

Comparison of amyloid PET measured in Centiloid units with neuropathological findings in Alzheimer's Disease

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Abstract

Background: We aimed to determine the Centiloid unit (CL) thresholds for sparse and moderate density neuritic plaques. **Methods:** Amyloid PET results in CL for 49 subjects were compared with post-mortem neuritic plaque density, visual read, and final clinicopathological diagnosis. A Youden Index was used to determine the optimal CL thresholds from receiver operator characteristic (ROC) curves. **Results:** A threshold of 20.1 CL yielded highest accuracy in detecting moderate or frequent plaque density (ROC AUC 0.97). A threshold of 9.5 CL was optimal for detecting sparse, moderate or frequent plaques (ROC AUC 0.96). Those cases with a final clinicopathological diagnosis of Alzheimer's disease yielded a median CL result of 87.7 (IQR \pm 42.2) with 94% > 45 CL. Positive visual read agreed highly with results >26 CL. **Conclusions:** In this cohort, values <9.5 CL accurately reflected the absence of any neuritic plaques, and >20.1 CL indicated the presence of at least moderate plaque density. Clinicopathological diagnosis of AD was rare with CL <45.

Background

Current standard-of-truth (SoT) diagnosis of Alzheimer's disease (AD) partly depends on neuropathological demonstration of brain amyloid-beta (A β) plaques and tau [1]. However, the invasive nature of brain biopsy requires antemortem diagnosis to be made using biomarkers. Recently, research criteria for AD have emphasized the importance of amyloid and tau positron emission tomography (PET) imaging biomarkers [2, 3].

¹¹C-Pittsburgh Compound B (¹¹C-PiB), ¹⁸F-florbetaben (FBB), ¹⁸F-florbetapir, ¹⁸F-flutemetamol and ¹⁸F-NAV4694 (NAV) are PET tracers that demonstrate binding to brain A β in AD from the preclinical AD stage onwards, with good sensitivity and specificity as biomarkers of antemortem AD pathology and predictors of progression to AD dementia [4-10]. Amyloid PET scans are used for inclusion and monitoring in AD-modifying clinical therapy trials, and to aid clinical diagnosis and prognostication [11].

Variability in tracers, PET scanners, procedural factors, and analysis methods across imaging centres have driven attempts for quantitative standardisation of amyloid PET results. Klunk and colleagues [12] derived a scale of 'Centiloid' units (CL) for standardised reporting of amyloid imaging. CL values range beyond the 'anchor-point' of 0, representing average young healthy controls, and 100, representing the amyloid burden present in average typical mild AD. This important work allows for amyloid PET scans across different sites to yield comparable results.

Comparison of neuropathological data with positive or negative amyloid PET scans based on expert visual read has been performed for ¹¹C-PiB, ¹⁸F-florbetapir, ¹⁸F-florbetaben and ¹⁸F-flutemetamol [7, 9, 13-17]. In-vivo biomarkers such as cerebrospinal fluid A β , tau PET, and volumetric MRI have been compared with amyloid PET in CL [18, 19]. Only three recent studies, to our knowledge, have examined the performance of amyloid PET CL thresholds compared with SoT neuropathology [20-22].

We aim to define the accuracy of CL values when compared with SoT post-mortem neuropathological data on neuritic amyloid plaque density, final clinicopathological diagnosis of AD, and visual reading threshold for a positive amyloid PET scan.

Methods

Compliance with Ethical Standards

Ethics approval was obtained from the Austin Health Human Research Ethics Committee (reference LNR/17/405).

Subject identification and demographic data collection

Fifty-one subjects in total were retrospectively identified from the databases of the Austin Health Molecular Imaging and Therapy department, Sydney Brain and Mind Centre, and the Victorian Brain Bank. These subjects, with prior informed consent, had undergone both an amyloid PET scan and post-mortem neuropathologic brain evaluation between all years recorded in the database (2004 to 2017). Exclusion criteria for these prior studies were history of stroke, significant medical illness, recent cancer, and substance use disorder. Cases with a diagnosis of familial AD were excluded, as different neuropathological processes in this condition, such as a significantly greater density of A β plaques in the cerebellum than in sporadic AD [23, 24], may have confounded amyloid PET quantification and interpretation. Data has been published on a proportion of these cases [25, 26]. Two of the fifty-one subjects were excluded due to a diagnosis of familial AD.

Amyloid PET Imaging and Centiloid determination

A β imaging was performed with either ^{11}C -PiB or FBB. The methodology for PET imaging with these tracers has been previously described [27, 28]. A 20-minute acquisition was commenced 50 minutes post-injection of ^{11}C -PiB or 90 minutes post-injection of FBB. A transmission scan was performed for attenuation correction. PET images were reconstructed using a 3D row-action maximum likelihood algorithm (RAMLA). The standard Centiloid cortical and whole cerebellar volumes of interest template were applied to the summed and spatially normalized PET images in order to obtain standardized uptake value ratios (SUVR). For this study we used the CapAIBL software package, with a validated lower transformation to correct for CapAIBL registration without MRI [29, 30]. This package has been validated against the standard Centiloid MRI based spatial normalization with SPM8. The SUVR were transformed into Centiloid units by linear transformation using the PET tracer specific equations published for conversion of Centiloid method SUVR to Centiloid units with a minor correction applied for the CapAIBL registration [12, 27, 28, 31].

Neuropathologic evaluation

Neuropathological evaluation was performed at the Victorian Brain Bank (Melbourne, Australia) and Sydney Brain Bank (Neuroscience Research Australia, Sydney, Australia) to determine a global C score from inferior temporal regions of fixed brain hemispheres based on the Consortium for Establish a Registry for Alzheimer's Disease (CERAD) neuropathologic assessment guidelines [32]. Frequency of neuritic plaques per 100x microscopic field were categorised as none, sparse, moderate or frequent with corresponding C scores of 0, 1, 2 or 3 respectively, as described in published guidelines [33].

Visual Read

One amyloid PET expert reader (author CR), blinded to CL values and neuropathological data, visually interpreted all scans using MedView v12 software, viewing images in greyscale and rainbow colour scale. The method used to visually read amyloid PET has been previously described [34]. Scans were classified positive when cortical activity was equal to or greater than white matter activity in one or more lobes.

Clinicopathological diagnosis

The clinicopathological diagnosis for each case factored in both neuropathological assessment and antemortem clinical diagnosis. Neuropathological diagnosis was made in accordance with published guidelines [33], and included morphological examination with immunohistochemistry analyses for A β , tau, TDP43, and alpha-synuclein. There were 17 AD and 32 non-AD cases. Non AD cases included diagnoses of frontotemporal dementia (n=12), normal controls (n=3), dementia with Lewy bodies (n=3), Parkinson's disease dementia with concurrent diffuse Lewy bodies (n=3), hippocampal sclerosis (n=2), Creutzfeldt-Jakob disease (n=2), progressive supranuclear palsy (n=2), motor neuron disease (n=1), hippocampal ischaemia (n=1), corticobasal degeneration (n=1), multisystem atrophy (n=1), and a case of mixed AD and dementia with Lewy bodies (n =1) was excluded from being categorised as clinicopathological AD.

Statistical analyses

Three aspects of CL performance were investigated. Firstly, CL values were compared with dichotomized neuropathological C score categories using two different approaches: "high vs low" plaque density ("high" = moderate and frequent, and "low" = none and sparse), and; "any vs none" ("any" = sparse, moderate and frequent, and "none" = none). A Youden Index [35] was used to determine the optimal CL thresholds from receiver operator characteristic curves. Secondly, CL values were compared with binary

visual read (positive or negative). Thirdly, CL values were compared with cases of AD as determined by clinicopathological diagnosis using descriptive statistics. To assess for the any contribution of interval from PET scan to time to death, analyses were repeated using adjusted CL values, after applying a sigmoidal adjustment with a linear-segment accumulation rate of 5.18% per year, based on previous work by Villemagne and colleagues [36]. It should be noted that very low or very high degrees of amyloid neuropathology do not change as much as intermediate levels over time.

Results

Case Characteristics

Of the forty-nine included subjects, thirty-three underwent ^{11}C PiB PET and sixteen underwent FBB PET. Thirty-eight (78%) cases were male. The mean age at death was 76 years, and the median interval between date of last amyloid PET scan and death was 1005 (IQR \pm 1114) days.

Centiloid results and neuropathological C score categories

There were 19 (39%) patients with “high”, and 30 (61%) with “low” C scores. The receiver operator characteristic curve (Figure 1) demonstrated an area under curve (AUC) of 0.97 and an optimal threshold of 20.1 CL for detection of a high level of amyloid plaque (i.e. moderate or frequent neuritic plaques). After applying the sigmoidal adjustment for interval from scan to post-mortem, the putative optimal threshold was 21.3 CL.

Fig. 1 Centiloid results and “high” vs “low” C score categories

Receiver Operator Characteristic curve for Centiloid units thresholds in determining “high” vs “low” amyloid plaque burden

When neuritic plaque scores were grouped as to “any” (i.e. sparse or more) vs “none”, there were 24 (49%) with “any” and 25 (51%) with no neuritic plaques. The optimal CL threshold found for detecting the presence of any amyloid plaques (i.e. sparse or more) was 9.5 CL (Figure 2), with an AUC of 0.96, yielding a sensitivity of 0.96 and a specificity of 0.88. After applying the sigmoidal adjustment for interval from scan to post-mortem, the putative optimal threshold was 9.6 CL.

Fig. 2 Centiloid results and “none” vs “any” C score categories

Receiver Operator Characteristic curve for Centiloid units thresholds in determining “none” vs “any” amyloid plaque burden

Centiloid results and amyloid PET visual read

Correlation of Centiloid values with amyloid PET expert visual read (positive /negative) yielded an AUC of 1.0 and optimal CL threshold of 26. Using this threshold there was 100% agreement between Centiloid (elevated/not elevated) and visual read (positive/negative), (Figure 3).

Fig. 3 Centiloid results and amyloid PET visual read

Scatterplot for Centiloid unit threshold testing against binary expert visual read categories. A 26 CL cut-off yielded a 100% match to expert visual read of “high” or “low”.

Centiloid results in clinicopathological AD diagnosis

CL values were reviewed for the 17 cases that were determined to have a clinicopathological diagnosis of AD. The median CL result was 87.7 (IQR \pm 42.2), and 16 of the 17 cases (>90%) had a value of > 45 CL.

Discussion

Improving accuracy of the detection of brain amyloid plaques is important for clinical trial enrichment in the quest to develop disease-modifying or curative treatment for the growing burden of Alzheimer’s disease. Contemporary clinical application can also better assist diagnosis, prognostication and planning for patients with cognitive disorders.

We have demonstrated that a threshold of 20.1 CL was optimal for the detection of “high” levels of neuritic plaque density, as determined by C score moderate or frequent classification. In other words, values of 20.1 CL or lower accurately reflected the absence of moderate or frequent plaques, with a high AUC. This threshold was not significantly altered when corrected for time between scan and post-mortem. CL results below this threshold should provide reassurance that patients are unlikely to have Alzheimer’s disease. This is reasonably concordant with the findings of Navitsky and colleagues, who determined a threshold of 24.1 CL with florbetapir for CERAD amyloid plaque classification of moderate or frequent vs sparse or none, in 59 individuals [21]. This is also concordant with the Centiloid analysis by Dore and colleagues of a florbetaban phase III post-mortem study in 66 individuals, which yielded a threshold of 19 CL for the same categorisation [22]. These thresholds are higher than those identified by La Joie et al, who determined a threshold of 12.2 CL for separating none or sparse plaques from moderate or frequent plaques; and the same threshold of 12.2CL for separating any plaques from no plaques in 179

individuals scanned with ^{11}C -PiB PET [20]. Those authors did, however, acknowledge a contribution from false positive results, and suggested that 24 CL may be a more appropriate threshold for “identifying clinically meaningful A β burden,” and suggested the cut point of 19 CL proposed by Jack and colleagues [37] as a threshold predictive of reliable worsening of amyloid burden with time would be a more reasonable threshold to use. These thresholds may be useful as cut-offs for clinical trial enrichment, or for guiding decision-making about commencing potential disease-modifying therapies when they become available.

For the detection of “any” amyloid plaque (sparse or more), the optimal threshold identified was 9.5 CL. Once again, this value only marginally increased when corrected for time between scan and post-mortem. This threshold is similar to two standard deviations of young controls as determined by Klunk and colleagues, equalling 8.68 CL [12] and 12.2 CL [20]. This suggests there are no differences in amyloid tracer binding between young and old individuals with no amyloid plaques, indicating no substantial increase in non-specific binding nor significant changes in tracer kinetics with normal ageing.

A threshold of 26 CL exactly matched expert visual read of positive vs negative scan. This is consistent with good concordance (97%) noted by Leuzy and colleagues [18] between visual read and PiB CL results. Larger numbers of more borderline cases to better assess the correlation of CL result with visual read and identify the reasons for discordant classification may be warranted.

Our results compare favourably with phase III trials of PET tracers that have compared blinded expert visual reads of amyloid imaging with post-mortem data, where moderate to frequent neuritic plaques (a “high” classification in our study) were considered positive. Florbetaben visual reads were reported as having sensitivity of 97.9% and specificity of 88.9% [15]. Florbetapir visual reads yielded sensitivity of 92% and specificity of 100% within a 2-year window between imaging and autopsy [14]. Flutemetamol visual reads in one study correlated with both original and modified CERAD criteria, yielded respective sensitivities of 91.9% and 90.8%, and specificities of 87.0% and 90.0% [16].

In our cases with clinicopathological AD, a median CL value of 87.7 was found at post-mortem, but with significant variability as demonstrated by the IQR of ± 42.2 CL. Only one of these cases, scoring 20.1 CL, returned a result under 45 CL, suggesting that a sensitivity cut-off for defining clinicopathological AD should be considerably higher than that for detecting “high” amyloid plaques alone. For comparison, an upper SUVR threshold of normal in a healthy control PiB PET group was calculated, using the upper 95% confidence interval, to be 1.41 by Nordberg et al [38], equating to 34 CL as converted by Leuzy et al [18]. The latter group demonstrated median PiB PET results of 47.5 CL for Mild Cognitive Impairment and 84.1 CL for AD in their group. These studies did not include post-mortem evaluation. Larger numbers in future studies would help confirm if a clinicopathological diagnosis of AD is indeed rare when under 45 CL.

The exclusion of mixed dementia only resulted in one case with AD neuropathology not being included in the analysis due to coexisting Lewy body disease; it is unclear what effect mixed dementia would have on the concordance of clinical diagnosis and CL values.

Limitations

A limitation of the study is the data distribution, in that only two subjects had results between 15 and 35 CL, consequently restricting the ability to tightly define thresholds. Specifically, these subjects had results of 20.1 and 30.9 CL.

Another limitation is the time elapsed between scan and death. This averaged approximately three years. We have accounted for this using established correction calculations. Of note, the only case of clinicopathological diagnosis of AD below 45 CL measured 20.1 CL, 6 years before death; this only increased to 21.3 CL after time-interval correction. Familial AD cases were excluded from this study due to potential presence of neuritic amyloid plaques in the cerebellum [23], and the lower affinity of PiB to “cotton wool” plaques found in some presenilin mutations [39]. These cerebellar plaques could interfere with the scaling to SUVR, and return misleading low CL results not applicable to typical sporadic AD [24]. Separate characterisation of CL performance in familial AD is warranted.

Conclusions

In our cohort, values <9.5 CL accurately reflected the absence of any neuritic plaques, and >20.1 CL indicated the presence of at least moderate plaque density. These neuropathology-based Centiloid thresholds may be used to exclude a diagnosis of AD and to define groups for early intervention and other disease specific trials.

List Of Abbreviations

^{11}C -PiB – ^{11}C -Pittsburgh Compound B

A β – amyloid-beta

AD – Alzheimer’s disease

AUC – Area Under Curve

CERAD – Consortium for Establish a Registry for Alzheimer’s Disease

CL – Centiloid

FBB – ^{18}F -florbetaben

IQR – Interquartile range

MRI – magnetic resonance imaging

NAV – ^{18}F -NAV4694 (NAV)

PET – Positron emission tomography

RAMLA – row-action maximum likelihood algorithm

ROC – Receiver Operator Characteristic

SoT – Standard of truth

SUVR - standardized uptake value ratio

TDP43 - TAR DNA-binding protein 43

Declarations

Ethical approval:

All procedures performed in studies involving human participants were in accordance with the Austin Health Human Research Ethics Committee (reference LNR/17/405) and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Consent for publication

All subjects had given consent for their data to be used for research purposes.

Availability of data

Data analysed in this study are available from the corresponding author on reasonable request.

Competing Interests and Funding:

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Author contributions

Name	Location	Role	Contribution
Sanka Amadoru, MBBS	Austin Health, Melbourne, Australia	Author	Study design, data acquisition and analysis; manuscript drafting and revision for intellectual content.
Vincent Dore, PhD	Austin Health and CSIRO, Melbourne, Australia	Author	Major role in data acquisition and analysis; revised the manuscript for intellectual content.
Catriona McLean, MD	Victorian Brain Bank, Melbourne Australia	Author	Major role in data acquisition and interpretation; revised the manuscript for intellectual content.
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Claire Shepherd, PhD	Sydney Brain Bank, Sydney, Australia	Author	Major role in data acquisition and interpretation; revised the manuscript for intellectual content.
Glenda Halliday, PhD	Sydney Brain Bank and University of Sydney, Sydney, Australia	Author	Major role in data acquisition and interpretation; revised the manuscript for intellectual content.
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Paul Yates, PhD	Austin Health, Melbourne, Australia	Author	Role in data acquisition, data analysis and interpretation; revised the manuscript for intellectual content.
John Hodges, MD	University of Sydney, Sydney, Australia	Author	Minor role in data acquisition; revised the manuscript for intellectual content.
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Victor L Villemagne, MD	Austin Health, Melbourne, Australia	Author	Major role in data acquisition, analysis and interpretation; revised the manuscript for intellectual content.
Christopher C Rowe, MD	Austin Health, Melbourne, Australia	Author	Major role in study design; data acquisition, analysis and interpretation; revised the manuscript for intellectual content.

Acknowledgments and author's information

Not applicable

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Figures

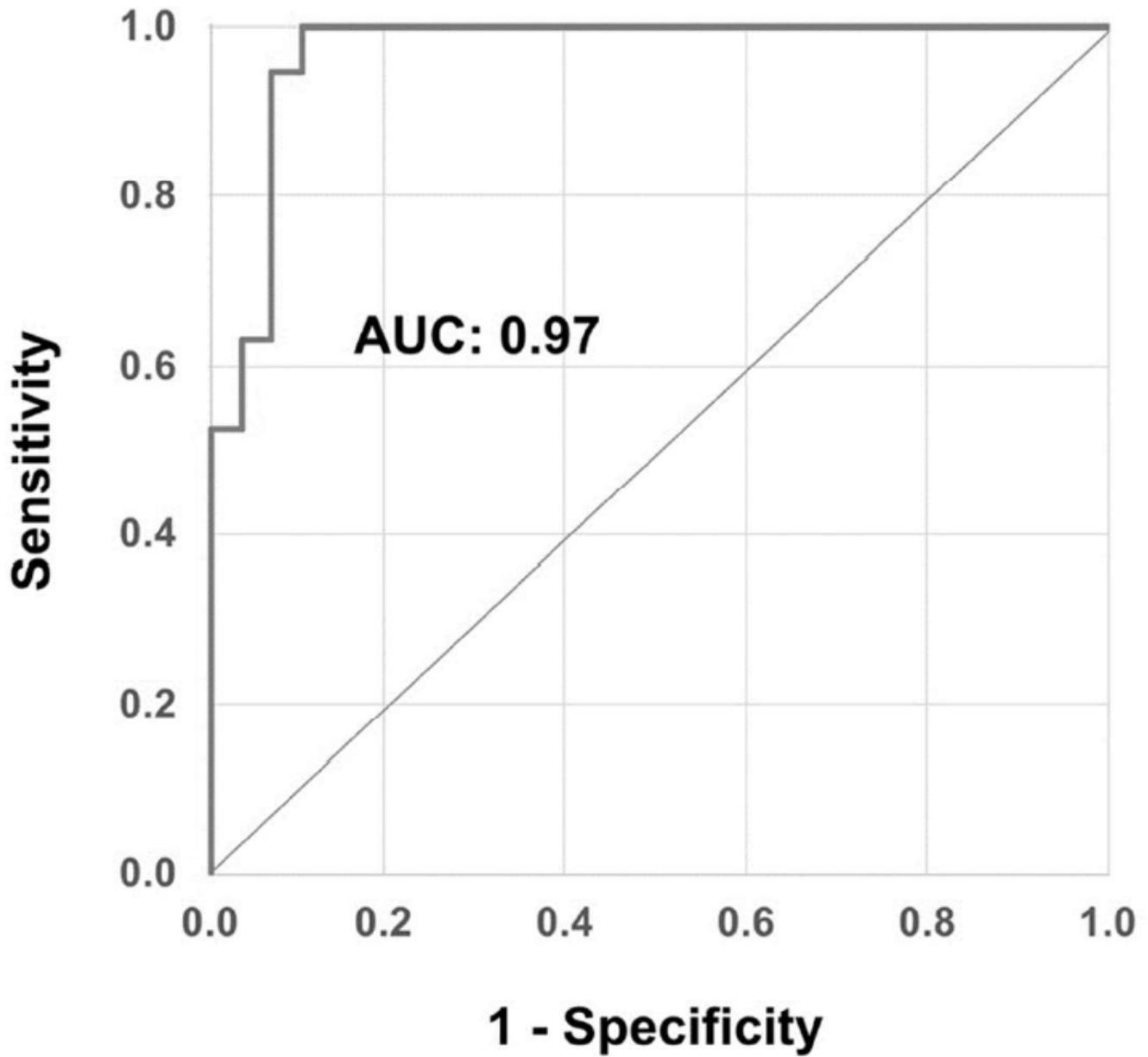


Figure 1

Centiloid results and "high" vs "low" C score categories Receiver Operator Characteristic curve for Centiloid units thresholds in determining "high" vs "low" amyloid plaque burden

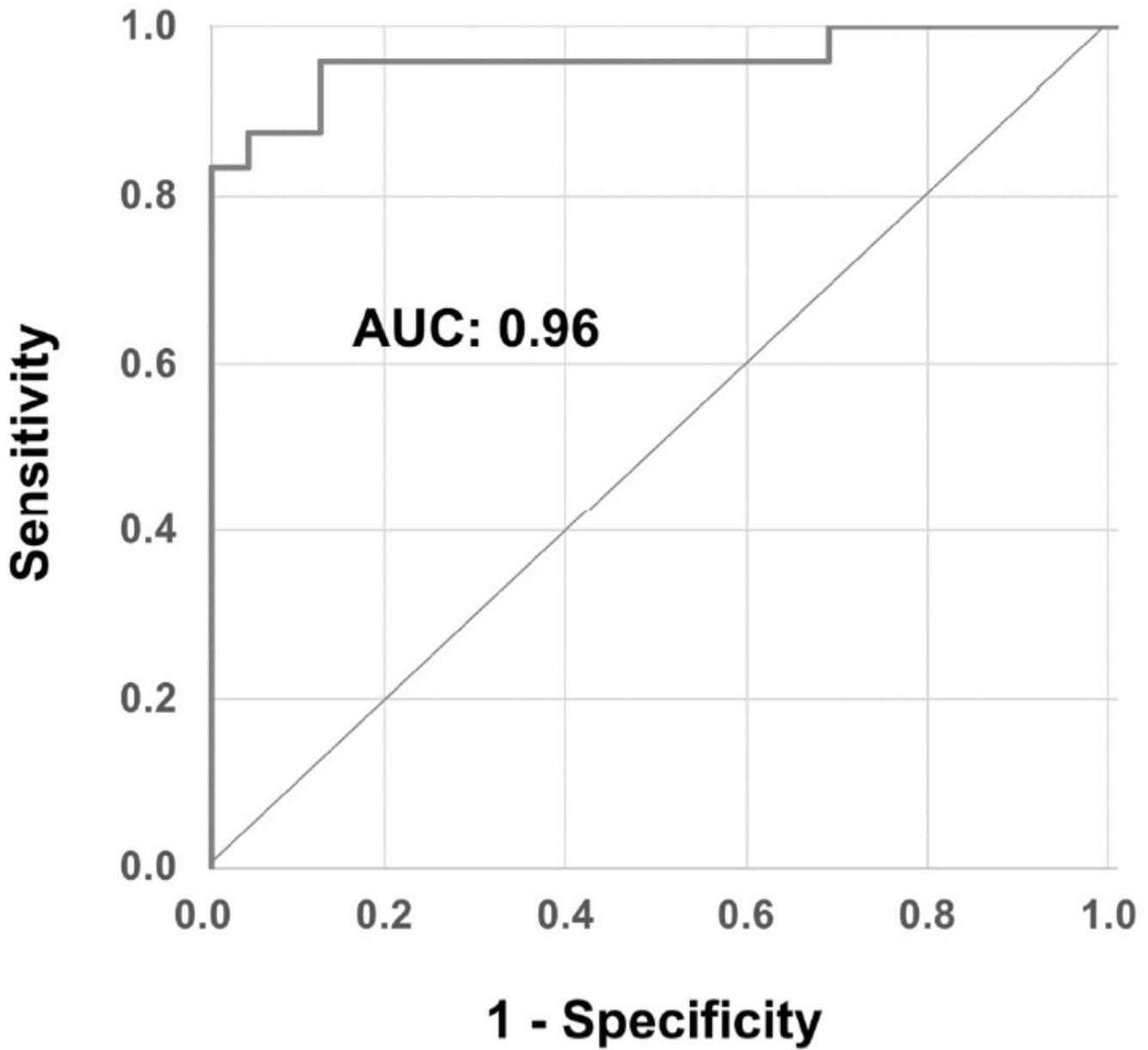


Figure 2

Centiloid results and "none" vs "any" C score categories Receiver Operator Characteristic curve for Centiloid units thresholds in determining "none" vs "any" amyloid plaque burden

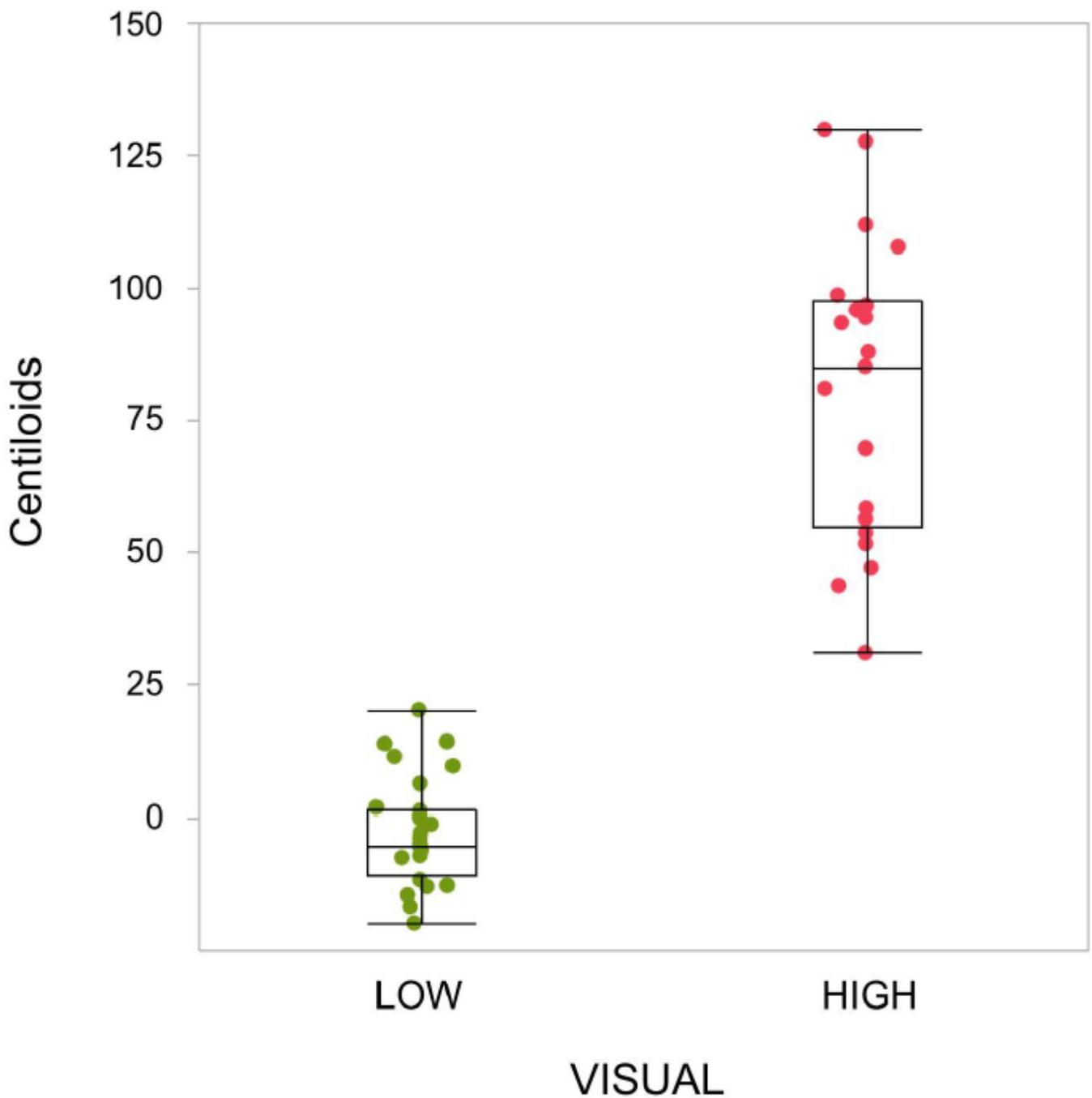


Figure 3

Centiloid results and amyloid PET visual read Scatterplot for Centiloid unit threshold testing against binary expert visual read categories. A 26 CL cut-off yielded a 100% match to expert visual read of “high” or “low”.