

A Novel Fluorescent Probe for Detecting Hydrogen Sulfide in Osteoblasts During Lipopolysaccharide-Mediated Inflammation Under Periodontitis

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1 *A novel fluorescent probe for detecting hydrogen sulfide in osteoblasts during*
2 *lipopolysaccharide-mediated inflammation under periodontitis*

3 **Title page**

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10 #These authors contributed equally to this study.

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12 **Author contributions**

13 Xiaoya Lu, Yi Chen and Yue Wu performed the bioimaging experiments together.

14 Hanchuang Zhu synthesized the H₂S fluorescence probe.

15 Shengyun Huang, Baocun Zhu and Dongsheng Zhang conceived the idea and directed
16 the work.

17 All authors contributed to data analysis, manuscript writing and participated in
18 research discussions.

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23 **Abstract**

24 *Periodontitis, one of the most common chronic inflammatory diseases, affects the*
25 *quality of life. Osteogenesis makes an important role of the disease. There is a*
26 *connection between hydrogen sulfide (H₂S) and periodontitis, but according to the*
27 *study has been published, the precise role of H₂S in inflammation remains in doubt.*
28 *The main reason of the lack of research is that H₂S is an endogenous gasotransmitter,*
29 *difficult to discern through testing. So, we synthesis a novel fluorescence probe which*
30 *can detective H₂S in vitro. By using the novel H₂S fluorescence probe, we found that*
31 *H₂S changes in osteoblasts mainly by cystathionine- γ -lyase, and H₂S increases under*
32 *LPS stimulation. H₂S could be a potential marker for diagnosis of inflammatory*
33 *diseases of bone, and might help deeper studies of the changes of H₂S level and promote*
34 *the progression on the researches about pathogenesis of periodontitis.*

35

36 **Keywords**

37 *Periodontitis, hydrogen sulfide, fluorescence probe, osteogenesis, LPS*

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45 **Introduction**

46 *Periodontitis as one of the most common chronic inflammatory diseases, afflicting man.*

47 *It can leading cause of bone resorption, even worse tooth loss. Under normal*

48 *physiologic conditions, the balance of osteoclasts and osteoblasts is tightly related to*

49 *avoid the loss of bone. The breakdown of the balance will cause diseases. Avoiding*

50 *alveolar bone destruction is an important problem to control the periodontitis.*

51 *However, the detailed mechanism of periodontitis is still largely unknown.*

52 *Lipopolysaccharide (LPS), a major toxic factor of gram-negative bacteria, plays a*

53 *main role in periodontitis. It can cause periodontitis by modulating the activity of the*

54 *host defenses [1], inducing a hypoxic phase [2] etc., and it eventually stimulates bone*

55 *resorption [3]. LPS may leads to inflammatory response in osteoclasts and osteoblast,*

56 *which may results in a disorder in the balance of osteoclasts and osteoblasts even cell*

57 *death, leading to accelerating bone loss [4]. For experimental researches, LPS*

58 *stimulated the rat gingival sulcus every day in order to obtain an experimental*

59 *periodontitis model by immunizing it with the antigen [5]. LPS treated cells are in a*

60 *similar situation as well. Halitosis is one of the clinical features of periodontitis, and*

61 *Hydrogen sulfide (H₂S) is the main unbearable stinky smell of periodontitis and may*

62 *play a significant role in its development.*

63 *Biothiols are indispensable in human physiology, which are in a vital branch of reactive*

64 *sulfur species (RSS) family. H₂S is an endogenous gasotransmitter, which is*

65 *well-known for its stinky smell like rotten eggs. H₂S is produced by the*

66 *sulfur-containing materials cysteine, homocysteine or*

67 3-mercaptopyruvate. H_2S is transformed by cystathionine- β -synthase (CBS),
68 cystathionine- γ -lyase (CSE) and 3-mercaptopyruvate sulfurtransferase (3-MST)[6].
69 Most researchers previously believe that H_2S can promote the pathogenesis of
70 periodontitis, and hugely harm to their periodontal tissue [7]. But recently, there is
71 evidence shows that H_2S might be useful in cell protection. For exogenous H_2S , it can
72 promote LPS-induced apoptosis of osteoblast cells, which might represent a new
73 direction in the treatment of osteomyelitis [8]. When oxidative damage occurs, H_2S can
74 increase cell viability and reduce cell apoptosis. H_2S might have a advantageous
75 effect, because according to the research, NaHS treatment can produce
76 anti-inflammatory effects via NO and TNF- α [9]. Besides, H_2S can protect cell injury
77 by regulating oxidative stress, mitochondrial function, and inflammation. It also have
78 the ability to potentially prevent bone loss in periodontitis [10]. So, there is a
79 connection between H_2S and periodontitis, but until now, the precise role of H_2S in
80 inflammation remains unknown.

81 Most of the studies focus on the effect of the H_2S , not many about H_2S changes under
82 stimulation. Researchers often use Western blot, immunohistochemical staining, and
83 some other methods to detect the H_2S changes indirectly. Recently, there are some
84 direct techniques to detect H_2S , such as chromatography, electrochemistry and
85 colorimetry [11]. But a technique that can detect H_2S directly in living cells is still
86 needed. H_2S -fluorescence probes, which is high-speed developing, is considered as one
87 of the most helpful instrument areas in the field of H_2S biology [12]. We previously
88 designed a H_2S probe, which consists of a 4-chloro-1,8-naphthalimide as fluorophore

89 and introduces hydrophobic chains (dodecylamine) and hydrogen sulfide recognition
90 groups (azide groups) (Figure 1 a). The introduction of the electron-withdrawing azide
91 group changes the push-pull system and quenches the fluorescence. It is noteworthy
92 that the reaction is easy to carry out and the yield is high. When the probe reacts with
93 hydrogen sulfide, the azide group is reduced to an amino group. Because the amino
94 group acts as an electron-donating group, the effect of intramolecular charge transfer
95 is enhanced, and the fluorescence is recovered. The probe is able to directly measure
96 the real time H₂S level in living cells. Overall, because of high resolution and sensitivity
97 of the H₂S probe make it a helpful tool. There are some studies showing that H₂S
98 fluorescence probe can detect endogenous H₂S in real-time and in situ. However, most
99 of them use tumor cells instead of somatic cells. If the probe could be used in
100 somatic cells, it can broaden the diagnosis and treatment applications of H₂S. By using
101 a novel H₂S fluorescence probe, we found that H₂S changes in osteoblast mainly by
102 CSE, and H₂S increases under LPS stimulation.

103

104 **Materials and methods**

105 1. Regents

106 The hydrogen sulfide fluorescent probe was provided by Professor Baocun Zhu (School
107 of Resources and Environment, University of Jinan, Jinan, China). The mother
108 solution of the probe was prepared with DMSO (Sigma-aldrich, USA) and
109 dichloromethane. The probe concentration was 1 mM. The test concentration was
110 10 μM and the experiment was carried out at room temperature (25°C).

111 *DL-propargylglycine (PAG) (cystathionine γ -lyase inhibitor, Sigma-Aldrich),*
112 *Cysteine (Cys), NaHS, lipopolysaccharide (LPS) (Sigma-aldrich, USA), cell counting*
113 *kit-8 (CCK-8; Dojindo Molecular Technologies, Tokyo, Japan).*

114 2. *MC3T3-E1 cell culture*

115 *The murine calvaria-derived MC3T3-E1 osteoblast-like cell line (Procell CL-0378,*
116 *subclone 14) was provided by Procell Life Science and Technology CO.,Ltd. Cells were*
117 *seeded at 5×10^4 cells/ml into 25 cm² flasks and maintained in α -MEM, supplemented*
118 *with 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin. Cells were*
119 *maintained in an incubator containing a 5% carbon dioxide/air environment at 37°C.*

120 3. *Toxicity Analysis:*

121 *The influence of the H₂S probe on MC3T3-E1 cell was examined by CCK-8. Briefly,*
122 *MC3T3-E1 cells, seeded at a density of 5×10^4 cells/ml on a 96-well plate, were*
123 *maintained at 37°C in a 5% CO₂, 95% air incubator for 24 h. Then the cells were*
124 *incubated with different concentrations (0, 5, 10, 20, 25, 37.5, 50, 75 and 100 μ M) of*
125 *probe suspended in culture medium for 24 h. Same as the probe group, the other plate*
126 *of cells incubated with same concentrations (0, 5, 10, 20, 25, 37.5, 50, 75 and 100 μ M)*
127 *of solvent. Subsequently, CCK-8 solution was added into each well for 2 h, and*
128 *measured the absorbance at 450 nm was measured.*

129 4. *Application of H₂S probe to access exogenous H₂S levels: The cells were pre-treated*
130 *with NaHS (50, 100, 150, 500 μ M) for 30 minutes, then, treated with the H₂S*
131 *probe (10 μ M) for 30 minutes. Fluorescence and bright field images were collected after*
132 *PBS washing for three times. Green fluorescence was observed under the confocal*

133 microscope at excitation wavelengths of 405 nm. In order to control exposure, Smart
134 Gain was kept at the same voltage in every photographs.

135 5. Application of H₂S probe to access endogenous H₂S levels:: In the periodontium of
136 mammalian host, H₂S is produced using Cys mainly by CSE and CBS. The cells were
137 pre-treated with Cys (100 μ M, 200 μ M) for 30 minutes, then, treated with the H₂S
138 probe (10 μ M) for 30min. Fluorescence and bright field images were collected after
139 PBS washing for three times. Green fluorescence was observed under the confocal
140 microscope at excitation wavelengths of 405 nm. In order to control exposure, Smart
141 Gain was kept at the same voltage in every photographs.

142 PAG is an irreversible inhibitor of CSE. It can blocking-up the produce of endogenous
143 H₂S in MC3T3-E1. Therefore, we pre-treated cells with 50 μ M PAG, 30min, then
144 cells were treated with or without Cys for 30 min. Last, fluorescence was examined as
145 before, Smart Gain was kept at the same voltage in every photographs.

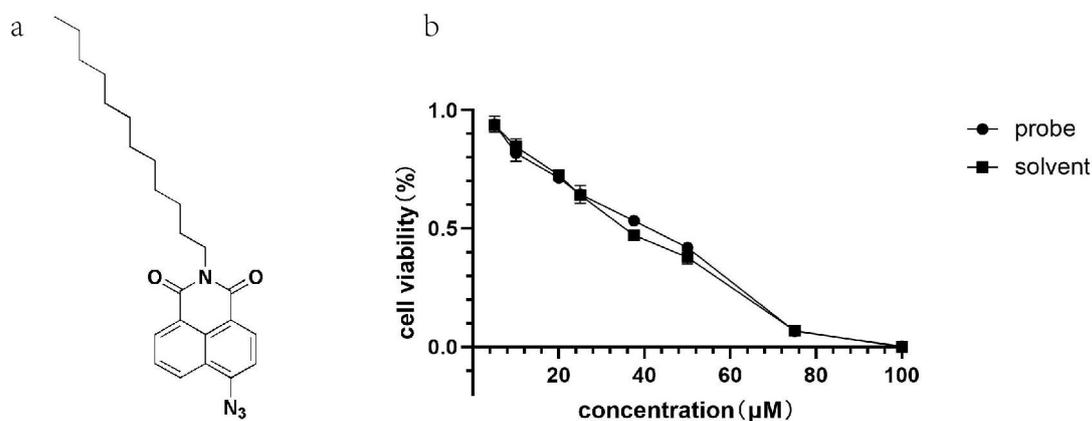
146 6. Addition of lipopolysaccharide (LPS) for inducing inflammation and assessment
147 with H₂S probe: The cells were incubated with 1, 2 μ g/ml LPS for one day.
148 Subsequently, the culture dish was washed with PBS for three times and incubated
149 with 10 μ M probe for 30 min. Then, the cells were washed with PBS, then the
150 fluorescence imaging was examined by confocal microscope, Smart Gain was kept at
151 the same voltage in every photographs.

152 **Results**

153 Toxicity analysis: The cell's viable and healthy during the detection is a key concern.

154 Figure 1 b showed that cell viability was almost not affected by the probe at 10 μ M.

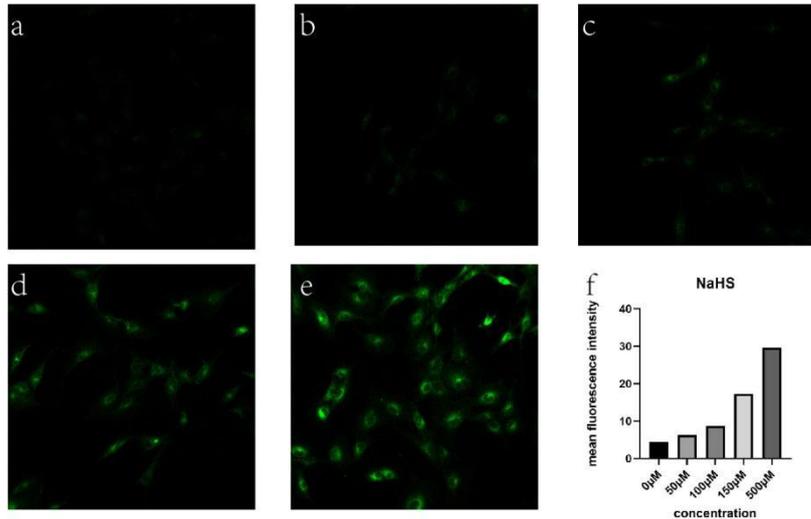
155 Toxicity is mainly introduced by solvent, DMSO and dichloromethane. The result
156 verify that the H₂S probe is harmless to the cell. Thus, the H₂S probe can be used in
157 living cells for fluorescence imaging analysis.



158
159 Figure 1 a. Structure of H₂S probe. b. toxicity Analysis.

160

161 Cell fluorescence imaging of different concentrations exogenous H₂S: As shown by
162 figure 2, with the different concentrations (0, 50, 100, 150, 500 μ M) of NaHS, a gradual
163 increase of intensive green fluorescence was observed using 405nm as an excitation
164 wavelength. Consistent with previous studies, the amount of H₂S is one third of
165 exogenous of NaHS. Thus, the probe was estimated detection of accuracy to 10 μ M.
166 Fluorescent intensity is stable during the progress of takeing pictures under the
167 confocal laser scanning microscopy. That indicated that our probe is sensitive to H₂S,
168 and it also prove that H₂S probe was cell membrane permeable and can used in the
169 normal cells for detecting intracellular H₂S.



170

171 *Figure 2 Cell fluorescence imaging of different concentrations exogenous H₂S. a-e.*

172 *Fluorescence imaging of cells incubated with different concentration of NaHS (0, 50,*

173 *100, 150, 500 μ M) f. Fluorescence intensity analysis.*

174

175 *Cell fluorescence imaging of endogenous H₂S: According to the previous research, for*

176 *osteoblasts, CSE-H₂S might be the major path for the H₂S produced [13]. As shown by*

177 *Figure 3, the incubation of cells with 100 μ M Cys produced intensive green*

178 *fluorescence, but the fluorescence decreased when cells were incubated with 200 μ M*

179 *Cys. That means that low dose of Cys could increase H₂S production, but high dose of*

180 *Cys inhibited H₂S production. In order to verify whether the CSE-H₂S pathway is the*

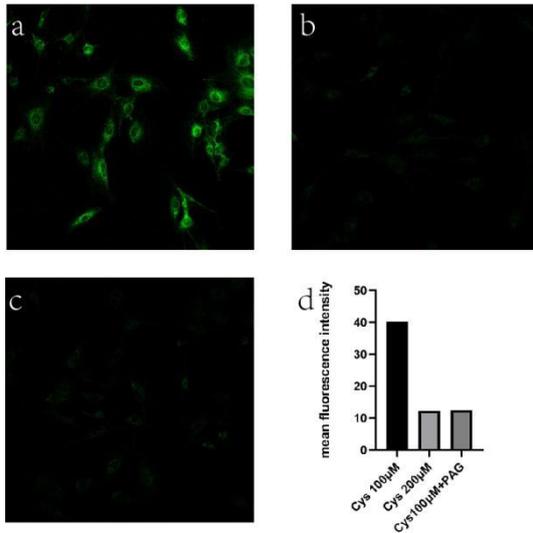
181 *main pathway to produce the H₂S, we used PAG as the irreversible inhibitor to CSE.*

182 *Figure 3 showed that the intensity of fluorescence was decreased, which means the H₂S*

183 *was decreased, because of the pretreatment of the inhibitor, and the intensity of PAG*

184 *group was as weak as the control group, indicating that the production of endogenous*

185 *H₂S was significantly inhibited with CSE inhibitor.*



186

187 *Figure3 Cell fluorescence imaging of endogenous H₂S. a. Cys 100 μ M, b. Cys 200 μ M,*

188 *c. Cys 100 μ M + PAG 50 μ M, d. Fluorescence intensity analysis.*

189

190 *Cell fluorescence imaging of LPS induced endogenous H₂S: when cells were treated*

191 *with LPS (2 μ g/ml) to produce inflammation, as shown by Figure 4, intensive green*

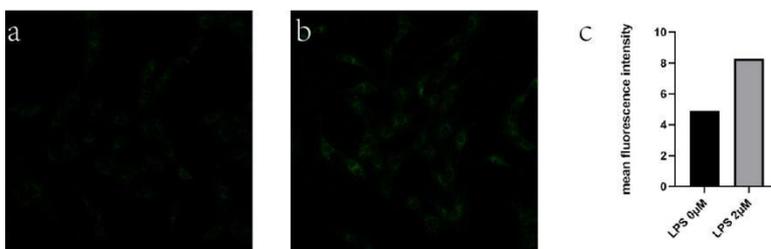
192 *fluorescence were produced compared to the control group. This indicates that when*

193 *inflammation occurs, a lot of H₂S was produced. In other words, the increase of H₂S*

194 *level can serve as an indicator for cells that are under the inflammation state. The*

195 *production of endogenous H₂S induced by lipopolysaccharide-mediated inflammation*

196 *was successfully monitored with this H₂S probe.*



197

198 *Figure4 Cell fluorescence imaging of LPS induced endogenous H₂S. a. LPS 0 μ M, b.*

199 *LPS 2 μ M, c. Fluorescence intensity analysis.*

200

201 **Discussion**

202 *The main aim of the experiment is to solve the problem of detection of the*
203 *inflammation of osteoblast, furthermore, we found that H₂S produced by osteoblast is*
204 *mainly via CSE-H₂S pathway. In our study, we proved that our probe can be used in*
205 *the normal cell to detect the H₂S changes, which is rarely studied. There are already a*
206 *lot of fluorescent probes that have been devised to detect intracellular H₂S levels,*
207 *however, to our knowledge, most of these probes were successfully applied to show*
208 *alteration of H₂S levels of tumor cells or living animals [14; 15; 16]. But in our study,*
209 *we used a novel fluorescent probe to detect alteration of H₂S levels in living osteoblast*
210 *cells with exogenous or endogenous H₂S for the first time. The H₂S probe possesses*
211 *high sensitivity, selectivity, and an ultrafast response to H₂S, rendering it suitable for*
212 *detection of H₂S concentration in living cells. In order to determine whether the cell*
213 *could translate Cys to H₂S, and whether the probe could visualize endogenous H₂S, we*
214 *treated the osteoblast cells with irreversible inhibitor, PAG. The result proved that*
215 *H₂S is produced mainly by CSE-H₂S pathway, which had not been proved in a visual*
216 *way before. Other researchers have proved that (CSE) majorly contributed to*
217 *endogenous H₂S production in the primary osteoblast by overexpression and*
218 *knockdown CSE [13]. This is consistent with our results.*

219 *For the inflammation of bone, there are two proved sources of H₂S: bacteria and*
220 *macrophage. When inflammation occurs, some bacteria produced and released H₂S,*
221 *including various common gram-negative pathogens in osteomyelitis such as*

222 *Escherichia coli*, *Enterococcus faecalis*, *Enterobacter cloacae*, and *Klebsiella*
223 *Pneumoniae*. For macrophage, research shows that the level of H_2S was improved and
224 the expression of CSE mRNA increased because of the stimulate of LPS[17]. Our study
225 shows that osteoblasts is the third source of H_2S . Different sources of H_2S might have
226 interaction effect, for example, H_2S production by osteoblast might modulate
227 macrophage polarization and contribute to bone reparation. Keeping physiological
228 level of endogenous H_2S in PDLSCs/periodontal tissue is beneficial to maintain the
229 homeostasis of periodontal tissue [18]. A appropriated level of H_2S may play an vital
230 role in maintaining the homeostasis of the bone marrow system. A previous study have
231 clarify that BMSCs can produce H_2S , regulate osteogenic differentiation and cell
232 self-renewal, and that the lack of H_2S could lead to defects in their differentiation [19].
233 Exogenous H_2S could protect cell injury by regulating oxidative stress, mitochondrial
234 function, and inflammation. While when inflammation occurs, H_2S from bacteria
235 disturbs the endogenous H_2S of osteoblast cells, leads to a negative effect. In
236 periodontitis studies, drugs that can release H_2S have been used for the treatment,
237 such as ATB-352, a kind of ketoprofen that can releasing H_2S . The main aim is to
238 minimize the presence of side-effect at the gastrointestinal tract. Meanwhile they
239 found that the reduction of the inflammation even had a beneficial effect on bone
240 resorption or tissue damage. ATB-346, releasing H_2S like ATB-352, is beneficial for
241 improving bone quality too [20]. Since H_2S also can promote the development of
242 periodontitis, there are still many questions about the biological mechanisms of H_2S . It
243 is well-know that there are many kinds of cell playing important roles in periodontitis,

244 such as periodontal ligament stem cells, osteoclasts, and immune cells. Independent
245 detection of H_2S changes in living cell might facilitate the study of the role of H_2S in
246 diseases.

247 It was found previously that CBS and CSE were both increased in human gingival
248 tissue during periodontitis through the technology of PCR and Western blot. However,
249 H_2S level or H_2S synthesis in gingivitis and periodontitis was detected not increase
250 after tissue homogenate [21]. This can be problematic for many reasons, such as the
251 synthesis capacity decreased or consume increased of H_2S in inflammation. But as a
252 gasotransmitter, half of H_2S can escape from medium in five minutes in tissue culture
253 wells, which makes it hard to detect [22]. Under physiological conditions, H_2S presents
254 in three chemical ionization forms, about 18.5% H_2S , 81.5% HS^- and minute quantities
255 of S^{2-} [23]. Different detection methods might lead to different results. H_2S is more
256 permeable in plasma membranes, the solubility of H_2S in lipophilic solvents is
257 quintuple greater than in water [24], thus, fluorescence probe in theory could detect
258 H_2S more precisely. Our H_2S probe might help deeper studies of the changes of H_2S
259 level and promote the progression on the researches about pathogenesis of
260 periodontitis.

261 Fluorescence techniques is gaining widespread attention as sensors offering excellent
262 sensitivity, good selectivity, and rapid response to changes. First of all, our probe has
263 been shown to be sensitive for endogenous H_2S detection and real-time monitoring of
264 the changes in H_2S in living cells, and it reacts quickly under physiological conditions.

265 There are some things that can be improved, for example, a more precise target of

266 probes to certain subcellular organelles, certain cells, tissues, or organs, which may be
267 achieved by using near-infrared emit to get a greater tissue penetration and minimize
268 the interference from background auto-fluorescence [25]. For clinical use, H₂S has a
269 potential to be used as an appropriate biomarker for the related investigations of
270 inflammation response. However, it still requires further development.

271

272 **Conclusion**

273 In conclusion, it is the first experiment using H₂S probe to detect H₂S changes under
274 stimulation in osteoblast in real time. We used a new hypotoxic H₂S probe for
275 exogenous and endogenous H₂S detection in living osteoblast cells. Moreover, the
276 results indicate that in osteoblast cells, H₂S is produced mainly by CSE-H₂S pathway
277 directly, it also shows that under inflammation stimulation, endogenous H₂S
278 production will increase. The results suggest that H₂S could be a potential marker for
279 diagnosis of inflammatory diseases of bone, and it might help further studies for
280 understanding the synthesis and change of H₂S level in pathogenesis of periodontal
281 disease.

282

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292 **Competing interests**

293 *The authors declare that they have no competing interests.*

294

295

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