S55,**  
 8 **S58)**. In this sense, the ratio that defines the ‘shrunk’ partial correlation in **Equation (10)** does not diverge  
 9 when removing the shrinkage. For a general proof of the existence of this limit, see **Supplementary materi-**  
 10 **al S5**. We propose this limit as a generalization of the classical partial correlation. The idea resembles the  
 11 classical example from Calculus, where the limits to zero of  $x^{-1}$  and  $x^{-2}$  are both infinite; while the limit of  
 12 their ratio  $x^{-1}/x^{-2}$  is finite (i.e. zero). For illustration, let us consider a  $3 \times 3$  correlation matrix of ones (all  
 13 variables are maximally correlated). The matrix is singular as all  $\alpha_k = 0$  and **Equation (2)** gives

$$\mathbf{R}^{[\lambda]} = \begin{pmatrix} 1 & (1-\lambda) & (1-\lambda) \\ (1-\lambda) & 1 & (1-\lambda) \\ (1-\lambda) & (1-\lambda) & 1 \end{pmatrix} \quad (11)$$

15 using **Equation (3)** we obtain

$$\mathbf{P}^{[\lambda]} = \frac{(1-\lambda)}{\frac{1}{\lambda}} \begin{pmatrix} 1 & (2-\lambda)^{-1} & (2-\lambda)^{-1} \\ (2-\lambda)^{-1} & 1 & (2-\lambda)^{-1} \\ (2-\lambda)^{-1} & (2-\lambda)^{-1} & 1 \end{pmatrix} \quad (12)$$

17  
 18 and we see that  $\mathbf{P}^{[0]}_{ij} = \lim_{\lambda \rightarrow 0} \mathbf{P}^{[\lambda]}_{ij} = 1/2$ . Here it is worth noting that a simple linear re-scaling by  $(1-\lambda)$  is  
 19 *not sufficient* to remove the shrinkage effect (see **Figure S1**). In this example,  $\mathbf{P}^{[\lambda]}_{ij}$  would become  $1/(2-$

1  $\lambda$ ) which for  $\lambda = 1/3$  gives  $\mathbf{P}^{[1/3]}_{ij} = 3/5 \approx 0.6$ , and for  $\lambda = 2/3$  is  $\mathbf{P}^{[2/3]}_{ij} = 3/4 \approx 0.75$ . More toy exam-  
2 ples can be found in **Supplementary material S6**.

### 3 2.4 Practical implementation

4 From a mathematical perspective, **Equation (8)** can be computed by means of the analytical results in **Equa-**  
5 **tions (S43, S48, S52, S55, S58)**. However, as numerical inaccuracies make the elements of  $\mathbf{V}$  unreliable, and  
6 often render zero eigenvalues (slightly) positive/negative, these results are un-practical. To circumvent nu-  
7 merical issues, we apply a simple approximation. We compute the ‘shrunk’ partial correlation  $\mathbf{P}^{[\lambda]}_{ij}$  for dif-  
8 ferent  $\lambda \in (0, 1)$  values, and then we fit a polynomial function to the points. By extrapolating the polynomial  
9 to  $\lambda = 0$  the limit in **Equation (8)** is approximated. For more details we refer to **Supplementary material**  
10 **S7**. All computations are performed with the R package *GeneNet* version 1.2.13.

## 11 3 Data

### 12 3.1 *Escherichia coli* microarray data

13 This data set consists of *Escherichia coli* microarray gene-expression measurements [19] from a study of the  
14 temporal stress response upon expression of recombinant human superoxide dismutase (SOD). SOD expres-  
15 sion was induced by isopropyl  $\beta$ -D-1-thiogalactopyranoside (IPTG), which is a lactose analogue inducer of  
16 the lac operon, and measured at 8, 15, 22, 45, 68, 90, 150, and 180 minutes. The authors identified 102 out of  
17 4289 protein coding genes as differentially expressed in one or more samples after induction. Data pre-  
18 processing included  $\log_2$ -ratio transformation with respect to the first time point. The final data set consists  
19 of expression values of 102 genes with 9 time points and was obtained from the R package *GeneNet* version  
20 1.2.13. Accessed May 15, 2020.

### 21 3.2 *Mus musculus* RNA sequencing data

22 This dataset corresponds to single end RNA-Seq reads from 21 male mice from two strains (B6,  $n = 10$  and  
23 D2,  $n = 11$ ), and is available at ReCount: [http://bowtie-](http://bowtie-bio.sourceforge.net/recount/ExpressionSets/bottomly_eset.RData)  
24 [bio.sourceforge.net/recount/ExpressionSets/bottomly\\_eset.RData](http://bowtie-bio.sourceforge.net/recount/ExpressionSets/bottomly_eset.RData) [20]. Accessed May 15, 2020. Genes with  
25 low count’s averages across samples (less than 5) were excluded. After correcting by strain type, 223 genes

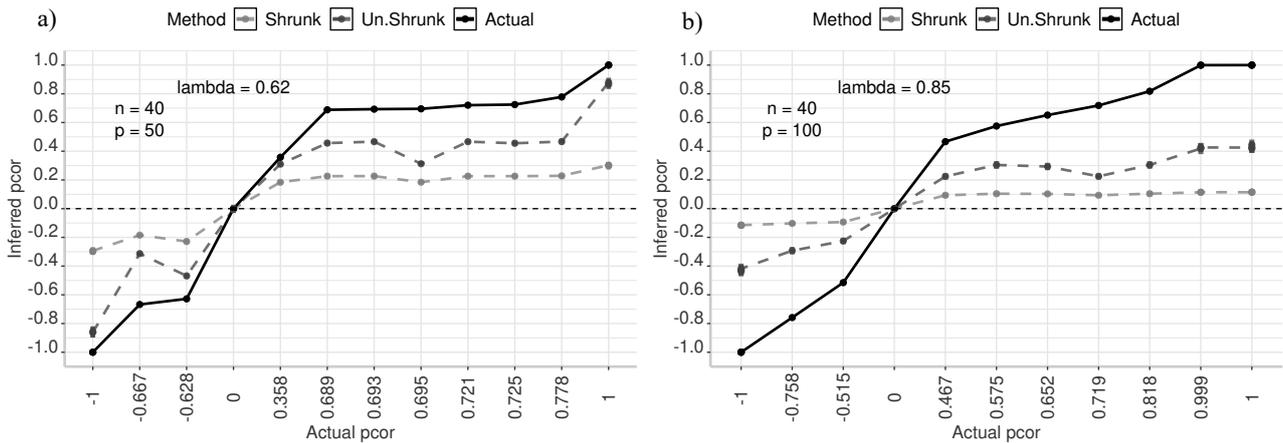
1 out of 9431 were identified as differentially expressed using the R package *limma* and Benjamini-Hochberg  
 2 (BH) adjusted  $p$ -values  $< 0.05$  [21, 22]. We applied upper quartile normalization,  $\log_2$ -transformation and a  
 3 correction by strain type using linear models. The final data set consists of 223 genes with 21 samples.

## 4 Results

### 4.1 Analysis of simulated data

6 Here we demonstrate the advantages of the new ‘un-shrunk’ estimator over the ‘shrunk’ estimator that uses a  
 7 fixed shrinkage value  $\lambda$ . The evaluation consists of comparing the reconstructed partial correlations to their  
 8 actual (population) values. In total, 1251 data sets were simulated (see **Table S2**).

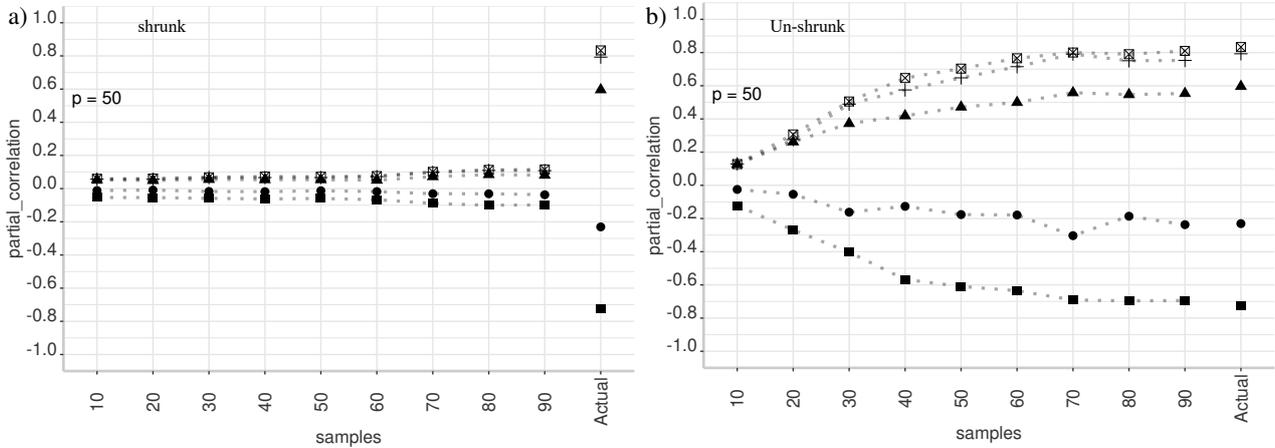
9 First, we study the shrinkage distortion for partial correlations coefficient with magnitudes between -1 and  
 10 1. To this end, we simulate two networks and 25 datasets in the high dimensional (or  $n \ll p$ ) scenario ( $p =$   
 11 50, 100, and  $n = 40$ ). **Figure 2** presents the partial correlations estimated with (i) the optimal shrinkage, (ii)  
 12 the new ‘un-shrunk’ approach, and (iii) their actual values. It can be seen that the new ‘un-shrunk’ coeffi-  
 13 cients are consistently closer to the actual values than the ‘shrunk’ coefficients. In **Figure S2** we see how a  
 14 GGM structure (i.e. its edges) changes with varying  $\lambda$ . Here we used  $p = 10$  and  $n = 1000$  (a large sample  
 15 size) to reduce the sampling variability, such that the observed effect on the edges order can be attributed to  
 16  $\lambda$ . **Table S1** lists the edges sorted by their magnitudes.



17

18 **Fig. 2.** Partial correlations versus their actual (population) values. We simulated 25 datasets from two network structures ( $n = 40$ ) and  
 19 reconstructed the ‘shrunk’ and ‘un-shrunk’ partial correlations. Panel a) corresponds to  $p = 50$ , and panel b) to  $p = 100$ . In black: the  
 20 actual partial correlations. In light grey (dashed): the average of the ‘shrunk’ method. In dark grey (dashed): the average of the new  
 21 ‘un-shrunk’ method. Error bars represent 2 standard errors.  
 22

1 Second, we study whether GGMs from different experimental conditions are comparable. We create a  
 2 (random) network structure and simulate 10 datasets for each  $n = 10, 20, \dots, 90$ . **Figure 3** shows the ‘shrunk’  
 3 and ‘un-shrunk’ partial correlations as a function of  $n$ .



4  
 5 **Fig. 3.** Partial correlation versus sample size. We create a fixed network structure for  $p = 50$ , and simulated data for sample size  $n$   
 6 ranging from 10 to 90. Panels a, b) show the average ‘shrunk’ and ‘un-shrunk’ partial correlations (over 10 simulations). The pro-  
 7 posed ‘un-shrunk’ partial correlations are closer to their actual values.  
 8

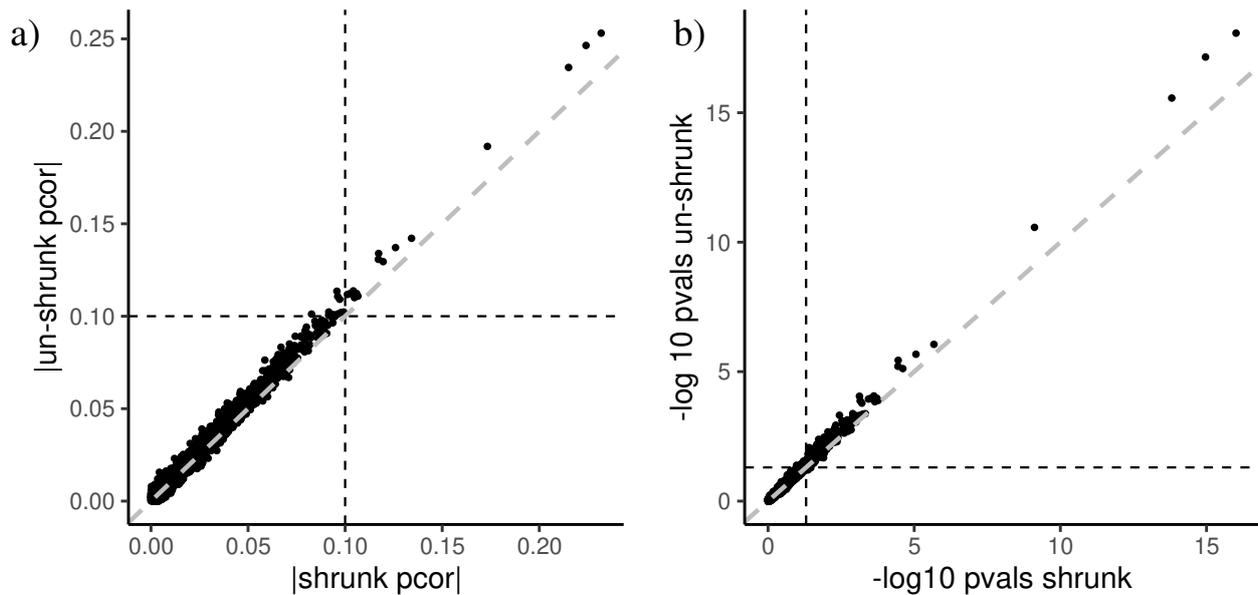
9 The new ‘un-shrunk’ partial correlation approaches the actual (population) values as the sample size increas-  
 10 es, while the shrunk counterpart stays far away from the actual values. **Figure S3** compares the performance  
 11 of both methods for different combinations of  $p$  and  $n$  for an actual partial correlation of 0.5. The results are  
 12 presented in the form of a heatmap over a  $p$ - $n$  grid. The color scale shows the L1 distances to the population  
 13 value of 0.5. We see that the proposed ‘un-shrunk’ estimation is consistently closer to the population values.  
 14 In **Figure S4** we illustrate how, in general, partial correlations are deflated when the samples size is very  
 15 small, e.g.  $n = 10$ .

## 16 4.2 Analysis of experimental data

### 17 Effects of human superoxide dismutase (SOD) protein expression on transcript expression in *E. coli*

18 Following previous works, the dataset is treated as static and nominal p-values are considered significant at  
 19 the 0.05 level [17, 18]. The ‘shrunk’ approach infers an optimal shrinkage of 0.18. A scatter plot of the par-  
 20 tial correlations in absolute value is presented in **Figure 4a, b**. The panels are segmented into four regions  
 21 using a threshold of  $|pcorr| = 0.1$  or p-values = 0.05. Clustering of points around the diagonal indicates that  
 22 the ‘shrunk’ and ‘un-shrunk’ coefficients (as well as the p-values) are similar.

1 The ‘shrunk’ network has 238 edges (involving 74 genes), and the ‘un-shrunk’ network has 34 additional  
 2 edges, for a total of 272 edges (involving 78 genes). We evaluate the enrichment of the connected genes us-  
 3 ing the protein-protein interaction STRING database version 10 [23, 24]. The enriched Gene Ontologies  
 4 (GOs), and Kyoto Encyclopedia of Genes and Genomes (KEGGs) pathways are reported in **Supplementary**  
 5 **Tables S4 a-d**. The complete set of 102 genes was set as background, which mapped 96 proteins.  
 6



7  
 8 **Fig. 4.** Comparison of partial correlations from *Escherichia coli* microarray data. Panel a) Scatter plot of the partial correlation in  
 9 absolute value |pcorr|. Panel b) Scatter plot of  $-\log_{10}(\text{p-values})$ . P-values were computed with the shrunk approach (Bernal et al.  
 10 2019). The panels are segmented into four regions using a threshold of  $|\text{pcorr}| = 0.1$  or  $\text{p-values} = 0.05$  (in dashed lines). The ‘un-  
 11 shrunk’ model has 34 additional significant edges at  $\text{p-values} < 0.05$ .  
 12

13 The expected number of interactions in STRINGdb for a random set of proteins of similar size was 581. The  
 14 102 genes retrieved 668 interactions ( $p\text{-value} = 0.00025$ ). The connected genes for the ‘shrunk’ method  
 15 mapped onto 69 proteins with 332 interactions (expected = 286,  $p\text{-value} = 0.00438$ ). For the ‘un-shrunk’ it  
 16 mapped onto 73 proteins with 372 interactions (expected = 320,  $p\text{-value} = 0.00272$ ). **Tables S4 a-b** show the  
 17 GO enrichment, where the new the ‘un-shrunk’ method has 9 additional GOs and one KEGG pathway (all  
 18 related to pyrimidine’s metabolism). **Tables S4 c-d** show that the connected genes from both methods are  
 19 enriched for (i) galactitol catabolic process (GO:0019404, FDR = 0.048) and (ii) galactitol metabolic process  
 20 (GO:0019402, FDR = 0.048) and (iii) the galactose metabolism KEGG pathway (FDR =  $1.96 \cdot 10^{-7}$ ,  $1.16 \cdot 10^{-7}$ ).  
 21 The network structures can be found in **Figure S6**.  
 22

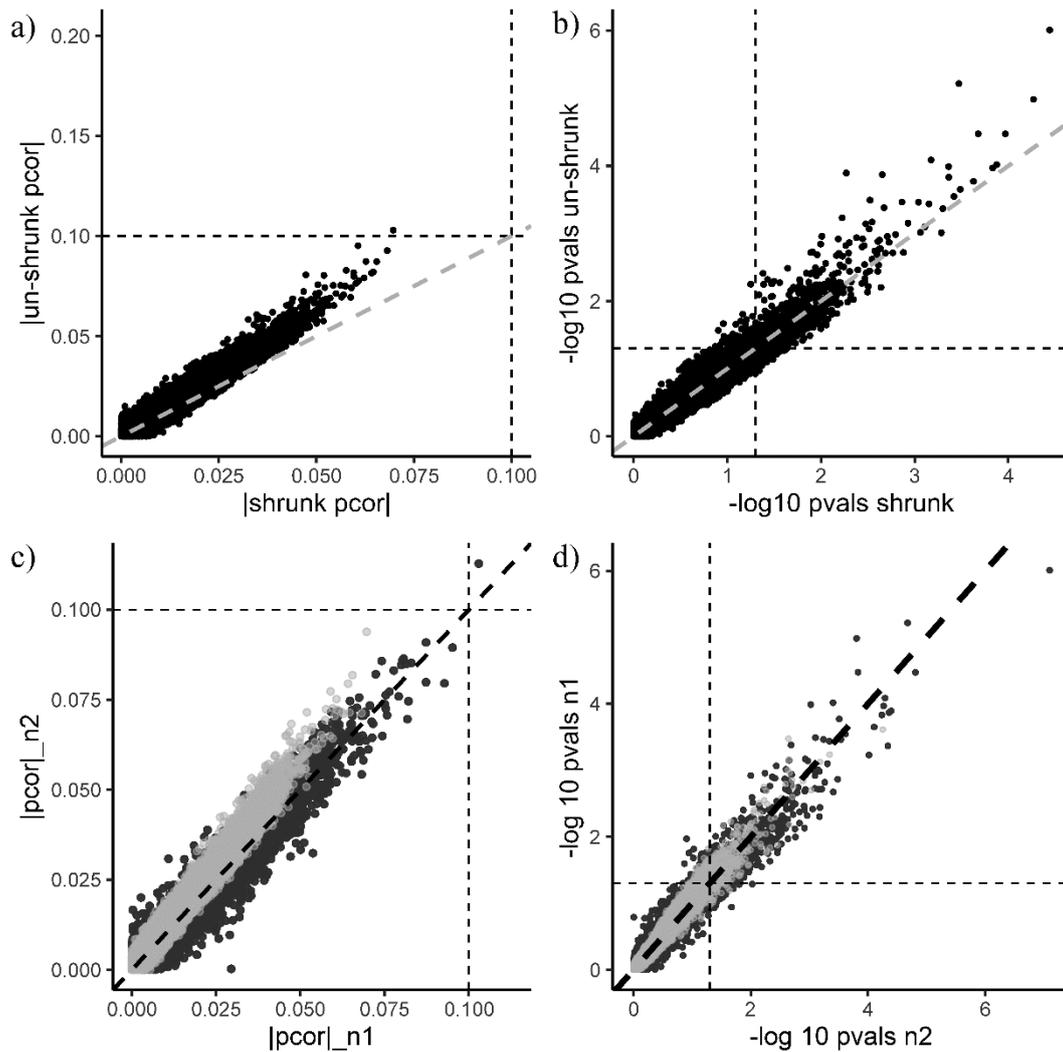
1 **Analysis of *M. musculus* RNA-seq dataset**

2 **Figure 5** presents scatter plots of the partial correlation (in absolute values) and their p-values. The ‘shrunk’  
3 approach infers an optimal shrinkage of 0.69. Points clustering near the diagonal indicate that the partial  
4 correlations are similar in magnitude, while scattering away from it reflects changes in their order.

5

6 The ‘shrunk’ network has no connections, and the ‘un-shrunk’ network has one connection using BH ad-  
7 justed *p-values* < 0.05. As the networks are empty, we proceed with nominal *p-values* at 0.001, where the  
8 ‘shrunk’ and ‘un-shrunk’ networks have 19 and 33 edges, involving 29 and 40 genes, respectively. The 223  
9 genes mapped onto 208 proteins (STRINGdb) with 478 interactions (*p-value* = 0.09), while the expected  
10 number of interactions (for a random set of proteins of similar size) was 449. The connected genes in the  
11 ‘shrunk’ network mapped onto 29 proteins with 3 interactions (expected = 4, *p-value* = 0.84). The new ‘un-  
12 shrunk’ method mapped 36 proteins with 5 interactions (expected = 6, *p-value* = 0.78). The Gene Ontologies  
13 (GOs), and Kyoto Encyclopedia of Genes and Genomes (KEGGs) pathways are reported in Supplementary  
14 **Tables S5 a-d**. The ‘shrunk’ and ‘un-shrunk’ connected genes are enriched for 562 and 647 GOs, and for 7  
15 and 13 KEGGs pathways at FDR 0.05, respectively. The GOs belong mainly to regular cellular processes.  
16 The ‘un-shrunk’ method retrieves (additional) KEGGs related to degenerative diseases (5010: Alzheimer and  
17 5012: Parkinson, both with FDR =  $7.17 \cdot 10^{-4}$ ). The network structure can be found in **Figure S7**.

18



1

2 **Fig. 5.** Comparison of partial correlations from *Mus musculus* RNA-seq data. Panel a) Scatter plot of the partial correlation in absolute value  $|pcorr|$ . Panel b) scatter plot of  $-\log_{10}$  (p-values). Panel c, d) scatter plots of  $|pcorr|$ , and of  $-\log_{10}$  (p-values) for different sample sizes ( $n_1 = 21$ ,  $n_2 = 31$ ). To generate an additional dataset ( $n_2 = 31$ ) from the original one ( $n_1 = 21$ ), we use a sampling approach. The dashed lines show the threshold of  $|pcorr| = 0.1$  and p-values = 0.05, respectively. P-values were computed with the shrunk approach (Bernal et al. 2019). In light grey: the ‘shrunk’ method with  $\lambda = 0.69$  and 0.42. In dark grey: the ‘un-shrunk’ method. The ‘shrunk’ partial correlations suffer a slope shift for different sample sizes and are not comparable. The ‘un-shrunk’ partial correlations are concentrated around the diagonal, what reflects that they are comparable.

3

4 To compare the methods for different shrinkage values, we generate a second dataset as follows: (i) we  
 5 randomly choose 10 samples out of the original 21 samples, (ii) we add Gaussian noise (mean = 0, sd = 0.05)  
 6 to all the expression levels, and (iii) we merged the ten newly generated samples to the original dataset,  
 7 which results in 31 samples. The ‘shrunk’ approach retrieves a new optimal shrinkage of 0.42. Both methods  
 8 were applied to the new data, and plotted against the previous results as shown in **Figure 5c-d**. While the  
 9 shrinkage change (from 0.69 to 0.42) made the ‘shrunk’ method deviate from the diagonal line (and hard to  
 10 compare directly), the new method provides coefficients that are more similar.

11

## 1 **5 Discussion**

2 GGMs are undirected graphical models that represent pairwise partial correlations in the form of a network.  
3 They are widely used in many fields, because they are computationally fast and simple to interpret. Despite  
4 of that, the estimation of partial correlations from gene-expression data is challenging whenever there are  
5 fewer samples than genes. This motivated the development of estimators based on shrinkage; however, we  
6 have observed some unexpected effect of the shrinkage and we have investigated it.

7 In particular, we have identified a bias in the ‘shrunk’ partial correlations. The bias is a non-linear effect  
8 caused by the shrinkage value, which modifies the magnitudes and order of the partial correlations, see **Fig-**  
9 **ure 1, S1, S2**. As the order of the edges is affected non-linearly, re-scaling the ‘shrunk’ partial correlations  
10 (e.g. dividing it by  $1 - \lambda$ ) is not sufficient (see **Figure S1** and **Section 2.3**). Consequently, as GGMs learnt  
11 from different experimental conditions use different shrinkage values, they infer networks that are not com-  
12 parable.

13 To correct for this bias, we have introduced the concept of ‘un-shrinking’ the partial correlation. On the  
14 theoretical side, the ‘un-shrunk’ partial correlation is a generalization of the classical partial correlation, and  
15 it is defined even for singular matrices. On the applied side, the ‘un-shrunk’ partial correlation is easy to  
16 interpret, because its magnitude is between -1 and 1, and it is significantly closer to its actual (population) val-  
17 ue (**Figures 2, 3, and S3**). For the well-conditioned case, we have also shown that the shrinkage can be con-  
18 sidered as a transformation of the original dataset into a new ‘shrunk’ dataset, see **Figure 1b**. Hence, ‘un-  
19 shrinking’ could be interpreted as the limit of the partial correlations when the ‘shrunk’ data points approach  
20 their original values.

21 Our empirical results show that they are local (edge-wise) shrinkage distortions. These can be identified by  
22 comparing the scatter plots of the partial correlation coefficients differing in their shrinkage value. For in-  
23 stance, (i) the slope reflects whether the scales/magnitudes of the coefficients are different, and (ii) points  
24 scattering away from the diagonal line reflect that their order has changed. For the *E. coli* dataset, the strong-  
25 est connections (in both models) were *lacA-lacZ*, *lacY-lacZ*, and *lacA-lacY*, all related to the lac operon (that  
26 was induced by IPTG in the experiment). The result is in agreement with previous analyses [17, 18], howev-  
27 er, it includes 34 additional edges. For the *E. coli*'s and *M. musculus*'s networks in **Figure 4-5** the edges are  
28 highly correlated, but their relative order is different. **Figure 5c** shows that the slope of the ‘shrunk’ coeffi-

1 coefficients (in light grey) is shifted away from the diagonal, and thus the two networks are not comparable. In  
2 contrast, the new ‘un-shrunk’ coefficients (in dark grey) are closer to the diagonal line.

3 A non-linear effect on the partial correlations translates into a non-linear effect on the *p-values*, and conse-  
4 quently affects the inferred network structure. The ‘shrunk’ *p-values* [18] account for the change in the mag-  
5 nitudes, but do not include a correction for the non-linear effects that alter the edge order. This is apparent  
6 from **Figure 5d**, where the *p-values* are around the diagonal line in both cases, but their deviations are dif-  
7 ferent. One may be tempted to say that the ‘shrunk’ points (in light grey) are less dispersed (or more similar)  
8 in **Figure 5d**, however, this is an effect of the variance-bias trade-off. Due to the shrinkage, partial correla-  
9 tions are less variable, but their order is biased.

10 In **Figure S3** we see that for every  $p$  and  $n \geq 30$ , the ‘un-shrunk’ coefficients are closer to their actual val-  
11 ues. From this figure the new method seems particularly superior when  $p / n > 2$ . For very small samples,  
12 e.g.  $n = 10$  or  $20$ , we see that both methods are suboptimal because the model’s assumptions are not neces-  
13 sarily fulfilled, see **Figure S4a**. Very often in bioinformatics the data is transformed e.g. log transformed,  
14 scaled. The transformed data is *approximately* a sample from a Gaussian distribution, where the approxima-  
15 tion improves for larger samples. This mismatch is not necessarily negligible for very small samples. For  
16 instance, the *Law of large numbers* ensures that the sample mean converges to the population mean when  $n$   
17 approaches infinity. The *Central limit theorem* states that the scaled sample mean is *asymptotically* normal,  
18 with an error that depends on  $n$ , see *Berry–Esseen theorem* [25]. The same applies to the sample correlation  
19 [26], and consequently for very small samples the distributional assumptions of GGMs are not always met.

20 To the best of our knowledge, this is the first study aiming to *de-regularize* the estimates. While the ‘un-  
21 shrunk’ partial correlation can be found algebraically (see **Section 2.1**), we employed a polynomial approxi-  
22 mation in order to achieve feasible computational costs for large scale applications. In principle, large sam-  
23 ples can cause weak associations to be statistically significant, and small samples can cause strong associa-  
24 tions to be non-significant. A proper discussion should therefore report the magnitude and significance of the  
25 estimates, as they provide different pieces of information. In this case, it must be accounted for the fact that  
26 the partial correlations are ‘shrunk’ (biased), before concluding about the result. Ignoring the shrinkage value  
27 would divorce the analysis from the original data (and its biological meaning) what could obscure the inter-  
28 pretation, and impede the validation of earlier reported results (e.g. in biomarker’s discovery).

29

1 **Declarations**

2

3 **Ethics approval and consent to participate:** Not applicable

4 **Consent for publication:** Not applicable

5 **Availability of data and materials:** The R code that generates the results is available in  
6 <https://github.com/V-Bernal/UnShrunk>. The datasets supporting the conclusions of this article are available  
7 in the [R package GeneNet version 1.2.13](#) (*E. coli* dataset) and at [http://bowtie-](http://bowtie-bio.sourceforge.net/recount/ExpressionSets/bottomly_eset.RData)  
8 [bio.sourceforge.net/recount/ExpressionSets/bottomly\\_eset.RData](http://bowtie-bio.sourceforge.net/recount/ExpressionSets/bottomly_eset.RData). (*M. musculus* dataset).

9 **Competing interests:** None

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11 versity of Groningen. This research was part of the Netherlands X-omics Initiative and partially funded by  
12 NWO, project 184.034.019.

13 **Authors' contributions:** VB, PH, VG, MG designed the study. VB developed the concept and implemented  
14 the code. VB and MG wrote the mathematical proofs and other theoretical results. VG, RB, and PH contrib-  
15 uted to the analysis and interpretation of the results. All authors participated in the writing of the manuscript.

16 **Acknowledgements:** Not applicable

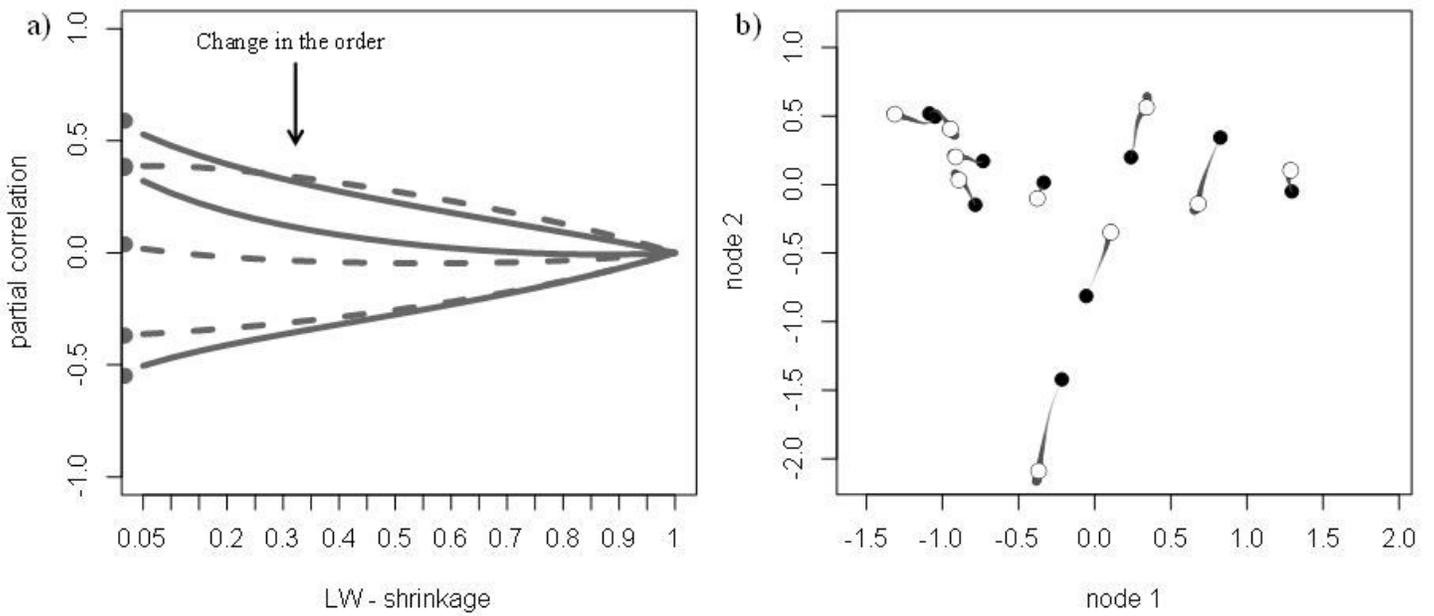
17 **References**

- 18 1. Butte AJ, Kohane IS. Relevance Networks: A First Step Toward Finding Genetic Regulatory Networks  
19 Within Microarray Data. In: Parmigiani G., Garrett E.S., Irizarry R.A. ZSL, editor. The Analysis of  
20 Gene Expression Data. NY: Springer, New York, NY; 2003. p. 428–46.
- 21 2. Friedman N, Linial M, Nachman I, Pe'er D. Using Bayesian Networks to Analyze Expression Data. J  
22 Comput Biol. 2000;7:601–20.
- 23 3. Edwards D. Introduction to Graphical Modelling 2nd edition. 2nd edition. Springer-Verlag New York;  
24 2000.
- 25 4. Huynh-Thu VA, Sanguinetti G. Gene Regulatory Network Inference: An Introductory Survey. In: Gene  
26 Regulatory Networks. 2019. p. 1–23.
- 27 5. Werhli A V., Grzegorzczak M, Husmeier D. Comparative evaluation of reverse engineering gene  
28 regulatory networks with relevance networks, graphical gaussian models and bayesian networks.

- 1        Bioinformatics. 2006;22:2523–31.
- 2    6. Opgen-Rhein R, Strimmer K. Learning causal networks from systems biology time course data: An  
3        effective model selection procedure for the vector autoregressive process. *BMC Bioinformatics*. 2007.
- 4    7. Opgen-Rhein R, Strimmer K. From correlation to causation networks: A simple approximate learning  
5        algorithm and its application to high-dimensional plant gene expression data. *BMC Syst Biol*.  
6        2007;1:37.
- 7    8. Beerenwinkel N, Antal T, Dingli D, Traulsen A, Kinzler KW, Velculescu VE, et al. Genetic progression  
8        and the waiting time to cancer. *PLoS Comput Biol*. 2007;3:e225.
- 9    9. Benedetti E, Pučić-Baković M, Keser T, Wahl A, Hassinen A, Yang JY, et al. Network inference from  
10       glycoproteomics data reveals new reactions in the IgG glycosylation pathway. *Nat Commun*.  
11       2017;8:1483.
- 12   10. Keller MP, Choi Y, Wang P, Davis DB, Rabaglia ME, Oler AT, et al. A gene expression network model  
13       of type 2 diabetes links cell cycle regulation in islets with diabetes susceptibility. *Genome Res*.  
14       2008;:gr. 074914.107.
- 15   11. Imkamp K, Bernal V, Grzegorzcyk M, Horvatovich P, Vermeulen CJ, Heijink IH, et al. Gene network  
16       approach reveals co-expression patterns in nasal and bronchial epithelium. *Sci Rep*. 2019;:1–13.  
17       doi:10.1038/s41598-019-50963-x.
- 18   12. Das A, Sampson AL, Lainscsek C, Muller L, Lin W, Doyle JC, et al. Interpretation of the precision  
19       matrix and its application in estimating sparse brain connectivity during sleep spindles from human  
20       electrocorticography recordings. *Neural Computation*. 2017.
- 21   13. McNally RJ, Robinaugh DJ, Wu GWY, Wang L, Deserno MK, Borsboom D. Mental disorders as causal  
22       systems: A network approach to posttraumatic stress disorder. *Clinical Psychological Science*. 2015.
- 23   14. Ledoit O, Wolf M. A well-conditioned estimator for large-dimensional covariance matrices. *J Multivar*  
24       *Anal*. 2004;88:365–411.
- 25   15. Friedman J, Hastie T, Tibshirani R. Sparse inverse covariance estimation with the graphical lasso.  
26       *Biostatistics*. 2008.
- 27   16. Ledoit O, Wolf M. Improved estimation of the covariance matrix of stock returns with an application to  
28       portfolio selection. *J Empir Financ*. 2003;10:603–21.
- 29   17. Schäfer J, Strimmer K. A Shrinkage Approach to Large-Scale Covariance Matrix Estimation and

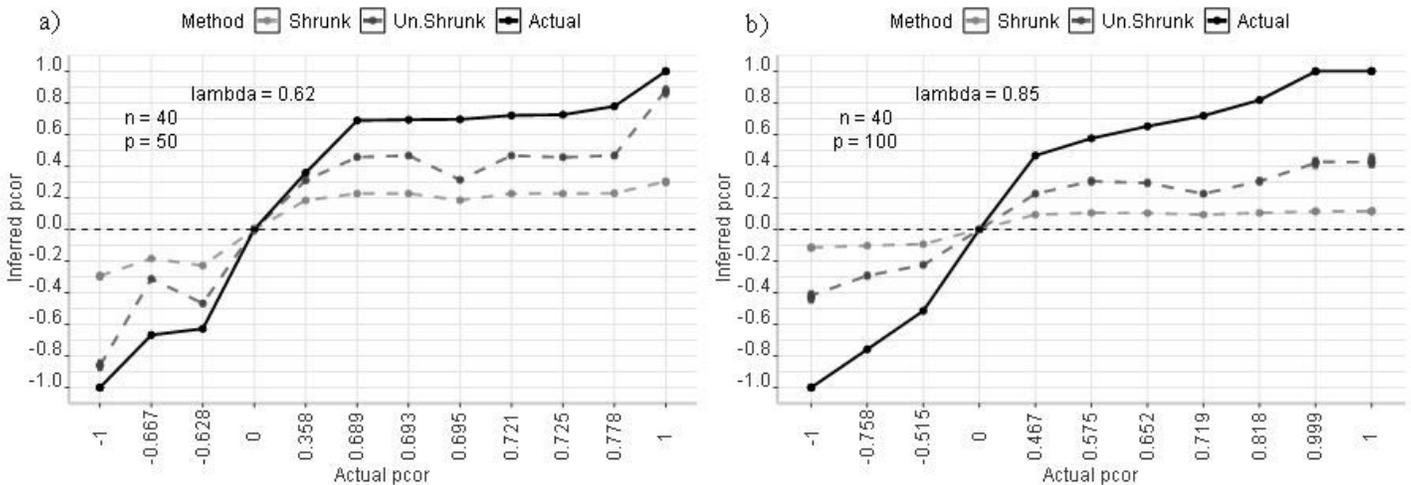
- 1 Implications for Functional Genomics. *Stat Appl Genet Mol Biol*. 2005;4:1175–89. doi:10.2202/1544-  
2 6115.1175.
- 3 18. Bernal V, Bischoff R, Guryev V, Grzegorzczak M, Horvatovich P. Exact hypothesis testing for shrinkage-  
4 based Gaussian graphical models. *Bioinformatics*. 2019; May:1–7.
- 5 19. Schmidt-Heck W, Guthke R, Toepfer S, Reischer H, Duerrschmid K, Bayer K, et al. Reverse engineering  
6 of the stress response during expression of a recombinant protein. *Proc EUNITE Symp*. 2004;;10–12.  
7 <http://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.129.539&rep=rep1&type=pdf>.
- 8 20. Bottomly D, Walter NAR, Hunter JE, Darakjian P, Kawane S, Buck KJ, et al. Evaluating gene  
9 expression in C57BL/6J and DBA/2J mouse striatum using RNA-Seq and microarrays. *PLoS One*.  
10 2011;6.
- 11 21. Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, et al. Limma powers differential expression  
12 analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res*. 2015.
- 13 22. Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to  
14 multiple testing. *Journal of the Royal Statistical Society*. 1995;57:289–300. doi:10.2307/2346101.
- 15 23. Szklarczyk D, Morris JH, Cook H, Kuhn M, Wyder S, Simonovic M, et al. The STRING database in  
16 2017: Quality-controlled protein-protein association networks, made broadly accessible. *Nucleic Acids*  
17 *Res*. 2017;;gkw937.
- 18 24. Franceschini A, Simonovic M, Roth A, Von Mering C, Szklarczyk D, Frankild S, et al. STRING v9.1:  
19 Protein-protein interaction networks, with increased coverage and integration. *Nucleic Acids Res*. 2013.
- 20 25. Berry AC. The Accuracy of the Gaussian Approximation to the Sum of Independent Variates. *Trans Am*  
21 *Math Soc*. 1941.
- 22 26. Zimmerman DW, Zumbo BD, Williams RH. Bias in Estimation and Hypothesis Testing of Correlation.  
23 *Psicológica*. 2003;24.
- 24

# Figures



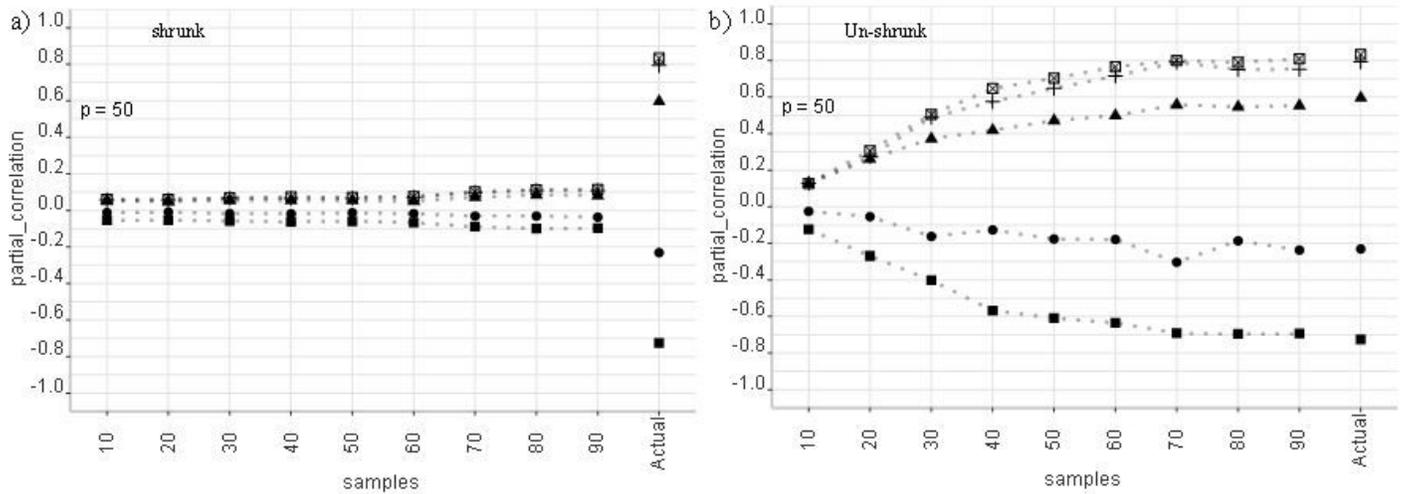
**Figure 1**

Non-linear effects of the shrinkage. Panel a) partial correlations obtained from Equations (3-4) while varying the shrinkage value. Lines represent 'shrunk' partial correlations, and their intersection reflects the changes in their order. Panel b) scatter plot of 'shrunk' data points (for two variables) that change as the shrinkage increases. We simulated data ( $n = 10$ ) from a random network ( $p = 8$ ), and the effect of the shrinkage at the data level is obtained via the Singular Value Decomposition of the data matrix, see Supplementary material S3. In black: the original data points. In grey: the data points changing their positions for  $\lambda \in (0,1)$ . In white: the 'shrunk' data points for the optimal shrinkage of 0.65.



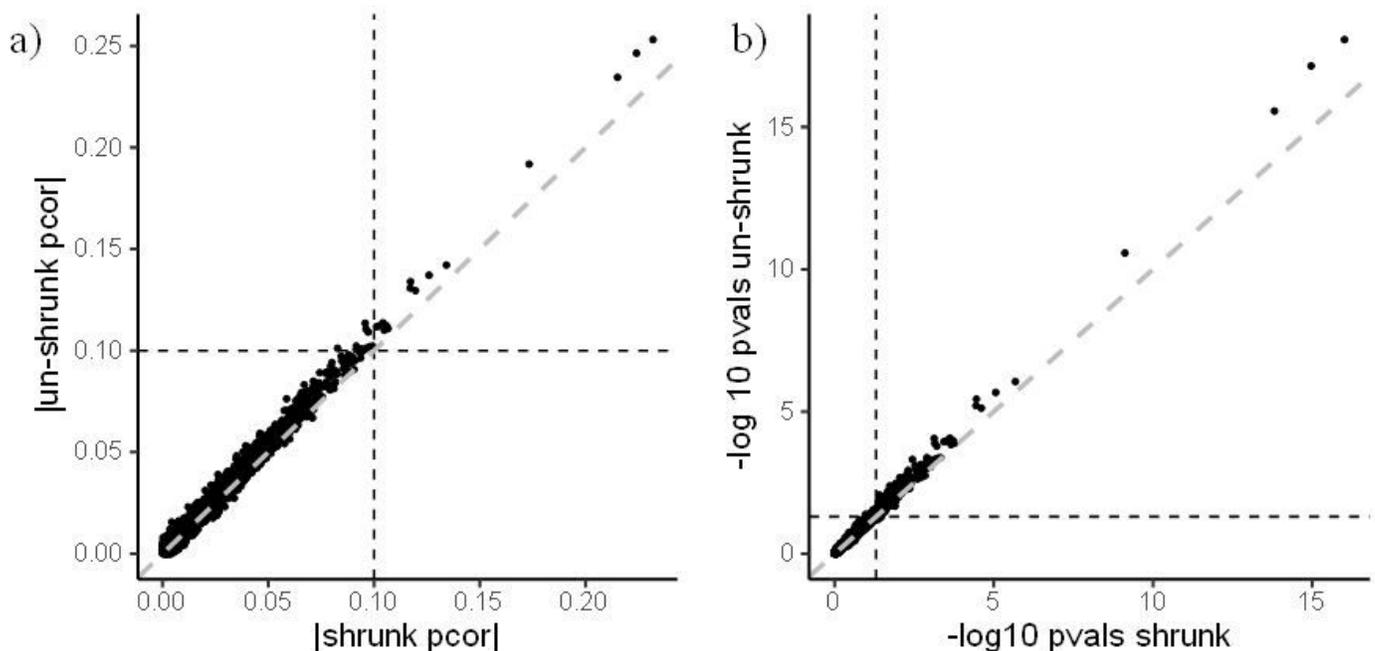
**Figure 2**

Partial correlations versus their actual (population) values. We simulated 25 datasets from two network structures ( $n=40$ ) and reconstructed the 'shrunk' and 'un-shrunk' partial correlations. Panel a) corresponds to  $p=50$ , and panel b) to  $p=100$ . In black: the actual partial correlations. In light grey (dashed): the average of the 'shrunk' method. In dark grey (dashed): the average of the new 'un-shrunk' method. Error bars represent 2 standard errors.



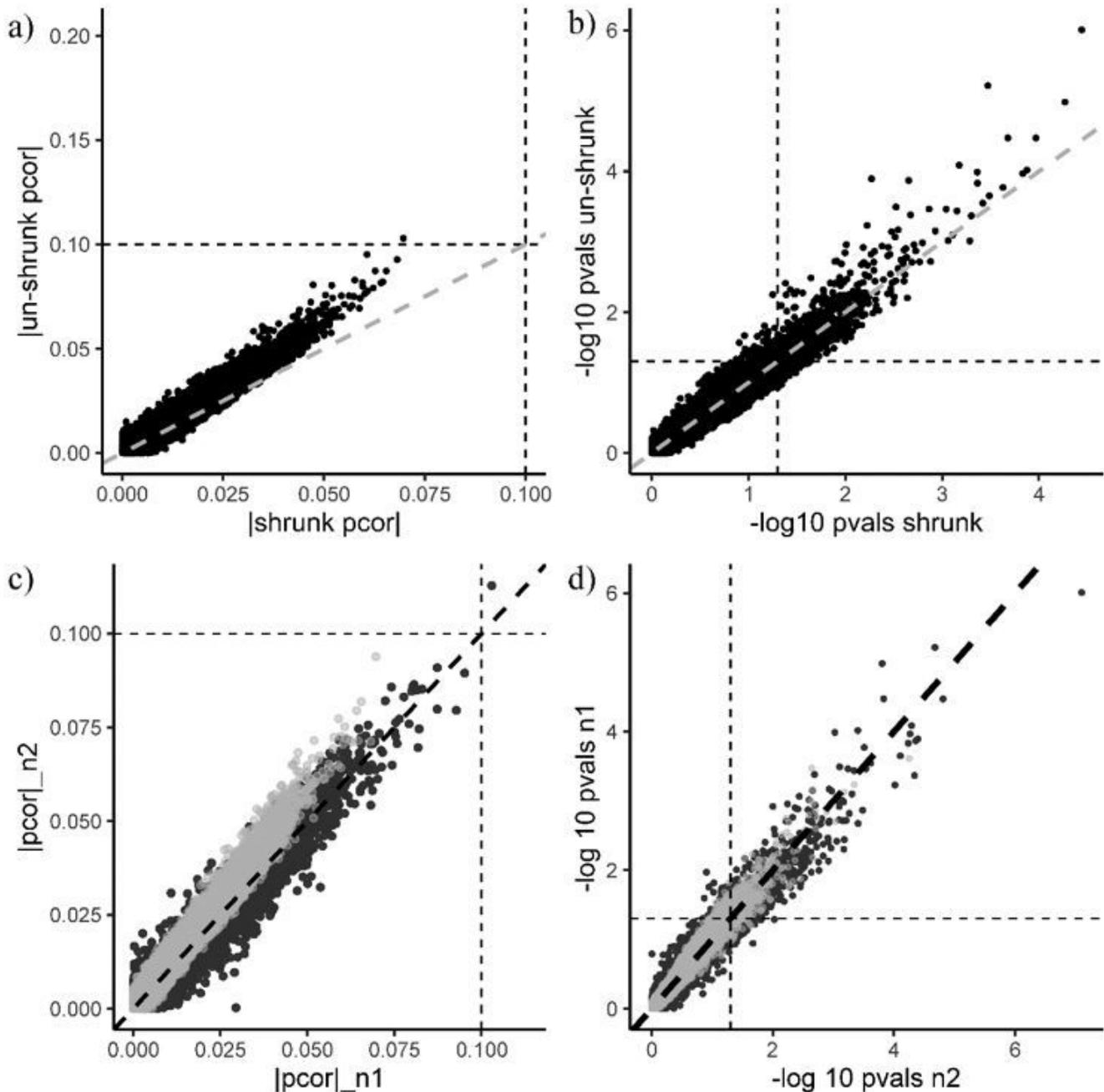
**Figure 3**

Partial correlation versus sample size. We create a fixed network structure for  $p=50$ , and simulated data for sample size  $n$  ranging from 10 to 90. Panels a, b) show the average 'shrunk' and 'un-shrunk' partial correlations (over 10 simulations). The proposed 'un-shrunk' partial correlations are closer to their actual values.



**Figure 4**

Comparison of partial correlations from *Escherichia coli* microarray data. Panel a) Scatter plot of the partial correlation in absolute value  $|pcorr|$ . Panel b) Scatter plot of  $-\log_{10}(p\text{-values})$ . P-values were computed with the shrunk approach (Bernal et al. 2019). The panels are segmented into four regions using a threshold of  $|pcorr| = 0.1$  or  $p\text{-values} = 0.05$  (in dashed lines). The 'un-shrunk' model has 34 additional significant edges at  $p\text{-values} < 0.05$ .



**Figure 5**

Comparison of partial correlations from *Mus musculus* RNA-seq data. Panel a) Scatter plot of the partial correlation in absolute value  $|pcorr|$ . Panel b) scatter plot of  $-\log_{10}(p\text{-values})$ . Panel c, d) scatter plots of  $|pcorr|$ , and of  $-\log_{10}(p\text{-values})$  for different sample sizes ( $n_1 = 21$ ,  $n_2 = 31$ ). To generate an additional dataset ( $n_2 = 31$ ) from the original one ( $n_1 = 21$ ), we use a sampling approach. The dashed lines show

the threshold of  $|\text{pcorr}| = 0.1$  and  $p\text{-values} = 0.05$ , respectively. P-values were computed with the shrunk approach (Bernal et al. 2019). In light grey: the 'shrunk' method with  $\lambda = 0.69$  and  $0.42$ . In dark grey: the 'un-shrunk' method. The 'shrunk' partial correlations suffer a slope shift for different sample sizes and are not comparable. The 'un-shrunk' partial correlations are concentrated around the diagonal, what reflects that they are comparable.

## Supplementary Files

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