Continuous Facial Nerve Stimulation During Cochlear Implantation – An Electrophysiological Technique to Control the Intracochlear Electrode Contact Position

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Abstract

Purpose: This proof of concept describes the use of evoked electromyographic (EMG) activation of the facial nerve for intraoperative monitoring of the electrode insertion during cochlear implantation (CI).

Methods: Intraoperative EMG measurements from the facial nerve were conducted in nine patients undergoing CI implantation. Electric pulses were emitted from contacts on the CI array during and immediately after electrode insertion. For control, the results of EMG measurements were compared to postoperative flat panel volume computed tomography scans with secondary reconstruction (fpVCT_{SECO}).

Results: During insertion, the EMG response evoked by the electrical stimulation from the CI was growing with the stimulating contact approaching the facial nerve and declined with increasing distance. After full insertion, contacts on the apical half of the CI array stimulated higher EMG responses compared with those on the basal half. Comparison with postoperative imaging demonstrated that electrode contacts stimulating high EMG responses had the shortest distances to the facial nerve.

Conclusion: It could be demonstrated that electrically evoked EMG activation of the facial nerve can be used to monitor the progress during CI electrode insertion and to control the intracochlear electrode position after full insertion.

1. Introduction

Cochlear implantation (CI) has developed as the most effective rehabilitation method for patients with high degrees of hearing loss over the last decades¹. Presupposition for a good speech understanding is the correct position of the electrode in the cochlea. Thus, an unfavorable intracochlear placement of the electrode like dislocation or a tip fold over has to be avoided, since this will lead to an inappropriate stimulation of the neuronal structures in the cochlea^{2,3}. In a recent review it was described, that incorrect electrode position, like incomplete insertion or kinking occurs more often in straight flexible electrodes, while electrode tip-fold over can more often be detected in pre-formed electrodes⁴. On the other hand the correct positioning of the electrode in combination with an anatomically based selection of electrode stimulation can lead to a better speech understanding, since an optimized cochlear coverage can be obtained⁵⁻¹⁰. To control the position of the electrode, different techniques are available either real-time during insertion or after the insertion process. Real-time control is possible using intraoperative imaging. The only technique, which can directly detect electrode position, is the fluoroscopy¹¹⁻¹³. Disadvantages of this technique are that the scan has to be performed in the Stenver's projection and the ionizing radiation exposure at least to the surgeon and the patient. In addition, real-time monitoring of the electrode insertion process is possible by electrocochleography (ECochG). During ECochG recording of signals elicited via hair cells and auditory nerve fibers the relative position of the electrode can be measured¹⁴. Increasing signal amplitudes in intracochlear ECochG recordings could be observed during CI electrode insertion¹⁵. However, as ECochG recordings depend on responses of residual functional

neural elements in the cochlear¹⁴, patients can show different patterns of signal behavior¹⁶ impeding the usage of ECoChG as a tool to locate CI array's position.

Different tools have been developed to intraoperatively detect electrode dislocation or malposition either using electrode-neuronal interactions or systems, which use the electric activity of the implant. By the measurement of electrically induced compound action potentials the scalar location¹⁷ and a tip fold over¹⁸ can be detected. In addition, the impedances¹⁹ or the spread of excitations²⁰ can be used to detect an unfavorable electrode position. After the insertion or after the operation radiological imaging is normally used to examine the position of the electrode. Various systems are available, which mostly use ionizing radiation to control the electrode position^{3,10,11}, but there is ongoing research to use magnetic resonance imaging (MRI) for control after cochlear implantation²¹.

Facial nerve monitoring (FNM) is widely used in neurosurgical²², neurotological^{23,24} and other head and neck interventions like e.g. parotid surgery²⁵. By FNM, the electromyographic activity (EMG) of the muscles innervated by the facial nerve is monitored to measure the function of the seventh cranial nerve indirectly. In CI, FNM is widely used to avoid unintended nerve injury during drilling in the mastoid and the posterior tympanotomy²⁶, but in some studies it was shown, that the application of FNM does not reduce the risk of harming the facial nerve²⁷. FNM is also used for navigation in robotic cochlear implantation²⁸.

CIs can in principle stimulate the facial nerve electrically. Postoperative facial nerve stimulation (FNS) is rare but may be facilitated by a number of known conditions, such as otosclerosis^{29,30}, cochlear malformations³¹ or in cases with a significantly reduced thickness of bone separating the upper basal turn of the cochlea and the labyrinthine segment of the facial nerve^{32,33}. Unintended FNS can be reduced by using triphasic pulse stimulation^{34,35}, electrode repositioning, change of implant type or finally by explantation³⁶.

The aim of the presented study was to use intentional FNS as a tool to detect the intracochlear electrode positioning in real time and after implantation. The basic hypothesis is that the EMG signal induced by the stimulation of electrode contacts in close proximity to the facial nerve is different to distant electrode contacts. Thus, the different signals can possibly be used to estimate the relative distance of the respective electrode contact to the facial nerve. To test this, a concept was developed in which the most apical electrode contact was used for FNS during insertion. Hypothetically, the EMG signal will rise in real-time, when the electrode contact approximates to the facial nerve in its labyrinthine portion. After passing the facial nerve and during increase of the distance the EMG signal should decline. By this procedure a pass of the electrode contact at the area of the facial nerve can be measured, which is a sign of a relative progress of insertion. After full insertion, stimulation is performed at every electrode contact. Hypothetically, the EMG signal should be highest for the electrode contact closest to the facial nerve. Thus, a map can be generated, which shows the distance between every contact and the facial nerve, which could be used to measure full insertion.

2. Methods

2.1. Subjects

The study was performed in 9 adult patients (Hereafter referred to as subjects and abbreviated as S1-S9), who underwent cochlear implantation at the Department of Oto-Rhino-Laryngology, Plastic, Aesthetic and Reconstructive Head and Neck Surgery at University of Wuerzburg. 7 women and 2 men were included, ranging in age from 29 to 84 (mean = 53.67, SD = 22.72) years (see Table 1 for demographic details). All subjects fulfilled the indication for cochlear implantation regarding to the German Sk2 guideline for cochlear implantation and were implanted with a SNYCHRONY2® cochlear implant system from the manufacturer MEDEL GmbH (Innsbruck, Austria). This manufacturer offers a range of electrode types of different array lengths to provide an optimal fit of the implant to the patient's anatomy. The participating subjects were implanted with 4 different electrode types (see Table 1). Table 2 lists the electrode types used, their respective array length and the spacing between the individual contacts. The electrode contacts on the arrays were numbered from apical to basal in ascending order.

The study was performed following the declaration of Helsinki and approved of the Ethics Committee of the University of Wuerzburg (315/15_z). Prior to surgery, all subjects were informed about the study and gave their written informed consent.

Table 1
Demographic details of the subjects. Subjects are numbered according to the order in which they participated in the study.

Subject	Age	Sex	Electrode type	Etiology
S1	34	F	Standard	Otosclerosis
S2	29	F	FLEX28	Neurofibromatosis type II
S3	84	F	FLEX28	Progressive hearing loss
S4	71	M	FLEX28	Progressive hearing loss
S5	57	F	Standard	Chronic otitis media
S6	58	F	FLEX28	Sudden hearing loss
S7	76	F	FLEXsoft	Sudden hearing loss
S8	32	M	FLEX24	Progressive hearing loss
S9	32	F	FLEX24	Progressive hearing loss

Table 2
Details of the different electrode types included.

Electrode type	Electrode array length [mm]	Active stimulation length [mm]	Intercontact spacing [mm]
Standard	31.5	26.4	2.4
FLEXsoft	31.5	26.4	2.4
FLEX28	28.0	23.1	2.1
FLEX24	24.0	20.9	1.9

2.2. Experimental Setup

The experimental setup (see Fig. 1) was based on the systems introduced by Bahmer et al.³⁴. It consisted of a research interface box 2 (RIB2; Department of Ion Physics and Applied Physics at the University of Innsbruck, Austria) and a g.USBamp biosignal amplifier (g.tec GmbH, Schiedlberg, Austria) connected to a laptop computer (2.5 GHz Dual Core Intel CPU and 8 GB RAM). All stimulation and recording parameters were operated via a graphical user interface programmed with the software Matlab R2017b (The MathWorks Inc., Natick, USA). The RIB2 converted the stimulation parameters into commands that were sent to the subject's CI via a telemetric transmitter coil. The biosignal amplifier recorded the EMG response to the stimulation pulses via subcutaneous needle electrodes, which were inserted ipsilaterally into two different groups of muscles innervated by the facial nerve. The EMG channel recording the EMG activity of the M. orbicularis oculi measured the potential differences between an electrode above the eyebrow and one below the eye. The electrodes of the channel recording the EMG activity of the M. orbicularis oris were inserted above the upper lip and below the lower lip. The reference electrodes were placed in inactive areas on the forehead and the chin. It was verified that all impedances were below 5 k Ω . To synchronize stimulation and EMG recordings, the RIB2 emitted a trigger signal on the onset of each pulse. The trigger signal was fed to an input of the biosignal amplifier. The inputs of the biosignal amplifier were sampled at a rate of 38.4 kHz. The digitized signals were stored on the hard drive of the computer.

2.3. Stimulation parameters

The stimulation consisted of biphasic pulses with cathodic-leading phase applied in monopolar mode. The phase duration of each pulse was set to 150 μ s and the interphase gap to 2.1 μ s. For each measurement, 2 subsequent pulses were emitted with a rate of 100 pulses per second.

2.4. Real time measurements

During the insertion of the electrode, the insertion process was stopped after insertion of two electrode contacts. Stimulation was performed from the most apical electrode contact with rising current level until a clear EMG signal could be detected. Thereafter, the next two electrode contacts were inserted, and the procedure was repeated. This was continued gradually until full insertion was achieved. Figure 2

describes three theoretical key stages of the measurement procedure during stepwise insertion. In Fig. 2A, only a small number of electrode contacts are inserted into the cochlea. The distance between the most apical contact and the FN is still far, thus stimulation evokes only minor EMG responses. In Fig. 2B, the insertion has already advanced. Because of the immediate proximity, the most apical contact can stimulate the FN most effectively and evoke a high EMG response. In Fig. 2C, the most apical contact has already passed the FN and the distance between the two is increasing again. As a result, the stimulation loses its effectiveness and the EMG response decreases with each insertion step.

2.5. Post insertion measurements

After full insertion of the electrode, stimulation was performed from every electrode contact on the array and the subsequent EMG response was recorded from both EMG channels. For each subject, multiple measurement runs with different stimulation levels were conducted to gain data with optimal signal-to-noise ratio.

2.6. Radiological measurements

For control of the insertion flat panel volume computed tomography (fpVCT) scans were performed on an angiographic system (Axiom Artis; Siemens Healthcare AG, Erlangen, Germany) with commercially available software Syngo DynaCT (Siemens Healthcare AG) resulting in a slice thickness of 466 μm . Subsequently, secondary reconstructions (fpVCT_{SECO}) with a slice thickness of 99 μm were created³⁷. All CT scans were converted to the DICOM format and imported to the open-source medical image viewer Horos (Version 3.3.5; Nimble Co LLC d/b/a Purview, Annapolis, Maryland USA). First, the “Cochlear View”³⁸ was set, which is an axial view of the cochlea generated by rotation against the three body axes. In this two-dimensional view, the electrode facial nerve distance (EFND) for each electrode was measured between the center of the metal artifact and the center of the facial nerve canal in cross section. Figure 3A is a schematic illustration of the EFND measurement. Figure 3B is an example of an fpVCT_{SECO} scan in cochlear view depicting the ENFDs as colored lines.

2.7. Statistics

The data were analyzed with the R programming language (version 4.0.2) and the RStudio (version 1.3.1073) integrated developments environment. The ggplot2 package (3.3.5) was used for graphical representation of the data. In order to investigate the correlation between EFNDs and normalized EMG amplitudes, the nonparametric Spearman’s rank correlation was calculated using the stats package (4.0.2).

3. Results

3.1. Measurement of the electrode facial nerve distance

As can be seen in Fig. 4, fpVCT_{SECO} of subjects implanted with the electrode type Standard, FLEXsoft, or FLEX28 showed the shortest EFND for the medial electrode contact number 6. The shortest EFND of S8 and S9, both implanted with the short electrode type FLEX24, was measured for electrode contact number 3 and 4, respectively.

3.2. Real-time insertion of the electrode

During insertion, multiple current levels were tested to stimulate the EMG response. Only those responses evoked at a current level, which had sufficiently high amplitudes, were used for analysis. In eight of the nine subjects, an EMG response was continuously recorded on both channels during the stepwise insertion of the electrode. S2 was excluded from the analysis because no coherent data set could be generated during insertion. In subject S1, recording began when four contacts had already been inserted instead of two. EMG response strength differed greatly between subjects as well as between the periocular and perioral EMG channels. Thus, to achieve better comparability of the results, the data of each subject and each EMG channel were normalized. The normalization process assigned the value 0 to the lowest EMG response and 1 to the maximum EMG response. Figure 5 shows the normalized EMG response during the stepwise insertion for both EMG channels. With few exceptions, the measurements of both EMG channels demonstrated a stable concordance for all of the 8 subjects, an initial increase and subsequent decrease in the normalized EMG response can be seen as the insertion progressed. Figure 5 shows that in the cases of S1, S3, and S6 the EMG response increases again towards the end, i.e. when the array was fully inserted. The measurements in S4 demonstrated peaks of the EMG signal at two different numbers of inserted contacts (periocular: 6; perioral: 8). For the remaining subjects, the peaks appearing at the medial portion of the cochlea revealed a stable concordance between both EMG channels. Under the condition of an exactly constant step size during insertion, it can be assumed that the position P of the stimulating apical contact can be calculated as the difference between the number of all contacts on the array (all MED-EL electrode types feature 12 contacts) and the number of already inserted contacts $N_{inserted}$. During the stepwise insertion, the EMG amplitude could be continuously tracked in eight out of nine subjects.

$$P = 12 - N_{inserted} \quad (1)$$

Taking into account the deflection of the EMG signal in respect to P , the contact that is closest to the facial nerve after full insertion can now be estimated. Table 3 lists the values of P calculated by formula 1 which show the highest EMG response in each channel and compares them with the respective shortest measured EFND. The comparison revealed a clear concordance between imaging and EMG measurement in five of seven (EMG periocular) and four of seven (EMG perioral) subjects. With respect to S8, contact 3 shows the shortest EFND. Since step sizes of 2 contacts were used during insertion, an exact agreement cannot be determined for odd contact numbers. However, at S8 the peak at the neighboring contact 2 is visible.

Table 3
Comparison between the shortest radiologically measured EFND per subject and the estimated position of the contact showing a clear peak in both EMG channels during insertion.

Subject	Shortest EFND	P _{oculi}	P _{oris}
S1	6	6	6
S3	6	6	6
S4	6	6	4
S5	6	2	2
S6	6	6	6
S7	6	4	4
S8	3	2	2
S9	4	4	4

3.3. Post insertion measurements

Similar to the real-time measurements during insertion, multiple current levels were used in the post insertion measurements. Each subject showed an individual stimulation threshold, above which a sufficiently strong EMG response could be observed. However, too high stimulation levels could result in no clear peaks depending on the electrode contact used. Such a "breakdown" consequently caused strong EMG responses on several contacts of the array, which prevent estimations of the position of the electrode with respect to the facial nerve by means of the EMG amplitudes. Therefore, only those data were included in the analysis, which were recorded using stimulation amplitudes that elicited a clear EMG response and allowed the best possible spatial differentiation. EMG responses of the M. orbicularis oculi are depicted in Fig. 6, those of the M. orbicularis oris in Fig. 7 after stimulation from every contact of the fully inserted electrode array for each of the nine subjects. As an additional dimension, the radiologically measured EFNDs per contact of the respective subject were plotted as a relative color gradient to the curves in Fig. 6 and Fig. 7. As for the real-time measurements, the EMG response of each subject and each EMG channel was normalized for better comparability. Normalization assigned the value 0 to the lowest EMG response and 1 to the highest. As evaluation criterion for the presence of a spike in EMG data, a threshold at 75% normalized EMG response was arbitrarily defined. As can be seen in Fig. 6 and Fig. 7, the selected current levels on single and multiple electrode contacts, respectively, achieved response strengths that exceeded 75% normalized EMG amplitude. The EMG responses that exceeded the threshold are subsequently referred to as peaks. Table 4 summarizes at which electrode contact the

shortest EFND was measured radiologically, and at which contacts EMG peaks were measured after complete insertion.

Table 4

Listing of electrode contacts per subject with the shortest EFND and EMG peaks in the periocular and perioral EMG channels, which exceeded the threshold at 75% normalized EMG amplitude.

Subject	Shortest EFND [electrode contact]	Peaks periocular [electrode contact]	Peaks perioral [electrode contact]
S1	6	6	6
S2	6	6, 7	5, 6, 7
S3	6	4	2, 4, 5
S4	6	2, 5	2, 5
S5	6	6	4, 5, 6
S6	6	6	2
S7	6	2, 4, 10	2, 4, 6, 7
S8	3	1, 3, 4, 5	1, 3, 4, 5
S9	4	5	3

The scatter plots in Fig. 8 show the relationship between the postoperatively determined EFND and the normalized amplitudes of the EMG responses of the orbicularis oculi (left side) and oris (right side) muscles measured for each electrode contact of all subjects after full insertion. The linear regression function of the data (red straight line) and the corresponding confidence interval (95%) show a clear decrease in normalized EMG responses with increasing EFND in both muscle groups. The Spearman's rank correlation coefficient R revealed a negative moderate correlation for the periocular EMG channel ($R(106) = -0.39$, $p < 0.001$) and a negative strong correlation for the perioral EMG channel ($R(106) = -0.6$, $p < 0.001$). Both correlations were statistically significant.

4. Discussion

In the present study, it was investigated whether the FNS evoked by electrical stimulation via the CI electrode can be used as a marker of the intracochlear position of the electrode array during and after insertion. It was hypothesized that electrode contacts with shorter distance to the facial nerve (EFND) can stimulate the nerve more effectively than contacts further away. The first part of this study included recordings of EMG responses during stepwise insertion of CI electrode arrays. The step size was two contacts at a time. After each step, stimulation pulses were emitted from the most apical contact on the

array and the evoked EMG responses of the orbicularis oculi and oris muscles were recorded. In the second part of the study, stimulation pulses were sequentially emitted from each contact on the fully inserted electrode array and the evoked EMG responses were recorded. EFNDs of each patient were measured in postoperative fpVCT_{SECO} scans and compared with the intraoperatively recorded EMG responses.

The measurements of the EFND in the "cochlear view" of fpVCT_{SECO} scans showed sufficient feasibility. Interestingly, two types of slopes could be distinguished based on their minima, i.e. the shortest EFND (Fig. 4). One slope type showed the shortest EFND at contact 3 or 4, and only included the shortest electrode type FLEX24. This electrode type was implanted in cases, in which the residual hearing should be preserved. Accordingly, the electrode should only partially cover the cochlear length to replace damaged inner hair cells in the range of the high frequencies, while keeping more apical hair cells functional. Therefore, the contacts closer to the apical end of the electrode were nearest to the facial nerve.

The other type of slope showed the shortest EFNDs at contact 6 and included all electrode types used for a full cochlear coverage (FLEX28, FLEXsoft, and Standard). The fact that the shortest EFND was measured at contact 6 for each of the three electrode types despite the difference in length between FLEX28 and FLEXsoft or Standard (see Table 2) can be explained by the fact that FLEXsoft and Standard were implanted in cochleae with longer CDL and both types have longer distances between contacts than the FLEX28.

In addition, the difference in length from the basal contact to contact 6 between the 28 mm and the 31.5 mm electrodes is relatively short (1.5 mm (5 x 0.3 mm)) and smaller than the distance between the contacts (2.1 or 2.4 mm). Consequently, the difference might not be large enough, that another contact other than number 6 will be closer to the facial nerve and though has a lower EFND. This knowledge might be used for further development and research. If further measurements of EFNDs confirm the present result, that electrode 6 has always the lowest EFND in cases of full cochlear coverage by long electrode types this can possibly be used to check correct insertion of the respected array in clinical routine by only measuring the EMG signal after insertion. In cases with partial cochlear coverage (e.g. for electro-acoustic stimulation) it might be possible to preoperatively determine, which contact should have the smallest EFND and though this contact should than be implanted closest to the facial nerve with consideration of the EMG signal.

In 8 of 9 patients, an electrically evoked EMG signal was successfully monitored during CI array insertion. As the insertion depth progressed, the distance between the most apical contact and the facial nerve initially decreased and the stimulation efficacy increased. This could be observed by the rise in EMG amplitudes as function of the number of inserted contacts. Thereafter, as insertion depth progressed, the distance between the electrode and the facial nerve increased again, which was reflected in the decrease of the EMG amplitudes. After the last insertion step, 4 of the subjects showed a new increase in EMG amplitudes. This occurred exclusively in subjects implanted with longer electrode types (Standard,

FLEX28) and not in the 2 patients provided with a short FLEX24. Therefore, it can be assumed that this increase was due to a second approximation of the most apical contact with the facial nerve in the second cochlear turn. In 5 out of 8 subjects, there was concordance between the estimated number of inserted contacts, which showed an EMG peak above the threshold of 75% normalized EMG amplitude and the radiologically measured EFNDs. In one patient, whose shortest EFND was measured for an odd contact number, the EMG peak appeared at one of the even neighboring contacts. These results support the hypothesis that during increasing insertion passing of the electrode at the area of the facial nerve can be measured electromyographically.

ECochG is another electrophysiological measurement that can be used for real-time monitoring during cochlear implantation¹⁴. Studies in humans and animals reported of increasing signal amplitudes with increasing electrode insertion depths, which were assumed to reflect the decreasing distance to residual functional neurons in the cochlear apex^{15,39}. However, previous studies attempting to associate CI electrode translocations and intraoperative ECochG showed either no correlation⁴⁰ or only a weak positive predictive value⁴¹. The advantage of the electromyographic measurement presented in this study over ECochG is that the facial nerve can be used as an anatomical landmark to estimate the relative location of the electrode.

After full insertion, stimulation was performed from each electrode contact and the evoked EMG responses were recorded. The EMG profiles were normalized for better comparability between each subject and the 2 EMG channels used. To identify electrode contacts, which exhibited EMG peaks along the array an arbitrarily chosen threshold of 75% normalized EMG amplitude was used. The electrode contacts thus identified were compared with the electrode contacts that showed the shortest EFNDs in the postoperative fpVCT_{SECO} images.

Both EMG channels showed agreement between the shortest EFND and EMG peaks in 5 of 9 subjects (periocular: S1, S2, S5, S6, S8; perioral: S1, S2, S5, S7, S8). Besides the peaks that exceeded the threshold, the results of the post insertion measurements showed higher EMG responses for contacts lying on the more apical half of the electrode array. As observed in the data from the insertion measurements, this may reflect the decreasing EFND of those contacts, which are closer to the facial nerve within the second cochlear turn than the more basal contacts. This assumption is also supported by the statistically significant results of the correlation analysis, which showed moderate (periocular) and strong (perioral) negative correlation between the normalized EMG amplitudes and the EFNDs of each contact.

The aim of this study was to prove the feasibility of the approach. The measurement presented is still under development and requires further improvements to increase precision. The following section is intended to provide starting points for future studies.

The "cochlear view" provided only EFNDs in a two-dimensional representation of the patient's anatomy. Close to the cochlea, however, the facial nerve exhibits a complex three-dimensional course. In addition, the CI array changes its position in three dimensions as it passes through the cochlear turns. The EFND

measurements proposed in the present study should be considered as an estimate of the actual distance. Future studies should therefore investigate whether the EFND can be determined more precisely by considering the three-dimensional anatomy. Recent studies reported that patients with postoperative FNS had significantly lower distance and bone density between the upper basal turn of the cochlea and the labyrinthine segment of the facial nerve^{32,33}. Therefore, besides the EFND future studies should also include parameters such as bone density, which can influence the propagation of the electric current. The stepwise insertion in the present study was performed manually. Thus, the number of contacts already inserted could only be estimated. In addition, an anatomically defined reference was missing for a precise indication of the length of the already inserted electrode array. The threshold 75% normalized EMG amplitude was chosen arbitrarily and is only a rough method to detect peaks. Modern methods of peak detection could achieve a better differentiation of different peaks (contact in close proximity to facial nerve, most apical contact in second cochlear turn). Future studies should consider changes to the stimulation paradigm that allow a higher number of repetitions which cause less fatigue to the facial nerve and the muscles innervated by it. This should improve the signal-to-noise ratio and additionally reduce the influence of outliers (electrical artefacts from the operating room, spontaneous movements of the patient) on peak detection. The used higher stimulation amplitudes resulted in broader electric fields in the cochlea and its surrounding anatomical structures, which might have influenced the accuracy of the system. Measurements in the orbicularis oris muscle yielded EMG responses with higher amplitudes, which resulted in a better signal-to-noise ratio. Therefore, compared with measurements in the orbicularis oculi muscle, lower current levels can be applied to stimulation, which should result in better spatial resolution of the EMG. Therefore, the authors suggest future studies to focus on EMG measurements in the orbicularis oris muscle.

5. Conclusion

To the best of our knowledge, this is the first study to demonstrate the potential of using induced FNS for real-time monitoring of the insertion status during and after cochlear implantation. This system might be helpful to the field of cochlear implantation, since by applying FNS stimulation, the position of the electrode can be determined quite precisely for at least the first six implanted contacts in the basal turn, which equals about 200 degrees of the cochlea. Because there is evidence that the EMG signal rises again as the stimulating contact approaches the FN in the mid turn at about 540 degrees, this method could hypothetically detect an electrode tip-fold over as well. The system can easily be used to perform further measurements after surgery and during all wearing-time of the implant. After wound closure, the system can control, if manipulations during suturing of the periosteum, the subcutaneous tissue, and the skin might have led to an electrode displacement. In addition, if there is a suspicion for electrode extrusion after surgery, a measurement of the evoked EMG can be performed again. If the electrode has not been displaced, the highest EMG responses should be measured at the same stimulating contact as before.

On the long term, the system might be used in a radiation free cochlear implantation. Omitting radiation is of high benefit, especially for children⁴², since there is growing evidence from epidemiological studies that radiation exposure of the brain poses further substantial risks^{43–45}. In this concept preoperative planning of the implantation, like measuring the CDL^{46,47}, the examination of the course of facial nerve⁴⁸ and the size of the temporal bone⁴⁹, will be performed in MRI imaging. For intra- and postoperative control the presented system, may be used in combination with electrocochleography⁵⁰ or electrically evoked compound action potentials¹⁸. If required, after the waiting time defined by the manufacturer, an MRI scan could be performed with special protocols to investigate the electrode position²¹.

Abbreviations

CDL, cochlear duct length; CI, cochlear implant; ECochG, electrocochleography; EFND, electrode facial nerve distance; EMG, electromyography; FNM, facial nerve monitoring; FNS, facial nerve stimulation; fpVCT, flat-panel volume computer tomography; MRI, magnet resonance imaging; RIB2, research interface box 2

Declarations

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Author contributions

K.R. and D.P.H. conceived and designed the project. R.H. and K.R. supervised the study. A.K. preselected suitable patients for participation in the study. K.R. and S.K. performed CI surgery. D.P.H. built the EMG recording setup with support from M.C. and conducted the intraoperative EMG measurements. T.N. performed medical imaging (fpVCT) and supported F.T.M.G. and D.P.H. in secondary reconstructions of fpVCT scans. D.P.H. evaluated the fpVCTSECO and EMG data and performed the analysis including figures and statistics. D.P.H. and K.R. wrote the manuscript. All authors reviewed the manuscript.

Competing interests

The authors declare no conflict of interest.

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Figures

Figure 1

Experimental setup for intraoperative EMG recordings. The setup consisted of a PC for stimulation and measurements control, a research CI interface (RIB2) for stimulation and a g.USBamp biosignal amplifier to record two EMG channels.

Figure 2

Schematic illustration of the real-time measurements. EMG activity is recorded during the stepwise insertion of the cochlear implant electrode array. Panels A, B, and C illustrate three different stages of the measurements procedure.

Figure 3

A: Schematic illustration of the measurement of the electrode facial nerve distance with color-coding of the relative distances (red: shortest distance; black: longest distance). B: Example of the measurement of the electrode facial nerve distance in the cochlear view of an fpVCTSECO scan.

Figure 4

Electrode facial nerve distance as function of the electrode contact number. Distances were measured from fpVCTSECO scans in cochlear view. Colors correspond to different implanted electrode types.

Figure 5

Normalized EMG responses during the stepwise electrode insertion in eight subjects. The EMG responses are shown as function of the number of inserted electrode contacts. The solid and dashed lines correspond to the periorcular and perioral EMG channel, respectively. Normalization assigned the value 0 to the lowest amplitude and 1 to the maximum amplitude.

Figure 6

Normalized EMG responses of the M. orbicularis oculi as function of the contacts on the fully inserted electrode array. The color gradient depicts the radiologically determined electrode facial nerve distance (red: shortest distance; black: longest distance) for each subject. The horizontal dashed line indicates the arbitrarily chosen EMG threshold at 75% normalized EMG response.

Figure 7

Normalized EMG responses of the M. orbicularis oris as function of the contacts on the fully inserted electrode array. The color gradient depicts the radiologically determined electrode facial nerve distance (red: shortest distance; black: longest distance) for each subject. The horizontal dashed line indicates the arbitrarily chosen EMG threshold at 75% normalized EMG response.

Figure 8

Correlation between the normalized EMG responses of the M. orbicularis oculi (left side) and oris (right side) after full insertion in all subjects and the radiologically measured electrode facial nerve distances. The red line indicates the linear regression of the respective results, the gray shaded area around the regression line indicates the 0.95 confidence interval. R is the Spearman's correlation coefficient and p is the significance level of the correlation.